

Handbook of Experimental Pharmacology 200

Bertil B. Fredholm

*Editor*

# Methylxanthines



Springer

# Handbook of Experimental Pharmacology

Volume 200

Editor-in-Chief

F.B. Hofmann, München

Editorial Board

J.A. Beavo, Seattle, WA

A. Busch, Berlin

D. Ganten, Berlin

J.-A. Karlsson, Singapore

M.C. Michel, Amsterdam

C.P. Page, London

W. Rosenthal, Berlin

For further volumes:

<http://www.springer.com/series/164>



Bertil B. Fredholm  
Editor

# Methylxanthines

 Springer

*Editor*

Bertil B. Fredholm  
Department of Physiology and Pharmacology  
Karolinska Institute  
Nanna Svartz väg 2  
177 17 Stockholm  
Sweden  
bertil.fredholm@ki.se

ISSN 0171-2004

e-ISSN 1865-0325

ISBN 978-3-642-13442-5

e-ISBN 978-3-642-13443-2

DOI 10.1007/978-3-642-13443-2

Springer Heidelberg Dordrecht London New York

Library of Congress Control Number: 2010935195

© Springer-Verlag Berlin Heidelberg 2011

This work is subject to copyright. All rights are reserved, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilm or in any other way, and storage in data banks. Duplication of this publication or parts thereof is permitted only under the provisions of the German Copyright Law of September 9, 1965, in its current version, and permission for use must always be obtained from Springer. Violations are liable to prosecution under the German Copyright Law.

The use of general descriptive names, registered names, trademarks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

*Cover design:* SPi Publisher Services

Printed on acid-free paper

Springer is part of Springer Science+Business Media ([www.springer.com](http://www.springer.com))

## Dedication to Klaus Starke

This volume of the *Handbook of Experimental Pharmacology* was one of the last that Klaus Starke initiated as Editor-in-Chief. We dedicate this 200th volume in the series to him.

Klaus was born in 1937. Since his father was an apothecary, it was natural for him to study pharmacy, but he also graduated in medicine. He combined the two, and devoted himself to pharmacology, where he has left a very important mark. Despite calls from many other universities, he remained in Freiburg from 1977. He was one of the scientists who simultaneously realized that neurotransmitters can regulate their own release via presynaptic receptors. This discovery has been extremely important in our understanding of the fine-tuning of neuronal activity. The pharmacology of these presynaptic receptors was recently summarized in volume 184 of the handbook, which was edited by Klaus together with Thomas Südhof. Klaus is deeply respected internationally for his solid science. He is a member of the Academia Europea, the Heidelberger Akademie der Wissenschaften, and the National German Academy “Deutsche Akademie der Naturforscher Leopoldina.” He received the Ernst Jung-Preis and the Wilhelm Feldberg-Preis.

In addition to his scientific accomplishments, Klaus has influenced and shaped German pharmacology by his teaching and his dedicated work on numerous committees and in grant-awarding agencies. He has also been deeply involved in many pharmacology journals. The extreme care he took with each manuscript submitted to *Naunyn-Schmiedeberg's Archives of Pharmacology* during his period as managing editor is very memorable to all of us who benefited from his penetrating, but benevolent editing. This devoted attention to both detail and strategic goals was brought by him to the *Handbook of Experimental Pharmacology*. He had, for example, several excellent suggestions on how to organize this volume and he even suggested that we include two poems. One of them was used in the introduction to this volume. The other is printed below. It shows his interests in cultural activities outside pharmacology.

Given the care with which Klaus always dealt with everything, it is with some trepidation that we submit this volume to his scrutiny.



Bertil Fredholm  
Volume Editor



Franz Hofmann  
Editor-in-Chief

For lo! the Board with Cups and Spoons is crown'd,  
The Berries crackle, and the Mill turns round;  
On shining Altars of *Japan* they raise  
The silver Lamp, and fiery Spirits blaze:  
From silver Spouts the grateful Liquors glide,  
And *China's* earth receives the smoking Tyde.  
At once they gratify their Scent and Taste,  
While frequent Cups prolong the rich Repast. . .  
*Coffee* (which makes the Politician wise,  
And see through all things with his half-shut Eyes)  
Sent up in Vapours to the *Baron's* Brain  
New Stratagems, the radiant Lock to gain.

Alexander Pope (1688–1744) *The Rape of the Lock*

# Preface

Methylxanthines are doubtless the most widely consumed of all pharmacologically active agents. The reason for this is, of course, that caffeine-containing beverages are consumed on a daily basis by the majority of humans. The human use of coffee and tea was limited until surprisingly recently. Now the global use means that coffee and tea are very important products commercially. Indeed, the sale of tea and coffee has been an important source of national income and for a long time provided the main source of income of the greatest nation in the world at the time, China.

Methylxanthines are found in several plants, from many parts of the world. Coffee beans were probably discovered in Africa, tea leaves in East Asia, mate and cocoa in South America, but it is also found in some 100 other plant species. To make these compounds the plants have developed sophisticated enzymatic machinery. The reason for the investment in methylxanthine synthesis is possibly because methylxanthines can act as a chemical defense, and hence because methylxanthines can have toxic effects. Caffeine is taken up well and distributed throughout the body and elimination depends on a series of enzymatic steps. These differ between species and ages of the same species, including man.

At the beginning of human use of both coffee and tea, the focus was on the medicinal effects, which were both lauded as beneficial and deplored as being detrimental. Now the major interest is perhaps in the public health consequences of the widespread use. Over the years, considerable effort has been spent in population studies to elucidate the risks of caffeine use. One of the surprising things in recent years has been the realization that the evidence for health benefits in, e.g., Parkinson's disease and type II diabetes, has been easier to document than that for possible detrimental effects in, e.g., cardiovascular disease. There are also some possibilities to use methylxanthines or derivatives as drugs. While this is good news, the bad news is that we are still not clear how these effects are brought about. There have been concerns that caffeine may be a major reproductive hazard, but provided that women limit their intake, this may not be a real concern.

Methylxanthines were early shown to cause muscle contractions in high doses, an effect we now know is due to mobilization of intracellular caffeine. In somewhat lower doses, caffeine and theophylline were found to prevent the enzymatic



hydrolysis of cyclic AMP. At still lower doses, they block the actions of adenosine at its receptors. All these actions, and some others, contribute to give methylxanthines a complex pharmacological profile, where utmost care must be taken with dosing.

In this volume of the *Handbook of Experimental Pharmacology*, well-known experts describe the facts alluded to above in detail with a focus on caffeine and theophylline. A special chapter is devoted to theobromine, an active component of chocolate, the actions of which are less well characterized. We also present the pharmacology of one xanthine derivative, propentofylline, as an example of a xanthine that has gone through extensive development for a novel therapeutic area.

The powerful effects caffeine exerts on the nervous system are covered. The ability of methylxanthines to influence the physiological processes involved in sleep and the pathophysiological processes involved in pain are described as largely secondary to adenosine antagonism. Methylxanthines can provoke epileptic seizures, and prevent neurodegenerative disease, but the possible mechanisms, involving actions on one or more adenosine receptors, on both neuronal and nonneuronal cells have not yet been fully elucidated. There are interesting therapeutic possibilities, and novel xanthine derivatives are being examined. The fact that caffeine-containing beverages have so rapidly established themselves in a variety of cultural settings raises the possibility that caffeine may actually be a dependence-producing drug. Indeed, there are important interactions with some of the neural systems involved in dependence, but caffeine is not a typical drug of addiction, despite the fact that in the famous coffee cantata of Bach (see below) the heroine is almost willing to forego the pleasures of sex for coffee.

It has also been well known for a long time that caffeine (and some of its metabolites) can influence respiration and can be used to treat asthma, that there are increases in cardiac activity and blood pressure, and that methylxanthines have marked renal effects. In all these instances, a major explanation for the effects is blockade of the actions of endogenous adenosine. This is also the reason why methylxanthines can influence cells of the immune system, an action with therapeutic implications, which has been realized for a much shorter time. By contrast, there is evidence that the metabolic effects of coffee and tea may not be entirely explained by adenosine receptor blockade, or by the caffeine content for that matter.

It has been a pleasure to work with world experts in a common effort to produce an up-to-date and authoritative account of the pharmacology of methylxanthines. We have aimed to give more than just a description of facts or findings, and instead to present ideas, concepts, and open questions.

“Ei! wie schmeckt der Coffee süße,  
Lieblicher als tausend Küsse,  
Milder als Muskatwein.  
Coffee, Coffee muß ich haben,  
Und wenn jemand mich will laben,  
Ach, so schenkt mir Coffee ein!”

Lieschens Aria (fourth movement) from Bach's Coffee Cantata BWV 211, “Schweigst stille, plaudert nicht.”

# Contents

<b>Notes on the History of Caffeine Use</b> .....	1
Bertil B. Fredholm	
<b>Distribution, Biosynthesis and Catabolism of Methylxanthines in Plants</b> .....	11
Hiroshi Ashihara, Misako Kato, and Alan Crozier	
<b>Pharmacokinetics and Metabolism of Natural Methylxanthines in Animal and Man</b> .....	33
Maurice J. Arnaud	
<b>Inhibition of Cyclic Nucleotide Phosphodiesterases by Methylxanthines and Related Compounds</b> .....	93
Sharron H. Francis, Konjeti R. Sekhar, Hengming Ke, and Jackie D. Corbin	
<b>Methylxanthines and Ryanodine Receptor Channels</b> .....	135
Serge Guerreiro, Marc Marien, and Patrick P. Michel	
<b>Xanthines as Adenosine Receptor Antagonists</b> .....	151
Christa E. Müller and Kenneth A. Jacobson	
<b>Theobromine and the Pharmacology of Cocoa</b> .....	201
Hendrik Jan Smit	
<b>Propentofylline: Glial Modulation, Neuroprotection, and Alleviation of Chronic Pain</b> .....	235
Sarah Sweitzer and Joyce De Leo	
<b>Methylxanthines, Seizures, and Excitotoxicity</b> .....	251
Detlev Boison	

<b>Impacts of Methylxanthines and Adenosine Receptors on Neurodegeneration: Human and Experimental Studies</b> .....	267
Jiang-Fan Chen and Yijuang Chern	
<b>Methylxanthines and Pain</b> .....	311
Jana Sawynok	
<b>Methylxanthines and Sleep</b> .....	331
Tarja Porkka-Heiskanen	
<b>Methylxanthines and Reproduction</b> .....	349
Alba Minelli and Ilaria Bellezza	
<b>Methylxanthines During Pregnancy and Early Postnatal Life</b> .....	373
Ulrika Ådén	
<b>Methylxanthines and the Kidney</b> .....	391
Hartmut Osswald and Jürgen Schnermann	
<b>The Cardiovascular Effects of Methylxanthines</b> .....	413
Niels P. Riksen, Paul Smits, and Gerard A. Rongen	
<b>Methylxanthines in Asthma</b> .....	439
Stephen L. Tilley	
<b>Methylxanthines and Inflammatory Cells</b> .....	457
György Haskó and Bruce Cronstein	
<b>Methylxanthines, Inflammation, and Cancer: Fundamental Mechanisms</b> .....	469
Akio Ohta and Michail Sitkovsky	
<b>Methylxanthines and Drug Dependence: A Focus on Interactions with Substances of Abuse</b> .....	483
Micaela Morelli and Nicola Simola	
<b>Methylxanthines and Human Health: Epidemiological and Experimental Evidence</b> .....	509
Marie-Soleil Beaudoin and Terry E. Graham	
<b>Index</b> .....	549

# Contributors

## **Ulrika Ådén**

Department of Woman and Child Health, Karolinska Institute, 171 77, Stockholm, Sweden, [ulrika.aden@ki.se](mailto:ulrika.aden@ki.se)

## **Maurice J. Arnaud**

Nutrition and Biochemistry, Bourg-Dessous 2A, 1814, La Tour-de-Peilz, Switzerland, [mauricearnaud@hotmail.com](mailto:mauricearnaud@hotmail.com)

## **Hiroshi Ashihara**

Department of Biological Sciences, Graduate School of Humanities and Sciences, Ochanomizu University, Otsuka, Bunkyo-ku, Tokyo 112-8610, Japan, [ashihara.hiroshi@ocha.ac.jp](mailto:ashihara.hiroshi@ocha.ac.jp)

## **Marie-Soleil Beaudoin**

Department of Human Health and Nutritional Sciences, University of Guelph, 50 Stone Road East, Guelph, ON N1G 2W1, Canada

## **Ilaria Bellezza**

Dipartimento di Medicina Sperimentale e Scienze Biochimiche, Università degli Studi di Perugia, Via del Giochetto, 06123 Perugia, Italy

## **Detlev Boison**

R.S. Dow Neurobiology Laboratories, Legacy Research, Portland, OR 97232, USA

## **Jiang-Fan Chen**

Department of Neurology, Boston University School of Medicine, 715 Albany Street, Boston, MA 02118, USA, [chenjf@bu.edu](mailto:chenjf@bu.edu)

**Yijuang Chern**

Institute of Biomedical Sciences, Academia Sinica, Taipei 11529, Taiwan

**Jackie D. Corbin**

Department of Molecular Physiology and Biophysics, Vanderbilt University School of Medicine, Light Hall Room 702, Nashville, TN 37232-0615, USA

**Bruce Cronstein**

Department of Medicine, New York University School of Medicine, 550 First Avenue, New York, NY 10016, USA

**Alan Crozier**

Plant Products and Human Nutrition Group, Division of Developmental Medicine, Faculty of Medicine, University of Glasgow, Graham Kerr Building, Glasgow G12 8QQ, UK, a.crozier@bio.gla.ac.uk

**Sharron H. Francis**

Department of Molecular Physiology and Biophysics, Vanderbilt University School of Medicine, Light Hall Room 702, Nashville, TN 37232-0615, USA  
sharron.francis@vanderbilt.edu

**Bertil B. Fredholm**

Department of Physiology and Pharmacology, Karolinska Institute, Nanna Svartz väg 2, 171 77 Stockholm, Sweden, bertil.fredholm@ki.se

**Terry E. Graham**

Department of Human Health and Nutritional Sciences, University of Guelph, 50 Stone Road East, Guelph, ON N1G 2W1, Canada, terrygra@uoguelph.ca

**Serge Guerreiro**

Université Pierre et Marie Curie-Paris 6, Centre de Recherche de l'Institut du Cerveau et de la Moelle Epinière, UMR-S975, Paris, France; Institut National de la Santé et de la Recherche Médicale, U-975, Paris, France; Centre National de la Recherche Scientifique, UMR 7225, Paris, France

**György Haskó**

Department of Surgery, University of Medicine and Dentistry of New Jersey-New Jersey Medical School, 185 South Orange Avenue, Newark, NJ 07103, USA

**Kenneth A. Jacobson**

Molecular Recognition Section, Laboratory of Bioorganic Chemistry, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Building 8A, Room BIA-19, 8 center Dr. 2089–0810 NIH, NIDDK, LBC, Bethesda, MD, USA

**Misako Kato**

Department of Biological Sciences, Graduate School of Humanities and Sciences, Ochanomizu University, Otsuka, Bunkyo-ku, Tokyo 112-8610, Japan, kato.misako@cha.ac.jp

**Hengming Ke**

Department of Biochemistry and Biophysics, University of North Carolina at Chapel Hill, 306 Mary Ellen Jones Building, Chapel Hill, NC 27599-7260, USA

**Joyce De Leo**

Department of Pharmacology and Toxicology, Dartmouth Medical School, Hanover, NH 03755, USA, joyce.a.deleo@dartmouth.edu

**Marc Marien**

Institut de Recherche Pierre Fabre, Castres, France

**Patrick P. Michel**

Université Pierre et Marie Curie-Paris 6, Centre de Recherche de l'Institut du Cerveau et de la Moelle Epinière, UMR-S975, Paris, France; Institut National de la Santé et de la Recherche Médicale, U-975, Paris, France; Centre National de la Recherche Scientifique, UMR 7225, Paris, France, patrick-pierre.michel@upmc.fr

**Alba Minelli**

Dipartimento di Medicina Sperimentale e Scienze Biochimiche, Università degli Studi di Perugia, Via del Giochetto, 06123 Perugia, Italy, aminelli@unipg.it

**Micaela Morelli**

Department of Toxicology, University of Cagliari, Via Ospedale 72, 09124 Cagliari, Italy; Centre of Excellence for Neurobiology of Dependence, University of Cagliari, 09124 Cagliari, Italy; CNR Institute of Neuroscience, Cagliari, Italy, morelli@unica.it

**Christa E. Müller**

Pharma Center Bonn, Pharmaceutical Sciences Bonn (PSB), University of Bonn, Pharmaceutical Institute, An der Immenburg 4, 53121 Bonn, Germany, christa.mueller@uni-bonn.de

**Akio Ohta**

Department of Pharmaceutical Sciences, New England Inflammation and Tissue Protection Institute, Northeastern University, 134 Mugar Building, 360 Huntington Avenue, Boston, MA 02115, USA, a.ohta@neu.edu

**Hartmut Osswald**

Department of Pharmacology and Toxicology, University of Tübingen, Wilhelmstrasse 56, 72074 Tübingen, Germany

**Tarja Porkka-Heiskanen**

Institute of Biomedicine/Physiology, University of Helsinki, P.O. Box 63, Haartmaninkatu 8, 00014 Helsinki, Finland, porkka@cc.helsinki.fi

**Niels P. Riksen**

Department of Pharmacology-Toxicology and Internal Medicine, Radboud University Nijmegen Medical Centre, PO Box 9101, 6500 HB Nijmegen, The Netherlands, n.riksen@aig.umcn.nl

**Gerard A. Rongen**

Department of Pharmacology-Toxicology and Internal Medicine, Radboud University Nijmegen Medical Centre, PO Box 9101, 6500 HB Nijmegen, The Netherlands

**Jana Sawynok**

Department of Pharmacology, Dalhousie University, Halifax, NS B3H 1X5, Canada, sawynok@dal.ca

**Jürgen Schnermann**

National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Building 10, Room 4D51, 10 Center Drive MSC 1370, Bethesda, MD 20892, USA, jurgens@intra.niddk.nih.gov

**Konjeti R. Sekhar**

Department of Radiation Biology, Vanderbilt University School of Medicine, DD-1205 MCN, Nashville, TN 37232, USA

**Nicola Simola**

Department of Toxicology, University of Cagliari, Via Ospedale 72, 09124 Cagliari, Italy

**Michail Sitkovsky**

Department of Pharmaceutical Sciences, New England Inflammation and Tissue Protection Institute, Northeastern University, 134 Mugar Building, 360 Huntington Avenue, Boston, MA 02115, USA; Cancer Vaccine Center, Dana-Farber Cancer Institute, Harvard Institutes of Medicine, 77 Avenue Pasteur, Room 418, Boston, MA 02115, USA

**Hendrik Jan Smit**

Functional Food Centre, Oxford Brookes University, Headington Campus, Gypsy Lane, Oxford OX3 0BP, UK, [hsmit@brookes.ac.uk](mailto:hsmit@brookes.ac.uk)

**Paul Smits**

Department of Pharmacology-Toxicology and Internal Medicine, Radboud University Nijmegen Medical Centre, PO Box 9101, 6500 HB, Nijmegen, The Netherlands

**Sarah Sweitzer**

Department of Pharmacology, Physiology and Neuroscience, University of South Carolina, USC School of Medicine, Columbia, SC 29208, USA

**Stephen L. Tilley**

Department of Medicine, Division of Pulmonary and Critical Care Medicine, and Center for Environmental Medicine, Asthma, and Lung Biology, University of North Carolina, Chapel Hill, NC 27599, USA, [stephen\\_tilley@med.unc.edu](mailto:stephen_tilley@med.unc.edu)





# Notes on the History of Caffeine Use

**Bertil B. Fredholm**

## Contents

1	Mythological Origins .....	2
2	Early History of Coffee .....	3
3	Early History of Tea .....	5
4	The History of Cocoa and Maté .....	7
5	Methylxanthines and Health .....	8
6	The Early Science of Methylxanthines .....	8
	References .....	9

**Abstract** As behooves something so deeply entrenched in culture, the historical origins of the use of methylxanthines are unknown and dressed in myth. This is true for coffee as well as tea, and for both it is interesting to note that their common use is really very recent. For coffee we know that its use became more widespread in the fifteenth and sixteenth centuries, and in Europe this occurred in the eighteenth and nineteenth centuries. The use of tea became more common during the Ming Dynasty in China and during the eighteenth century in Britain. Coffee was mostly an upper-class drink in Arabia, and remained a relative luxury in Europe until quite recently. The use of other methylxanthine-containing beverages, such as maté, is even less well known. It is interesting to note that before these drinks were commonly used on a daily basis they were used for medicinal purposes, indicating that their pharmacological actions had long been noted.

**Keywords** Cocoa · Coffee · Maté · Tea

---

B.B. Fredholm

Department of Physiology and Pharmacology, Karolinska Institute, Nanna Svartz väg 2, 171 77 Stockholm, Sweden

e-mail: bertil.fredholm@ki.se

## 1 Mythological Origins<sup>1</sup>

The coffee bush grows wild in many parts of Africa and it may also have been indigenous in Arabia. There is, however, little solid evidence that people had knowledge that the coffee bush could yield a useful drink before Islamic times. Solid evidence is replaced by good mythological stories. One myth is that the archangel Gabriel offered coffee brewed in heaven to Muhammad when he was overcome by sleepiness. A sip of this heavenly coffee was sufficient to make him so vigorous that he could “unhorse 40 men and make 40 women happy.” A more often told myth is about an Ethiopian (sometimes Arabian) goatherd, Kaldi, who observed that his goats became very agitated, indeed dancing, when they ate the berries of a certain bush. Having tried them himself, he informed the abbot of a local (perhaps Sufi) monastery, and from there the conquest of the earth was supposed to start. The story of Kaldi appears to occur first in a book by a Maronite professor of oriental languages, Antoine Faustus Nairon, but local sources for the legend are lacking (Nairon 1617). There are also other stories involving the role of the civet cat in spreading the coffee bush, which has been exploited to market very, very expensive kopi luwak coffee (Mair and Hoh 2009), where beans have passed through the gastrointestinal tract of the animal. In a manuscript published in Paris in 1699 by Antoine Galland, the so-called Abd-el-Kadir manuscript, a sheik Omar is credited with the discovery in AD 1258 during his banishment to a mountainous region near Ousab. There he detected a bush with white flowers and red berries and being famished, he cooked a soup of the berries and leaves and was very invigorated. The brew was also able to heal all illness in the surrounding villages.

The history of tea is similarly clouded in myth and national pride (Mair and Hoh 2009) – the Chinese maintain that tea has “always” been part of Chinese culture. A variant of “always” is the story that the nonhistorical second emperor Shen Nung discovered tea when tea leaves blew into his cup of hot water in 2737 BC. The credibility of this is reduced by the “fact” that this emperor was born with the head of a bull and body of a man: he spoke after 3 days, walked within a week, and could plow a field at age three. Many Chinese associate the widespread use of tea with the introduction and spread of Buddhism. According to one account, a Buddhist monk named Gan Lu brought tea back with him when he returned from a pilgrimage to India during the first century. He is supposed to have planted seven “fairy tea trees.” They are still shown to tourists on Mt Mengding in Sichuan.

Another, pretty grim, story says tea sprang from the eyelids of Bodhidharma, the first patriarch of Zen. He had sailed from India to China but after he arrived he merely sat down facing a wall at the Shaolin Temple and did not stir for 9 years. Unsurprisingly, the determined saint once drowsed off, so far forgetting himself that his eyes closed momentarily. Without hesitation he sliced off his eyelids to make sure they would never

---

<sup>1</sup>The author is heavily indebted to some excellent earlier treatises and the reader is referred to these for references (Ukers 1922; Weinberg and Bealer 2001; Elgklou 1993; Mair and Hoh 2009).

again close and interrupt his wakefulness. Where they fell the compassionate deity Quan Yin caused tea plants to grow to serve Bodhidharma and all who came after him as an aid on the path to enlightenment. The story may have gained particular popularity in Japan, because the Japanese characters for tea leaf and eyelid are the same.

## 2 Early History of Coffee

It seems clear that coffee beans were first eaten as such; later they were ground up and mixed with fat paste as a stimulating travel snack. Only around AD 1000 did infusions with boiling water start to be used. The word “coffee” derives from *qahva* (or *qahwah*), which is simply a word denoting a drink made from plants. From the initial cultures in Ethiopia, cultivation of coffee bushes soon came to be dominated by Yemen. The city of Mocha became a center – and its name came to denote the drink. Now coffee is grown in 50 different countries around the world (Ukers 1922; Weinberg and Bealer 2001; Elgklou 1993).

Although the Arabs cultivated the plants and prepared drinks from coffee beans, it was only by the fourteenth century that the process of roasting was discovered. And only when this happened did the use of coffee rapidly spread in the Arab world. In the Muslim world the need for a social drink was filled by coffee and the beverage was consumed both at home and in coffee bars. The fact that these coffee houses developed into independent intellectual centers was perceived as a threat to the authorities, and sometimes they were forced to close. Already in the sixteenth century health arguments were used, for example, when Kair Bey, the governor of Mecca, prohibited the use of coffee in 1511. The coffee bars started in the mid sixteenth century in Constantinople, and soon became very popular. They were called *quaweh khaneh* and developed into centers for cultural and intellectual activity. For that reason they also got the name *mekteb-i-irfan* (“the school of the wise”). In Turkey the use of coffee tended to decrease the use of opiates and was promoted by most officials. Since the Arabs controlled a vast territory at the time, the use of coffee spread to Spain, North Africa, India, Turkey, and the Balkans. The permeation into popular culture is revealed by its common use among women, such that a failure to provide sufficient coffee for the wife (or wives) was reason for dissolution of a marriage.

The use of coffee was described in travelogues. The first may have been that of Leonhard Rauwolf (1535–1596) from Augsburg, who published an account in 1582–1583. One particularly well-known description was in *De Plantis Aegypti liber* (Venice, 1592) by Prospero Alpini (1553–1617). He was born in the republic of Venice. After a time in the army, he went to Padua to study medicine, and settled as a physician outside Paduan territory. He traveled to Egypt in 1580 as physician to the Venetian consul in Cairo. He is reputed to have deduced that “the female date-trees or palms do not bear fruit unless the branches of the male and female plants are mixed together; or, as is generally done, unless the dust found in the male sheath or male flowers is sprinkled over the female flowers.” He returned to Padua and

became professor of botany. Although *De Plantis Aegypti liber* is his best-known work, an earlier work, *De Medicina Egyptiorum* (Venice, 1591), that describes Egyptian medical practices is said to contain the first account of the coffee plant published in Europe. He used the word *caova* to describe the drink.

Another influential writer was Pietro della Valle (1586–1652), who learned Arabic and traveled widely in Turkey, Egypt, Mesopotamia, and Persia. His name for the drink was *cahve*. He was well connected with the Vatican and this argues that it is probably incorrect that Pope Clemens VIII in 1605 (10 years before della Valle) was given coffee by members of the Curia who wanted him to ban this Muslim drink. He is reputed to have said: “This satanic drink is in truth so good that it would be a pity if only nonbelievers were allowed to drink it. We will fool Satan and baptize it so that it becomes a Christian drink, with no danger for the soul.”

According to one story, coffee was introduced to Europe in the seventeenth century. It is said that after their defeat at the gates of Vienna in 1683, the Turks left a large amount of coffee behind, and that the Viennese learned to prepare it and served it with half-moon-shaped cakes. It is, however, well established that coffee was introduced to Europe by Venetian merchants in early 1615. About 10 years later they had learned to roast and grind the imported green beans. The popularity of coffee was promoted by *botteghe del caffè* (coffee shops, originally called *botteghe delle acque e dei giacci*, because all kinds of drinks made with water and ice were served). They were modeled on the Arab establishments and also developed into gradually more sophisticated coffee establishments such as Caffé alla Venezia Trionfante, which opened in 1720. Here was the start of the typical café with newspapers and reviews that developed into cultural and commercial centers. By the mid eighteenth century Venice had more than 200 such establishments. The first Viennese establishment was established by Georg Kolshnitzky (according to legend using the Turkish leftover coffee sacks). In Paris, Café Procope, in rue de l’Ancienne Comédie, was opened in 1686 by the Sicilian Francesco Procopio dei Coltelli. It still exists and was given a name that alluded to hidden secrets in high places by alluding to the Byzantine historian Procopius, whose secret history had just been discovered and published for the first time ever in 1623. The café was a meeting place of the chic elite especially after Comédie Française opened across the street a few years later. It was a famous meeting place of the encyclopedists and the early revolutionaries, but also the literary elite after the political restoration and late nineteenth century met there. The first coffee house in London was apparently opened even earlier, in 1652.

Venice also became the transit port for shipment of coffee to other parts of Europe. By the end of the seventeenth century, the use of coffee in Europe was widespread. For some time the Arab countries maintained their monopoly of supply, but in the seventeenth century plants were smuggled to India and to Amsterdam and from there to the Dutch colonies in the East Indies. However, coffee had been grown outside the Arab countries and Ethiopia before that as small coffee plantations (and there had also been tea plantations of course) founded on Ceylon by the Portuguese in the early sixteenth century. By the beginning of the eighteenth century, Java and Sumatra were the main suppliers. The quality of the

Indonesian coffee was, however, for long inferior to that of the Yemenite coffee – at least if we are to believe Voltaire, who spoke of the “. . . mauvais café de Batavia et des îles” and explicitly preferred the Mocha coffee. One celebrated description of coffee farming is by Karen Blixen (pen name Isak Dinesen), whose book *Out of Africa* (Dinesen 1937) was filmed (albeit not following the book very closely) with Meryl Streep playing the author and Robert Redford playing her husband, Bror. The book starts, “I had a farm in Africa at the foot of the Ngong Hills...,” and in it she describes both ups and downs and the hard labor involved: “Coffee-growing is a long job. It does not all come out as you imagine, when, yourself young and hopeful, in the streaming rain, you carry the boxes of your shining young coffee plants from the nurseries...patiently, awaiting coming bounties” (Dinesen 1937; Lorenzetti and Lorenzetti 1999).

