

# Semen Analysis

**Collage of Veterinary Medicine and animal sciences  
University of Gondar,  
Gondar, Ethiopia**

# Objectives

- At the end of the sessions students will be able to determine and explain
  - General consideration of Semen
  - Indication and sample collection of semen
  - Volume and gross appearance of ejaculates
  - Microscopic evaluation of semen
  - Biochemical tests of semen

# INTRODUCTION

- Semen is a mixture of **fluids** and **cells**
- It is composed of **four fractions** that are contributed by **epididymis, seminal vessels, prostate** and **bulbourethral glands**.
- Each fraction differs in its contribution
- But the mixing of **all four fractions** is essential for the production of a normal semen.

# Formation of the sperm cell

- Sperm is produced in *seminiferous tubules* located in the testes.
- **Germ cells** located in the epithelial cells of the *seminiferous tubules* are responsible for the production of *spermatozoa*
- The phenomena is aided by specialized *Sertoli cells* that **provide support and nutrients** for the germ cells that undergo mitosis and meiosis (spermatogenesis).

- When spermatogenesis is complete, the **immature sperm (non-motile)** enter to the epididymis.
- In the epididymis, the **sperm mature and develop flagella** and remain **stored** until ejaculation.
- During **propelling** the sperm cell will **combine to seminal fluids** inside ductus deferens
- The **seminal vesicles** produce the **majority of the fluid** present in semen (60% to 70%).

- The fluid contains a **high concentration of fructose** that are metabolized for the energy needed for the **flagella to swim in the female reproductive tract.**
- Approximately **20% to 30%** of the semen volume is **acidic fluid** produced by the **prostate gland.**

- The acidic fluid contains **high concentrations of acid phosphatase, citric acid, zinc,** and proteolytic enzymes responsible for both the *coagulation* and *liquefaction* of semen
- The **bulbourethral glands** contribute about **5%** of the fluid volume in the **form of a thick, alkaline mucus** that helps to **neutralize acidity from the prostate secretions** and the vagina.

# Composition of Semen in volume

## Composition of Semen in volume

- Spermatozoa= 5%
- Seminal fluid= 60%–70% (Spermatozoa become mobile when exposed to seminal fluid)
- Prostate fluid =20%–30%
- Bulbourethral glands fluid = 5%

# Significance of semen analysis

- ♣ Soundness evaluation and investigation of fertility problems
- ♣ For selection of donors for artificial insemination
- ♣ To pass soundness decision for breeding program.
- ♣ To predict the fertilizing capacity of a semen
- ♣ To assess post **vasectomy**

# Collection and Handling of Semen

- There are three commonly-used techniques for collecting semen:
  - Use of an artificial vagina
  - Manipulation and
  - Electroejaculation.
- The technique used depends on the **species being collected** and the disposition of the individual male.

- Semen is **fragile** and **susceptible to damage** and killing by several environmental conditions.
- When collecting and handling semen it is critical to avoid exposing sperm to **two types of insults**:

### *1. Exposure to toxic chemicals:*

- Keeping collection **equipment clean** and free of **spermicidal element**.
- Best to use **deionized water** for cleaning

## *2. Thermal stress:*

- Sperm are sensitive to **environmental temperature**.
  - It has to be examined **at temp near to body T°** and stored at **-196 °C** in liquid nitrogen
  - **Handle semen with care** because it may contain **infectious pathogens**.