The spread of producers allowed coffee to become a mass product. It is culturally very much entrenched and it is relevant that the culture associated with the coffee house has been the subject of much intense research (Ellis 2007). It is beyond the scope of this historical sketch to outline the development of the coffee business as we now see it (Ukers 1922; Weinberg and Bealer 2001; Prendergast 1999). Suffice it to say here that it is remarkable that coffee has in a few hundred years become such an important part of everyday culture throughout the world that it has become one of the commercially most important traded commodities.

### 3 Early History of Tea

It is commonly stated that tea use was common in China for thousands of years before Christ. This is difficult to verify and there can be no one simple explanation for China’s nationwide adoption of the tea habit. Tea is (probably – but it could be other infusions) referred to in old texts from the first centuries BC. A clear unambiguous reference to the tea plant occurs only from AD 750. Alerting and mood-elevating effects are referred to in early literature. Indeed, this early literature mainly refers to the use of tea for medicinal purposes (Mair and Hoh 2009). However, by the Tang Dynasty, and particular Emperor Tai-tsung (627–649), the cultural and ceremonial aspects took a larger part. During this dynasty the famous *Chá Ching* (*The Classic of Tea*) was written as a manual of tea connoisseurship (Lu 1974) by Lu Yu. This developed into a ceremonial use of tea that incorporated aspects of both Taoism and Confucianism. Tea fell somewhat out of favor as a drink during the years of the Mongol Yuan Dynasty and it does not figure in Marco Polo’s descriptions of court life<sup>2</sup> (but he did describe how important taxes on tea were for the national economy). Tea clearly was of major importance in trade – in particular

---

<sup>2</sup>This has, however, also been interpreted as evidence that Marco Polo never visited the imperial court.

the trade of horses for tea, which represented a major part of the foreign/defense policy of China (Mair and Hoh 2009).

Tea use increased in popularity under the Chinese Ming Dynasty (1368–1644), which represented a return to power of the Han people, and it was in this period that tea began to be brewed by steeping cured loose leaves in boiling water. Because it was at this time that the tea was first tried by Europeans, it was this method of making tea that became popular in the West and remains so to this day. Also under the Ming Dynasty different types of teas, including fermented black teas, unfermented green teas, and the semifermented variety that is now known as oolong, were developed. During the Ming Dynasty there were tea houses that apparently functioned much as the coffee houses in Arab countries (Mair and Hoh 2009).

Tea was introduced to Japan around 800 BC and it came with all the ceremonial and quasi-religious overtones. However, it took many centuries until tea use became more popular, and it coincided in time with the spread of the Zen variant of Buddhism. The monk Myoan Eisai is credited with the popularization, including the demonstration for the shogun that tea can help you sober up a bit after too much sake. In Japan the institutions of the tea house and tea garden, as well as the Tea Ceremony, reached full development through a series of Zen monks (Mair and Hoh 2009). Of particular importance was Sen No Rikyu (1522–1591), Tea Master of the powerful political leader Hideyoshi, who incorporated the essence of Zen into the Tea Ceremony, and it is in the form he developed that the Way of Tea (*chado*) is practiced through the Tea Ceremony to this day. This classical ceremony is based on powdered green tea that is intensely whisked. By the mid sixteenth century traditions based on “boiled tea,” *sencha*, using intact tea leaves were established (Mair and Hoh 2009). A well-known description of tea use in Japan (with notes on the rest of the world) was written by Okakuzo Kakuzo (2000).

Tea arrived in Europe about the same time as coffee did. The first green tea leaves from China were brought to Amsterdam by the Dutch East India Company and tea was drunk in France by 1636. Tea in Russia was first offered by China as a gift to Czar Michael I in 1618. Tea appeared in Germany by 1657. In Britain tea was apparently first publicly distributed in the 1650s by Thomas Garraway and it was within the confines of his coffee house. Whereas tea never became very popular in mainland Europe (except Russia), by 1730 the use of tea in England had passed the use of coffee. The reason for this could be the strong position of the British East India Company. Despite the fact that tea is an indigenous Indian plant, culture of tea plants began in earnest only after the British rule. In particular, the teas producing the Darjeeling variety were from Chinese plants planted during British rule. The native Assam tea was also being cultivated more systematically, and cultivation was transferred to Ceylon.

Like the Arabs in the case of coffee, the Chinese wanted to monopolize tea trade, and, conversely, many wanted to get plants and grow them. One well-known story is how the Swedish naturalist Carl von Linné (1707–1778) attempted to get tea plants for his botanical garden in Uppsala. The idea was to test if tea could be grown locally and therefore improve the national economy. He made a deal with the Swedish East India Company to have one of his students travel with it and bring

home a plant. The student, Pehr Osbeck, managed to collect numerous plants and bring them home, but the one tea plant brought on board that survived most of the sea voyage during the first trip was knocked overboard when the crew fired a salute. He lost a plant on his second trip during a storm off the Cape of Good Hope. The third effort was a plant brought by a director of the East India Company directly to von Linné, but it soon proved not to be the proper plant at all but another species of *Camellia*. The fourth one was the proper thing, but was deemed so valuable that it was left in the commander's safe room – where it was devoured by rats. Finally, one of two plants brought as potted seeds was brought by the wife of the commander of the ship herself after the first one had been destroyed in transit. Finally, in October 1763 von Linné was the first person in Europe to have a tea plant of his own (Mair and Hoh 2009). The plants gradually died and by 1781 only one remained. By now tea is widely used throughout the world (but not very much in Sweden).

Even though coffee was the first methylxanthine-containing drink associated with Muslim countries<sup>3</sup>, tea has subsequently become the dominant drink in these parts of the world (Mair and Hoh 2009). In the UK the first such drink was again coffee, but by the nineteenth century there was a switch to tea; in the USA the opposite development is seen. In each of these cases there are interesting political overtones (Mair and Hoh 2009).

## 4 The History of Cocoa and Maté

These two methylxanthine-containing beverages have Latin American or Central American origins. Cocoa beans were used and the tree was cultured by the Olmec people (1500–400 BC), and was later used by the Maya culture. The Maya recorded their use in writing. After them Toltec and Aztec cultures used it, and via the latter the Spaniards (Hernán Cortés, 1485–1547) came in contact with the drink and its proper preparation. The huge consumption of chocolate by the Montezuma court was recorded by the historiographer Bernal Diaz del Castillo (Ukers 1922; Coe and Coe 1996).

In all probability the cocoa seeds brought by Columbus to King Ferdinand in 1502 were the first plant products containing methylxanthines brought to Europe, and the Spanish royalty was thus prepared when Cortes wrote a exuberant description calling cacao the “drink of goods.” Cortés returned to the Americas to set up cocoa plantations for King Charles V in Haiti, Trinidad, and Fernando Po (Weinberg and Bealer 2001; Coe and Coe 1996). The Indians probably mixed vanilla with the cocoa to reduce the bitter taste. In Europe sugar in different forms was added as were milk products. Now, of course, cocoa products are eaten as much as they are drunk (Coe and Coe 1996).

---

<sup>3</sup>However, tea export from China to muslim countries had been important for a long time.



Maté is prepared from the leaves of a holly (*Ilex paraguariensis*) harvested in pre-Hispanic times along the Paraná–Paraguay river system. It has oval, dark green leaves that are six to eight inches long and white flowers. As with tea, the best product is from young, unopened leaflets. The history of this use is clouded in mystery, but the use is often associated with the Guarani Indians, who have a myth of the maté being a present from a shaman to a traveling chief. It is clear that the Spanish invaders rapidly adopted the practice and saw economic possibilities. Jesuits developed maté into a plantation crop and before 1700 the use had spread along the Andes and the Rio de la Plata. However, the use has never spread much beyond South America.

## 5 Methylxanthines and Health

As already noted, in the earliest history of methylxanthine use the medicinal effects were important (Ukers 1922; Weinberg and Bealer 2001; Mair and Hoh 2009). For example, in India, where tea is indigenous, tea was long used for medicinal purposes, but its social use was limited. Just as there were overblown claims for the wholesome effects of the drinks made with coffee, tea, or cocoa, there were claims about the negative health consequences. A case in point is an apocryphal experiment of Gustaf III (1746–1792). This Swedish king was reported to be convinced that coffee was dangerous and in order to prove his opinion he is said to have given two condemned twins an alternative to death: they should take part in a long-term experiment. One twin was to drink coffee, the other tea. Reportedly, the experiment was ended when the tea-drinking twin died at age 83. Unfortunately much of the early literature of the health effects is of the same standard and the present author is not a great believer in the usefulness of mining old literature for clues about medicinal herbs in general. Only by the mid nineteenth century did the reports of health effects get a sufficiently scientific basis to be taken seriously. Some of the early literature is cited in relevant chapters of this volume. One very clear example is the reports of the usefulness of coffee in asthma (Salter 1860). There were also early correct reports on the negative effects of high doses of caffeine (Cole 1833). An attitude representative of much later literature is represented by the “discoverer” of homeopathy, Samuel Hahnemann (1755–1843), who found the alerting effects of caffeine to be very beneficial, but warned against its use because it involved the disruption of a natural balance (Hahnemann 1803).

## 6 The Early Science of Methylxanthines

It was early realized that the effects of coffee were due to some active principle. Prompted by Goethe, the young amateur chemist Friedrich Ferdinand Runge (1795–1865) was able to identify in a rather pure form a substance we call caffeine (Weinberg and Bealer 2001). The chemistry of caffeine and its relatives was

clarified by the great Hermann Emil Fischer (1852–1919) in a series of studies that were explicitly cited in his Nobel prize nomination and lecture in 1902. The study of caffeine was also his first major effort, published when he was 29–30 years of age (Fischer 1881a, b, 1882). Using oxidation with moist chlorine, he found that caffeine had a similar heterocyclic skeleton as uric acid. He then found that it was a trimethylxanthine, but he struggled for some time to clarify the structure of xanthine. However, it was only when Fischer turned to a synthetic approach to structure that he finally correctly realized that the xanthine structure proposed by Ludwig Medicus (1847–1915) prior to Fischer was the correct one (Fischer 1897).

Among the dimethylxanthines, theobromine was first identified in 1841. The name derives from the name of the cocoa plant and refers to the name given to it by Cortés, “the food of Gods” (*Theobroma*). Fischer discovered a synthesis in 1882. Theophylline was identified as a minor component in tea in 1888, and a synthetic pathway was devised by Fischer, who also established the structure. Already at the beginning of the twentieth century theophylline was introduced in the clinic, first as a diuretic, later in the treatment of asthma. The third dimethylxanthine, paraxanthine, is not a major component in plants but is the major metabolite of caffeine in several species, including man.

## References

- Dinesen I (1937) Out of Africa
- Coe SD, Coe MD (1996) The true history of chocolate. Thames and Hudson, London
- Cole J (1833) On the deleterious effects produced by drinking tea and coffee in excessive quantities. *Lancet* 2:274–278
- Elgklou L (1993) Kaffebooken. Wiken, Sverige
- Ellis M (ed) (2007) Eighteenth-century coffee house culture. Pickering and Chatto, London
- Fischer E (1881a) Über das Caffein. *Ber Dtsch Chem Ges* 14:637–644
- Fischer E (1881b) Über das Caffein. Zweite Mitteilung. *Ber Dtsch Chem Ges* 14:1905–1915
- Fischer E (1882) Über das Caffein. Dritte Mitteilung. *Ber Dtsch Chem Ges* 15:29–33
- Fischer E (1897) Über die Constitution des Caffeins, Xanthins. Hypoxanthins und verwandter Basen. *Ber Dtsch Chem Ges* 30:549–559
- Hahnemann S (1803) Der Kaffee und seine Wirkungen. Leipzig
- Kakuzo O (2000) The book of tea (first published 1906), new edition. Barnes and Noble, Boston
- Lorenzetti D, Lorenzetti LR (1999) ‘Out of Africa’: Karen Blixen’s coffee years. *Tea and Coffee Trade Journal*, 1 Sept
- Lu Y (1974) The classic of tea (translated and introduced by Francis Ross Carpenter). Brown, New Jersey
- Mair VH, Hoh E (2009) The true history of tea. Thames Hudson, London
- Nairon AF (1617) Referred to in Weinberg BA, Bealer BK (eds) *De Saluberrimá Cabue seu Café nuncupata Discoursus. The World of Caffeine*. Routledge, New York, 2001
- Prendergast M (1999) Uncommon grounds. The history of coffee and how it transformed our world. Basic Books, London
- Salter H (1860) Asthma: its pathology and treatment. Churchill, London
- Ukers WH (1922) All about coffee. The Tea and Coffee Trade Journal Company, New York
- Weinberg BA, Bealer BK (2001) The world of caffeine. The science and culture of the world’s most popular drug. Routledge, New York

# Distribution, Biosynthesis and Catabolism of Methylxanthines in Plants

Hiroshi Ashihara, Misako Kato, and Alan Crozier

## Contents

1	Introduction .....	12
2	Distribution of Methylxanthines in Plants .....	12
2.1	Coffee and Related <i>Coffea</i> Plants .....	13
2.2	Tea and Related <i>Camellia</i> Plants .....	14
2.3	Cacao and Related <i>Theobroma</i> and <i>Herrania</i> Plants .....	15
2.4	Maté, Guarana and Other Species .....	15
3	Methylxanthine Biosynthesis in Plants .....	15
3.1	Formation of 7-Methylxanthine .....	17
3.2	Formation of Theobromine .....	17
3.3	Conversion of Theobromine to Caffeine .....	18
3.4	Formation of Methyluric Acids .....	18
3.5	Supply of Xanthosine for Caffeine Biosynthesis .....	19
4	<i>N</i> -Methyltransferases Involved in Methylxanthine Biosynthesis .....	20
4.1	Gene Expression in Coffee and Tea Plants .....	20
4.2	Evolutionary Relationship of Caffeine Synthase and Related Enzymes .....	21
5	Catabolism of Methylxanthines in Plants .....	23
5.1	Conversion of Caffeine to Theophylline .....	23
5.2	Metabolism of Theophylline .....	25
5.3	Catabolism of Theobromine .....	25
6	Ecological Roles of Purine Alkaloids .....	26
6.1	Chemical Defence Theory .....	26
6.2	Allelopathy Theory .....	26

---

H. Ashihara (✉) and M. Kato

Department of Biological Sciences, Graduate School of Humanities and Sciences, Ochanomizu University, Otsuka, Bunkyo-ku, Tokyo 112-8610, Japan

e-mail: ashihara.hiroshi@ocha.ac.jp; kato.misako@ocha.ac.jp

A. Crozier

Plant Products and Human Nutrition Group, Division of Developmental Medicine, Faculty of Medicine, University of Glasgow, Graham Kerr Building, Glasgow G12 8QQ, UK

e-mail: a.crozier@bio.gla.ac.uk

7	Production of Decaffeinated Coffee .....	26
7.1	Production by Breeding .....	27
7.2	Production by Genetic Engineering .....	27
8	Summary and Perspectives .....	27
	References .....	28

**Abstract** Methylxanthines and methyluric acids are purine alkaloids that are synthesized in quantity in a limited number of plant species, including tea, coffee and cacao. This review summarizes the pathways, enzymes and related genes of caffeine biosynthesis. The main biosynthetic pathway is a sequence consisting of xanthosine → 7-methylxanthosine → 7-methylxanthine → theobromine → caffeine. Catabolism of caffeine starts with its conversion to theophylline. Typically, this reaction is very slow in caffeine-accumulating plants. Finally, the ecological roles of caffeine and the production of decaffeinated coffee plants are discussed.

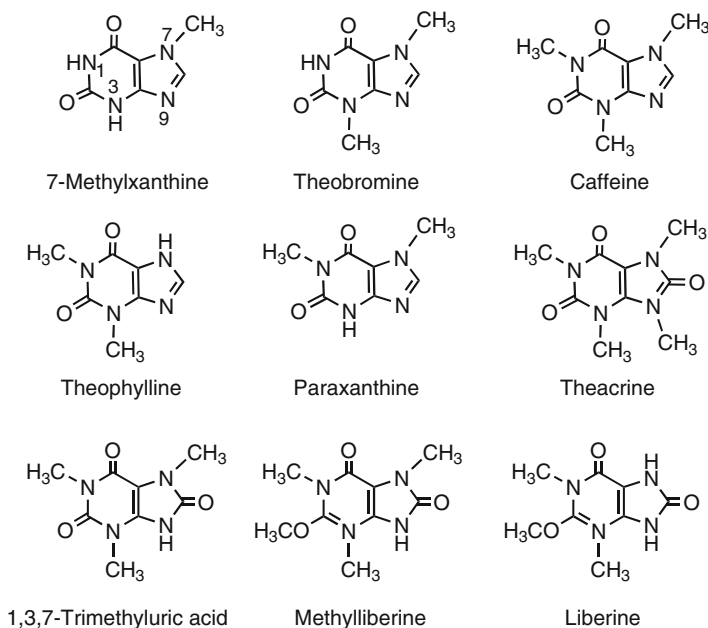
**Keywords** Biosynthesis · Caffeine · Catabolism · Coffee · *N*-Methyltransferase · Tea · Theobromine

## 1 Introduction

Methylxanthines and methyluric acids (Fig. 1) are secondary plant metabolites derived from purine nucleotides (Ashihara and Crozier 1999a). The most well known methylxanthines are caffeine (1,3,7-trimethylxanthine) and theobromine (3,7-dimethylxanthine), which occur in tea, coffee, cacao and a number of other non-alcoholic beverages of plant origin. Caffeine was isolated from tea and coffee in the early 1820s, but the main biosynthetic and catabolic pathways of caffeine were not fully established until recently, when highly purified caffeine synthase was obtained from tea leaves and a gene encoding the enzyme was cloned (Kato et al. 1999; Kato et al. 2000). In this chapter, the distribution, biosynthesis and catabolism of methylxanthines in plants are described. Furthermore, the roles of methylxanthines *in planta* and production of decaffeinated coffee plants are summarized.

## 2 Distribution of Methylxanthines in Plants

Methylxanthines have been found in nearly 100 species in 13 orders of the plant kingdom (Ashihara and Suzuki 2004; Ashihara and Crozier 1999a). Compared with other plant alkaloids, such as nicotine, morphine and strychnine, purine alkaloids are distributed widely throughout the plant kingdom although accumulation of high concentrations is restricted to a limited number of species, including *Coffea*



**Fig. 1** Structures of purine alkaloids present in plants constituting methylxanthines (7-methylxanthine, theobromine, caffeine, theophylline and paraxanthine) and methyluric acids (theacrine, 1,3,7-trimethyluric acid, methyllicberine and liberine)

*arabica* (coffee), *Camellia sinensis* (tea) and *Theobroma cacao* (cacao). All caffeine-containing plants, except *Scilla maritima*, belong to the Dicotyledoneae. In some species the main methylxanthine is theobromine or methyluric acids, including theacrine (1,3,7,9-tetramethyluric acid), rather than caffeine (Ashihara and Crozier 1999a).

## 2.1 Coffee and Related Coffee Plants

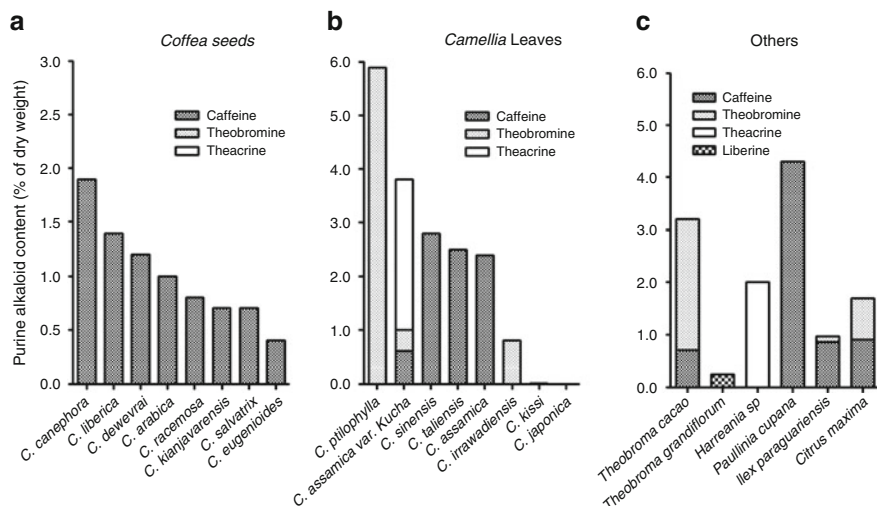
The caffeine content of seeds of different *Coffea* species varies from 0.4 to 2.4% dry weight (Mazzafera and Carvalho 1992). Green beans (as opposed to roasted beans, which are used to prepare the beverage) of current commercially cultivated coffee plants contain substantial quantities of caffeine; arabica coffee (*Coffea arabica*) beans usually contain 1.2–1.4% caffeine (Charrier and Berthaud 1975), while robusta coffee (*Coffea canephora*) contains 1.2–3.3% caffeine (Charrier and Berthaud 1975; Mazzafera and Carvalho 1992). There are also several wild coffee species where the green beans contain either no caffeine or extremely low levels of caffeine. Such low-caffeine species include *Mascarocoffea* sp. and *Coffea eugenioides* (Mazzafera and Carvalho 1992; Rakotomalala et al. 1992; Campa et al. 2005).

Caffeine is distributed mainly in the leaves and cotyledons of *Coffea arabica* seedlings, at concentrations ranging from 0.8 to 1.9% dry weight. Essentially, there

is no caffeine in roots or in the older brown parts of the shoot (Zheng and Ashihara 2004). Mature leaves of *Coffea liberica*, *Coffea dewevrei* and *Coffea abeokutae* contain the methyluric acids theacrine, liberine [*O*(2),1,9-trimethyluric acid] and methyltheacrine [*O*(2),1,7,9-tetramethyluric acid] (Fig. 1) (Baumann et al. 1976; Petermann and Baumann 1983). Examples of the purine alkaloid content in the seeds of *Coffea* species are illustrated in Fig. 2a.

## 2.2 Tea and Related Camellia Plants

The caffeine content of young leaves of first flush shoots of *Camellia sinensis*, *Camellia assamica* and *Camellia taliensis* is 2–3% of dry weight, while the level in *Camellia kissi* is less than 0.02%. Unusually, theobromine is the predominant purine alkaloid (5.0–6.8%) in young leaves of a Chinese tea, kekecha (cocoa tea) (*Camellia ptilophylla*) (Ye et al. 1997), and *Camellia irrawadiensis* (less than 0.8%) (Nagata and Sakai 1985). Theacrine and caffeine are the major purine alkaloids in the leaves of another Chinese tea called “kucha” (*Camellia assamica* var. *kucha*). The endogenous levels of theacrine and caffeine in expanding buds and young leaves of kucha are approximately 2.8 and 0.6–2.7%, respectively (Zheng et al. 2002). Some examples of the purine alkaloid content of the leaves of *Camellia* species are shown in Fig. 2b.



**Fig. 2** The methylxanthine and methyluric acid content of selected plant species. **a** Leaves of *Camellia* species, **b** seeds of *Coffea* species and **c** seeds of *Theobroma cacao* (cacao), *Theobroma grandiflorum* (cupu), *Herrania* sp. and *Paullinia cupana* (guarana), leaves of *Ilex paraguariensis* (maté) and anthers of *Citrus maxima* (pomelo). Values were obtained from references cited in the text

### 2.3 Cacao and Related *Theobroma* and *Herrania* Plants

Theobromine is the dominant purine alkaloid in seeds of cacao (*Theobroma cacao*). The cotyledons of mature beans contain 2.2–2.7% on a dry weight basis and 0.6–0.8% caffeine, while shells contain 0.6–0.7% theobromine and 0.5–0.6% caffeine (Senanayake and Wijesekera 1971). Examination of several cacao genotypes representing the three horticultural races Criollo, Forastero and Trinitario revealed considerable variations in the purine alkaloid content of the seed, with slightly higher levels found within the Criollo types (Hammerstone et al. 1994). Roasted seeds of *Theobroma cacao* are used to make cocoa and chocolate products (Duthie and Crozier 2003).

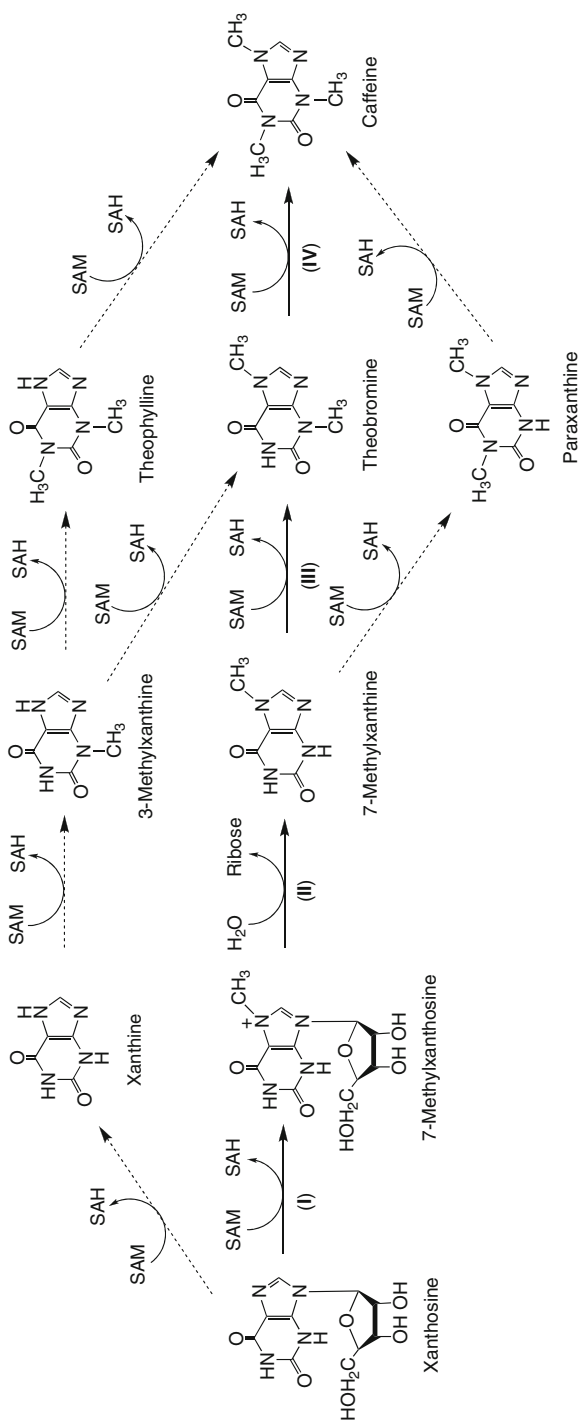
Cupu (*Theobroma grandiflorum*) contains 0.25% liberine in cotyledons and 0.08% in the nut shells (Baumann and Wanner 1980). Hammerstone et al. (1994) reported that theacrine is the principal purine alkaloid in seeds of 11 species of *Theobroma* and nine species of *Herrania*. Quantitative data on purine alkaloid levels in *Theobroma* and *Herrania* species are presented in Fig. 2c.

### 2.4 Maté, Guarana and Other Species

Maté (*Ilex paraguariensis*) leaves are used to make a beverage that is consumed widely in rural areas of Argentina, Paraguay and Brazil. Young maté leaves contain caffeine (0.8–0.9%), theobromine (0.08–0.16%) and theophylline (less than 0.02%). Methylxanthines have been detected in *Paullinia cupana* (guarana), *Paullinia yoco*, *Paullinia pachycarpa*, *Cola* species and *Citrus* species (Baumann et al. 1995; Kretschmar and Baumann 1999; Weckerle et al. 2003). In seeds of guarana, caffeine is located mainly in the cotyledons (4.3%) and testa (1.6%). Citrus flowers can accumulate up to 0.17% methylxanthines on a fresh weight basis; caffeine is the main methylxanthine, but theophylline is also present. Trace quantities of caffeine have also found in the nectar of citrus flowers (Weckerle et al. 2003). Quantitative data of selected samples are shown in Fig. 2c.

## 3 Methylxanthine Biosynthesis in Plants

Methylxanthines are formed from purine nucleotides in plants. Historically, there have been a number of proposals on the pathways involved in such conversions (see Ashihara and Crozier 1999a). However, data from studies on in situ metabolism of labelled precursors, as well as enzymes and genes have established that the main caffeine biosynthetic pathway is a four-step sequence consisting of three methylations and one nucleosidase reaction starting with xanthosine acting as the initial substrate (Fig. 3). Although the information has been obtained mainly from coffee



**Fig. 3** The biosynthetic pathways of caffeine from xanthosine. The major pathway consists of four steps from I to IV. The enzymes involved are as follows: 7-methylxanthosine synthase (EC 2.1.1.158) (I and II); N-methyltransferase (EC 3.2.2.25) (III); theobromine synthase (EC 2.1.1.159) (IV); caffeine synthase (EC 2.1.1.160) (III and IV). Minor pathways, shown with *dotted arrows*, may occur because of the broad substrate specificities of the N-methyltransferases. SAM S-adenosyl-L-methionine, SAH S-adenosyl-L-homocysteine



(*Coffea arabica*) and tea (*Camellia sinensis*), the available evidence suggests that the pathway is essentially the same in other methylxanthine-forming plants (Ashihara et al. 1998; Zheng et al. 2002; Koyama et al. 2003).

### 3.1 Formation of 7-Methylxanthine

The formation of monomethylxanthine in the main caffeine biosynthetic pathway is initiated by the conversion of xanthosine to 7-methylxanthosine (Fig. 3). This reaction is catalysed by 7-methylxanthosine synthase (xanthosine 7*N*-methyltransferase, EC 2.1.1.158). The genes encoding 7-methylxanthosine synthase, *CmXRS1* (AB034699) and *CaXMT* (AB048793), were isolated from *Coffea arabica* (Mizuno et al. 2003a; Uefuji et al. 2003). The second step involves a nucleosidase which catalyses the hydrolysis of 7-methylxanthosine. It was thought that *N*-methylnucleosidase (EC 3.2.2.25), which occurs in tea leaves, participates in this reaction (Negishi et al. 1988), but structural studies on coffee 7-methylxanthosine synthase suggested that the methyl transfer and nucleoside cleavage may be coupled and catalysed by a single enzyme (McCarthy and McCarthy 2007).

### 3.2 Formation of Theobromine

The third step in the caffeine biosynthesis pathways is also catalysed by *S*-adenosyl-L-methionine (SAM)-dependent *N*-methyltransferase(s). Highly purified caffeine synthase (EC 2.1.1.160) obtained from young tea leaves has broad substrate specificity and catalyses the two-step conversion of 7-methylxanthine to caffeine via theobromine (Kato et al. 1999). This enzyme is distinct from the *N*-methyltransferase that catalyses the first methylation step in the caffeine pathway. The isolated complementary DNA from young tea leaves, termed *TCS1* (AB031280), consists of 1,438 base pairs and encodes a protein of 369 amino acids (Kato et al. 2000). The function of *TCS2* (AB031281), which occurs as a paralogous gene to *TCS1* in the tea genome, has not yet been determined (Yoneyama et al. 2006). Plural genes encoding *N*-methyltransferases which have different substrate specificities have been isolated from coffee plants. *CCS1* (AB086414), *CtCS7* (AB086415) and *CaDXMT1* (AB084125) are caffeine synthase genes (Mizuno et al. 2003a; Uefuji et al. 2003). The recombinant caffeine synthases (EC 2.1.1.160) can utilize paraxanthine, theobromine and 7-methylxanthine as substrates. *CTS1* (AB034700), *CTS2* (AB054841), *CaMXMT1* (AB048794) and *CaMXMT2* (AB084126) were identified as genes encoding theobromine synthase (Mizuno et al. 2001; Ogawa et al. 2001). The activity of the recombinant theobromine synthase (EC 2.1.1.159) is specific for the conversion of 7-methylxanthine to theobromine.

Theobromine synthase, but not the dual-functional caffeine synthase, appears to participate principally in theobromine synthesis in theobromine-accumulating plants, such as *Theobroma cacao*, *Camellia ptilophylla* and *Camellia irrawadiensis* (Yoneyama et al. 2006).

### 3.3 Conversion of Theobromine to Caffeine

Conversion of theobromine to caffeine is performed by the dual-functional caffeine synthase discussed already. The methylation of N1 of 7-methylxanthine by caffeine synthase is much slower than that of N3, and as a consequence, theobromine is temporally accumulated in caffeine-synthesizing tissues. This is the final step in the main caffeine biosynthesis pathway, i.e., xanthosine  $\rightarrow$  7-methylxanthosine  $\rightarrow$  7-methylxanthine  $\rightarrow$  theobromine  $\rightarrow$  caffeine.

To date, three caffeine synthase genes have been identified in coffee plants (Mizuno et al. 2003b; Uefuji et al. 2003). Expression profiles of these genes in different organs are variable and the kinetic properties of each recombinant enzyme, such as  $k_m$  values, are different. Therefore, the enzymes participating in caffeine biosynthesis in organs and at different stages of growth may vary.

In addition to the main caffeine biosynthesis pathway, various minor routes may also operate (Fig. 3) which are mainly dependent upon the broad specificities of the *N*-methyltransferases, especially caffeine synthase. For example, caffeine synthase catalyses the synthesis of 3-methylxanthine from xanthine. Paraxanthine is synthesized from 7-methylxanthine. However, little accumulation of these compounds occurs in plant tissues. 3-Methylxanthine may be catabolized to xanthine, and paraxanthine appears to be immediately converted to caffeine. Paraxanthine is the most active substrate of caffeine synthase, but only limited amounts of paraxanthine accumulate in plant tissues, because the N1-methylation of 7-methylxanthine is very slow (Ashihara et al. 2008).

### 3.4 Formation of Methyluric Acids

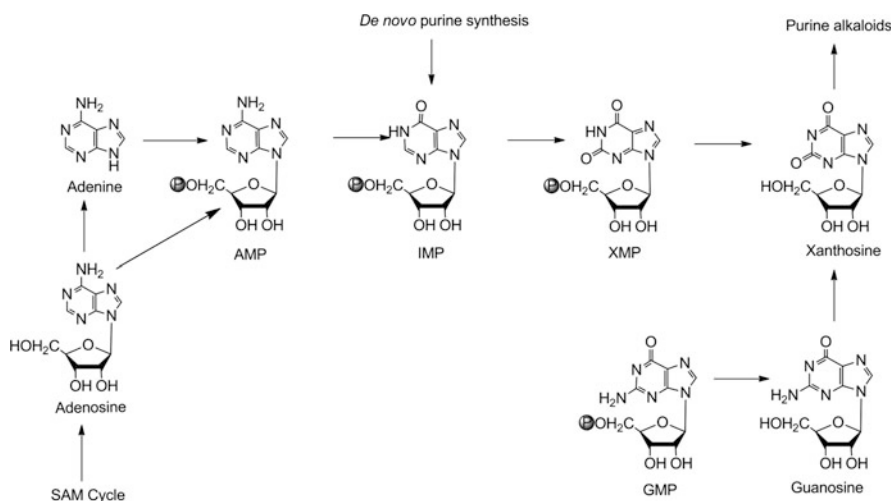
Formation of methyluric acids occurs in a limited number of plant species. As noted in Sect. 2.2, theacrine is found in kucha leaves in high concentrations (Zheng et al. 2002). Radiolabelled feeding experiments, indicate that theacrine is synthesized from caffeine. Conversion of caffeine to theacrine probably occurs by successive oxidation and methylation steps with 1,3,7-trimethyluric acid acting as the intermediate. Leaves of *Coffea dewevrei*, *Coffea liberica* and *Coffea abeokuta* convert caffeine to liberine probably via theacrine and methyl liberine (Petermann and Baumann 1983).

### 3.5 Supply of Xanthosine for Caffeine Biosynthesis

Xanthosine, the initial substrate of purine alkaloid synthesis, is supplied by at least four different pathways: de novo purine biosynthesis (de novo route), degradation of adenine nucleotides (AMP route), the SAM cycle (SAM route) and guanine nucleotides (GMP route) (Fig. 4).

#### 3.5.1 De Novo Route

Like mammals, plants synthesize purine nucleotides by de novo and salvage pathways (Ashihara and Crozier 1999a; Moffatt and Ashihara 2002; Stasolla et al. 2003), although some sections of the pathways are unique to plants. Utilization of IMP, formed by the de novo purine biosynthetic pathway, for caffeine biosynthesis was demonstrated in young tea leaves using  $^{15}\text{N}$ -glycine and  $^{14}\text{C}$ -labelled precursors and inhibitors of de novo purine biosynthesis (Ito and Ashihara 1999). Xanthosine is produced by an  $\text{IMP} \rightarrow \text{XMP} \rightarrow \text{xanthosine}$  pathway. IMP dehydrogenase (EC 1.1.1.205) and  $5'$ -nucleotidase (EC 3.1.3.5) catalyse these reactions. Ribavirin, an inhibitor of IMP dehydrogenase, reduces the rate of caffeine biosynthesis in tea and coffee plants (Keya et al. 2003).



**Fig. 4** Formation of xanthosine for caffeine biosynthesis from purine nucleotides and SAM. Xanthosine is produced via at least four routes: from IMP originating from de novo purine synthesis (de novo route), from the cellular adenine nucleotide pool (AMP route), from adenosine released from the SAM cycle (SAM route), and from the guanine nucleotide pool (GMP route)

### 3.5.2 AMP Route

A portion of the xanthosine used for caffeine biosynthesis is derived from the adenine and guanine nucleotide pools which are produced by the de novo and salvage pathways. There are several potential pathways for xanthosine synthesis from AMP, although the  $\text{AMP} \rightarrow \text{IMP} \rightarrow \text{XMP} \rightarrow \text{xanthosine}$  route is likely to predominate. All three enzymes involved in the conversion have been detected in tea leaves (Koshiishi et al. 2001).

### 3.5.3 SAM Route

The SAM route is a variation of the AMP route. SAM is the methyl donor for various methylation reactions in the caffeine biosynthetic pathway. In the process, SAM is converted to *S*-adenosyl-L-homocysteine (SAH), which is then hydrolysed to homocysteine and adenosine. Homocysteine is recycled via the SAM cycle to replenish SAM levels, and adenosine released from the cycle is converted to AMP and utilized for caffeine biosynthesis by the AMP route. Since 3 moles of SAH are produced via the SAM cycle for each mole of caffeine that is synthesized, in theory this pathway has the capacity to be the sole source of both the purine skeleton and the methyl groups required for caffeine biosynthesis in young tea leaves (Koshiishi et al. 2001).

### 3.5.4 GMP Route

Xanthosine utilized for caffeine biosynthesis is also produced from guanine nucleotides by a  $\text{GMP} \rightarrow \text{guanosine} \rightarrow \text{xanthosine}$  pathway. 5'-Nucleotidase (EC 3.1.3.5) and guanosine deaminase (EC 3.5.4.15) participate in this conversion (Negishi et al. 1994).

## 4 N-Methyltransferases Involved in Methylxanthine Biosynthesis

### 4.1 Gene Expression in Coffee and Tea Plants

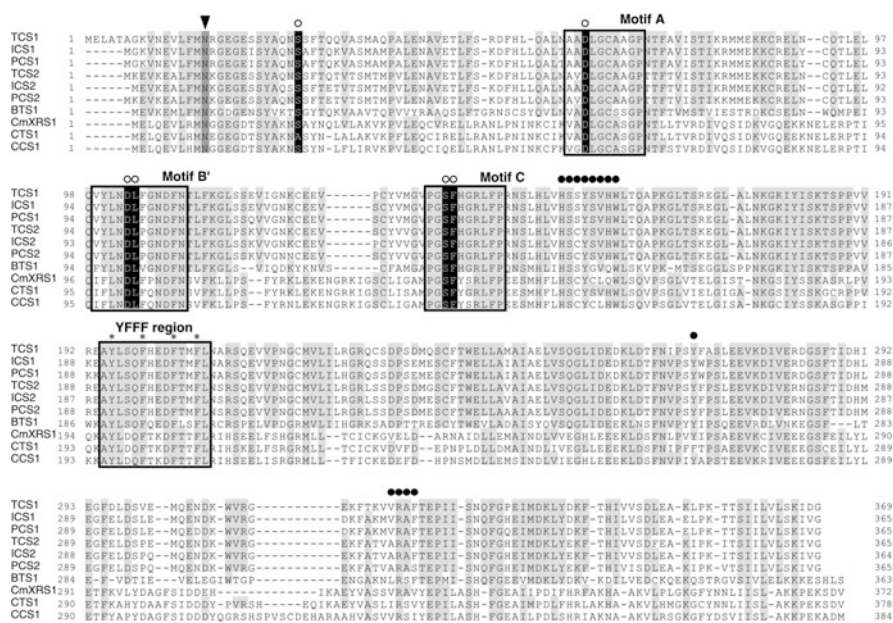
Expression of genes involved in caffeine biosynthesis has been demonstrated in young leaves, flower buds and developing endosperm of *Coffea arabica* (Mizuno et al. 2003a, b). The expression of *CmXRS1*, *CTS2* and *CCSI*, which encode 7-methylxanthosine synthase, theobromine synthase and caffeine synthase, respectively, was examined. Transcripts of *CmXRS1* and *CCSI* were observed in all organs, but the

highest level was found in developing endosperm. Significant expression of *CTS2* was found only in flower buds. The patterns of expression of *CmXRS1* and *CCS1* were synchronized. During development of *Coffea arabica* fruits, the transcripts of *CmXRS1* and *CCS1* are present in every stage of growth except in fully ripened tissues. The pattern of expression of these genes during growth is roughly related to the in situ synthesis of caffeine from adenine nucleotides, although exceptions were found in the very early and the later stages of fruit growth. Since the level of *CTS2* transcripts encoding the theobromine synthase is very low in fruits, the alternative *CCS1* gene encoding the dual-functional caffeine synthase may be operative for the last two steps of caffeine biosynthesis. In developing *Coffea arabica* fruits, the levels of transcripts of *CmXRS1* and *CCS1* are higher in seeds than in pericarp. Native caffeine synthase (3*N*-methyltransferase) activity is distributed in both organs in a similar manner. Therefore, caffeine accumulating in ripened coffee seeds appears to be synthesized within the developing seeds and is not transported from pericarp (Koshiro et al. 2006).

In *Camellia sinensis*, expression of *TCSI* encoding caffeine synthase is higher in young leaves than in mature leaves, stems or roots (Li et al. 2008). This is consistent with the fact that biosynthetic activity of caffeine occurs mainly in young leaves (Ashihara and Kubota 1986). Recent studies using *Camellia sinensis* tissue culture indicate that the expression of *TCSI*, and possibly the unidentified gene encoding 7-methylxanthosine synthase, represents the principal control mechanism for caffeine biosynthesis. Although increased caffeine content was observed when cultures were grown in media containing paraxanthine, addition of adenosine, guanosine or hypoxanthine did not have a similar impact. Thus, neither the supply of non-methylated purine precursors nor the availability of SAM appears to be an important factor in the regulation of caffeine biosynthesis (Li et al. 2008; Deng et al. 2008).

## 4.2 Evolutionary Relationship of Caffeine Synthase and Related Enzymes

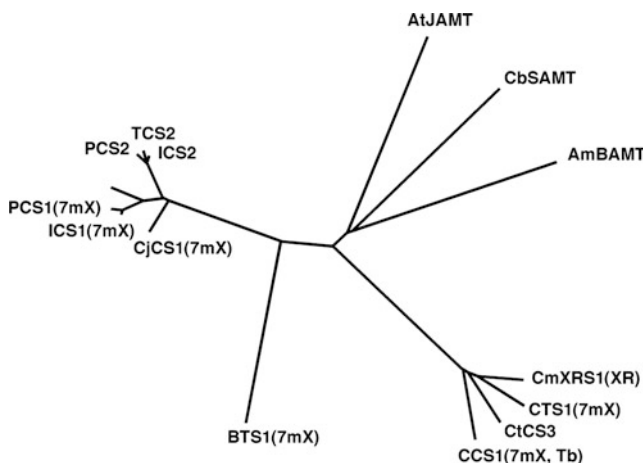
Figure 5 shows the amino acid sequences of caffeine synthase and related enzymes. There are four highly conserved regions: motif A, motif B', motif C and the YFFF region in the amino acid sequence of the caffeine synthase family (Kato and Mizuno 2004). Three conserved motifs, A, B and C, of the binding site of the methyl donor of SAM have been reported in the majority of plant SAM-dependent *O*-methyltransferases (Joshi and Chiang 1998). The motif B' and YFFF region contains many hydrophobic amino acids which are specific to the motif B' methyltransferase family. Most members of this newly characterized motif B' methyltransferase family catalyse the formation of small and volatile methyl esters by using SAM as a methyl donor and substrates with a carboxyl group as the methyl acceptor. Members of this family include salicylic acid carboxyl methyltransferase (SAMT) (Ross et al. 1999),



**Fig. 5** Comparison of the amino acid sequences of caffeine synthases and their related enzymes. Alignment of the amino acid sequences for TCS1 and TCS2 from tea, ICS1 and ICS2 from *Camellia irrawadiensis*, PCS1 and PCS2 from *Camellia ptilophylla*, BTS1 from cocoa, and CmXRS1, CTS1 and CCS1 from coffee is indicated. Shaded boxes represent conserved amino acid residues, and dashes represent gaps that have been inserted for optimal alignment. The proposed SAM-binding motifs (A, B' and C) and the conserved “YFFF region” are shown by open boxes (Mizuno et al. 2003a). Asterisks indicate tyrosine (Y) or phenylalanine (F) residues in the region. The nominated amino acids in substrate binding are indicated by closed circles, and additional active sites are indicated by arrowheads (Zubieta et al. 2003). The sources of the sequences are as follows: TCS1, AB031280 (Kato et al. 2000); TCS2, AB031281; BTS1, AB096699; PCS1, AB207817; PCS2, AB207818; ICS1, AB056108; ICS2, AB207816 (Yoneyama et al. 2006); CmXRS1, AB034699 (Mizuno et al. 2003b); CTS1, AB034700 (Mizuno et al. 2001); CCS1, AB086414 (Mizuno et al. 2003a). (Adapted from Yoneyama et al. 2006)

benzoic acid carboxyl methyltransferase (BAMT) (Dudareva et al. 2000), jasmonic acid carboxyl methyltransferase (JAMT) (Seo et al. 2001), farnesic acid carboxyl methyltransferase (FAMT) (Yang et al. 2006), indole-3-acetic acid methyltransferase (IAMT) (Zhao et al. 2008), gibberellic acid methyltransferase (GAMT) (Varbanova et al. 2007) and loganic acid carboxyl methyltransferase (LAMT) (Murata et al. 2008). The motif B' methyltransferase family is also referred to as the SABATH family, based on the initial letters of the names of the substrates (D'Auria et al. 2003). Crystallographic data on SAMT from *Clarkia breweri* suggest that members of this family exist as dimers in solution (Zubieta et al. 2003). Further structural analysis of 7-methylxanthosine synthase and caffeine synthase from *Coffea canephora* also revealed a dimeric structure (McCarthy and McCarthy 2007).

Amino acid sequences of the caffeine synthase family derived from coffee are more than 80% homologous but share only 40% homology with caffeine synthase



**Fig. 6** Evolutionary relationship of caffeine synthase and its related enzymes. Substrates of the enzymes involved in caffeine synthesis are shown in *parentheses*. The substrates of TCS2, ICS2, PCS2 and CtCS3 are not known. The sources of the sequences are as follows: CmXRS1, AB034699 (Mizuno et al. 2003b); CTS1, AB034700 (Mizuno et al. 2001); CtCS3, AB054842 (Mizuno et al. 2003a); CCS1 AB086414 (Mizuno et al. 2003a); CbSAMT, AF133053 (Ross et al. 1999); AtJAMT, AY008434 (Seo et al. 2001); BTS1, AB096699 (Yoneyama et al. 2006); and AmBAMT, AF198492 (Dudareva et al. 2000). The unrooted tree was created by using ClustalW through application of the neighbour-joining method (Thompson et al. 1994). (Adapted from Ishida et al. 2009)

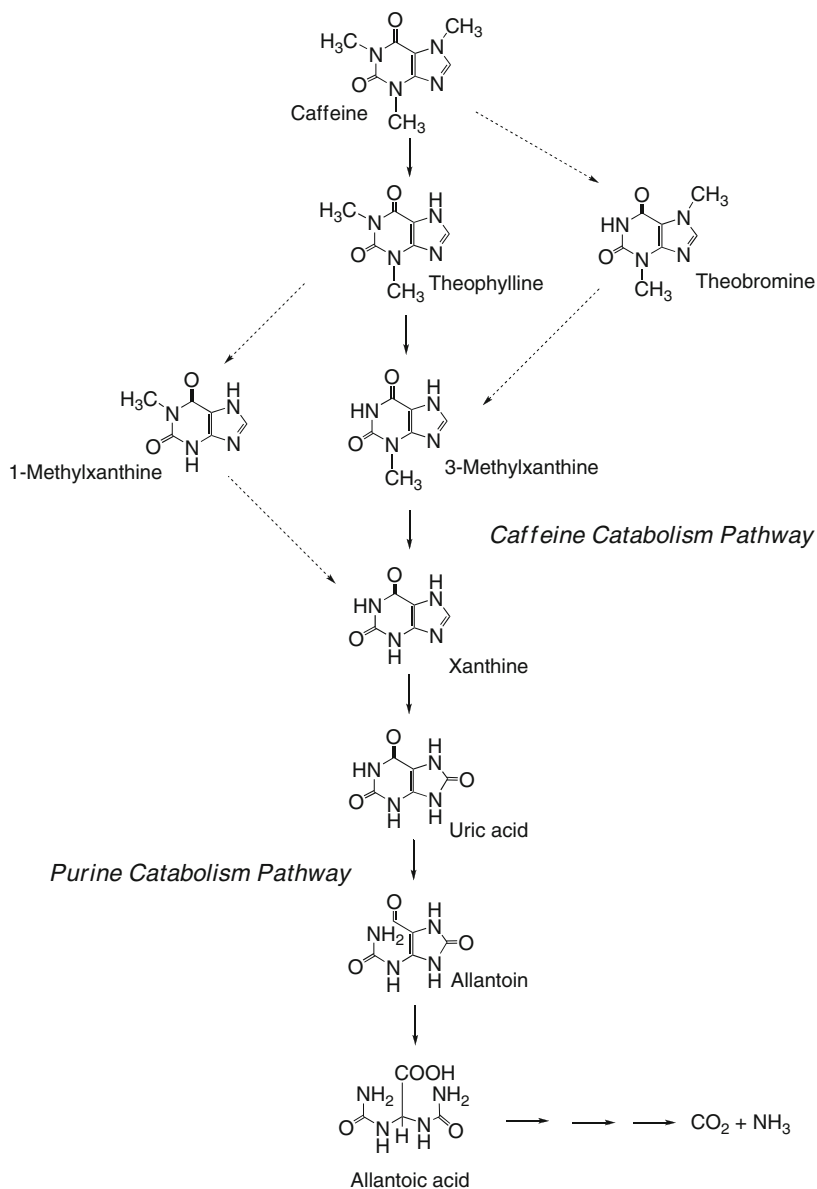
from tea. There is a similar homology between SAMT from *Clarkia breweri* and caffeine synthase from tea and coffee plants. That is to say, the amino acid sequences share a high degree of sequence identity within the same genus.

Figure 6 shows the phylogenetic tree analysis of the motif B' methyltransferase family. This implies that the caffeine biosynthetic pathways in coffee, tea and cacao might have evolved in parallel with one another, consistent with different catalytic properties of the enzymes involved. Recently, Ishida et al. (2009) reported the occurrence of theobromine synthase genes in purine alkaloid-free species of *Camellia*. This represents additional evidence that monophyletic genes occur in *Camellia* plants.

## 5 Catabolism of Methylxanthines in Plants

### 5.1 Conversion of Caffeine to Theophylline

Limited amounts of caffeine are very slowly degraded with the removal of the three methyl groups, resulting in the formation of xanthine in almost all caffeine-forming plant species (Fig. 7). Catabolism of caffeine has been studied using  $^{14}\text{C}$ -labelled



**Fig. 7** Catabolic pathways of caffeine and theobromine. Caffeine is catabolized mainly to xanthine via theophylline and 3-methylxanthine. Theobromine is catabolized to xanthine via 3-methylxanthine. Xanthine is further degraded to  $\text{CO}_2$  and  $\text{NH}_3$  by the conventional oxidative purine catabolic pathway. *Dotted arrows* indicate minor routes



caffeine (Ashihara et al. 1997; Ashihara et al. 1996; Mazzafera 2004; Suzuki and Waller 1984). Caffeine catabolism usually begins with its conversion to theophylline catalysed by *N7*-demethylase. This conversion is the rate-limiting step in purine alkaloid catabolism and provides a ready explanation for the high concentration of endogenous caffeine in species such as *Coffea arabica* and *Camellia sinensis*. The involvement of the P450-dependent monooxygenase activity for this reaction has been proposed (Huber and Baumann 1998; Mazzafera 2004), although the activity of this enzyme has not yet been demonstrated. In leaves of *Coffea eugenoides*, which contain low levels of caffeine, [8-<sup>14</sup>C]caffeine is catabolized rapidly primarily by the main caffeine catabolic pathway via theophylline. This suggests that the low caffeine accumulation in *Coffea eugenoides* is a consequence of rapid degradation of caffeine, perhaps accompanied by a slow rate of caffeine biosynthesis (Ashihara and Crozier 1999b).

## 5.2 Metabolism of Theophylline

In caffeine-producing plants such as tea, coffee and maté, [8-<sup>14</sup>C]theophylline is catabolized rapidly (Ito et al. 1997). The main route of theophylline degradation in higher plants involves a theophylline → 3-methylxanthine → xanthine → uric acid → allantoin → allantoic acid → CO<sub>2</sub> + NH<sub>3</sub> pathway (Fig. 7). In contrast, theophylline is catabolized at extremely low levels in non-methylxanthine-forming plants. Higher plants do not convert [8-<sup>14</sup>C]theophylline to either 1-methyluric acid or 1,3-dimethyluric acid, which are the main catabolites of theophylline in mammals (Scheline 1991). In tea and maté, large amounts of [8-<sup>14</sup>C]theophylline are also converted to theobromine and caffeine via a theophylline → 3-methylxanthine → theobromine → caffeine salvage pathway (Ito et al. 1997).

## 5.3 Catabolism of Theobromine

In contrast to theophylline, theobromine is a precursor, as opposed to a catabolite, of caffeine. However, degradation of theobromine has been observed in mature leaves (Koyama et al. 2003) and pericarp of the theobromine-accumulating plant *Theobroma cacao* (Zheng et al. 2004). Theobromine was degraded to CO<sub>2</sub> via 3-methylxanthine, xanthine and allantoic acid (Fig. 7). Although conversion of caffeine to theobromine was detected in *Theobroma cacao*, caffeine was catabolized principally to CO<sub>2</sub> via theophylline, which is the same degradation pathway that operates in *Coffea arabica* and *Camellia sinensis*.

## 6 Ecological Roles of Purine Alkaloids

The physiological role of purine alkaloids *in planta* is largely unknown. It appears not to act as a nitrogen reserve since considerable amounts remain in leaves after abscission. There are two hypotheses concerning the ecological roles of caffeine in plants.

### 6.1 Chemical Defence Theory

The chemical defence theory proposes that the high concentrations of caffeine in young leaves, fruits and flower buds of species such as tea and coffee act as a defence to protect young soft tissues from pathogens and herbivores. It has been shown that spraying tomato leaves with caffeine deters feeding by tobacco hornworms, while treatment of cabbage leaves and orchids with caffeine acts as a neurotoxin and kills or repels slugs and snails (Hollingsworth et al. 2003). This work has now been extended and convincing evidence for the chemical defence theory has recently been obtained with transgenic caffeine-producing tobacco plants (Kim et al. 2006; Uefuji et al. 2005).

### 6.2 Allelopathy Theory

The allelopathic or autotoxic function theory proposes that caffeine in seed coats and falling leaves is released into the soil to inhibit germination of seeds around the parent plants (Anaya et al. 2006). In caffeine-synthesizing cells, caffeine accumulates in vacuoles, so caffeine does not impact on cellular metabolism. Exogenously applied caffeine does, however, inhibit various aspects of metabolism in the cells. Although there is experimental evidence from laboratory studies to support this proposal, it is unclear to what extent caffeine is involved in allelopathy in natural ecosystems, especially as soil bacteria such as *Pseudomonas putida* can degrade methylxanthines (Hohnloser et al. 1980; Gluck and Lingens 1988).

## 7 Production of Decaffeinated Coffee

Demand for decaffeinated coffee has increased gradually since the early 1970s. Worldwide sales of “decaf” have achieved a 12% share of the market, estimated to be worth more than US \$4 billion (Heilmann 2001). Modern methods of decaffeination, such as supercritical fluid extraction with carbon dioxide, may have minimal effect on the organoleptic quality of the beverage if carried out

correctly (Vitzthum 2005). Nevertheless, coffee cultivars combining high cup quality with a low caffeine content may provide a superior, less expensive and ecofriendly alternative to meet the demand for decaffeinated coffee.

### 7.1 *Production by Breeding*

Silvarolla et al. (2004) discovered naturally decaffeinated mutant plants in the progeny of *Coffea arabica* accessions from Ethiopia. Three of these Ethiopian mutant plants were almost completely free of caffeine. The seeds of those plants had low caffeine content (mean caffeine content 0.076% dry weight), but significant amounts of theobromine (about 0.61%), another methylxanthine which is capable of causing physiological effects similar to those of caffeine (Eteng et al. 1997). It would, therefore, appear to be worth searching for mutant plants with a low theobromine and caffeine content.

Recently, Nagai et al. (2008) produced a new low-caffeine hybrid coffee which is a tetraploid interspecific hybrid developed in Madagascar from *Coffea eugenioides*, *Coffea canephora* and *Coffea arabica*. Green beans of selected hybrids contain 0.37% caffeine and no detectable theobromine. Low caffeine accumulation is due mainly to the low biosynthetic activity of purine alkaloids, possibly the extremely weak *N*-methyltransferase reactions in caffeine biosynthesis.

### 7.2 *Production by Genetic Engineering*

Attempts to use genetic engineering to produce transgenic caffeine-deficient coffee have to date had only limited success. Low-caffeine-containing transgenic *Coffea canephora* plants have been produced but the caffeine content of the leaves was variable, depending on the line; the most notable example yielded a reduction of up to 70% (Ogita et al. 2003; Ogita et al. 2004). Coffee produced from beans of *Coffea arabica* has a flavour superior to that of robusta coffee but as yet caffeine-deficient transgenic arabica beans have not been produced. When this is achieved, because of the substantial market for decaffeinated coffee, it is likely to have major commercial implications.