# Tests for semen

- There are several **Macroscopic** and **Microscopic** evaluations which give **useful diagnostic information** about the semen sample

## **Macroscopic**

- Appearance
- Odour
- Liquefaction
- Volume
- Viscosity
- pH

## **Microscopic**

- Motility
- Morphology
- Concentration
- Viability

# Macroscopic Evaluation

## 1. Appearance and odour

- Normal semen has a **gray-white color**, appears **translucent**, and has a characteristic **musty odor**.
- **Yellow** coloration may be **caused by urine contamination**, specimen collection following **prolonged abstinence**, and **medications**.
- Increased **white turbidity** indicates the presence of **white blood cells (WBCs)** and **infection** within the reproductive tract.
- If **blood** is present it may appear **pink to orange**

## 2. Liquefaction and Viscosity

- **Liquefaction** is the **breakdown of the gel portion** of the **seminal plasma** by the help of the enzyme known as Fibrinolysin
- Normal semen liquefy with in **30-60 minutes** after collection.
- **Failure to liquefy** is due to deficiency in **prostatic fluid**
- **Viscosity** is inversely related to specimen liquefaction
- **Increased viscosity** and **incomplete liquefaction** will impeded **sperm motility**

### 3. Volume

- Normal volume **vary from species to species**
- **Increased volume:** following periods of **extended abstinence**
- **Decreased volume:** associated with **infertility**; may indicate **improper functioning of one of the semen producing organ.**
- **Aspermia** – no semen production at all

	Bovine	Shoat	Swine	Horses	Man
Volume (ml)	4-6	1-2	225	60	2-5

## 4. pH

- Semen has a narrow pH range from 6.4-8.0.
- It can be measured by immersing lithmus paper or spread a drop of liquefied semen on the paper.
- When the pH is over 8.0 this may be due to infection in RT.
- Decreased pH; associated with increased prostatic fluid
- When the pH is below 6.4 and the semen is found to contain no sperm, this may indicate dysgenesis (failure to develop) of the vas deferens, seminal vesicles or epididymis.