## 8 Summary and Perspectives

The major route to caffeine in higher plants is a xanthosine  $\rightarrow$  7-methylxanthosine  $\rightarrow$  7-methylxanthine  $\rightarrow$  theobromine  $\rightarrow$  caffeine pathway. The precursors of caffeine are derived from purine nucleotides. The rate of caffeine biosynthesis appears to be regulated primarily by the induction and repression of *N*-methyltransferases,

especially 7-methylxanthosine synthase. The first paper on the cloning of caffeine synthase from tea appeared in 2000. Since then there has been a veritable explosion of research that has led to the successful cloning of a number of *N*-methyltransferase-encoding genes from coffee. The rate-limiting step in the caffeine biosynthetic pathway, the initial conversion of xanthosine to 7-methylxanthosine, is catalysed by 7-methylxanthosine synthase, and the encoding gene for this *N*-methyltransferase has been isolated from coffee. Although funding from industry has been very limited to non-existent, much of the extensive interest in this research has been fuelled by the possibilities of using genetic engineering to obtain transgenic, low-caffeine-containing coffee and tea that could be used to produce “natural” decaffeinated beverages. Although transgenic *Coffea canephora* seedlings with a 70% reduced caffeine content have been obtained, there is as yet no information on the caffeine content of beans produced by these plants. The real breakthrough in commercial terms will come with the production of transgenic caffeine-deficient *Coffea arabica* beans.

## References

- Anaya AL, Cruz-Ortega R, Waller GR (2006) Metabolism and ecology of purine alkaloids. *Front Biosci* 11:2354–2370
- Ashihara H, Crozier A (1999a) Biosynthesis and metabolism of caffeine and related purine alkaloids in plants. *Adv Bot Res* 30:117–205
- Ashihara H, Crozier A (1999b) Biosynthesis and catabolism of caffeine in low-caffeine-containing species of *Coffea*. *J Agric Food Chem* 47:3425–3431
- Ashihara H, Kubota H (1986) Patterns of adenine metabolism and caffeine biosynthesis in different parts of tea seedlings. *Physiol Plant* 68:275–281
- Ashihara H, Suzuki T (2004) Distribution and biosynthesis of caffeine in plants. *Front Biosci* 9:1864–1876
- Ashihara H, Gillies FM, Crozier A (1997) Metabolism of caffeine and related purine alkaloids in leaves of tea (*Camellia sinensis* L.). *Plant Cell Physiol* 38:413–419
- Ashihara H, Kato M, Ye C-X (1998) Biosynthesis and metabolism of purine alkaloids in leaves of cocoa tea (*Camellia pitlophylla*). *J Plant Res* 111:599–604
- Ashihara H, Monteiro AM, Moritz T, Gillies FM, Crozier A (1996) Catabolism of caffeine and related purine alkaloids in leaves of *Coffea arabica* L. *Planta* 198:334–339
- Ashihara H, Sano H, Crozier A (2008) Caffeine and related purine alkaloids: biosynthesis, catabolism, function and genetic engineering. *Phytochemistry* 69:841–856
- Baumann TW, Oechsli M, Wanner H (1976) Caffeine and methylated uric acids: chemical patterns during vegetative development of *Coffea liberica*. *Biochem Physiol Pflanz* 170:217–225
- Baumann TW, Wanner H (1980) The 1,3,7,9-tetramethyluric acid content of cupu (*Theobroma grandiflorum*). *Acta Amaz* 10:425
- Baumann TW, Schulthess BH, Hanni K (1995) Guarana (*Paulinia cupana*) rewards seed dispersers without intoxicating them by caffeine. *Phytochemistry* 39:1063–1070
- Campa C, Doubeau S, Dussert S, Hamon S, Noirot M (2005) Diversity in bean caffeine content among wild *Coffea* species: evidence of a discontinuous distribution. *Food Chem* 91:633–637
- Charrier A, Berthaud J (1975) Variation de la teneur en cafeine dans le genre *Coffea*. *Cafe Cacao* The 19:251–264

- D'Auria JC, Chen F, Pichersky E (2003) The SABATH family of MTs in *Arabidopsis thaliana* and other plant species. In: Romeo JT (ed) Recent advances in phytochemistry, vol 37. Elsevier, Oxford, pp 253–283
- Deng WW, Li Y, Ogita S, Ashihara H (2008) Fine control of caffeine biosynthesis in tissue cultures of *Camellia sinensis*. *Phytochem Lett* 1:195–198
- Dudareva N, Murfitt LM, Mann CJ, Gorenstein N, Kolosova N, Kish CM, Bonham C, Wood K (2000) Developmental regulation of methyl benzoate biosynthesis and emission in snapdragon flowers. *Plant Cell* 12:949–961
- Duthie GG, Crozier A (2003) Beverages. In: Goldberg G (ed) Plants: diet and health. British Nutrition Foundation/ Chapman and Hall, London, pp 147–182
- Eteng MU, Eyong EU, Akpanyung EO, Agiang MA, Aremu CY (1997) Recent advances in caffeine and theobromine toxicities: a review. *Plant Food Hum Nutr* 51:231–243
- Gluck M, Lingens F (1988) Heteroxanthine demethylase, a new enzyme in the degradation of caffeine by *Pseudomonas putida*. *Appl Microbiol Biotechnol* 28:59–62
- Hammerstone JF, Romanczyk LJ, Aitken WM (1994) Purine alkaloid distribution within *Herrania* and *Theobroma*. *Phytochemistry* 35:1237–1240
- Heilmann W (2001) Technology II: decaffeination of coffee. In: Clarke RJ, Vitzthum OG (eds) Coffee: recent developments. Blackwell, Oxford, pp 108–124
- Hohnloser W, Osswald B, Lingens F (1980) Enzymological aspects of caffeine demethylation and formaldehyde oxidation by *Pseudomonas putida* C1. *Hoppe Seylers Z Physiol Chem* 361:1763–1766
- Hollingsworth RG, Armstrong JW, Campbell E (2003) Pest control: caffeine as a repellent for slugs and snails. *Nature* 417:915–916
- Huber M, Baumann TW (1998) The first step of caffeine degradation in coffee – still a mystery. In: Symposium future trends in phytochemistry. The Phytochemical Society of Europe, Rolduc
- Ishida M, Kitao N, Mizuno K, Tanikawa N, Kato M (2009) Occurrence of theobromine synthase genes in purine alkaloid-free species of *Camellia* plants. *Planta* 229:559–568
- Ito E, Ashihara H (1999) Contribution of purine nucleotide biosynthesis de novo to the formation of caffeine in young tea (*Camellia sinensis*) leaves. *J Plant Physiol* 254:145–151
- Ito E, Crozier A, Ashihara H (1997) Theophylline metabolism in higher plants. *Biochim Biophys Acta* 1336:323–330
- Joshi CP, Chiang VL (1998) Conserved sequence motifs in plant *S*-adenosyl-L-methionine-dependent methyltransferases. *Plant Mol Biol* 37:663–674
- Kato M, Mizuno K, Crozier A, Fujimura T, Ashihara H (2000) A gene encoding caffeine synthase from tea leaves. *Nature* 406:956–957
- Kato M, Mizuno K, Fujimura T, Iwama M, Irie M, Crozier A, Ashihara H (1999) Purification and characterization of caffeine synthase from tea leaves. *Plant Physiol* 120:579–586
- Kato M, Mizuno K (2004) Caffeine synthase and related methyltransferases in plants. *Front Biosci* 9:1833–1842
- Keya CA, Crozier A, Ashihara H (2003) Inhibition of caffeine biosynthesis in tea (*Camellia sinensis*) and coffee (*Coffea arabica*) plants by ribavirin. *FEBS Lett* 554:473–477
- Kim YS, Uefuji H, Ogita S, Sano H (2006) Transgenic tobacco plants producing caffeine: a potential new strategy for insect pest control. *Transgenic Res* 15:667–672
- Koshiishi C, Kato A, Yama S, Crozier A, Ashihara H (2001) A new caffeine biosynthetic pathway in tea leaves: utilisation of adenosine released from the *S*-adenosyl-L-methionine cycle. *FEBS Lett* 499:50–54
- Koshiro Y, Zheng XQ, Wang M, Nagai C, Ashihara H (2006) Changes in content and biosynthetic activity of caffeine and trigonelline during growth and ripening of *Coffea arabica* and *Coffea canephora* fruits. *Plant Sci* 171:242–250
- Koyama Y, Tomoda Y, Kato M, Ashihara H (2003) Metabolism of purine bases, nucleosides and alkaloids in theobromine-forming *Theobroma cacao* leaves. *Plant Physiol Biochem* 41:977–984
- Kretschmar JA, Baumann TW (1999) Caffeine in *Citrus* flowers. *Phytochemistry* 52:19–23

- Li Y, Ogita S, Keya CA, Ashihara H (2008) Expression of caffeine biosynthesis gene in tea (*Camellia sinensis*). *Z Naturforsch* 63c:267–270
- Mazzafera P (2004) Catabolism of caffeine in plants and microorganisms. *Front Biosci* 9:1348–1359
- Mazzafera P, Carvalho A (1992) Breeding for low seed caffeine content of coffee (*Coffea* L.) by interspecific hybridization. *Euphytica* 59:55–60
- McCarthy AA, McCarthy JG (2007) The structure of two *N*-methyltransferases from the caffeine biosynthetic pathway. *Plant Physiol* 144:879–889
- Mizuno K, Tanaka H, Kato M, Ashihara H, Fujimura T (2001) cDNA cloning of caffeine (theobromine) synthase from coffee (*Coffea arabica* L.). In: International scientific colloquium on coffee 19. ASIC, Paris, pp 815–818
- Mizuno K, Kato M, Irino F, Yoneyama N, Fujimura T, Ashihara H (2003a) The first committed step reaction of caffeine biosynthesis: 7-methylxanthosine synthase is closely homologous to caffeine synthases in coffee (*Coffea arabica* L.). *FEBS Lett* 547:56–60
- Mizuno K, Okuda A, Kato M, Yoneyama N, Tanaka H, Ashihara H, Fujimura T (2003b) Isolation of a new dual-functional caffeine synthase gene encoding an enzyme for the conversion of 7-methylxanthine to caffeine from coffee (*Coffea arabica* L.). *FEBS Lett* 534:75–81
- Moffatt BA, Ashihara H (2002) Purine and pyrimidine nucleotide synthesis and metabolism. The *Arabidopsis* book. American Society of Plant Biologists, Rockville. doi:10.1199/tab.0018, <http://www.aspb.org/publications/arabidopsis/>
- Murata J, Roepke J, Gordon H, Luca VD (2008) The leaf epidermome of *Catharanthus roseus* reveals its biochemical specialization. *Plant Cell* 20:524–542
- Nagai C, Rakotomalala JJ, Katahira R, Li Y, Yamagata K, Ashihara H (2008) Production of a new low-caffeine hybrid coffee and the biochemical mechanism of low caffeine accumulation. *Euphytica* 164:133–142
- Nagata T, Sakai S (1985) Purine base pattern of *Camellia irrawadiensis*. *Phytochemistry* 24:2271–2272
- Negishi O, Ozawa T, Imagawa H (1988) *N*-Methylnucleosidase from tea leaves. *Agric Biol Chem* 52:169–175
- Negishi O, Ozawa T, Imagawa H (1994) Guanosine deaminase and guanine deaminase from tea leaves. *Biosci Biotechnol Biochem* 58:1277–1281
- Ogawa M, Herai Y, Koizumi N, Kusano T, Sano H (2001) 7-Methylxanthine methyltransferase of coffee plants. Gene isolation and enzymatic properties. *J Biol Chem* 276:8213–8218
- Ogita S, Uefuji H, Morimoto M, Sano H (2004) Application of RNAi to confirm theobromine as the major intermediate for caffeine biosynthesis in coffee plants with potential for construction of decaffeinated varieties. *Plant Mol Biol* 54:931–941
- Ogita S, Uefuji H, Yamaguchi Y, Nozomu K, Sano H (2003) Producing decaffeinated coffee plants. *Nature* 423:823
- Petermann J, Baumann TW (1983) Metabolic relations between methylxanthines and methyluric acids in *Coffea*. *Plant Physiol* 73:961–964
- Rakotomalala JJ, Cros E, Clifford MN, Charrier A (1992) Caffeine and theobromine in green beans from *Mascarocoffea*. *Phytochemistry* 31:1271–1272
- Ross JR, Nam KH, D'Auria JC, Pichersky E (1999) *S*-Adenosyl-L-methionine: salicylic acid carboxyl methyltransferase, an enzyme involved in floral scent production and plant defense, represents a new class of plant methyltransferases. *Arch Biochem Biophys* 367:9–16
- Scheline RR (1991) Handbook of mammalian metabolism of plant compounds. CRC, Boca Raton
- Senanayake UM, Wijesekera ROB (1971) Theobromine and caffeine content of the cocoa bean during its growth. *J Sci Food Agric* 22:262–263
- Silvarolla MB, Mazzafera P, Fazuoli LC (2004) A naturally decaffeinated arabica coffee. *Nature* 429:826
- Seo HS, Song JT, Lee YH, Iwang I, Lee JS, Choi YD (2001) Jasmonic acid carboxyl methyltransferase: a key enzyme for jasmonate-regulated plant responses. *Proc Natl Acad Sci USA* 98:4788–4793

- Stasolla C, Katahira R, Thorpe TA, Ashihara H (2003) Purine and pyrimidine nucleotide metabolism in higher plants. *J Plant Physiol* 160:1271–1295
- Suzuki T, Waller GR (1984) Biodegradation of caffeine: formation of theophylline and caffeine in mature *Coffea arabica* fruits. *J Sci Food Agric* 35:66–70
- Thompson JD, Higgins DG, Gibson TJ (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucl Acids Res* 22:4673–4680
- Uefuji H, Tatsumi Y, Morimoto M, Kaothien-Nakayama P, Ogita S, Sano H (2005) Caffeine production in tobacco plants by simultaneous expression of three coffee *N*-methyltransferases and its potential as a pest repellent. *Plant Mol Biol* 59:221–227
- Uefuji H, Ogita S, Yamaguchi Y, Koizumi N, Sano H (2003) Molecular cloning and functional characterization of three distinct *N*-methyltransferases involved in the caffeine biosynthetic pathway in coffee plants. *Plant Physiol* 132:372–380
- Varbanova M, Yamaguchi S, Yang Y, McKelvey K, Hanada A, Borochoff R, Yu F, Jikumaru Y, Ross J, Cortes D, Ma CJ, Noel JP, Mander L, Shulaev V, Kamiya Y, Rodermeier S, Weiss D, Pichersky E (2007) Methylation of gibberellins by *Arabidopsis* GAMT1 and GAMT2. *Plant Cell* 19:32–45
- Vitzthum OG (2005) Decaffeination. In: Illy A, Viani R (eds) Espresso coffee: the science of quality, 2nd edn. Elsevier, Amsterdam, pp 142–147
- Weckerle CS, Stutz MA, Baumann TW (2003) Purine alkaloids in *Paullinia*. *Phytochemistry* 64:735–742
- Yang Y, Yuan JS, Ross J, Noel JP, Pichersky E, Chen F (2006) An *Arabidopsis thaliana* methyltransferase capable of methylating farnesoic acid. *Arch Biochem Biophys* 448:123–132
- Ye C, Un Y, Zhou H, Cheng F, Li X (1997) Isolation and analysis of purine alkaloids from *Camellia pilophylla* Chang. *Acta Sci Nat Univ Sunyatseni* 36:30–33
- Yoneyama N, Morimoto H, Ye CX, Ashihara H, Mizuno K, Kato M (2006) Substrate specificity of *N*-methyltransferase involved in purine alkaloids synthesis is dependent upon one amino acid residue of the enzyme. *Mol Genet Genomics* 275:125–135
- Zhao N, Ferrer J-L, Ross J, Guan J, Yang Y, Pichersky E, Noel JP, Chen F (2008) Structural, biochemical, and phylogenetic analyses suggest that indole-3-acetic acid methyltransferase is an evolutionarily ancient member of the SABATH family. *Plant Physiol* 146:455–467
- Zheng XQ, Ashihara H (2004) Distribution, biosynthesis and function of purine and pyridine alkaloids in *Coffea arabica* seedlings. *Plant Sci* 166:807–813
- Zheng XQ, Ye CX, Kato M, Crozier A, Ashihara H (2002) Theacrine (1,3,7,9-tetramethyluric acid) synthesis in leaves of a Chinese tea, kucha (*Camellia assamica* var. kucha). *Phytochemistry* 60:129–134
- Zheng XQ, Koyama Y, Nagai C, Ashihara H (2004) Biosynthesis, accumulation and degradation of theobromine in developing *Theobroma cacao* fruits. *J Plant Physiol* 161:363–369
- Zubieta C, Ross JR, Koscheski P, Yang Y, Pichersky E, Noel JP (2003) Structural basis for substrate recognition in the salicylic acid carboxyl methyltransferase family. *Plant Cell* 15:1704–1716

# Pharmacokinetics and Metabolism of Natural Methylxanthines in Animal and Man

Maurice J. Arnaud

## Contents

1	Introduction .....	35
2	Caffeine .....	35
2.1	Absorption .....	35
2.2	Distribution .....	36
2.3	Excretion .....	38
2.4	Pharmacokinetics .....	39
2.5	Metabolism .....	41
2.6	Sources of Variation in Caffeine Pharmacokinetics and Metabolism .....	45
2.7	Metabolites and Metabolic Pathway .....	51
3	Theophylline .....	55
3.1	Absorption .....	55
3.2	Distribution .....	58
3.3	Excretion .....	59
3.4	Pharmacokinetics .....	59
3.5	Metabolism .....	61
3.6	Sources of Variation in Theophylline Pharmacokinetics and Metabolism .....	62
3.7	Metabolites and Metabolic Pathway .....	65
4	Theobromine .....	66
4.1	Absorption .....	66
4.2	Distribution .....	67
4.3	Excretion .....	68
4.4	Pharmacokinetics .....	69
4.5	Metabolism .....	70
4.6	Sources of Variation in Theobromine Pharmacokinetics and Metabolism .....	70
4.7	Metabolites and Metabolic Pathway .....	71
5	Paraxanthine .....	72
5.1	Absorption and Distribution .....	73
5.2	Excretion .....	74
5.3	Pharmacokinetics .....	74

---

M.J. Arnaud

Nutrition and Biochemistry, Bourg-Dessous 2A, 1814, La Tour-de-Peilz, Switzerland

e-mail: mauricearnaud@hotmail.com



5.4 Metabolism .....	75
5.5 Metabolites and Metabolic Pathways .....	76
References .....	77

**Abstract** Caffeine, theophylline, theobromine, and paraxanthine administered to animals and humans distribute in all body fluids and cross all biological membranes. They do not accumulate in organs or tissues and are extensively metabolized by the liver, with less than 2% of caffeine administered excreted unchanged in human urine. Dose-independent and dose-dependent pharmacokinetics of caffeine and other dimethylxanthines may be observed and explained by saturation of metabolic pathways and impaired elimination due to the immaturity of hepatic enzyme and liver diseases. While gender and menstrual cycle have little effect on their elimination, decreased clearance is seen in women using oral contraceptives and during pregnancy. Obesity, physical exercise, diseases, and particularly smoking and the interactions of drugs affect their elimination owing to either stimulation or inhibition of CYP1A2. Their metabolic pathways exhibit important quantitative and qualitative differences in animal species and man. Chronic ingestion or restriction of caffeine intake in man has a small effect on their disposition, but dietary constituents, including broccoli and herbal tea, as well as alcohol were shown to modify their plasma pharmacokinetics. Using molar ratios of metabolites in plasma and/or urine, phenotyping of various enzyme activities, such as cytochrome monooxygenases, N-acetylation, 8-hydroxylation, and xanthine oxidase, has become a valuable tool to identify polymorphisms and to understand individual variations and potential associations with health risks in epidemiological surveys.

**Keywords** Absorption · Age · Alcohol · Bioavailability · Caffeine · Cytochromes · Diet · Diseases · Distribution · Drugs · Excretion · Gender · Hormones · Interactions · Metabolism · Obesity · Paraxanthine · Pharmacokinetics · Physical exercise · Smoking · Theobromine · Theophylline

## Abbreviations

1,3,7DAU	6-Amino-5-( <i>N</i> -formylmethylamino)-1,3-dimethyluracil
1,3,7TMU	1,3,7-Trimethyluric acid
1,3DMU	1,3-Dimethyluric acid
1,7DAU	6-Amino-5-( <i>N</i> -formylmethylamino)-3-methyluracil
1,7DMU	1,7-Dimethyluric acid
1MU	1-Methyluric acid
1MX	1-Methylxanthine
3,7DAU	6-Amino-5-( <i>N</i> -formylmethylamino)-1-methyluracil

3,7DMU	3,7-Dimethyluric acid
3MU	3-Methyluric acid
3MX	3-Methylxanthine
7MU	7-Methyluric acid
7MX	7-Methylxanthine
AAMU	5-Acetylamino-6-amino-3-methyluracil
AFMU	5-Acetylamino-6-formylamino-3-methyluracil
AUC	Area under the concentration versus time curve
$C_{\max}$	Peak plasma concentration
CYP	Cytochrome P450
GSH	Glutathione
IBW	Ideal body weight
$K_a$	Absorption rate constant
$K_e$	Elimination rate constant
$K_m$	Michaelis–Menten constant
NAT2	<i>N</i> -Acetyltransferase 2
TBW	Total body weight
$T_{\max}$	Time to reach the peak plasma concentration

## 1 Introduction

Caffeine metabolism and pharmacokinetics have been reported in several reviews and monographs (Arnaud and Welsch 1982; Arnaud 1984, 1987, 1988, 1993a, b, 1998; Anonymous 1991). Reviews on theophylline (Arnaud and Welsch 1982; Hendeles et al. 1986; Anonymous 1991) were published because of its wide application as an active component of a variety of over-the-counter pharmaceutical products and drugs. A few reviews have been published on theobromine (Tarka 1982; Anonymous 1991), but there is no review on paraxanthine, perhaps because it can be found in plants only in trace amounts and is identified in human urine (Salomon 1883). This chapter highlights some of the present knowledge, including the most recently published studies on these four methylxanthines.

## 2 Caffeine

### 2.1 Absorption

Absorption and bioavailability of caffeine were generally similar between humans, dogs, rabbits, rats, and mice (Walton et al. 2001). In animals and man, absorption is characterized by rapid and complete gastrointestinal absorption (Arnaud 1976;

Arnaud and Welsch 1980a; Yesair et al. 1984). In man, 99% of the administered dose was absorbed in 45 min (Blanchard and Sawers 1983a), mainly from the small intestine but also 20% from the stomach (Chvasta and Cooke 1971). The absolute bioavailability of caffeine (5 mg/kg) in healthy adult male volunteers showed a rapid oral absorption with the time to reach peak the peak plasma concentration ( $T_{\max}$ ) of  $29.8 \pm 8.1$  min and a peak plasma concentration ( $C_{\max}$ ) of  $10.0 \pm 1.0$   $\mu\text{g}/\text{mL}$ . From comparison of the caffeine area under the concentration versus time curve (AUC) after intravenous and oral doses, a complete absolute bioavailability of caffeine was demonstrated (Blanchard and Sawers 1983a). Caffeine pharmacokinetics were independent of the route of administration as shown by superimposable plasma concentration curves, suggesting that there is no important hepatic first-pass effect. Caffeine absorption from food and beverages does not seem to be dependent on age, gender, genetics, and disease or consumption of drugs, alcohol, and nicotine. Caffeine absorption from cola and chocolate was delayed, with  $T_{\max}$  of 1.5–2 h instead of 0.5 h for a capsule and the  $C_{\max}$  values were 1.57 for cola, 1.50 for chocolate, and 2.05  $\mu\text{g}/\text{mL}$  for the capsule (Mumford et al. 1996). Pharmacokinetics of caffeine (100 mg) and its dimethylxanthine metabolites studied after inhalation in heroin users and compared with intravenous and oral administration in healthy volunteers showed a rapid and effective absorption after inhalation with an approximate bioavailability of inhaled caffeine of 60% in experienced smokers (Zandvliet et al. 2005). The efficacy of percutaneous caffeine absorption has been demonstrated in premature infants treated for neonatal apnea (Morisot et al. 1990).

## 2.2 Distribution

Caffeine enters the intracellular tissue water and is found in all body fluids – plasma, cerebrospinal fluid, saliva, bile, semen, breast milk, and umbilical cord blood as well – as in all tissue organs. There is no long-term accumulation of caffeine or its metabolites in the body as seen by whole-animal autoradiography using radiolabeled caffeine (Arnaud 1976).

### 2.2.1 Tissues

The tissue distribution 1 h after intravenous injection of caffeine into rabbits showed that the caffeine tissue-to-blood concentration ratio was approximately 1.0 with concentrations of  $3.32 \pm 0.47$   $\mu\text{g}/\text{g}$ . Exceptions included fat ( $0.79 \pm 0.21$   $\mu\text{g}/\text{g}$ ), adrenals ( $1.96 \pm 0.36$   $\mu\text{g}/\text{g}$ ), liver ( $4.88 \pm 0.48$   $\mu\text{g}/\text{g}$ ), and bile ( $8.97 \pm 0.701$   $\mu\text{g}/\text{g}$ ), in which the ratios were 0.2, 0.6, 1.5, and 2.7, respectively (Beach et al. 1985). Microdialysis applied with simultaneous subcutaneous infusion of caffeine and theophylline (20 mg/kg) to measure their concentrations in blood, adipose tissue, muscle, and liver of rats showed that caffeine was found to be evenly distributed in an in vitro test (Stähle et al. 1991). While there was no

difference between caffeine and theophylline for in vitro recovery, the in vivo recovery of theophylline was significantly less than the recovery of caffeine in brain, liver, muscle, and adipose tissue and this difference was significantly larger in the brain than in other tissues (Stähle 1991).

There is no blood–brain barrier limiting the passage of caffeine through tissues. Therefore, from mother to fetus and to the embryo, an equilibrium can be continuously maintained. The disposition of caffeine and its metabolites, theophylline, theobromine, and paraxanthine, in the 20-day fetal and adult brains following a single maternal dose of 5 or 25 mg/kg caffeine showed that fetal and adult caffeine AUC values did not differ between the brain and plasma at either dose (Wilkinson and Pollard 1993). The brain-to-plasma ratio was close to 1 for a dose of 100 mg/kg and was lower for 10 mg/kg (0.6–0.7) and changed with time for 1 mg/kg, from 0.8 up to 1.9 after 4 h. When the dose of caffeine administered orally in the rat changed by 10 or 100 times, the AUC changed by 22 and 385 times in brain (Latini et al. 1978). Caffeine was found to be evenly distributed with a free concentration of approximately 120  $\mu\text{M}$  and the rate of penetration into brain extracellular space was higher for caffeine than for theophylline (Stähle et al. 1991). The pharmacokinetics of caffeine in the blood and cerebrospinal fluid were similar (Vickroy et al. 2008).

Similarly, caffeine is readily distributed to the fetus (Kimmel et al. 1984). The amniotic fluid to maternal plasma concentration ratio was higher for caffeine than for its major metabolite, paraxanthine, throughout gestation, and increased near term for both compounds. Both compounds distributed nearly homogeneously to fluids and tissues of the 29-day fetus, with mean fetal-to-maternal concentration ratios of 0.7 for paraxanthine and 0.9 for caffeine. The free fraction of caffeine was constant during gestation (about 0.8), while that of paraxanthine increased from 0.25 to 0.4 (Dorrbecker et al. 1988a). Caffeine is also readily excreted in milk (Gilbert et al. 1985), but there are significantly lower concentrations of caffeine and dimethylxanthine metabolites in milk when compared with serum in rabbits (McNamara et al. 1992), while in milk of lactating dairy cows the caffeine concentration was similar to the serum concentration 1.5–24 h after caffeine administration (DeGraves et al. 1995).

The situation in man is similar. After oral or intravenous doses of 5–8 mg/kg, mean plasma concentrations of 8–10 mg/L are observed. The caffeine plasma concentrations then decrease more rapidly than those of its metabolite paraxanthine, so in spite of important interindividual differences, paraxanthine concentrations become higher than those of caffeine within 8–10 h after administration. A good correlation was observed between the concentrations of caffeine in serum and in saliva, so noninvasive salivary measurements may be used for determination of caffeine pharmacokinetics in man (Scott et al. 1984). The caffeine concentrations in saliva were 65–85% of those in plasma (Callahan et al. 1982). After a 200 mg caffeine oral load in healthy adults, the saliva concentrations of caffeine, paraxanthine, and theophylline were lower than the plasma concentrations ( $P < 0.001$ ), whereas the theobromine concentrations in plasma and saliva were similar. The saliva concentrations of these methylxanthines were higher than the free plasma concentrations ( $P < 0.001$ ) (Rodopoulos and Norman 1996). The median AUC

value for caffeine measured from saliva was 72% of that from serum, with variations of 56–95% between individuals (Spigset et al. 1999a).

In newborn infants, similar levels of caffeine concentration were found in plasma and cerebrospinal fluid (Anonymous 1991). Caffeine and its metabolites were detected in cerebrospinal fluid of patients with severe traumatic brain injury and increased concentration was associated with significant favorable outcomes (Sachse et al. 2008).

Urinary and umbilical cord blood analyses of caffeine have been correlated with reported intake throughout pregnancy ( $P < 0.0001$ ) (Grosso et al. 2008). Ex vivo perfusion of the human placenta showed that caffeine crossed the placenta by passive diffusion (Mose et al. 2008) and analyses of human fetal gonads found that the caffeine concentrations were the same as in plasma (Anonymous 1991; Arnaud 1993a).

Transcutaneous collection allowing quantitation of caffeine that diffuses directly through the skin from within the body in healthy volunteers taking caffeine orally showed that the amount of caffeine collected was linearly related to the plasma AUC. Increased sweating carried out on one arm of each subject maintained at 40°C to induce local sweating showed a larger contribution to transdermal collection (40%) in the first 5 h and much less (14%) after 10 h (Conner et al. 1991). Caffeine, paraxanthine, and theobromine measured from transdermal sweat patches that continuously collected and stored analytes lost through the skin showed caffeine and paraxanthine accumulated at comparable rates, while theobromine accumulated more slowly (Delahunty and Schoendorfer 1998).