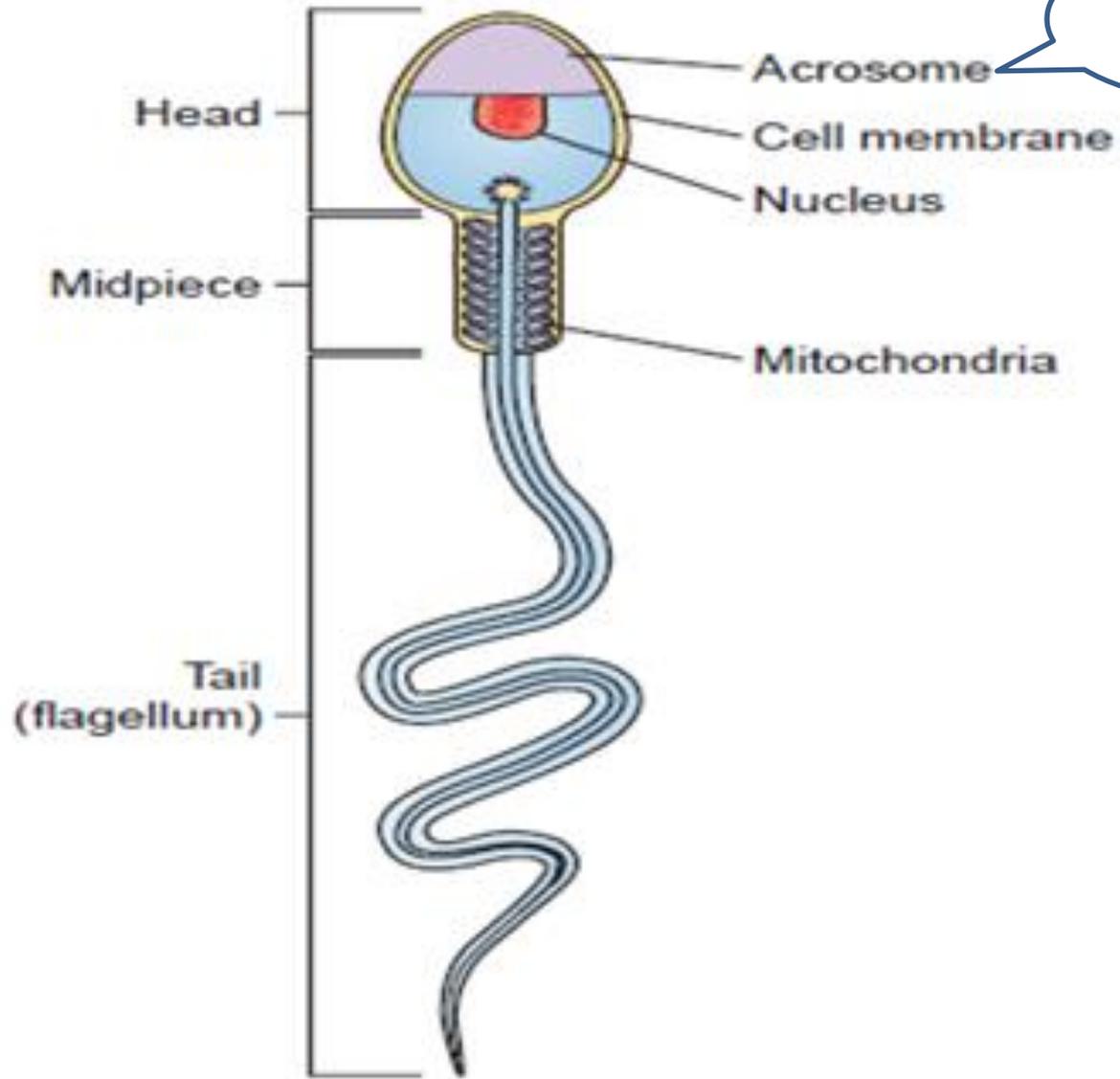
# Microscopic Examination

- It is performed to obtain estimates of sperm **morphology, motility, concentration, viability** and some times **wave pattern**
- It is done by placing **10 $\mu$ l** of thoroughly **mixed, liquefied semen** on a **Microscope slide** and coverslip with a **22x22mm** size

- The quality of sperm motility is affected by temperature
  - So great care must be taken to ensure that the slides and coverslips, as well as the pipette tips are kept around 37°C
  - The assessment must start as soon as the flow stops. if this is >1 minute, a new wet prep must be made

# 1. Morphology

- Sperm morphology is evaluated from a **thin smeared and stained slide under oil immersion.**
- Staining can be performed using **Carbol fuchsin, Wright's, Giemsa, HE stain** which is a matter of laboratory preference.
- The normal sperm has an **oval-shaped head, midpiece** located between **head and tail** and **flagellated tail** required for **motility**



Normal spermatozoa structure.

- **Reporting morphology of spermatozoa**
  - Examine the preparation for **normal** and **abnormal** spermatozoa using the **40X objective**
  - Use the **100X objective** to **confirm abnormalities**.
  - Count **100 spermatozoa** and **estimate the percentage** showing **normal morphology** and the percentage that **appear abnormal**.
  - In normal semen, at **least 50% of spermatozoa should show normal morphology**.

# The following abnormalities may be seen:

- **Head**
  - **Greatly** increased or decreased in **size**.
  - **Abnormal shape** and tapering head (pyriform)
  - **Acrosomal cap absent** or abnormally large.
  - **Bifurcated heads**.
- ***Tail***
  - **Absent or markedly reduced** in length.
  - **Double tail**.
  - **Bent or coiled tail**.

# Abnormalities of sperm on head and tail



Normal



Double head



Giant head



Amorphous head



Pinhead



Tapered head



Constricted head



Double tail



Coiled tail



Spermatid

## 2. Motility assessment

- The sperm cells **capable of forward and progressive movement** is critical for fertility.
- Because once presented to the **cervix**, the **sperm must propel themselves** through the cervical mucosa, uterus, fallopian tubes, and ovum for fertilization.

- Traditionally, clinical laboratory reporting of sperm motility has been a **subjective evaluation** and determining the **percentage of motile sperm** and the quality of its motility.
- **Assessment of sperm motility** should be performed on **well mixed, liquefied semen** soon after specimen collection.