Values of the milk-to-serum concentration ratio of 0.52 and 0.81 were found in breast milk. As the binding of caffeine to constituents of serum and whole breast milk was 25.1 and only 3.2%, respectively, it was suggested that all the binding in breast milk was accounted for by the butterfat content (Arnaud 1993a).

### 2.3 Excretion

In both animals and man, renal excretion dominates. The metabolic disposition of [1-Me-<sup>14</sup>C]caffeine studied and compared in the rat, the mouse, and the Chinese hamster showed no interspecies differences in urinary excretion of radioactivity, with 67–70% of the administered dose recovered (Arnaud 1985) and after the administration of [8-<sup>14</sup>C]caffeine to various mouse strains, 73–89% of the dose was recovered in urine (Arnaud et al. 1989). In rabbits, after the administration of [2-<sup>14</sup>C]caffeine, 82% of the administered radioactivity was recovered in urine (Beach et al. 1985). After oral administration of [2-<sup>14</sup>C]caffeine and [1-Me-<sup>14</sup>C] caffeine in rats, fecal excretion amounted to 2–7% of the administered dose. Most of the fecal excretion corresponded to caffeine metabolites secreted from enterohepatic cycling with intestinal and biliary secretion (Arnaud 1976). With [1-Me-<sup>14</sup>C]caffeine administered to rats, mice, and Chinese hamsters, no interspecies differences appeared in fecal excretion of caffeine metabolites and 3–6% of the

administered dose was recovered (Arnaud 1985). After the administration of [8-<sup>14</sup>C]caffeine to various mouse strains, 7–12% of the dose was recovered in feces (Arnaud et al. 1989). After oral administration of radiolabeled caffeine in man, the 48-h fecal excretion amounted to 2–5% of the dose. The products identified in the feces of human volunteers were 1,7-dimethyluric acid (1,7DMU), 1-methyluric acid (1MU), 1,3-dimethyluric acid (1,3DMU), 1,3,7-trimethyluric acid (1,3,7TMU), and caffeine, which amounted to 44, 38, 14, 6, and 2% of fecal radioactivity, respectively (Callahan et al. 1982).

Caffeine and its dimethylxanthine primary metabolites are extensively reabsorbed in the renal tubule and their renal clearances were highly urine flow dependent, so urinary excretion varied with urine output. About 70% of the administered oral dose of caffeine (7.5 mg/kg) was recovered in urine (Tang-Liu et al. 1983) but less than 2% of caffeine was excreted unchanged in the urine. This low caffeine urine excretion (0.5–2%) is explained by a 98% renal tubular reabsorption. For higher caffeine intake (1 g, 10–12 cups of coffee), the recovery of caffeine in urine was from 0.74 to 0.91% of the dose and the urinary concentration was 14 mg/L. A good correlation was found between urinary and plasma caffeine concentrations (Birkett and Miners 1991).

## 2.4 Pharmacokinetics

Important pharmacokinetic differences have been reported between animal species, making the extrapolation between species difficult. In most studies performed in animal species, dose-independent pharmacokinetics for caffeine were reported and analyzed according to a one-compartment open model, while at higher doses applied in toxicology, dose-dependent pharmacokinetics were observed with lower plasma clearances. When the dose of caffeine administered orally to rats changed by 10 or 100 times, the AUC by changed by 45 and 746 times in plasma (Latini et al. 1978). These dose-dependent kinetics effects reported in animals can be explained by a saturation of metabolic transformation of caffeine (Bortolotti et al. 1985; Arnaud 1993a, b). Linear or nonlinear caffeine pharmacokinetics may be observed depending on the route and the rate of administration (Lau et al. 1995). The systemic clearance of total caffeine was  $3.83 \pm 1.94$  and  $1.14 \pm 0.80$  mL/min/kg and the unbound systemic clearance was  $5.09 \pm 2.60$  and  $1.41 \pm 0.71$  mL/min/kg in rabbit adults and the pups, respectively. A significant decreased caffeine clearance in the pups is thus demonstrated when compared with the adults (McNamara et al. 1992).

There is minimal (Yesair et al. 1984) or no first-pass metabolism for caffeine in human and the caffeine elimination is a first-order process in healthy human (Arnaud 1993a) and is described by a one-compartment open model system in the dose range of intake of 2–10 mg/kg observed in the population (Blanchard and Sawers 1983a; Newton et al. 1981; Bonati et al. 1982). Dose-dependent kinetics were observed when caffeine plasma levels were higher than 30 mg/L in the case

of acute intoxication in an infant (Jarboe et al. 1986), but in adult subjects some metabolic transformations can be saturated in lower dose range of 1–4 mg/kg, particularly demethylation into paraxanthine, which is selectively catalyzed by CYP1A2 (Tang-Liu et al. 1983; Cheng et al. 1990; Denaro et al. 1990; Arnaud and Enslen 1992). To explain why epidemiology studies reported a nonlinear dose response between coffee consumption and health risks, the presence of a dose-dependent metabolism of caffeine was studied. Under chronic dosing conditions, healthy subjects received a placebo, a low dose of caffeine (4.2 mg/kg/day caffeine), or a high dose of caffeine (12 mg/kg/day caffeine) in decaffeinated coffee and in six divided doses spaced throughout the day. Caffeine clearance fell from 0.118 L/h/kg (placebo treatment) to 0.069 L/h/kg (low dose;  $P < 0.005$ ) and to 0.54 L/h/kg (high dose;  $P < 0.001$ ). The formation and metabolite clearances of paraxanthine, the major primary metabolite of caffeine, also decreased when comparing the low and high doses ( $P < 0.05$ ). These results suggest that caffeine metabolism is dose-dependent, resulting in nonlinear elimination (Denaro et al. 1990). When caffeine clearance was determined on separate occasions using a single oral caffeine (70-, 200-, and 300-mg) dose, caffeine exhibited dose-dependent pharmacokinetics, particularly in subjects who showed high initial clearance with the lowest dose of caffeine. This significant decrease in caffeine clearance with increasing dose from 70 to 300 mg ( $P < 0.01$ ) indicated a saturable caffeine metabolism in the dose range tested (Cheng et al. 1990). In addition to the dose, the plasma kinetics of caffeine can be influenced by the presence of food in the stomach and gastric emptying (Brachtel and Richter 1988). Both genetic and environmental factors are suggested as an explanation for the larger variability of caffeine clearance (Nagel et al. 1990). Fluid intake may also modify renal clearance and thus affect caffeine pharmacokinetics (Trang et al. 1985). Chronovariation in caffeine elimination appears to be small (–25 to 16%) in most of subjects (Levy et al. 1984). Measurements of caffeine clearance, acetylation phenotype, and urinary molar ratios of metabolites [5-acetylamino-6-formylamino-3-methyluracil (AFMU) plus 1-methylxanthine (1MX) plus 1MU to 1,7DMU] were not changed when caffeine was given orally at 10 a.m. and at 10 p.m. (Hashiguchi et al. 1992). Surprisingly, sleep deprivation in healthy subjects receiving 2.1, 4.3, or 8.6 mg/kg caffeine showed a significantly ( $P < 0.05$ ) and disproportional increase in the dose-normalized caffeine AUC. Clearance and the paraxanthine-to-caffeine ratio were significantly decreased with increasing dose, suggesting that under severe sleep deprivation caffeine exhibited dose-dependent pharmacokinetics (Kamimori et al. 1995).

After a single dose of caffeine (4 mg/kg) peak plasma concentrations were observed at 1–2 h with half-lives of 2.5–5 h (Anonymous 1991; Arnaud 1993a). Larger variations of caffeine plasma half-lives from 2.3 to 9.9 h were reported, indicating substantial intersubject variability in its elimination (Blanchard and Sawers 1983a). The half-lives of theophylline and theobromine (6.2–7.2 h) were significantly longer than those of caffeine and paraxanthine (4.1–3.1 h) (Lelo et al. 1986a). A peak serum level of  $13.5 \pm 2.9$  mg/L for caffeine occurred 1 h after the administration and was delayed 1 h later for theophylline when caffeine (10 mg/kg)

and theophylline (5 mg/kg) were given orally to asthmatic young patients. The half-life of caffeine was  $3.9 \pm 1.4$  h and was shorter than the half-lives of theophylline with a twofold higher dosage level (Becker et al. 1984). After oral administration, the total plasma clearance of caffeine was similar to that of paraxanthine (2.07–2.20 mL/min/kg) and approximately twofold higher than the total plasma clearances of theophylline and theobromine (0.93–1.20 mL/min/kg). The unbound plasma clearances of caffeine and paraxanthine were also similar in magnitude (3.11–4.14 mL/min/kg) and also higher than those of theophylline and theobromine (1.61–1.39 mL/min/kg) (Lelo et al. 1986a). In nonsmoking subjects, the mean partial clearance of caffeine to paraxanthine was approximately eightfold and 23-fold greater than that to theobromine and theophylline, respectively, confirming earlier reports that paraxanthine is the major metabolite of caffeine in humans (Lelo et al. 1986b). At the steady state, the volume of distribution of theophylline (0.44 L/kg) was lower than that of the other methylxanthines (0.63–0.72 L/kg) and the unbound volumes of distribution of theophylline and theobromine (0.79 L/kg) were lower than the unbound volume of distribution of caffeine (1.06 L/kg), which was similar to that of paraxanthine (Lelo et al. 1986a).

## 2.5 Metabolism

To assess the validity of the interspecies toxicokinetics of caffeine, theobromine, theophylline, and paraxanthine, absorption, bioavailability, and the route of excretion were generally similar between humans and dogs, rabbits, rats, and mice but there were interspecies differences in the route of metabolism, and the enzymes involved in this process (Walton et al. 2001). CYP1A2, which has been detected only in the liver, and accounts for about 15% of the total cytochromes P450 (CYPs) in the human liver, where its protein content corresponds to  $12.7 \pm 6.2\%$  of total CYP (Shimada et al. 1994), is responsible for more than 90% of caffeine clearance. The large interindividual variability of CYP1A2 activity influences the disposition of a substrate such as caffeine (Landi et al. 1999) and these variations may be due to factors such as gender, race, genetic polymorphisms, and exposure to inducers (Rasmussen et al. 2002). The molar ratios of metabolites of caffeine used as an index of CYP1A2 activity in populations are distributed according to bimodal or trimodal distributions, and normal or unimodal distributions have also been suggested (Landi et al. 1999). At least two distinct liver CYP enzymes with differing substrate affinities have the potential to catalyze caffeine N-demethylations and C8-hydroxylations in vitro but at the low concentrations routinely encountered in vivo, participation by the high-affinity site is expected to predominate (Campbell et al. 1987a). In vivo and in vitro evidence suggests that CYPs involved in the demethylation pathways are distinct from isozymes involved in the hydroxylation pathways, but these different isozymes seem to be under common regulatory control (Robson 1992).



The ratios of urinary concentrations of AFMU to 1MX or AFMU to 1MX plus 1MU (Grant et al. 1984) or the corresponding ratios with the complete conversion of AFMU into 5-acetylamino-6-amino-3-methyluracil (AAMU) (Tang et al. 1986; Kilbane et al. 1990) give markers of acetylator status in man. In addition, the ratio of 1MU to 1MX represents an index of xanthine oxidase, that of 1,7DMU to paraxanthine represents an index of microsomal 8-hydroxylation, that of AFMU plus 1MX plus 1MU to paraxanthine represents an index of microsomal 7-demethylation, and the caffeine metabolic ratio, AFMU plus 1MX plus 1MU to 1,7DMU, reflects microsomal 3-demethylation and also systemic caffeine clearance as well as polycyclic aromatic hydrocarbon-inducible CYP activity (Arnaud and Enslen 1992; Campbell et al. 1987a, b). The molar ratio of paraxanthine to caffeine in urine taken 3–4 h after caffeine administration was proposed as an alternative to evaluate hepatic CYP1A2 activity (Kadlubar et al. 1990). The ratio of paraxanthine to caffeine or the ratio of paraxanthine plus 1,7DMU to caffeine, has been used as an indicator of CYP1A2 activity and the AFMU-to-1MX ratio indicated *N*-acetyltransferase 2 (NAT2) activity; both appear to be polymorphically distributed in human populations with slow and rapid phenotypes (Butler et al. 1992). These ratios have been tested and validated (Spigset et al. 1999a; Butler et al. 1992; Tang et al. 1994; Carrillo et al. 2000; Doude van Troostwijk et al. 2003b; Derby et al. 2008) but a more detailed analysis of the literature is beyond the scope of this review. No association was found between acetylation activity and sex; race; age; education; smoking; physical activity; weight; consumption of coffee, alcohol, red meat, processed meat, and cruciferous vegetables; or use of estrogens, after taking the genotype into account (Le Marchand et al. 1996). Drug cocktails have been developed for simultaneous phenotyping of CYP1A2, CYP2A6, CYP2C9, CYP2E1, CYP2C19, CYP2D6, CYP3A, NAT2, and xanthine oxidase (Streetman et al. 2000; Zhu et al. 2001; Christensen et al. 2003; Fuhr et al. 2007; Ryu et al. 2007).

CYP1A2 was responsible for caffeine 3-demethylation and paraxanthine 7-demethylation and may catalyze virtually all reactions related to caffeine and its metabolites. Caffeine biotransformation by CYP1A2 averaged 81.5% for paraxanthine, 10.8% for theobromine, and 5.4% for theophylline, while CYP2E1 had major influences on the formation of theophylline and theobromine (Gu et al. 1992). Whereas CYP1A2 accounts for the high-affinity component of all three human hepatic caffeine *N*-demethylations, CYP2E1 appears to be the main enzyme involved in the low-affinity components of caffeine *N*1- and *N*7-demethylation, while 8-hydroxylation of caffeine was suggested to be catalyzed predominantly by a CYP3A isoform (Tassaneeyakul et al. 1994). CYP2D6-Met also had high intrinsic clearance and catalyzed caffeine demethylation and 8-hydroxylation. CYP2E1 played a less important role *in vitro* and CYP3A4, which predominantly catalyzed 8-hydroxylation, may contribute significantly to the *in vivo* formation of 1,3,7TMU, owing to its high abundance in human liver. Thus at least four CYP isoforms are involved in caffeine metabolism at 3 mmol/L caffeine concentration, but at concentrations below 0.1 mmol/L, CYP1A2 and CYP1A1 are the most important isoenzymes (Ha et al. 1996).

In humans no gender differences in caffeine metabolism were observed from urinary metabolite patterns or metabolite ratios (Grant et al. 1983), although higher activity of CYP1A2 has been shown in men than in women (Landi et al. 1999). This general conclusion is supported by other studies (Vistisen et al. 1992; Campbell et al. 1987b; Kall and Clausen 1995; Rasmussen et al. 2002; Chung et al. 2000; Ghotbi et al. 2007; Begas et al. 2007; Djordjevic et al. 2008). During pregnancy, the excretion of 1MX and of 1MU were increased (Scott et al. 1986). This observation is in agreement with a caffeine study showing a significantly increased hydroxylation activity during pregnancy. Late pregnancy was also characterized by a decrease in CYP1A2, xanthine oxidase, and acetyltransferase activities (Bologa et al. 1991). In nonsmoking pregnant women and in smoking women using oral contraceptives, the caffeine metabolic ratio was reduced by 29 and 20%, respectively, compared with a control group, demonstrating an inhibition of CYP1A2 (Vistisen et al. 1991). Metabolic ratios for the CYP1A2 index during early, middle, and late pregnancy were significantly lower than the ratio after delivery ( $P < 0.0001$ ). A lower metabolic ratio for NAT2 was also observed during pregnancy ( $P < 0.01$ ) but there was no significant difference in the metabolic ratios for xanthine oxidase during pregnancy and after delivery (Tsutsumi et al. 2001). Oral contraceptive users had lower ( $P < 0.05$ ) ratios of paraxanthine 7-demethylation to 8-hydroxylation products than women not taking oral contraceptives (Campbell et al. 1987b). Upon administration of oral contraceptives, the urinary excretion of caffeine, paraxanthine, and 1,7DMU was increased at the expense of 1MX, 1MU, and the acetylated metabolites AFMU and AAMU. A 33% decrease of the caffeine metabolic ratio was reported in women using oral contraceptives (Kalow and Tang 1991a).

The caffeine ratio AFMU plus 1MU plus 1MX to 1,7DMU in a 6-h urine sample was significantly higher in women not taking oral contraceptives compared with women taking oral contraceptives, thus confirming that CYP1A2 is inhibited by oral contraceptives (Rasmussen et al. 2002). As a marker of CYP1A2 activity, the plasma caffeine-to-paraxanthine ratio was 2.8 times higher ( $P < 0.001$ ) in the oral contraceptive (ethinylestradiol) users than in the control subjects, suggesting an inhibition of CYP1A2 activities (Granfors et al. 2005).

Analysis of caffeine metabolites revealed two interethnic variations, one pertaining to the acetylation polymorphism and the other consisting of a difference in paraxanthine excretion, which might indicate an ethnic difference in renal function (Kalow 1986). A nonsignificant higher proportion of rapid acetylator was observed in the Oriental compared with the European population and a 6.3-fold range variation was observed (Kalow and Tang 1991a). The NAT2 activity showed a typically bimodal distribution with 47% fast acetylators and 53% slow acetylators, consistent with a Danish population (Vistisen et al. 1992). However, only 11.0% of Japanese men and women residents of Kyushu were slow acetylators (Saruwatari et al. 2002). With use of the urinary caffeine metabolic ratio AFMU to 1X (less than 0.6) to classify subjects as slow acetylators, a prevalence of this phenotype of 92.2 and 74.5% was noted in two studies in a population of Minnesota Hmong, but a significant discordance between phenotype and genotype was identified

(Straka et al. 2006). The combined low-risk phenotype (slow CYP1A2/rapid NAT2) was more common in blacks than in whites (25 vs. 15%,  $P < 0.02$ ), but there were no significant racial differences in slow and rapid CYP1A2 phenotypes, and in the combined slow NAT2/rapid CYP1A2 phenotype (Muscat et al. 2008). The ratios reflecting CYP1A2 activities were described as log-normal-distributed (Vistisen et al. 1992). CYP1A2 activity was not normally distributed in subjects from Arkansas, Italy, and China and appeared trimodal with arbitrary designation of slow, intermediate, and rapid phenotypes, which ranged from 12–13% slow, 51–67% intermediate, and 20–37% rapid (Butler et al. 1992). Slow and intermediate CYP1A2 metabolizers represent about 50% of Caucasians, while their frequency in Japanese subjects seems to be much lower (Landi et al. 1999). The distribution of CYP1A2 measured with the plasma paraxanthine-to-caffeine ratio in a Chinese population showed a 16-fold variation of CYP1A2 activity and a coefficient of variation of 62.9%. Nonnormal CYP1A2 activity ( $P < 0.001$ ) with a bimodal distribution ( $P < 0.01$ ) was observed. The percentage of poor metabolizers was 5.24% in this Chinese population (Ou-Yang et al. 2000). The metabolic ratio for CYP1A2 was not polymorphic in Japanese subjects and decreased 1,7DMU formation from caffeine in poor metabolizers of CYP2A6 appeared to affect the metabolic ratio used for the assessment of CYP1A2 activity (Saruwatari et al. 2002). CYP1A2 enzyme activity determined using the 4-h plasma paraxanthine-to-caffeine ratio was 1.54-fold higher in Swedes than in Koreans ( $P < 0.0001$ ) despite them having the same CYP1A2 genotype, smoking habit, and oral contraceptive use. Four known (CYP1A2\*1A, CYP1A2\*1D, CYP1A2\*1F, and CYP1A2\*1L) and two novel (CYP1A2\*1V and CYP1A2\*1W) haplotypes were found (Ghotbi et al. 2007). The mean CYP2A6 activity measured by the caffeine metabolite ratio (1,7DMU to paraxanthine) was significantly lower in Japanese Americans than in native Hawaiians ( $P = 0.001$  and  $P < 0.0001$ , respectively) or whites ( $P < 0.0001$ ) (Derby et al. 2008). The xanthine oxidase index was not different between Chinese and European populations and showed a 1.7-fold range variation (Kalow and Tang 1991a). The ratios reflecting xanthine oxidase activities were normally distributed (Vistisen et al. 1992). Low xanthine oxidase activities exist in a Japanese population corresponding to 11% of the subjects with a mean urinary uric acid concentration 53% lower than that of the other subjects ( $P < 0.0001$ ) (Saruwatari et al. 2002).

As noted in Sect. 2.5, drug intake is expected to alter caffeine metabolism when competitive inhibition or induction of the relevant enzymes is observed. Such interactions can involve smoking and Chinese herbal medicines, but St John's wort, garlic oil, *Panax ginseng*, and *Ginkgo biloba* showed no effect on CYP1A2 activity measured from the paraxanthine-to-caffeine serum ratio (Gurley et al. 2002). As expected, allopurinol treatment caused a specific, dose-dependent inhibition of the conversion of the caffeine metabolite 1MX to 1MU, thus validating an in vivo index of xanthine oxidase activity in man (Grant et al. 1986; Lelo et al. 1989). The proton pump inhibitor omeprazole induces hepatic CYP1A2 activity, as shown by the increased N3-demethylation of [3-Me-<sup>13</sup>C]caffeine. In extensive metabolizers there was a 8–17% CYP1A2 induction after administration of

40 mg omeprazole and a 25–32% increase ( $P < 0.002$ ) was observed with 120 mg/day. In poor metabolizers a higher increase of 40–41% was observed and there was a good correlation between the caffeine breath test and plasma caffeine clearance (Rost and Roots 1994).

## 2.6 Sources of Variation in Caffeine Pharmacokinetics and Metabolism

Caffeine metabolism is affected by genetic determinants, age, pregnancy, diet, and lifestyle such as smoking, environmental factors, medications, including contraceptive use, and disease states.

### 2.6.1 Age

The pharmacokinetics of caffeine studied in young dogs aged 1 day, and 7, 14, and 30–45 days and adult dogs showed that the plasma elimination half-life was  $47.5 \pm 5.35$  h in 1-day-old puppies, as opposed to  $6.66 \pm 0.85$  h in adult dogs. A rapid decrease in plasma half-lives occurred during the first 2 weeks of life and at about 14 days of age the caffeine plasma half-life was similar to that of adults. The volume of distribution was greatest and the total body clearance was smaller in the 1-day-old dogs (Warszawski et al. 1977). The time needed to reach the plateau of the cumulative excretion of radioactivity in the urine decreased with age. All these results are consistent with the slow plasma elimination of caffeine in the newborn as compared with the adult (Warszawski et al. 1982). The elimination of caffeine is impaired in neonates because of their immature metabolizing hepatic enzyme systems (Pons et al. 1988) and plasma half-lives of 65–103 h in neonates have been reported, decreasing rapidly to 14.4 h in 3–5-month-old infants, 2.6 h in 5–6-month-old infants, and 3–6 h in adults and the elderly. The clearance of 31 mL/kg/h in 1-month-old infants increases to a maximum of 331 mL/kg/h in 5–6-month-old infants, and is 155 mL/kg/h in adult subjects. A mean distribution volume of 0.7 L/kg (0.5–0.8 L/kg) was found in newborn infants, adult subjects, or aged subjects. The pharmacokinetics of caffeine in healthy young men aged  $20.5 \pm 2.0$  years and in healthy elderly men aged  $71.2 \pm 3.9$  years showed that  $T_{\max}$ ,  $C_{\max}$ , and caffeine bioavailability were essentially identical. The apparent volume of distribution was significantly lower but the larger clearances and the greater elimination rate constant in the elderly subjects were not significant because of the wide intersubject variability as shown in the caffeine half-lives ranging from 2.27 to 9.87 h. In this study, pharmacokinetic parameters of caffeine were similar in young and elderly men (Blanchard and Sawers 1983b). The renal clearance of caffeine calculated following both oral and intravenous doses of caffeine in young and elderly, healthy human volunteers showed a highly statistically significant positive correlation ( $P < 0.001$ ) between the renal clearance of both unbound and total clearance of caffeine and the mean urine flow rate (Blanchard and Sawers

1983c). Thus, the comparative pharmacokinetics in the young and elderly shows no significant differences in half-life, suggesting that aging does not alter caffeine elimination in contrast to the rat model, where an age-dependent increase of caffeine half-life has been observed.

## 2.6.2 Gender and Hormones

The caffeine apparent volume of distribution and the caffeine elimination rate constant were influenced by the different modes of maternal caffeine ingestion during the pre-mating and pregnant periods (Nakazawa et al. 1985). The disposition of caffeine given as single oral dose of 5 and 25 mg/kg to 20-day pregnant and nonpregnant rats showed a significantly longer plasma half-life in the pregnant than in the nonpregnant rats for the highest dose, while the elimination rate was similar at the lowest dose (Abdi et al. 1993). In pregnant rabbits, the pharmacokinetics of caffeine received by continuous intravenous infusion through 29 days of gestation exhibited increased plasma concentrations of caffeine and its major metabolite paraxanthine in the last half of gestation. Rabbits exhibited caffeine AUC at 29 days of gestation that were 85–165% greater than those observed before mating, suggesting that the elimination of caffeine is diminished in late gestation in the rabbit (Dorrbecker et al. 1988a).

Similarly, the caffeine half-life was prolonged during the last trimester in pregnant women and returned to the prepregnant value a few weeks after they had given birth (Arnaud 1993a). From a cohort study of normal third-trimester pregnancies with significant “high”(H) and “low”(L) long-term maternal caffeine ingestion ( $P < 0.0002$ ), it was shown that the maternal serum caffeine levels in group H were significantly higher ( $P < 0.05$ ) at each week of gestation than those in group L and increased until 36 weeks ( $P < 0.0039$ ) but did not increase significantly in group L until 40 weeks (Devoe et al. 1993). The half-life of caffeine increases during pregnancy, reaching 11.5–18 h by the end of pregnancy, leading to an accumulation with regular daily consumption as neither the fetus nor the placenta can metabolize caffeine (Grosso and Bracken 2005). Comparisons of the follicular and luteal phases revealed that systemic clearance of caffeine was slower in the luteal phase, an effect related to the proximity to onset of menstruation and to levels of progesterone although the half-life did not differ (Lane et al. 1992). All pharmacokinetic parameters were similar between women taking no oral contraceptives and men except for the volume of distribution, which was significantly larger in the women ( $P < 0.05$ ) (Patwardhan et al. 1980), and gender had no significant effect on caffeine pharmacokinetics (McLean and Graham 2002). Oral contraceptive use has been shown to double the caffeine half-life (Abernethy and Todd 1985; Patwardhan et al. 1980; Arnaud 1993a). As compared with women taking no oral contraceptives, the half-life of caffeine was significantly prolonged in women taking oral contraceptives from  $6.2 \pm 1.6$  to  $10.7 \pm 3.0$  h ( $P < 0.001$ ), showing impaired elimination of caffeine. Women taking oral contraceptives had a significantly lower total plasma clearance ( $0.79 \pm 0.21$  vs.  $1.3 \pm 0.35$  mL/min/kg)

and free clearance ( $1.12 \pm 0.28$  vs.  $1.97 \pm 0.57$  mL/min/kg) than women not taking oral contraceptives, while the volumes of distribution and plasma binding were similar in both groups (Patwardhan et al. 1980). Oral contraceptive steroids increased twofold the residence time of caffeine in young women. The effect was already observed during the first cycle 2 weeks after starting to take oral contraceptive steroids and was slightly increased in the second cycle, after 6 weeks on oral contraceptive steroids (Rietveld et al. 1984). The effect of chronic administration of low-dose estrogen-containing (less than 50  $\mu$ g estrogen) oral contraceptives on the pharmacokinetics of caffeine confirmed that the elimination half-life of caffeine was prolonged to 7.88 h versus 5.37 h in the controls, as a result of impairment of the plasma clearance of caffeine (1.05 vs. 1.75 mL/min/kg, respectively), with no change in the apparent volume of distribution (Abernethy and Todd 1985). The plasma caffeine clearances and elimination half-lives after ingestion of a guarana-containing supplement were lower ( $0.34 \pm 0.01$  vs.  $0.99 \pm 0.41$  mL/min/kg) and longer ( $15.5 \pm 0.3$  vs.  $5.6 \pm 1.7$  h), respectively, in subjects taking oral contraceptives (Haller et al. 2002).

### 2.6.3 Physical Exercise

The effect of moderate exercise (30% of maximum O<sub>2</sub> uptake) on the kinetics of caffeine in healthy volunteers showed that exercise significantly raised  $C_{\max}$  and reduced both the half-life and the volume of distribution (Collomp et al. 1991), but other studies show minimal effects on caffeine pharmacokinetics (Kamimori et al. 1987; McLean and Graham 2002).

### 2.6.4 Obesity

Obesity increases the apparent volume of distribution ( $69.9 \pm 5.9$  vs.  $43.6 \pm 2.8$  L;  $P < 0.001$ ), with no significant change in clearance and a trend toward a prolonged elimination half-life (Abernethy et al. 1985). At rest, obese subjects (more than 30% body fat) had a significantly higher absorption rate constant ( $K_a$  0.0757 vs. 0.0397/min), a lower elimination rate constant ( $K_e$  0.0027 vs. 0.0045/min), and a longer serum half-life (4.37 vs. 2.59 h) in comparison with lean subjects. In exercise as well as at rest, lean and obese subjects had a large difference in the volume of distribution, 43.2 versus 101 L in exercise and 54.1 versus 103 L at rest. Exercise consistently resulted in a decrease in caffeine  $C_{\max}$  and AUC in obese subjects (Kamimori et al. 1987). In severely obese subjects, the caffeine half-life and oral clearance rate were not altered significantly, but it was confirmed that obese individuals exhibited an increased volume of distribution and this volume was decreased with weight reduction. The effect was more important in females and it was suggested that the caffeine distribution was incomplete into the adipose tissue representing 70–80% excess of body weight in obese subjects (Caraco et al. 1995). After oral administration of caffeine as coffee in obese subjects (body mass index

28.01 ± 0.92) and control subjects, there was no significant difference in the caffeine and theobromine levels in saliva but significantly lower levels of theophylline ( $P < 0.05$ ) and higher levels of paraxanthine ( $P < 0.01$ ) were found in obese subjects, suggesting that obesity alters caffeine metabolism and modifies the urinary metabolite concentration ratios used as indexes of enzyme activities (Bracco et al. 1995).