- The motility movement can then be estimated after evaluating approximately **20 high-power fields**.
- **Motility is evaluated** by both **speed** and **direction**.
- **Grading can** be done using a scale ranging from **0** to **4**,
  - **4** indicating **rapid, straight-line movement** and
  - **0** indicating **no movement**.
- A **minimum** motility of **50%** with a **rating of 2** is considered **as normal**.

# Motility assessment - types

Grade	Motility type	Percentage
4	Rapid and straight-line motility	80-100%
3	Slower straight line speed with some lateral movement	60-80%
2	Slow forward progression with noticeable lateral movement	40-60%
1	No forward progression but slow lateral movement	20-40%
0	No movement	0-20%

### 3. Sperm Concentration/Count

- Even though fertilization is accomplished by **one spermatozoon**, the **actual number of sperm present in a semen specimen** is a **valid measurement** for fertility.
- Normal values for sperm **concentration varies among species**.
- The **total sperm count for the ejaculate** can be calculated by **multiplying the sperm concentration in ml** by the **specimen volume**.

	<b>Cattle</b>	<b>Shoat</b>	<b>Swine</b>	<b>Horses</b>	<b>Man</b>
<b>Sperm conc. (<math>10^9</math>/ml)</b>	<b>1–1.2</b>	<b>3.0</b>	<b>0.2</b>	<b>0.15</b>	<b>0.02</b>
<b>Sperm/ejac. (<math>10^9</math>/ml)</b>	<b>4–7</b>	<b>4</b>	<b>45</b>	<b>9</b>	<b>0.04</b>

- Sperm concentration is usually performed using the **Neubauer counting chamber**.
- The sperm are counted in the **same manner as cells in RBC counting**.

- Dilution of the semen is essential because it immobilizes the sperm prior to counting.
- The traditional diluting fluid contains *sodium bicarbonate* (5g) and *formalin* (1ml) to 100 ml of water which immobilize and preserve the cells
- However, good results can also be achieved using saline and distilled water.