### 2.6.5 Drugs

Given the *major* role of the liver in the metabolism of caffeine and many drugs, a few examples of these interactions are described from a very large literature. Clinical studies have reported frequent drug interactions leading to impaired caffeine elimination and decreased clearance both for caffeine and for its metabolites (Lelo et al. 1989). Traditional medicine as well as supplements prepared from plant extracts may affect caffeine pharmacokinetics. With use of caffeine as a probe drug, the effect of sodium tanshinone IIA sulfonate, a water-soluble derivative of the Chinese medicine Danshen, on the activity of CYP1A2 in humans has been tested on healthy volunteers. CYP1A2 activity monitored by the ratio of paraxanthine to caffeine at 6 h in plasma significantly increased by 41.1%, the AUC of caffeine significantly decreased by 13.3%, and the AUC of paraxanthine significantly increased by 17.4% (Chen et al. 2009a). After administration of St John's wort (*Esbericum* capsules; 240 mg/day of extract, 3.5 mg hyperforin) or a placebo, no statistically significant differences of the primary kinetic parameter, the AUC of caffeine and paraxanthine, between the placebo group and the St John's wort group were observed (Arold et al. 2005). Interaction between the selective serotonin reuptake inhibitor fluvoxamine (50–100 mg/day) and caffeine (200 mg orally) in healthy volunteers showed a decreased total clearance of caffeine from 107 to 21 mL/min and an increased half-life from 5 to 31 h. The N3-, N1-, and N7-demethylation clearance of caffeine decreased from 46 to 9 mL/min, from 21 to 9 mL/min, and from 14 to 6 mL/min, respectively (Jeppesen et al. 1996). However, fluvoxamine (50 mg/day orally) disposition studied in healthy nonsmoking male volunteers who also received caffeine (200 mg orally) showed no significant correlations between caffeine and fluvoxamine clearance or between the paraxanthine-to-caffeine ratio in serum 6 h after caffeine intake and fluvoxamine oral clearance (Spigset et al. 1999b), in contrast to previous *in vitro* (Brøsen et al. 1993) and *in vivo* (Sperber 1991; Jeppesen et al. 1996) studies. Other drugs that may interfere include the antipsychotic drug clozapine (Doude van Troostwijk et al. 2003a), the anti-inflammatory drugs idrocilamide (Brazier et al. 1980a) and rofecoxib (Backman et al. 2006), and tacrine (Fontana et al. 1998). Caffeine metabolism has been shown to be inhibited by quinolone antibiotics. *In vitro* tests ranked the likelihood of these interactions as follows: enoxacin, 74.9%; ciprofloxacin, 70.4%; nalidixic acid, 66.6%; piperimidic acid, 59.3%; norfloxacin, 55.7%; lomefloxacin, 23.4%; pefloxacin, 22.0%; amifloxacin, 21.4%; difloxacin, 21.3%; ofloxacin, 11.8%; temafloxacin, 10.0%; fleroxacin, no effect. *In vivo* studies showed that



the likelihood of an interaction with caffeine is as follows: enoxacin > ciprofloxacin = norfloxacin > ofloxacin = lomefloxacin (Fuhr et al. 1992). Among fluoroquinolones, enoxacin and to a lesser extent ciprofloxacin and pefloxacin inhibit the metabolic clearance of caffeine (Kinzig-Schippers et al. 1999; Granfors et al. 2004) and it was suggested to use noninteracting quinolones such as ofloxacin and norfloxacin.

Other antidepressants and drugs for the management of anxiety disorders such as venlafaxine, alprazolam, zolpidem, and trimethadione as well as the wakefulness-promoting agent armodafinil did not significantly alter the pharmacokinetics of caffeine and its metabolites (Amchin et al. 1999; Schmider et al. 1999; Cysneiros et al. 2007; Tanaka et al. 1993; Darwish et al. 2008).

### 2.6.6 Disease

Several animal models of liver disease show reduced total body clearance of caffeine (Tanaka et al. 1992a, 1995; Schaad et al. 1995; Kokwaro et al. 1993). Similarly, humans with several types of liver disease, including cirrhosis (Wietholtz et al. 1981), noncirrhotic, chronic hepatitis B or C, and subjects with cirrhosis, showed a highly significant reduction in plasma clearance correlating with the severity of the disease ( $P < 0.001$ ) (Park et al. 2003; Tanaka et al. 1992b; Scott et al. 1989). The reduced plasma disappearance rate of caffeine in cirrhotics was related to the delayed formation of paraxanthine (Holstege et al. 1989; Jodynis-Liebert et al. 2004). Chronic consumption of alcohol leading to cirrhosis was shown to increase the caffeine half-life up to 50–160 h (Statland and Demas 1980; Desmond et al. 1980; Renner et al. 1984; Scott et al. 1988). A study performed in patients with decompensated type I and type II diabetes mellitus showed that the caffeine half-life, apparent clearance and distribution volume, and paraxanthine-to-caffeine ratio for the CYP1A2 index were similar to those of controls (Zysset and Wietholtz 1991; Matzke et al. 2000).

### 2.6.7 Smoking

Although some studies showed no effect (Oliveto et al. 1991), most studies found that caffeine clearance was stimulated by smoking (Brown and Benowitz 1989; Parsons and Neims 1978; Wietholtz et al. 1981; Arnaud and Welsch 1982; Kotake et al. 1982; Caraco et al. 1995; Zevin and Benowitz 1999; Bchir et al. 2006). Cigarette smoking nearly doubles the rate of caffeine metabolism owing to the enzyme-inducing effects of polycyclic aromatic hydrocarbons, known to increase liver enzyme activity (Kalow and Tang 1991b; Parsons and Neims 1978). Multivariate analysis revealed that with disease state, smoking ( $P < 0.001$ ) was a significant predictor of the caffeine breath test, thus showing it to be a valid indicator of plasma caffeine clearance and hepatic function (Park et al. 2003). Cigarette smoking increases the elimination of caffeine, whereas cessation of



cigarette smoking significantly reduces caffeine clearance (Murphy et al. 1988) and changes the pattern of caffeine metabolism (Brown et al. 1988). The time-course changes of CYP1A2 activity measured from the paraxanthine-to-caffeine ratio in plasma after cessation of smoking in heavy smokers showed that the initial caffeine clearance decreased significantly ( $P < 0.01$ ) by 36.1% and the apparent half-life of the CYP1A2 activity decrease was 38.6 h (Faber and Fuhr 2004).

## 2.6.8 Diet and Alcohol

During the treatment of neonatal apnea, formula-fed infants, compared with breastfed infants, show a nearly threefold increase in the clearance of caffeine. In HepG2 cells, messenger RNA and protein expression of CYP1A1/CYP1A2 were significantly induced by cow-milk-based formula, but not by human milk. The enhanced in vitro CYP1A expression via an AhR-mediated pathway by infant formula but not human milk provides a potential mechanistic basis for the increased caffeine elimination in formula-fed infants (Xu et al. 2005). The caffeine elimination rate constant was low 2 weeks after birth and displayed a significant positive linear correlation with age ( $P < 0.001$ ). A significantly greater elimination rate constant was observed in formula-fed than in breast-fed infants ( $P < 0.001$ ). This occurred concomitantly with a significant increase in the levels of urinary paraxanthine and 1MX ( $P < 0.001$ ), suggesting increased CYP1A2 activity in formula-fed infants. The urinary molar ratio of paraxanthine plus 1MX to caffeine and age strongly predicted the caffeine elimination rate constant ( $P < 0.001$ ) irrespective of feeding type (Blake et al. 2006).

The influence of nutritional status was investigated in elderly institutionalized patients with either malnutrition or adequate nutrition. The plasma paraxanthine-to-caffeine metabolic ratio was similar in both groups and was not correlated to the body mass index, serum albumin, or renal clearance (Hamon-Vilcot et al. 2004). Daily consumption of at least three cups of coffee significantly increased CYP1A2 enzyme activity (Djordjevic et al. 2008). These results confirmed the findings of a previous study showing that multiple ingestions of dietary caffeine (two to seven cups of coffee) in healthy subjects increased the theophylline serum concentrations given as a single oral dose when compared after deprivation of dietary caffeine. The theophylline half-life was prolonged by 32% ( $P < 0.01$ ) and the total body clearance was reduced by 23% ( $P < 0.001$ ) (Sato et al. 1993). When subjects resumed coffee drinking, interindividual variations preclude a clear answer about the time period required for deinduction to occur. However, regular caffeine intake in high doses for 1 week failed to alter caffeine pharmacokinetics (George et al. 1986).

Grapefruit juice beverage consumption decreased the oral clearance of caffeine by 23% and prolonged its half-life by 31% (Fuhr et al. 1993, 1995). The pharmacokinetics of caffeine and its metabolite paraxanthine were affected by the flavonoid quercetin as shown by their significantly decreased AUC (16%) and decreased urinary excretion of paraxanthine (32%) and 1MX (156%), while urinary excretion of 1,7DMU and of 1MU were both increased by 90% (Chen et al. 2009b).

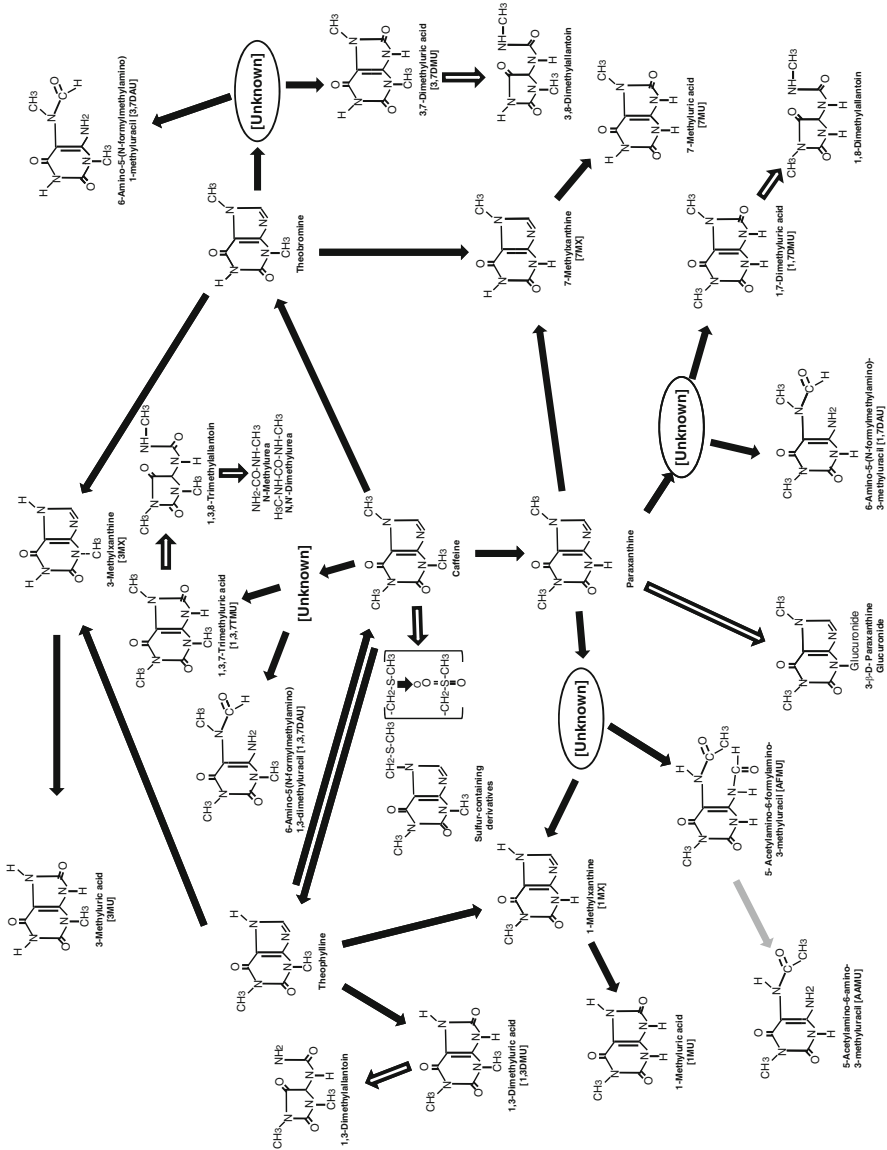
Alcohol intake in amounts commonly consumed significantly prolonged the caffeine half-life by 72% ( $P < 0.005$ ) and diminished the caffeine clearance by 36% ( $P < 0.0005$ ) (George et al. 1986), while the AUC for caffeine was significantly higher when caffeine was administered with 0.8 g/kg alcohol (Azcona et al. 1995).

## 2.7 Metabolites and Metabolic Pathway

Metabolites specific to animal species were identified in urine, such as trimethylallantoin (Arnaud et al. 1986a), sulfur-containing metabolites (Kamei et al. 1975) and *N*-methylurea and *N,N'*-dimethylurea (Arnaud 1976), and may be produced by the intestinal flora. These sulfur-containing metabolites of caffeine were detected in the urine of the horse, rabbit, rat, and mouse and were isolated and identified as  $\alpha$ -[7-(1,3-dimethylxanthinyl)]methyl methyl sulfoxide, while two other new metabolites were isolated from the urine of the mouse and identified as  $\alpha$ -[7-(1,3-dimethylxanthinyl)]methyl methyl sulfide and  $\alpha$ -[7-(1,3-dimethylxanthinyl)]methyl methyl sulfone, respectively (Kamei et al. 1975). Bacterial degradation through C8 oxidation results in the formation of 1,3,7TMU, which is further degraded to trimethylallantoin, *N,N'*-methylurea and *N*-methylurea (Madyastha and Sridhar 1998). In urine; a larger fraction of 6-amino-5(*N*-formylmethylamino)-1,3-dimethyluracil (1,3,7DAU) was excreted in rat (30%) in contrast with monkey and man (2%) (Latini et al. 1981). When [Me-<sup>14</sup>C]-1,3,7DAU was administered orally or intravenously to rats, no further metabolites could be found (Arnaud et al. 1983). After identification and quantification in rat urine of 1,3,8-trimethylallantoin (1–14%) (Rao et al. 1973; Arnaud 1976), its formation from caffeine was demonstrated in rat liver slices and all *N*-demethylation, oxidation to uric acids, and formation of uracil derivatives were also demonstrated in vitro (Arnaud et al. 1986a). In rat liver slices only primary metabolites were detected and *N*1-demethylation was the most important pathway, with theobromine representing 51% of total dimethylxanthines produced and 1,3,7DAU was an important metabolite, corresponding to 9.7% of total caffeine metabolites (Bienvenu et al. 1990). In mice strains paraxanthine glucuronide was identified, a metabolite not found in other animal species and in humans (Arnaud et al. 1989). In beagle dogs, the most important metabolic pathway of caffeine (2.8% of the dose) was the 7-methyl demethylation to theophylline (8% with paraxanthine) with further metabolism to 1,3DMU (13%), 3-methylxanthine (3MX) (21%), and 1MU (8%) excreted in urine. Minor metabolites were theobromine (5%), 1,3,7TMU (2.5%), 1,7DMU (2%), 1MX (1%), and 7-methyluric acid (7MU) (2.5%) (Aldridge and Neims 1979). In 2-day-old puppies, urinary caffeine metabolites derived, respectively, from paraxanthine, theophylline, and theobromine accounted for 42, 33 and 14%. Between 2 and 22 days of age, this metabolic pattern changed, with metabolites derived from theophylline increasing from 33 to 82% (Aldridge and Neims 1980). The metabolism of [2-<sup>14</sup>C]caffeine (4 mg/kg intravenously) studied in rabbits showed that the major urinary metabolites were 1MX (22%), 1MU (19%),

7-methylxanthine (7MX) (16%), and paraxanthine (14%), with other minor metabolites such as 3-methyluric acid (3MU) (4.4%), theobromine (4.0%), 1,7DMU (3.9%), 3MX (3.8%), 1,3DMU (2.7%), 1,3,7TMU (2.0%), and theophylline (1.6%), while the uracil derivative AAMU amounted to 4.9% (Beach et al. 1985). After caffeine administration, similar hepatic capacity to clear caffeine was observed, but 7-demethylation was the preferred pathway in sheep and 3-demethylation in cattle, suggesting different species-specific expression of the CYP1A subfamily (Danielson and Golsteyn 1996).

Caffeine metabolism in humans includes multiple and separate pathways with demethylation to dimethylxanthines and monomethylxanthines, C8 oxidation of these methylxanthines into methylurates, and ring opening yielding substituted uracil derivatives. The reverse biotransformation of theophylline to caffeine was first shown in infants but later also in adult subjects. From the metabolic pathways of caffeine (Fig. 1) it is apparent that each metabolite may be derived from more than one precursor and assessment of caffeine demethylations from urinary metabolite profiles is not accurate (Lelo et al. 1986b). In Fig. 2 the various ratios of metabolites that have been used to measure activities of enzymes involved in caffeine metabolism are shown, particularly CYP1A2, NAT2, and xanthine oxidase. The analysis of urinary caffeine metabolites in man shows the presence of uracil derivatives produced from caffeine, 1,3,7DAU (Arnaud and Welsch 1980a), from theobromine, 6-amino-5-(*N*-formylmethylamino)-1-methyluracil (3,7DAU) (Arnaud and Welsch 1979a, 1980a), and from paraxanthine, 6-amino-5-(*N*-formylmethylamino)-3-methyluracil (1,7DAU) (Arnaud and Welsch 1980a). The amount of 1,3,7DAU found in the urine of adult subjects is about 1% of the administered dose, while its excretion increased in the urine of a premature infant in the case of caffeine overload (Gorodischer et al. 1986a). In this study, the 1,3,7DAU identified in urine was of neonatal and not of maternal origin as it was not present in the urine from the infant obtained prior to administration of caffeine. The acetylated uracil derivative AAMU detected in man (Callahan et al. 1982) has not been identified in animal species. Its precursor was detected, isolated, purified, and identified as AFMU, a structure confirmed by chemical synthesis (Tang et al. 1983). AFMU was unstable in the presence of dilute base and/or methanol, giving rise to a deformylated compound, AAMU, which was reported in the literature as a major metabolite of caffeine in man (Arnaud 1980; Arnaud and Welsch 1980a; Callahan et al. 1982). The production and excretion rates of AAMU and AFMU were shown to be related to the acetylation polymorphism (Grant et al. 1983) with a bimodal distribution of the general population into fast and slow acetylators. Paraxanthine is the precursor of AFMU, which accounts for 67% of paraxanthine clearance. The rate of AFMU production and clearance approximates and changes according to the rates for the production of 1MX and 1MU (Yesair et al. 1984), suggesting that its formation occurs through a common precursor of AFMU and 1MX. This intermediate has not yet been identified. A major difference between humans and rats is the total excretion of caffeine and metabolites without demethylation, which amounts to 5 and 42% of the dose administered, respectively (Arnaud and Welsch 1980a; Arnaud 1985). From the analyses of urine metabolites



**Fig. 1** Metabolic pathways of caffeine, theophylline, theobromine, and paraxanthine in human (→) and animals (⇨)

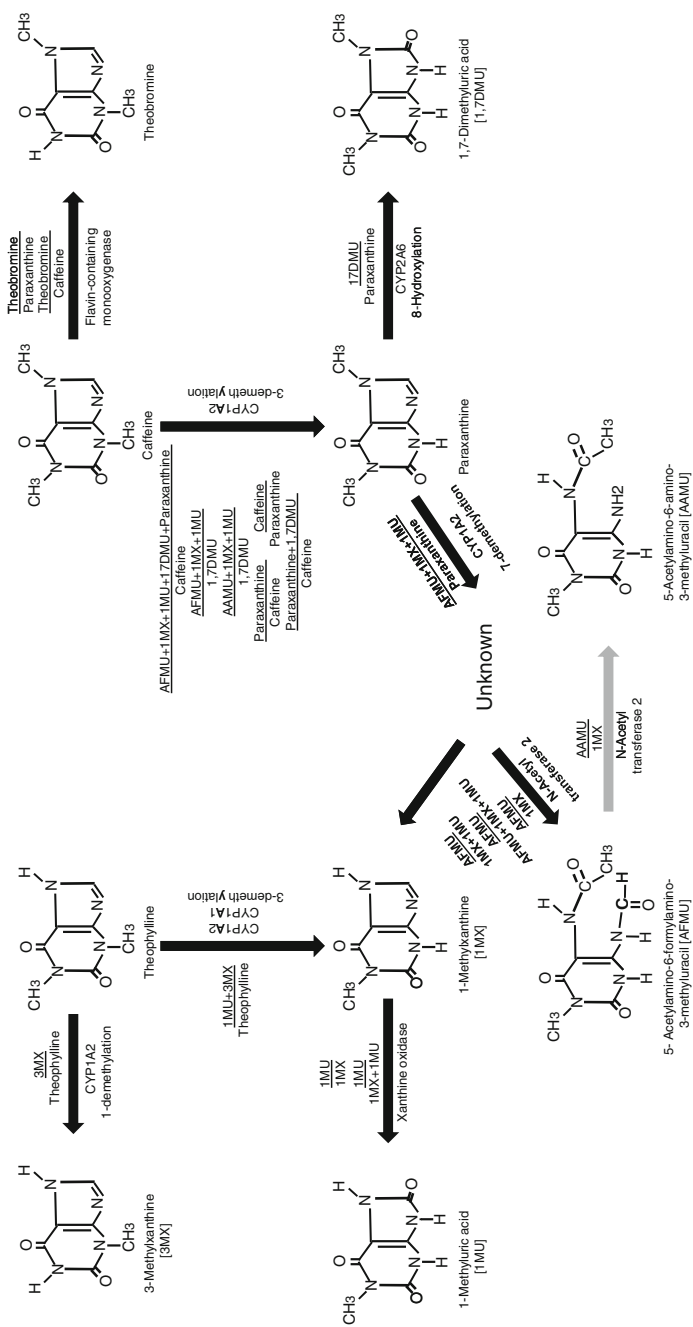


Fig. 2 Metabolite ratios measured in urine and plasma used as indexes of enzyme activities

in humans, the quantitative importance of the metabolic pathways through paraxanthine (72%) was the greatest followed by theobromine (20%) and theophylline (8%) (Arnaud and Welsch 1980a, 1982). These results were confirmed where  $78 \pm 11\%$  of the excreted metabolites were metabolized through the paraxanthine pathway,  $14 \pm 8\%$  through theobromine, and  $9 \pm 4\%$  through theophylline. However, the plasma AUC for dimethylxanthines underestimates the formation of paraxanthine, overestimates the formation of theobromine, and gives a similar formation for theophylline from caffeine, when compared from the urinary metabolites (Rodopoulos and Norman 1996). From the plasma AUC of caffeine and each dimethylxanthine, the mean fractional conversion of caffeine to paraxanthine, theobromine, and theophylline was  $79.6 \pm 21.0$ ,  $10.8 \pm 2.4$ , and  $3.7 \pm 1.3\%$ , respectively (Lelo et al. 1986b). Another study found that paraxanthine accounted for  $63 \pm 13\%$  of the dimethylxanthines in plasma, theobromine  $27 \pm 15\%$ , and theophylline  $10 \pm 2.6\%$  (Rodopoulos and Norman 1996). For demethylation, 3-demethylation represents 52% of all metabolites, 7-demethylation 35%, and 1-demethylation 13% (Arnaud and Welsch 1980a; Yesair et al. 1984). When demethylation pathways are considered from plasma AUC results, paraxanthine, theobromine, and theophylline accounted for  $83.9 \pm 5.4$ ,  $12.1 \pm 4.1$ , and  $4.0 \pm 1.4\%$ , respectively, of the caffeine demethylations (Lelo et al. 1986b). From clearance values of caffeine and its primary metabolites, it was calculated that approximately 37% of a caffeine dose was biotransformed to paraxanthine (Tang-Liu et al. 1983), a lower value when compared with excreted metabolite. To quantify the total demethylation process, the administration of [1,3,7-Me- $^{13}\text{C}$ ] caffeine to volunteers and continuous collection of expired  $^{13}\text{CO}_2$  showed that 21–26% of the total  $^{13}\text{C}$  administered was recovered in expired  $\text{CO}_2$  over 24 h, corresponding to a mean percentage of demethylation for each methyl group (Arnaud et al. 1980). The quantitative urinary excretion of caffeine metabolites in man and in various animal species, expressed as the percentage of the administered dose, is shown in Table 1. However, large individual variations in urinary metabolite excretion have been reported and the caffeine metabolites recovered, expressed as the percentage of the dose, in young and elderly men showed significantly higher excretion in the elderly for 1,7DMU ( $P < 0.05$ ), 1MU ( $P < 0.03$ ), and 7MU ( $P < 0.03$ ) for both oral and intravenous administration, but lower urine recoveries were observed in young men (Blanchard et al. 1985).

### 3 Theophylline

#### 3.1 Absorption

Theophylline transfer across rat jejunum *in vitro* showed that its clearance was directly proportional to the fraction unionized at various pH values (Perry et al. 1984), and there were small but nonsignificant differences in absorptive capacity between the

**Table 1** Urinary excretion of caffeine, theophylline, theobromine, and paraxanthine metabolites in human and animals

	Caffeine		Theophylline		Theobromine		Paraxanthine	
	Human	Animals	Human	Animals	Human	Animals	Human	Animals
Caffeine	1.2	2 <sup>a</sup> , 3 <sup>b</sup> , 0.9 <sup>c</sup> , 4.5 <sup>d</sup>						
Theophylline	1	0.7 <sup>a</sup> , 6 <sup>b</sup> , 1.6 <sup>c</sup> , 12.5 <sup>*,d</sup>	16	38 <sup>b</sup>				
Theobromine	2	4 <sup>a</sup> , 8 <sup>b</sup> , 4 <sup>c</sup> , 7.5 <sup>d</sup>	20		26 <sup>a</sup> , 53 <sup>b</sup> , 19.5 <sup>c</sup> , 50 <sup>a</sup>			
Paraxanthine	6.5	14.5 <sup>a</sup> , 12.5 <sup>b</sup> , 14.5 <sup>c,d</sup>					11	52 <sup>b</sup>
Trimethyluric acid	1.4	4 <sup>a</sup> , 8 <sup>b</sup> , 2 <sup>c</sup> , 4 <sup>d</sup>						
Trimethylallantoin		0.4 <sup>a</sup> , 7 <sup>b</sup>						
5-Acetylamino- 6-formylamino- 3-methyluracil	16	—					20	
5-Acetylamino-6-amino- 3-methyluracil	—	5 <sup>c</sup>						
6-Amino-5-[N- formylmethylamino]- 1,3-dimethyluracil	1.2	9.5 <sup>a</sup> , 21 <sup>b</sup>						
1,7-Dimethyluric acid	6	6.3 <sup>a</sup> , 5 <sup>b</sup> , 4 <sup>c</sup> , 3 <sup>d</sup>					17	7 <sup>b</sup>
6-Amino-5[N- formylmethylamino]- 3-methyluracil	2.5	1.4 <sup>a</sup> , 2.5 <sup>b</sup>					5	7 <sup>b</sup>
1,3-Dimethyluric acid	2.6	7.5 <sup>a</sup> , 4 <sup>b</sup> , 2.5 <sup>c</sup> , 20 <sup>d</sup>	47	38 <sup>b</sup>				
6-Amino-5[N- formylmethylamino]- 1-methyluracil	2	5 <sup>a</sup> , 6 <sup>b</sup>			12	44 <sup>a</sup> , 29 <sup>b</sup> , 14 <sup>c</sup> , 10 <sup>d</sup>		
3,7-Dimethyluric acid	0.8	1 <sup>a</sup> , 1 <sup>b</sup>	1			4 <sup>a</sup> , 4.5 <sup>b</sup> , 2 <sup>c</sup> , 0.5 <sup>d</sup>		
Dimethylallantoin		T <sup>b</sup>				T <sup>b</sup>		
1-Methylxanthine	19		1	<1 <sup>b</sup>			17	11 <sup>b</sup>

7-Methylxanthine	7.5	6.3 <sup>a</sup> , 5 <sup>b</sup> , 22 <sup>c</sup> , 1.5 <sup>d</sup>	36	12.5 <sup>b</sup> , 4 <sup>b</sup> 49 <sup>c</sup> -4.5 <sup>d</sup>	5.5	2 <sup>b</sup>
3-Methylxanthine	3	2 <sup>a</sup> , 1 <sup>b</sup> , 4 <sup>c</sup> , 31 <sup>d</sup>	14	3.5 <sup>b</sup>	19	21 <sup>b</sup>
1-Methyluric acid	26.5	8.5 <sup>a</sup> , 6.5 <sup>b</sup> , 19 <sup>c</sup> , 12.5 <sup>d</sup>	20	20 <sup>b</sup>	4.5	T <sup>b</sup>
7-Methyluric acid	-	1 <sup>a</sup> , 0.8 <sup>b</sup> , 3.5 <sup>d</sup>	1	<1 <sup>b</sup>	8.5	9-25 <sup>a</sup>
3-Methyluric acid	0.1	2 <sup>a</sup> , 0.3 <sup>b</sup> , 4.5 <sup>c</sup>	1		0.6	
3-β-D-Paraxanthine glucuronide	-	20 <sup>a</sup> , 0 <sup>b</sup>				
α-[7-(1,3- Dimethylxanthinyl)] methyl methyl sulfoxide	-	T <sup>a-c</sup>				
α-[7-(1,3- Dimethylxanthinyl)] methyl methyl sulfide	-	T <sup>a</sup>				
α-[7-(1,3- Dimethylxanthinyl)] methyl methyl sulfone	-	T <sup>a</sup>				T <sup>b</sup>
N-Methylurea	-	T <sup>b</sup>				
N,N'-Dimethylurea	-	T <sup>b</sup>				

The results are expressed as the percentage of metabolites excreted in urine (at 48 h in human). From Arnaud and Welsch (1980a, b), Callahan et al. (1982), Shively and Tarka (1983), Tarka et al. (1983), Miller et al. (1984), Arnaud (1984, 1985), Beach et al. (1985), Birket et al. (1985), Arnaud et al. (1989), Rodopoulos et al. (1996), and Rodopoulos and Norman (1997).