## 4. Viability

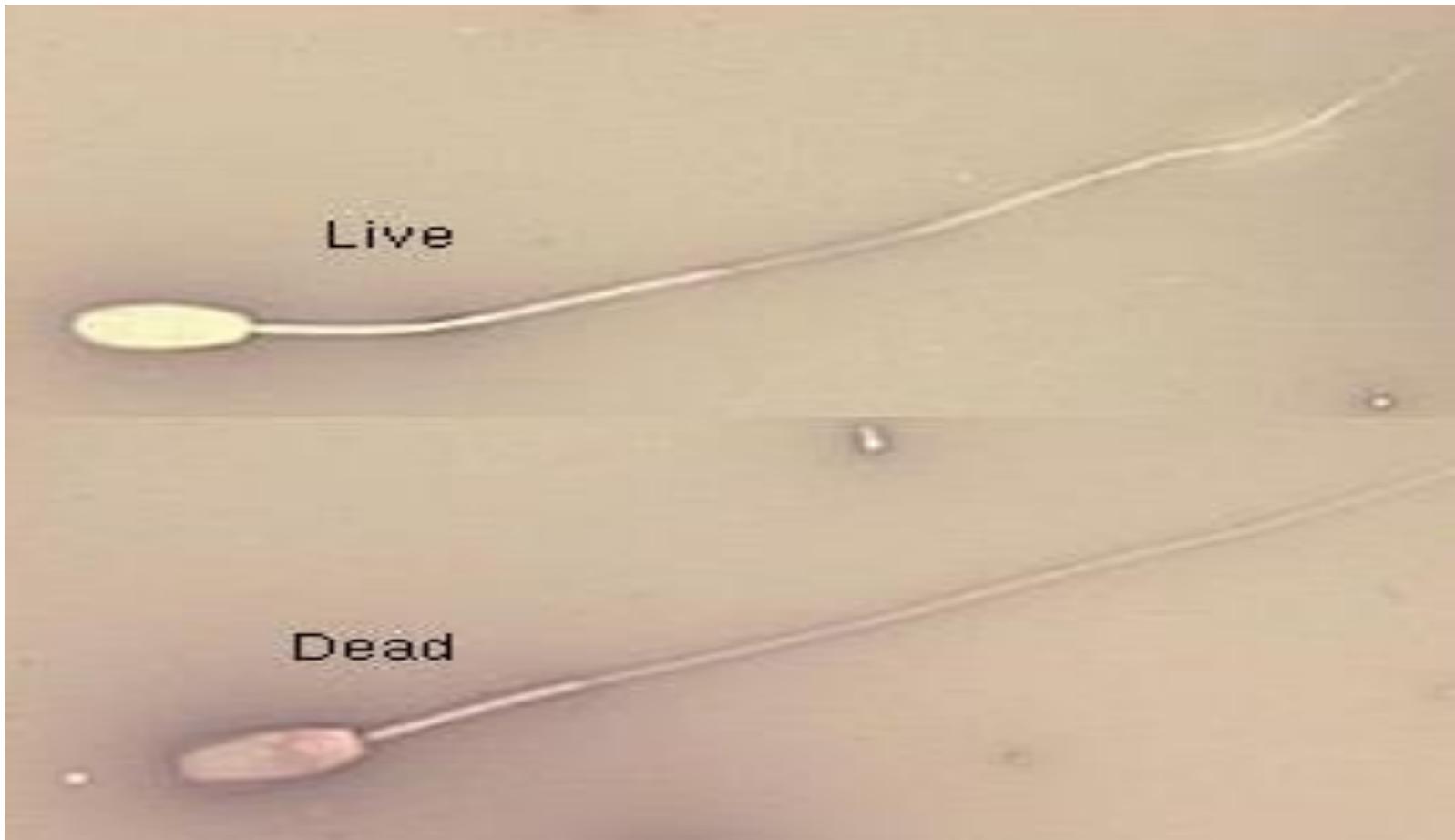
- **Decreased sperm viability** may be suspected if there is a **marked decreased of motility**.
- Viability is **evaluated** by **mixing the specimen** with an **eosin- aniline stain**.
- The **membranes of dead sperms** are **damaged** and can **easily take up eosin stain**.

- The viable sperms do not allow the stain to penetrate leaving a colorless (bluish) sperm.
- Normal viability requires 75% living cells and should correspond to the evaluated motility.

# Procedure

- **Mix one drop** of semen with **1 drop of 0.5% eosin** solution on a slide.
- Make a smear using mixture
- After **2 minutes** examine microscopically using **40X objective**
- Count **100 sperm cells** and **compute the percentage** of viable and non-viable spermatozoa.
  - **Viable** spermatozoa remain **unstained**
  - **Non-viable** spermatozoa **stain red**.

# Viability and Nonviability spermatozoa demonstrated by the eosin stain



## 5. Wave pattern

- It is an **alternative test** for motility
- It is determined by **placing a thick drop** of semen on a slide **under a microscope** with **low power** and **reduced light**.

- The result is reported as
  - ✓ Very good (4) - **Dark, distinct waves** moving rapidly
  - ✓ Good(3)- **Waves apparent**, but with moderate motion
  - ✓ Fair(2) - **Waves barely** distinguishable
  - ✓ Poor(1) - **No waves, but motile sperm** are present
  - ✓ Very poor(0)- **No waves and no sperm motility**

# Semen biochemistry

- *Acid phosphatase*: marker for **prostatic function** and important in rape cases
- *Citric acid*: can indicate prostatic function - **low levels** may **indicate dysfunction** or a **prostatic duct obstruction**
- *Zinc*: marker for **prostatic function** - *colorimetric assay*
- *Fructose*: marker for **seminal vesicle function**, and is a substrate for energy metabolism – *spectrophotometric assay*

# QUESTIONS