T traces

<sup>a</sup>Mouse urine (24-36 h)

<sup>b</sup>Rat urine (24 h)

<sup>c</sup>Rabbit urine (36 h)

<sup>d</sup>Dog urine (48 h)

\*Value of theophylline + paraxanthine



intestinal segments studied (Murray et al. 1993). The rhythmicity in plasma levels found in theophylline disposition was not due to diurnal variation in the passive transport of the mucosa, but may be caused by differences in food intake between morning and evening, in the transit time or gastric emptying, or in the amount or composition of the gastric or intestinal fluid (Tukker and Meulendijk 1991). Exsorption of theophylline from blood to the gastrointestinal tract corresponded to 12–15%, while the extent of the drug excreted into the bile varied from 0.17 to 0.30% (Arimori and Nakano 1988). Rectal absorption of theophylline is slow but complete.

Also in humans, theophylline is rapidly and completely absorbed (Ogilvie 1978; Yesair et al. 1984). It appeared to be almost completely absorbed before it reached the jejunum and the jejunal concentrations were lower than 10% of the maximal duodenal concentrations (Brouwers et al. 2005). The absolute bioavailability of theophylline was investigated by comparing the AUC after intravenous and oral administration of theophylline. The fraction of the dose absorbed averaged  $0.99 \pm 0.02$ , thus showing a bioavailability close to 100% (Hendeles et al. 1977) as well as in neonates and young infants (Moore et al. 1989). Food decreased significantly the absorption rate of theophylline, prolonged  $T_{\max}$ , and decreased  $C_{\max}$ , but the AUC was slightly but not significantly smaller, indicating that theophylline bioavailability was not modified (Jonkman et al. 1985). Oral administration of activated charcoal is a well-established therapy for treatment of theophylline intoxication (Cooling 1993).

### 3.2 *Distribution*

Plasma theophylline concentrations in guinea pigs could be quantitatively described by a two-compartment model with nonlinear elimination kinetics and individual volume distribution of theophylline at each dose (Sato et al. 2007). Protein binding in blood was  $48.8 \pm 6.2\%$  in the rats (Ingvast-Larsson et al. 1992). The theophylline dose required to achieve the narrow therapeutic concentrations (10–20 mg/mL) varies among subjects, largely because of differences in metabolism. Another important pharmacokinetic parameter is protein binding. Theophylline binds mainly to albumin and the protein binding of approximately 50% was shown to be nonlinear (Fleetham et al. 1981; Trnavská 1990), with little variations in healthy subjects but important changes for physiological (Shaw et al. 1982) and disease (Lesko et al. 1981; Siegel et al. 1990; Korrapati et al. 1995) states. Measured saliva levels allow predictions of the unbound serum theophylline levels. The therapeutic range for saliva, which corresponds to the accepted total serum concentration range of 10–20  $\mu\text{g/mL}$ , is approximately 5.55–11.3  $\mu\text{g/mL}$  (Blanchard et al. 1992).

Recovery of theophylline is much less than that of caffeine in brain, liver, muscle, and adipose tissue (Ståhle 1991), reflecting the lower lipid solubility of theophylline. Lower theophylline concentrations were found in the brain (91  $\mu\text{M}$ ) than in other tissues (120  $\mu\text{M}$ ) and this rate of penetration into the brain extracellular

space was higher for caffeine than for theophylline (Stähle et al. 1991). With microdialysis methods applied in anesthetized rats, striatum-to-blood ratios at the steady state of approximately 0.5 were shown (Sjöberg et al. 1992). These results were confirmed and compared with fetal brain AUC values of theophylline; those found in the brains of adults were lower compared with those found in plasma after a dose of 25 mg/kg, suggesting that theophylline might be selectively excluded from the adult brain (Wilkinson and Pollard 1993). Theophylline crosses the placenta and distributes in the organs of the rat fetus and the pregnant animal, except for the brain, where the exposure of the fetal brain was twice that of the adult brain (Arnaud et al. 1982a). Placental clearance of theophylline averaged 0.62 mL/min in the rabbit but it was difficult to extrapolate these results as human and rabbit placentas are structurally dissimilar (Omarini et al. 1991).

### 3.3 Excretion

There were no significant differences in the elimination and metabolism of [8-<sup>14</sup>C] theophylline when given orally or intravenously to rats. Fecal excretion amounted to  $5 \pm 3\%$  of the dose after 7 h and increased to  $18 \pm 2\%$  after 24 h. About 25% of the dose was secreted in the bowels (Arnaud and Welsch 1980b; Arnaud et al. 1982a, b). After 1 day,  $70 \pm 7\%$  of the dose was excreted in urine and  $6 \pm 1\%$  in CO<sub>2</sub> and less than 1% remained in the body (Arnaud and Welsch 1980b). In pregnant rats, labeled caffeine was found in the fetus and only traces were detected in the urine. At the 18th day of pregnancy, unchanged theophylline corresponded to  $70 \pm 10\%$  of urine activity, suggesting impaired metabolism when compared with  $35 \pm 3\%$  in nonpregnant rats (Arnaud and Bracco 1981). Theophylline in humans is completely absorbed and fecal excretion has not been reported.

In premature infants with postconception ages of 28–42 weeks, the urinary percentages of unchanged theophylline decreased from 61 to 43%, respectively, suggesting an increased theophylline metabolism into 1,3DMU with age (Tserng et al. 1983). In 10–12-year-old asthmatic children, the percentage of unchanged theophylline excreted in the urine was  $11.6 \pm 1.75\%$ . Metabolites found in urine in addition to theophylline were 3MX, 1,3DMU, and 1MU (Monks et al. 1979; Wijnands et al. 1990). Theophylline was shown to be extensively reabsorbed in the renal tubule and its renal clearance was highly urine flow dependent and urinary excretion varied with urine output (Tang-Liu et al. 1983).

### 3.4 Pharmacokinetics

In rats, the plasma concentration decayed according to a first-order process with an apparent half-life of about 4 h, but after 4–8 h the slope of the curves declined, resulting in elimination half-lives of about 70 min, a value similar that for lower

doses. The AUC increased disproportionately with dose, indicating capacity-limited elimination, but there was no capacity-limited elimination of 1,3DMU and 1MU with the dose. These results showed that linear pharmacokinetics of theophylline in rats can be applied only to doses not exceeding 10 mg/kg (Teunissen et al. 1985). The plasma theophylline concentrations in rat declined in a monoexponential manner, while those of 1MU and 1,3DMU declined in a biexponential manner upon their injection. The total body clearances of the metabolites were fourfold to sixfold larger and their distribution volumes were 40–50% smaller than that of theophylline (Kuh and Shim 1994). The pharmacokinetics of theophylline were investigated in Cyp1(+/+) wild-type mice, Cyp1a1(-/-) and Cyp1a2(-/-) knockout mice, and humanized hCYP1A1\_1A2 mice lacking either the mouse Cyp1a1 or the mouse Cyp1a2 gene. The half-life of elimination from plasma was more than 4 times longer in Cyp1a2(-/-) mice than in Cyp1(+/+) mice. In humanized hCYP1A1\_1A2 mice lacking the mouse Cyp1a2 gene, the half-life of elimination from plasma was 2–3 times longer than that in Cyp1(+/+) mice (Derkenne et al. 2005). A pharmacokinetics study conducted in dogs showed the bioequivalence of the two injectable forms containing theophylline and aminophylline (ethylenediamine salt of theophylline) and thus the lack of influence of ethylenediamine on the pharmacokinetics of theophylline (Kawai et al. 2000).

The demethylation of theophylline at high concentrations shows biphasic kinetics in the production of individual metabolites with human microsomes (Campbell et al. 1987a). In children with chronic asthma receiving two dosage levels of theophylline, the steady-state serum concentrations increased to a greater degree than predicted with a significantly lower clearance at the higher dose ( $P < 0.02$ ). These results showed the nonlinear nature of the relationship between dose and theophylline serum concentration in these children with asthma (Weinberger and Ginchansky 1977). Theophylline elimination from blood in a 10-month-old female acutely intoxicated had a half-life of 10.0 h, an elimination that was anomalously long for a child of this age (Jarboe et al. 1986). Theophylline pharmacokinetics in asthmatic patients of 8–18 years of age showed that  $C_{\max}$  of  $8.4 \pm 1.7$  mg/L occurred 2.2 h after oral ingestion, with a mean serum half-time for theophylline of  $5.8 \pm 1.7$  h (Becker et al. 1984). The volume of distribution of theophylline depends primarily on age; it is twofold greater in newborns than in adults (Tröger and Meyer 1995). These results were confirmed in healthy male volunteers with total plasma clearances of theophylline of 0.93 mL/min/kg, unbound plasma clearances of 1.61 mL/min/kg, half-lives of 6.2 h, a volume of distribution at steady state of 0.44 L/kg, and an unbound volume of distribution of 0.77 L/kg (Lelo et al. 1986a). Studies have established relationships between renal clearance and urine flow rate for caffeine and theophylline (Trang et al. 1985). Theophylline frequently exhibits nonlinear pharmacokinetics with a relatively large inpatient variability in clearance over time (Pan et al. 2000). A study confirmed the intrasubject variability reported for theophylline clearance in healthy male volunteers but no significant dose dependency was observed for doses of 1 and 6 mg/kg (Fleetham et al. 1981).

### 3.5 Metabolism

Several CYP isoenzymes, including CYP1A2, CYP2E1, and CYP3A4, are involved in the hepatic metabolism of theophylline (Pan et al. 2000). Theophylline has (as caffeine) been used as a marker of CYP1A2 activity (Obase et al. 2003). In human adults, approximately 90% of theophylline is metabolized in the liver by CYPs, while unchanged theophylline is excreted via the kidneys (Tröger and Meyer 1995). CYP1A is responsible for theophylline N-demethylation to 3MX and 1MX (Sarkar and Jackson 1994). The positive relationship between clearance of 1MU and of 3MX in both smokers and nonsmokers ( $P < 0.001$ ) suggests that the two N-demethylation pathways for theophylline metabolism are under common regulatory control and involve a CYP distinct from that mediating 8-hydroxylation of theophylline to 1,3DMU (Grygiel and Birkett 1981). Theophylline is metabolized by 8-hydroxylation to 1,3DMU, which accounts for about half of the clearance of the drug in humans (Ogilvie 1978), and by N-demethylation to 3MX and 1MX. Although theophylline 8-hydroxylation is catalyzed by several CYP subfamilies (Zhang and Kaminsky 1995; Gu et al. 1992; Sarkar et al. 1992), CYP1A2 is reported to play a major role only at lower substrate concentrations (Zhang and Kaminsky 1995). A 30-fold individual difference was observed for the 1MU plus 3MX to theophylline ratio in patients receiving theophylline therapy, and in healthy volunteers a 70-fold difference was found for the 1MX plus 1MU plus AFMU to 1,7DMU ratio. The CYP1A2 activities were not significantly influenced by CYP1A2\*1C or CYP1A2\*1F polymorphism, suggesting that these CYP genotypes are not major factors for the variability of CYP1A2 activity. The CYP1A2\*1K haplotype seems to show a very low frequency in this Japanese population (Takata et al. 2006). Pretreatment in rats with an inhibitor or inducer of CYPs such as troleandomycin, 3-methylcholanthrene, orphenadrine or dexamethasone suggested that theophylline was metabolized via CYP1A1/CYP1A2, CYP2B1/CYP2B2, and CYP3A1/CYP3A2, and that 1,3DMU is primarily formed via CYP1A1/CYP1A2, and possibly CYP3A1/CYP3A2 (Yang et al. 2008).

Theophylline was metabolized in cultured hepatocytes and in liver slices of young and adult rats into 1MU, 1MX, 1,3DMU and/or 3MX, caffeine, a uracil derivative, and two unknown polar compounds. Although the same metabolites were identified in young and adult rats, the development pattern was not uniform and formation of caffeine from theophylline was not dependent on a lack of activity of other pathways. Preincubation with caffeine or theobromine inhibited theophylline metabolism (Gorodischer et al. 1986b). In human liver microsomes, the formation of 3MX, 1MX, and 1,3DMU from theophylline was reported and the two demethylation pathways seemed to be performed by the same enzyme (Robson et al. 1988). In addition,  $\alpha$ -naphthoflavone inhibited theophylline demethylations, whereas 8-hydroxylations were generally less inhibited (Campbell et al. 1987a). In microsomes prepared from different human livers, the formation of 3MX and 1MX correlated best with amounts of the immunoreactive protein HLd (P-IA2) ( $P < 0.05$ ), whereas formation of 1,3DMU correlated with the microsomal content

of HLp (P-IIIa3) and HLj (P-IIe1). In immunoinhibition experiments, incubations conducted with a polyclonal antirat P-c/d antibody, the formation of all three theophylline metabolites was significantly inhibited ( $P < 0.05$ ). However, addition of isoform-specific antirat-CYP-d antibodies to the microsomal mixture significantly and selectively inhibited 1-N-demethylation, with little inhibition of 3-N-demethylation or 8-hydroxylation (Sarkar et al. 1992). 1MX seemed to be mediated by CYP1A1/CYP1A2 and 3MX specifically by CYP1A2 (Sarkar and Jackson 1994). CYPs expressed in human B-lymphoblastoid cell lines showed that at high theophylline concentration (10 mM) four CYPs (CYP1A1, CYP1A2, CYP2D6, CYP2E1) catalyzed the metabolism of theophylline, but the highest affinity was for the CYP1A subfamily. CYP2E1, responsible for a relatively high intrinsic clearance by 8-hydroxylation, may be the low-affinity high-capacity isoform involved in theophylline metabolism. The affinity of theophylline for CYP1A1 was comparable with that of its homologue CYP1A2 and when induced, the participation of CYP1A1 in theophylline metabolism may be important. CYP2D6 played only a minor role and CYP3A4 was not active in the *in vitro* metabolism of theophylline. These results confirm the major role of CYP1A2 in theophylline metabolism and explain why the elimination kinetics of theophylline *in vivo* are nonlinear (Ha et al. 1995). In microsomes, at low theophylline concentrations the metabolism of theophylline to 1,3DMU was catalyzed primarily by CYP1A2, while at high substrate concentrations CYP2E1 was primarily responsible for 1,3DMU formation. At theophylline concentrations achieved *in vivo*, its metabolism must thus be catalyzed primarily by CYP1A2 (Zhang and Kaminsky 1995). In human, rabbit, and rat liver microsomes 1,3DMU accounted, respectively, for 59, 77, and 94% of the total metabolites formed. In both human and rabbit liver microsomes the N-demethylation of theophylline to 1MX accounted for 20% of the total metabolites formed. In human microsomes N-demethylation of theophylline to 3MX accounted for 21% of theophylline metabolism, but it was a minor pathway in rabbit and rat microsomes (McManus et al. 1988).

### ***3.6 Sources of Variation in Theophylline Pharmacokinetics and Metabolism***

*Age.* Differences in the half-life and total clearance were found among the age groups with a linear correlation between age and the clearance of theophylline (Kearns et al. 1986). The average mean residence time of theophylline was significantly longer in 20-month-old rats than in 2- and 14-month-old rats. A greater elimination rate constant was observed in 14-month-old rats and the apparent volume of distribution decreased from 0.71 to 0.57 L/kg in the 2- and 20-month-old rats, respectively (Jung and Nanavaty 1990). The ability of the rat fetus to methylate theophylline into caffeine was demonstrated when [8-<sup>14</sup>C] theophylline was administered to pregnant rats (Arnaud et al. 1982a, b). However, the biotransformation of theophylline to caffeine reported for human neonates and the rat fetus was not observed in neonatal piglets (Kearns et al. 1986).

In premature neonates, weighing less than 1,500 g at birth, and under 32 weeks of gestational age, theophylline clearance was lower (12 mL/h/kg) and the volume of distribution (0.8–0.9 L/kg) was higher than previously reported for less premature neonates, term babies, and older children (Lee et al. 1996). The weight-normalized value of the volume of distribution in premature neonates during the first week of life was 0.63 L/kg (du Preez et al. 1999). In contrast to older children and adults, in whom theophylline disposition follows zero-order kinetics at high concentrations, a monoexponential function best described theophylline elimination in the premature newborn, with half-lives ranging from 24.7 to 36.5 h and estimated clearance ranging from 0.02 to 0.05 L/kg/h (Lowry et al. 2001). In premature neonates, only unchanged theophylline and caffeine were found in urine, indicating the absence of oxidative pathways for theophylline metabolism. In both adults and children, there was high positive correlation between urinary excretion of 3MX and 1MU. Both 3MX and 1MU correlated negatively with urinary excretion of 1,3DMU (Grygiel and Birkett 1980). The metabolism of theophylline in premature infants showed that the urinary percentages of unchanged theophylline decreased from 61% at a post-conception age of 28–32 weeks to 43% at 38–42 weeks. This increased metabolism of theophylline is explained by the production of 1,3DMU (20–34%). It was hypothesized that methylation of theophylline to caffeine is equally active in adults and premature infants and the absence of caffeine in adults is due to the maturing caffeine-metabolizing enzymes (Tserng et al. 1983).

Postnatal age was the most powerful predictor for theophylline half-life in the neonatal period, while gestational age, duration of treatment, and weight did not correlate significantly with any pharmacokinetic parameters (Dothey et al. 1989). Theophylline clearance reached adult values at 55 weeks of postconceptional age and the disappearance of serum caffeine concentrations and the maturation of theophylline clearance were primarily related ( $P < 0.001$ ) to development of the demethylation pathway to 3MX. Postconceptional age was the major factor ( $P < 0.001$ ) explaining the interpatient variability in theophylline clearance (Kraus et al. 1993). The total clearance of theophylline was 87–100 mL/h/kg in children and 57 mL/h/kg in adults, with important interindividual differences in the biological half-life (1.42–7.85 h) and a higher elimination rate constant ( $0.49 \pm 0.30$ /h) in the children (Ellis et al. 1976; Gardner and Jusko 1982; Kolski et al. 1987; Berdel et al. 1987). There was a linear decrease in clearance with increasing age (1.3–30.0 years) regardless of the sex (Gardner and Jusko 1982).

In healthy volunteers and patients with asthma, 20–87-years old and receiving theophylline, although clearance did not fall with increasing age during younger adult life, there was a fall during late adult life, becoming apparent in the seventh, eighth, and ninth decades of age with a reduction in the basal rate of theophylline metabolism (Crowley et al. 1988; Jackson et al. 1989) and plasma clearance of theophylline was 30% lower in elderly male subjects than in young male subjects (Loi et al. 1997). A considerably higher interindividual variability in the disposition of theophylline was observed in frail elderly women (Groen et al. 1993).

*Gender and hormones.* There were statistically significant differences ( $P < 0.01$ ) in the theophylline kinetic parameters, such as the elimination half-life,  $8.70 \pm 0.60$  h

during proestrus,  $4.61 \pm 0.16$  h during estrus, and  $5.01 \pm 0.85$  h during diestrus, and the AUC were  $214.61 \pm 3.58$ ,  $128.64 \pm 9.64$ , and  $165.57 \pm 23.86$   $\mu\text{g h/mL}$ , respectively (Bruguerolle 1987). In pregnant rats, theophylline was eliminated at a slower rate than in both lactating rats and virgin controls, resulting in a longer half-life and lower clearance, while the volumes of distribution in pregnant, lactating, and control rats were not different (Brandstetter et al. 1986) and the impaired theophylline metabolism in late pregnancy exhibited increased excretion of unchanged theophylline with decreased formation ( $-68\%$ ) of 1,3DMU and ( $-30\%$ ) of 1MU (Arnaud et al. 1982a, b).

The elimination of theophylline does not differ between men and women (Jusko et al. 1979; Powell et al. 1977). Several other studies looking at the effect of gender on theophylline clearance in children (8 years) reported that gender had no effect on theophylline clearance (Ellis et al. 1976; Yano et al. 1993), but male children aged 4–20 years were shown to have significantly higher theophylline clearances (31 and 22%, respectively) than female children in other studies (Gardner and Jusko 1982; Driscoll et al. 1989; Igarashi and Iwakawa 2009). In healthy men and premenopausal women, statistically significant gender-related effects were seen for the theophylline half-life and clearance (Jennings et al. 1993). The disposition of theophylline throughout pregnancy and in the postpartum period showed that theophylline clearance was slightly affected during the first two trimesters ( $2.61$ – $2.85$  L/h), while a statistically significant reduction was observed late in pregnancy ( $2.05$  L/h). The postpartum clearance values suggest an ongoing suppression relative to prepregnancy levels. A significant higher half-life of  $13.00 \pm 2.31$  h was observed during the third trimester when compared to  $9.53 \pm 3.53$  h in the postpartum period. The absolute volume of distribution increased with gestation (Gardner et al. 1987). Chronic oral contraceptive users exhibited significantly lower total plasma theophylline clearance ( $-30\%$ ) and the half-life was also significantly prolonged from 7.3 to 9.8 h, while the volume of distribution was unchanged (Tornatore et al. 1982; Teichmann 1990). In contrast, acute oral contraceptive exposure failed to induce significant changes (Gardner et al. 1983).

*Physical exercise.* The volumes of theophylline distribution decreased significantly after exercise in the heat, apparently due to dehydration (Schlaeffer et al. 1984; Lenz et al. 2004).

*Obesity.* Age was the most important determinant of theophylline clearance in pediatric patients and weight had less effect than age and did not statistically improve the model ( $P > 0.005$ ) when combined with age (Driscoll et al. 1989). In obese and normal subjects, the apparent volume of distribution measured from the total body weight (TBW) or the ideal body weight (IBW) averaged  $0.482$  L/kg TBW in normal subjects and  $0.382$  L/kg TBW in obese subjects and  $0.77$  L/kg IBW in obese subjects. Clearance averaged  $63.0$  mL/h/kg IBW in normal subjects and  $32.8$  mL/h/kg TBW and  $64.1$  mL/h/kg IBW in obese subjects. The mean half-lives were longer in obese subjects than in normal subjects,  $8.6 \pm 2.0$  and  $6.0 \pm 2.1$  h, respectively (Gal et al. 1978).

*Drugs.* As expected from the metabolism described already, theophylline pharmacokinetics can be influenced by drugs, herbal supplements, and diet. The extensive



literature on the interactions of drugs with theophylline will not be described here. A review on the interaction of drugs has been published describing an increase or a decrease of theophylline clearance (Upton 1991). One aspect that needs mentioning is that theophylline is demethylated to 1MX and 1MU was produced from a rapid xanthine oxidase mediated 8-oxidation, while no 1MU was formed by 3-demethylation of 1,3DMU (Birkett et al. 1983).

Cigarette smoking appeared to induce theophylline metabolism as reflected by the mean theophylline half-life in smokers (5.4 h) versus nonsmokers (8.3 h) (Jusko 1979; Jusko et al. 1979; Powell et al. 1977). Cigarette smoking significantly altered the theophylline clearance processes (Schrenk 1998; Teichmann 1990; Jennings et al. 1993; Zevin and Benowitz 1999) and a 40% elevation in theophylline clearance was observed in women who smoked (Gardner et al. 1983).

Daily caffeine intake significantly altered the theophylline clearance processes (Gardner et al. 1983). On caffeine administration, the theophylline steady-state concentration and AUC increased by 23 and 40%, respectively, and the reduction in the apparent total body clearance and elimination rate constant of theophylline reached 29 and 31%, respectively, indicating a pronounced influence on theophylline of concomitant ingestion of caffeine in normal consumers (Jonkman et al. 1991). Abstention from methylxanthine-containing foods and beverages led to a significant decrease in the elimination half-life ( $P < 0.02$ ) owing to increases in the elimination constants for theophylline, 3MX, and 1,3DMU (Monks et al. 1979). However, in contrast to the effect of deprivation of dietary methylxanthines, the addition of extra methylxanthines from six bottles per day of a cola beverage to the diet did not influence the disposition of theophylline (Monks et al. 1981).

*Diseases.* As well as for caffeine, diseases that compromise liver function, especially cirrhosis, reduce theophylline clearance in animals and man (Park et al. 1999; Nam et al. 1997; Amodio et al. 1991). There is also a small effect in diabetes mellitus rats induced by alloxan or streptozotocin (Kim et al. 2005). In patients with insulin-dependent diabetes mellitus and in sex-, age-, and weight-matched healthy nonsmokers, the pharmacokinetic parameters of theophylline, plasma clearance, elimination half-life, and volume of distribution were similar, but there was a positive correlation between hemoglobin A1c values and plasma theophylline clearance ( $P < 0.05$ ), formation clearance of 1,3DMU ( $P < 0.05$ ), and formation clearance of 1MU ( $P < 0.05$ ) (Korrapati et al. 1995). Among animal and human studies, renal disease, Down syndrome, psoriasis, endotoxin-induced fever, acidosis, hypoxia, hyperlipidemia, and hypoalbuminemia were shown to alter the pharmacokinetics of theophylline.

### 3.7 Metabolites and Metabolic Pathway

The methylation of theophylline to caffeine was shown in rat (Gorodischer et al. 1986b) and in rat fetus (Arnaud et al. 1982a, b), in premature infants (Boutroy et al. 1979; Bory et al. 1979), and in vitro in the human fetal liver (Aranda et al. 1979).



In premature infants, plasma concentrations of caffeine increased from 1.8 mg/L at day 1 to 3.7 mg/L 7 days after initiation of theophylline therapy. Labeled caffeine, paraxanthine, and theobromine were found in plasma and urine of preterm newborns receiving [1,3-<sup>15</sup>N],[2-<sup>13</sup>C]theophylline for the treatment of primitive apneas, showing that theophylline was converted to caffeine by N7-methylation (Brazier et al. 1980b). Several studies confirmed this methylation pathway in the newborn (Soyka et al. 1981; Simons et al. 1981). This pathway was believed to be specific to the neonatal period, explained by the immaturity of liver enzymes. However, caffeine (0.21–0.75 mg/L) and its major metabolite, paraxanthine, were observed in plasma following oral administration of theophylline (8.1–21.5 mg/L) in a multiple-dose study in healthy subjects. In adult subjects, about 6% of the theophylline dose was converted to caffeine (Tang-Liu and Riegelman 1981; Arnaud 1984). Only 7–19% of theophylline is excreted unchanged in the urine with other metabolites, including 1,3DMU (35–55%), 1MU (13–26%), 3MX (9–18%), 1MX (0.3–4%), and 3MU (1%) (Arnaud 1984; Birkett et al. 1985; Anonymous 1991). The N3-demethylation of theophylline accounted for  $34 \pm 6\%$  of the urinary metabolites, N1-demethylation of theophylline for  $15 \pm 3\%$ , and C8-oxidation of theophylline for  $51 \pm 9\%$ . The C8-oxidation of 1MX and 3MX corresponded to  $93 \pm 4$  and  $9 \pm 4\%$ , respectively, of the excreted monomethylxanthine and urate. In addition to theophylline, 1,3DMU and 1MU were consistently found in plasma and saliva. Theophylline accounted for  $91 \pm 4\%$  of the total plasma AUC, with 1,3DMU accounting for  $3.1 \pm 1.4\%$ , 3MX for  $3.4 \pm 1.8\%$ , and 1MU for  $2.5 \pm 1.5\%$  (Rodopoulos and Norman 1997). Urinary excretions of 1,3DMU and 1MU exhibited the highest correlations, while the poorest correlations were observed for 1MX compared with those of 1MU and 1,3DMU, suggesting that 1MU did not derive solely from 1MX and implicating 1,3DMU as an alternative precursor (Bayar and Ozer 1997). However, previous results on oral administration of 1,3DMU in healthy male volunteers showed that 1,3DMU was recovered unchanged in urine and was not demethylated to 1MU (Birkett et al. 1983). The quantitative urinary excretion of theophylline metabolites in man and in various animal species, expressed as the percentage of the administered dose, is shown in Table 1.

## 4 Theobromine

When compared with caffeine and theophylline, fewer studies have been performed on theobromine.

### 4.1 Absorption

In rats there is complete absorption of theobromine, with only 1% of the dose excreted in feces as unchanged theobromine (Arnaud and Welsch 1979a; Bonati et al. 1984) and 94–106% was recovered in urine (Shively and Tarka 1983).

A marked decrease of the absorption rate constant was observed with increased dose, but the absolute bioavailability of theobromine remained 100%. As a consequence, the peak blood level tends to appear later with larger doses (Bonati et al. 1984). Theobromine bioavailability after an oral administration in healthy, non-medicated, nonsmoking men and after 14 days' abstention from all methylxanthine sources was  $0.96 \pm 0.02$  (Tarka et al. 1983; Miners et al. 1982; Yesair et al. 1984). Both the rate and the extent of absorption of theobromine in chocolate were less than those of theobromine in solution and the relative bioavailability of theobromine in chocolate was 80%, suggesting food interaction with chocolate ingredients (Shively et al. 1985). Theobromine absorption after oral administration of capsules and chocolate candy was compared in volunteers who abstained from methylxanthines. A theobromine plasma  $C_{\max}$  of 6.72  $\mu\text{g/mL}$  was measured 3 h after ingestion of a capsule containing 370 mg theobromine and absorption of the same dose from chocolate was more rapid and produced a higher  $C_{\max}$  of 8.05  $\mu\text{g/mL}$  after 2 h (Mumford et al. 1996).

## 4.2 Distribution

One day after the oral administration of [7-Me- $^{14}\text{C}$ ]theobromine to rats, no organ accumulation of theobromine and metabolites could be seen by whole-animal autoradiography and the most labeled organ was the liver, with 0.4% of the administered radioactivity and 2% was present in the cecum and the colon coming from intestinal and bile secretion (Arnaud and Welsch 1979a). Theobromine was shown to cross the placenta in the pregnant rat (Arnaud and Gétaz 1983).

In rats, blood samples taken at various intervals, from 0.5 to 3 h, showed that the mean value of the unbound theobromine fraction was 0.88 (Bonati et al. 1984). In plasma of pregnant and nonpregnant rats, theobromine corresponds to about 99% and metabolites to less than 1% (Shively and Tarka 1983).

When [8- $^{14}\text{C}$ ]theobromine was administered to newborn rats and on the following days, it was shown that the brain/blood theobromine concentrations ratio decreased continuously from  $0.96 \pm 0.02$  at birth to  $0.60 \pm 0.02$  in 30-day-old rats, while the liver/blood ratio remained constant at  $1.18 \pm 0.05$ . These results have been interpreted as a postnatal blood–brain barrier for theobromine in the rat (Arnaud and Gétaz 1982). The theobromine concentration was shown to be in equilibrium between blood, brain, and liver of the fetus and blood of the pregnant rat (Arnaud and Gétaz 1983). In man, theobromine is distributed throughout the total body water (Yesair et al. 1984). Milk chocolate containing theobromine (240 mg) was ingested by nursing mothers and peak theobromine concentrations of 3.7–8.2 mg/L were found in all fluids, including plasma, saliva, and breast milk, at 2–3 h after ingestion (Resman et al. 1977). In *in vitro* and *in vivo* studies, the fraction of theobromine unbound to plasma proteins averaged 0.90 over a wide range of concentrations (Bonati et al. 1984).

Compared with the fetus, the AUC values of theobromine were lower in the brains of adults compared with plasma, confirming that theobromine might be selectively excluded from the adult brain (Wilkinson and Pollard 1993). Theobromine and caffeine milk-to-serum concentration ratios were twofold higher compared with those of paraxanthine and theophylline (McNamara et al. 1992). The mean concentration ratio of theobromine in nursing mothers was  $0.82 \pm 0.17$  for milk/plasma and if a mother ate a 4-oz chocolate bar every 6 h and the infant nursed when the theobromine concentration in milk was at its peak, the infant could ingest about 10 mg of theobromine per day (Resman et al. 1977).

The theobromine concentrations in plasma and saliva were similar, after a 500-mg oral dose, whereas the saliva concentrations for 7MX and 3MX were found to be  $63 \pm 17\%$  of the plasma concentrations for 7MX and  $74 \pm 13\%$  for 3MX, respectively (Rodopoulos et al. 1996).

### 4.3 Excretion

In rats, urine excretion was the main excretory route and amounted to  $84 \pm 8\%$  of the administered dose (Arnaud and Welsch 1979a; Bonati et al. 1984). Urinary excretion was compared in rats, mice, hamsters, and male rabbits and dogs after oral administration of  $[8-^{14}\text{C}]$ theobromine and about 60–89% of the dose was recovered in urine (Miller et al. 1984).

Theobromine was shown to be extensively reabsorbed in the renal tubule and its renal clearance was highly urine flow dependent and thus urinary excretion varied with urine output (Tang-Liu et al. 1983). After the ingestion of 1 g theobromine, 62% of the dose was recovered in 48-h urine collected in adult subjects and unchanged theobromine, 3MX, 7MX, and 7MU were identified (Cornish and Christman 1957). After a single oral dose of theobromine with a trace amount of  $[8-^{14}\text{C}]$ theobromine had been administered to healthy nonsmoking men, 50% of the radioactivity was recovered in urine after 8–12 h and the entire radioactivity administered was found after 3 days (Tarka et al. 1983). The cumulative urinary excretion of radioactivity from  $[8-^{14}\text{C}]$ theobromine in subjects who maintained 14 days of methylxanthine abstinence was 86.4% (80–96%) and in the same subjects not limited in their methylxanthine consumption the excretion in urine amounted to 81.1% (81–93%) (Shively et al. 1985).

After the administration of  $[7\text{-Me-}^{14}\text{C}]$ theobromine (1–6 mg/kg) to male rats, fecal excretion amounted to  $11 \pm 1\%$  of the administered dose, but only 10% was unchanged theobromine and incubation of theobromine into the gastrointestinal content showed no metabolic transformation, suggesting that the metabolites found were excreted through gastrointestinal secretion (Arnaud and Welsch 1979a). After oral administration of  $[8-^{14}\text{C}]$ theobromine to rats, similar amounts of radioactivity (2.5%) were recovered in feces as after intravenous administration (Bonati et al. 1984). In pregnant rats, the oral administration of  $[8-^{14}\text{C}]$ theobromine showed that fecal excretion amounted to 31% of the ingested dose (Arnaud and Gétaz 1983).

Fecal excretion was compared in rats, mice, hamsters, and male rabbits and dogs after oral administration of [8-<sup>14</sup>C]theobromine and from 2 to 38% of the dose was recovered in feces. In male and female rats,  $38.2 \pm 0.8$  and  $16.2 \pm 1.3\%$  were excreted in feces, respectively, and the values were  $8.8 \pm 1.1$  and  $11.5 \pm 1.8\%$  for mice,  $15.0 \pm 6.0\%$  and  $14.3 \pm 3.3\%$  for hamsters, and  $1.6 \pm 0.2\%$  male rabbits and  $4.5 \pm 0.1\%$  for male dogs (Miller et al. 1984). However, bentonite was often added to the rodent diet as a pellet binder and may explain the higher fecal excretion reported in rats (Arnaud 1983). Fecal elimination of [8-<sup>14</sup>C]theobromine-derived radioactivity after the oral administration of a tracer dose in subjects who maintained or did not maintain 14 days of methylxanthine abstinence was 0.56–0.54 and 0.15–1.42%, respectively (Shively et al. 1985).

#### 4.4 Pharmacokinetics

The half-life of theobromine in rats exhibited large variations from 1.9 to 6.4 h, with an average value similar to that reported for man of  $6.1 \pm 0.7$  h (Drouillard et al. 1978). The kinetics of theobromine in rats after a dose ranging from 1 to 100 mg/kg and chronic intake showed no significant difference in the pharmacokinetic profile except for a reduction in the absorption rate constant as the dose increased. Linear pharmacokinetics was observed up to the dose of 100 mg/kg and the AUC values increased in proportion to the dose (Bonati et al. 1984). Pregnancy in rats on day 19 of gestation did not affect the pharmacokinetics of theobromine (15–100 mg/kg orally) and similar values were obtained in nonpregnant rats. No dose-dependent kinetics was observed in the theobromine plasma half-life, volume of distribution, systemic clearance, dose-normalized AUC, or  $T_{\max}$  (Shively and Tarka 1984). A decrease in the elimination rate constant of theobromine was observed at the highest dose of 50 mg/kg (1–100 mg/kg/day) in rabbits, suggesting saturation (Latini et al. 1984).

The half-life of theobromine in nursing mothers after ingestion of milk chocolate containing 240 mg of theobromine averaged  $7.1 \pm 2.1$  h, body clearance was  $65 \pm 20$  mL/h/kg, and the apparent volume of distribution was  $0.62 \pm 0.13$  L/kg (Resman et al. 1977). A similar mean value of the half-life was reported 1 year later from measurements in man and was  $6.1 \pm 0.7$  h (Drouillard et al. 1978). Theobromine disposition follows first-order kinetics with a one-compartment open model and the mean theobromine half-life was  $9.28 \pm 0.7$  h, plasma clearance was  $0.87 \pm 0.06$  mL/min/kg, the AUC was  $117 \pm 7.9$  mg h/L, and the volume of distribution was  $0.68 \pm 0.03$  L/kg (Tarka et al. 1983). In healthy volunteers the total plasma clearance and renal clearance for theobromine were 46 and 67% greater than those for theophylline, respectively, but most of the difference was due to the lower protein binding of theobromine with the free fraction of 0.86 compared with 0.58 for theophylline. Clearance by 3-methyl demethylation was 3.7-fold higher for theobromine than for theophylline. There were high degrees of correlation between theophylline and theobromine plasma clearances, partial metabolic clearances, and renal clearances (Birkett et al. 1985; Lelo et al. 1986a). Correlation between renal

clearance of theobromine and the urine flow rate was reported (Trang et al. 1985). A supplement of theobromine (6 mg/kg) given to healthy men did not modify significantly theobromine pharmacokinetics and a similar half-life, apparent volume of distribution, and clearance were reported (Shively et al. 1985).

#### 4.5 *Metabolism*

In human liver microsomes, at least two distinct liver enzymes, isozymes of CYP, with differing substrate affinities have the potential to catalyze theobromine N-demethylations and C8-hydroxylations. At the low theobromine concentrations encountered in vivo, the high-affinity site is expected to predominate (Campbell et al. 1987a). The identification of the CYP isoforms responsible for the conversion of theobromine to its primary metabolites was studied in human liver microsomes using various specific inhibitors. Furafylline variably inhibited 7MX formation from theobromine, but had no effect on other pathways. Diethyldithiocarbamate and 4-nitrophenol, probes for CYP2E1, inhibited the formation of 3MX, 7MX, and 3,7-dimethyluric acid (3,7DMU) by approximately 55–60, 35–55, and 85%, respectively. Recombinant CYP1A2 and CYP2E1 enzymes exhibited similar values of the apparent Michaelis–Menten constant ( $K_m$ ) for 7MX formation, and CYP2E1 was further shown to have the capacity to convert theobromine to both 3MX and 3,7DMU (Gates and Miners 1999). The total plasma and partial metabolic and renal clearances of theobromine determined in healthy volunteers supported the view that theobromine was metabolized by a common group of CYPs under similar regulatory control and it was proposed to use theobromine to assess the activity of these enzymes in man (Birkett et al. 1985). It was suggested that 3,7DAU and 3,7DMU are derived from a common oxidized intermediate of theobromine which is the precursor of 3,7DMU, but in the presence of glutathione (GSH) or some other cellular thiol it may be reduced to give 3,7DAU (Lelo et al. 1990). The involvement of GSH and CYPs in the conversion of theobromine to 3,7DAU and 3,7DMU has been demonstrated in rat liver microsomes, showing that the ratio of formation of 3,7DAU to 3,7DMU increased with increasing GSH concentration to a maximum of 12:1 for 2 mM. When compared with untreated animals, 3,7DAU and 3,7DMU formation was increased approximately 12- and 1.6-fold in liver microsomes of rats treated with 3-methylcholanthrene and phenobarbitone, respectively (Lelo et al. 1990).

#### 4.6 *Sources of Variation in Theobromine Pharmacokinetics and Metabolism*

*Gender and hormones.* The kinetic parameters of pregnant and nonpregnant rats were similar at all theobromine dose levels studied (Shively and Tarka 1983). In rabbits there was also no significant difference due to either gender or pregnancy

(Latini et al. 1984). The most important theobromine metabolite excreted by mice was 3,7DAU and male mice converted theobromine to this metabolite more extensively than did female mice. There was significantly more 3,7DMU in female rats than in male rats (Miller et al. 1984).

*Drugs.* Allopurinol had no effect on the clearance of theobromine, suggesting that the elimination of theobromine is not dependent on xanthine oxidase (Miners et al. 1982).

*Smoking.* As expected, theobromine plasma clearance was 33% higher in smokers than in nonsmokers owing to induction of all metabolic pathways, but 7-demethylation was induced to a greater extent than the other pathways (Miners et al. 1985; Gates and Miners 1999).

*Diet.* The mean theobromine half-life, apparent volume of distribution, and clearance were unaffected by abstinence from all methylxanthines or receiving high daily doses of theobromine from chocolate for 1 week (Shively et al. 1985). However, a previous study (Drouillard et al. 1978) suggested that immediately after five daily doses of theobromine, an impairment of theobromine clearance occurred that was reversible by 4 days of dietary abstention from methylxanthines.

#### 4.7 *Metabolites and Metabolic Pathway*

After the administration of [7-Me-<sup>14</sup>C]theobromine (1–6 mg/kg) to male rats, the radioactivity collected in <sup>14</sup>CO<sub>2</sub> corresponding to the formation of 1-methyl and 3-methyl derivatives amounted to 6 ± 1% of the administered dose. In urine unchanged theobromine (49 ± 4% of excreted metabolites), 3,7DAU (36 ± 4%), 7MX (6 ± 1%), 7MU (3.9 ± 0.5%), 3,7DMU (2.7 ± 0.2%) and trace amounts of *N*-methylurea and dimethylallantoin were found (Arnaud and Welsch 1979a). Urinary excretion of unchanged theobromine increased in the pregnant rat from 47 ± 4 to 74 ± 3%, while urinary excretion of 3,7DAU decreased from 35 ± 4% to 22 ± 2%, thus showing that pregnancy impaired theobromine metabolism (Arnaud and Gétaz 1983; Bonati et al. 1984). After an oral dose of 5 and 100 mg/kg theobromine, pregnant and nonpregnant rats showed similar qualitative metabolic patterns and the metabolites identified in the urine were theobromine (39–62%), 3,7DAU (20–32%), 3MX and 7MX (8–15%), 3,7DMU (5–10%), and 7MU (5–7%) (Shively and Tarka 1983). Unchanged theobromine corresponded to about 50% of urinary metabolites in rat and dog, 32.4 ± 2.2% of the administered dose in rat and 36.8 ± 5.9% in dog, about 30% of urinary metabolites for mouse and hamster, 22.1 ± 3.6% of the dose in mouse and 20.0 ± 2.8% in hamster, and less than 20% of urinary metabolites for rabbit, 13.9 ± 2.7% of the dose (Latini et al. 1984). 3,7DAU was the most important metabolite after theobromine and corresponded to 25% of urinary metabolites in rat (16.5 ± 0.9% of the dose), mouse (13.9 ± 2.1% of the dose) and hamster (14.6 ± 1.6% of the dose), while it represented only 10% of urinary metabolite in rabbit (10.0 ± 1.1% of the dose)

and dog ( $7.5 \pm 3.1\%$  of the dose). In rabbit, 7MX ( $35.5 \pm 3.2\%$  of the administered dose) and 3MX ( $8.4 \pm 0.5\%$  of the dose) corresponded to about 50 and 11% of urine metabolites while 3,7DMU ( $1.5 \pm 0.1\%$ ), 7MU ( $1.6 \pm 0.2\%$ ), and 3MU ( $0.6 \pm 0.1\%$ ) were minor metabolites. In hamster, 7MX ( $11.4 \pm 1.0\%$  of the administered dose) corresponded to 20% of urine metabolites and 3MX ( $2.3 \pm 0.2\%$ ), 7MU ( $2.9 \pm 0.4\%$ ), 3MU ( $0.3 \pm 0.1\%$ ), and 3,7DMU ( $2.3 \pm 0.6\%$ ) were minor metabolites. In dog, 3MX ( $19.9 \pm 2.7\%$  of the administered dose) corresponded to 25% of urine metabolites and 7MX ( $3.4 \pm 0.9\%$ ), 7MU ( $4.4 \pm 2.2\%$ ), and 3,7DMU ( $0.4 \pm 0.1\%$ ) were less important. In mouse, 7MX ( $8.2 \pm 0.4\%$  of the administered dose) and 7MU ( $5.3 \pm 0.2\%$  of the dose) both corresponded to less than 10% of urine metabolites and 3MX ( $3.2 \pm 0.2\%$ ), 3MU ( $0.4 \pm 0.1\%$ ), and 3,7DMU ( $2.5 \pm 0.3\%$ ) were minor metabolites. In rat, 3MX ( $3.9 \pm 0.3\%$  of the administered dose), 3,7DMU ( $2.1 \pm 0.2\%$  of the dose), and 7MX ( $2.5 \pm 0.1\%$  of the dose) corresponded to about 5% of urine metabolites and 7MU ( $1.3 \pm 0.1\%$ ) was less important and 3MU could not be quantified. Demethylation of theobromine was greatest in rabbits and lowest in rats and 3-methyl demethylation predominated over 7-methyl demethylation in all species except the rat and the dog. In dog, demethylation of theobromine was most important on 7-methyl, while in rat there was no specific demethylation activity. Oxidation to uric acids was a minor metabolic pathway in all species, with the greatest activity in mice. In addition to these identified metabolites, an unidentified but apparently unique metabolite was detected in dog (Miller et al. 1984).

In healthy, nonmedicated, nonsmoking men after 14 days' abstinence from all methylxanthine sources, the urine metabolites measured were theobromine ( $18.2 \pm 2.1\%$ ), 7MX ( $33.6 \pm 1.6\%$ ), 7MU ( $7 \pm 0.7\%$ ), 3MX ( $19.9 \pm 0.9\%$ ), 3,7DAU ( $5.7 \pm 0.9\%$ ), and 3,7DMU ( $1.0 \pm 0.2\%$ ) (Tarka et al. 1983). After the oral administration of a dose of [8- $^{14}$ C]theobromine, the percentage of urinary metabolites recovered consisted of 42% 7MX, 20% 3MX, 18% theobromine, 10% 7MU, and 10% 3,7DAU (Shively et al. 1985). In urine collected 48 h after administration of a 500-mg theobromine dose, unchanged theobromine accounted for  $21 \pm 4\%$  of total urine excretion and  $36 \pm 5\%$  for 7MX,  $21 \pm 4\%$  for 3MX,  $11 \pm 4\%$  for 3,7DAU,  $10 \pm 2\%$  for 7MU,  $1.3 \pm 0.6\%$  for 3,7DMU and  $0.5 \pm 0.4\%$  for 3MU. The N3-demethylation of theobromine accounted for  $58 \pm 7\%$  of the urinary metabolites, the N7-demethylation of theobromine for  $27 \pm 6\%$ , the C8-oxidation of 7MX for  $22 \pm 4\%$ , the C8-oxidation of 3MX for  $2 \pm 2\%$ , and the formation of 3,7DAU for  $13 \pm 4\%$  (Rodopoulos et al. 1996). The quantitative urinary excretion of theobromine metabolites in man and in various animal species, expressed as the percentage of the administered dose, is shown in Table 1.

## 5 Paraxanthine

Plant biosynthesis leads to the accumulation of caffeine, theobromine, and theophylline, while only trace amounts of paraxanthine were identified as a constituent of *Coffea arabica* (Arnaud and Enslen 1992). The absence of food containing



paraxanthine has limited the number of studies investigating its metabolic fate. As 80% of caffeine ingested by humans is transformed through paraxanthine and the average serum levels of paraxanthine are two thirds those of caffeine, the contribution of paraxanthine to the pharmacological activity of caffeine needs to be considered in understanding the clinical pharmacological activity of caffeine, particularly with chronic, repetitive caffeine consumption (Benowitz et al. 1995).

### 5.1 Absorption and Distribution

It is believed that paraxanthine absorption from the gastrointestinal tract, like the other methylxanthines, was complete after an oral dose (Lelo et al. 1989).

The concentration of radioactivity in the blood and the liver 2 h after oral administration was the same, indicating a complete equilibrium between blood and the tissues except for the brain (Arnaud and Enslin 1992). Twenty-four hours after intravenous administration of [1-Me-<sup>14</sup>C]paraxanthine to rats, there was no accumulation in the body as seen by whole-animal-body autoradiography and the cumulated dose measured in organs reached 0.2% of the dose (Arnaud and Welsch 1979b). Whole-animal-body autoradiography showed a wide distribution throughout the body in liver, heart, muscle, thymus, lungs, and the gastrointestinal tract 0.5 h after [8-<sup>14</sup>C]paraxanthine had been given orally to rats. Higher concentrations were observed in the stomach, the kidney, and the bladder, showing that a fraction of the dose had still not emptied from the stomach and that another fraction had already been excreted in the urine. After 10 h, only traces of radioactivity were detected in the body (Arnaud and Enslin 1992).

*Brain.* The radioactivity from [8-<sup>14</sup>C]paraxanthine 2 h after oral administration was lower in the brain, which is protected by a blood–brain barrier, and the brain-to-blood concentrations ratio was 0.30 for paraxanthine, while this ratio is 1 for caffeine (Arnaud and Enslin 1992). Subcutaneous injection of caffeine into rats resulted in higher concentrations of methylxanthines, particularly paraxanthine, in the striatum than in the rest of the brain and it was observed that the clearance of paraxanthine was faster in serum than in brain structures (Parra et al. 1991). However, an alternative explanation is that paraxanthine binding to abundant A<sub>2A</sub> receptors in the striatum delayed elimination. Compared with the fetus, the AUC value of paraxanthine was found to be lower in the brains of adults compared with plasma after a dose of 25 mg/kg. This suggests that paraxanthine might be partly excluded from the adult brain (Wilkinson and Pollard 1993).

The distribution into milk of paraxanthine measured in lactating New Zealand white rabbits following a bolus dose of caffeine showed that the milk-to-serum paraxanthine concentration ratio was  $0.358 \pm 0.019$ . Paraxanthine and theophylline have the lowest ratios, about half those of caffeine and theobromine (McNamara et al. 1992).



## 5.2 Excretion

A systematic survey of individual pollutants in a sewage treatment plant receiving urban wastewater recently showed that paraxanthine was one of the main product found in concentrations over 20 ppb (Rosal et al. 2010). Paraxanthine has been identified in human urine (Salomon 1883) and then recovered unchanged in wastewater. The most frequently detected compounds in surface water on the coastline within the western Lake Erie basin were caffeine (88%) and paraxanthine (56%), which was detected with a maximum concentration of 1.8  $\mu\text{g/L}$  (Wu et al. 2009). Intravenous administration of [1-Me- $^{14}\text{C}$ ]paraxanthine to rats showed that 1 day after injection  $7 \pm 1\%$  was recovered in feces (Arnaud and Welsch 1979b).

Intravenous administration of [1-Me- $^{14}\text{C}$ ]paraxanthine to rats showed that the main excretory pathway was urine, where  $85 \pm 3\%$  of the administered dose was recovered (Arnaud and Welsch 1979b). In man, approximately 60% of orally administered paraxanthine may be recovered as unchanged in the urine (Arnaud and Welsch 1980a) and after caffeine administration to healthy subjects, paraxanthine was excreted in the urine in amounts sixfold higher than caffeine (Arnaud and Welsch 1980a; Callahan et al. 1982). Paraxanthine was extensively reabsorbed in the renal tubule and its renal clearance was highly urine flow dependent and thus urinary excretion varied with urine output (Tang-Liu et al. 1983).

## 5.3 Pharmacokinetics

The pharmacokinetics of paraxanthine given intravenously in different doses to rats showed that the fraction bound was 15% and remained constant in the plasma for a concentration range of 1–100  $\mu\text{g/mL}$ . Paraxanthine elimination followed first-order kinetics for a dose up to 10 mg/kg and the blood concentrations versus time data were described by a one-compartment, open model system. The mean half-life and elimination rate constant were 1 h and 0.70/h, respectively. The average apparent volume of distribution was 1.50 L/kg and the total clearance was 0.90 L/h/kg. After larger doses (15 and 30 mg/kg), the kinetics were nonlinear and the AUC increased, but not in proportion to the dose. These findings indicated that paraxanthine in the rat is eliminated by a saturable process with an apparent  $K_m$  of about 31  $\mu\text{g/mL}$  and an apparent maximum rate of metabolism of about 0.40  $\mu\text{g/mL/min}$  (Bortolotti et al. 1985; Arnaud and Enslin 1992). In rabbit, paraxanthine clearance was not dose-dependent (Dorrbecker et al. 1987).

The elimination of paraxanthine after its formation has been shown not to follow linear kinetics (Tang-Liu et al. 1983). A relationship has been established between renal clearance of paraxanthine and the urine flow rate (Trang et al. 1985). The partial clearance of caffeine to paraxanthine was eightfold and 23-fold greater than that of theobromine and theophylline, respectively (Lelo et al. 1986b). The

clearances of paraxanthine and caffeine were similar, 2.20 and 2.07 mL/min/kg, respectively and were twofold lower for theophylline and theobromine (Lelo et al. 1986a). After a single dose of 3–4 mg/kg paraxanthine, a mean half-life of  $3.9 \pm 0.7$  h was reported (Lelo et al. 1986a, 1989). The volume of distribution at the steady state of paraxanthine was similar to that of caffeine and theobromine (0.63–0.72 L/kg) and was higher than that of theophylline (0.44 L/kg) (Lelo et al. 1986a).

The administration of [8-<sup>14</sup>C]caffeine (40 mg/kg) to various mouse strains showed higher plasma paraxanthine concentrations in the CBA/J strain compared with the SJL/J, A/J, and SWR/J strains. This effect may be explained by differences in the capacity for paraxanthine glucuronidation. The ratio of the plasma concentration of paraxanthine to total methylxanthine was 7–12% after 0.5 h and increased after 8 h to 29% for the SJL/J and A/J strains and to 44–52% for the CBA/J and SWR/J strains. Paraxanthine was the most important methylxanthine 4 h after caffeine administration for the CBA/J and SWR/J strains but not for the SJL/J and A/J strains, where similar concentrations of paraxanthine and caffeine are observed (Arnaud et al. 1989). There were no differences in the blood pharmacokinetics of paraxanthine between CYP1A2-null and wild-type mice (Labeledzki et al. 2002).

Allopurinol pretreatment had no effect on paraxanthine plasma clearance but decreased 1MU excretion and increased 1MX excretion, with the combined excretion of these metabolites remaining constant (Lelo et al. 1989). Among the high degree of interliver variation in metabolic rates, smokers showed the second highest activity among a 20-fold range in paraxanthine demethylation rates (Campbell et al. 1987a).

## 5.4 Metabolism

The main metabolite of paraxanthine was 1MX both in human and in wild-type mice liver microsomes. In contrast, in CYP1A2-null murine liver microsomes, the main paraxanthine metabolite was 7MX (Labeledzki et al. 2002). It was suggested that the formation of paraxanthine may be a better indicator of *in vivo* CYP1A2 activity than caffeine levels (Bapiro et al. 2005). The high value of the CYP1A2 index defined as urinary AFMU plus 1MX plus 1MU to 1,7DMU could be explained by a low urinary concentration of 1,7DMU and was suggested to be affected by the whole deleted allele of CYP2A6 (CYP2A6\*4) in healthy Japanese volunteers. It was shown that CYP2A6 and CYP1A2 exhibited high catalytic activities for the paraxanthine 8-hydroxylation, which was significantly associated with coumarin 7-hydroxylase activities ( $P < 0.01$ ) in liver microsomes. Tranylcypropane, an inhibitor of CYP2A6, reduced the paraxanthine 8-hydroxylase activities of human liver microsomes. Paraxanthine 8-hydroxylase activities were also found to be low in liver microsomes from individuals possessing deletion of or mutations in the CYP2A6 gene, suggesting that CYP2A6 is a main paraxanthine

8-hydroxylase and this activity is reduced by the genetic polymorphisms of the CYP2A6 gene (Kimura et al. 2005). Children had a higher urine metabolite ratio of paraxanthine 7-demethylation to 8-hydroxylation ( $P < 0.001$ ) than adults (Campbell et al. 1987b).

## 5.5 Metabolites and Metabolic Pathways

Paraxanthine is the main pathway (75–80%) of the first step of caffeine metabolism in man (Arnaud and Welsch 1980a), while it represents a maximum of 40% in rats, 65% in mice, 55% in Chinese hamsters, and less than 10% in monkeys. Rabbits seems the closest model to man, but with a relative inefficiency for further metabolizing paraxanthine (Dorrbecker et al. 1987). The mean fractional conversion of caffeine to paraxanthine was  $79.6 \pm 21\%$ , while it was only  $10.8 \pm 2.4\%$  for theobromine and  $3.7 \pm 1.3\%$  for theophylline. Demethylation pathways accounted for  $83.9 \pm 5.4\%$  for paraxanthine and only  $12.1 \pm 4.1\%$  for theobromine and  $4.0 \pm 1.4\%$  for theophylline (Lelo et al. 1986b). The combined formation of the 7-demethylated products (1MX, 1MU) and AFMU was found to account for 67% of paraxanthine clearance. Formation of 7MX and 1,7DMU and renal excretion of unchanged paraxanthine corresponded to 6, 8, and 9% of paraxanthine clearance, respectively. Data from the effect of allopurinol and cimetidine are consistent with 1MX and AFMU being derived from a common intermediate (Lelo et al. 1989).

Unchanged paraxanthine was the most important urine metabolite in rats,  $52 \pm 3\%$ , and 1,7DMU with the uracil derivative 1,7DAU both corresponded to  $15 \pm 2\%$  of the dose excreted. Paraxanthine 1-methyl demethylation with the formation of 7MX and 7MU is a minor pathway ( $2.3 \pm 0.2\%$ ), while 7-methyl demethylation was the most important with urine excretion of 1MX ( $11 \pm 1\%$ ) and 1MU ( $21 \pm 3\%$ ) (Arnaud and Welsch 1979b). Important species differences were shown for paraxanthine metabolism (Arnaud 1985) and a glucuroconjugate of paraxanthine was identified only in mice (Arnaud et al. 1986b). Paraxanthine and paraxanthine glucuronide urine excretion in mice showed an inverse relationship in CBA/J ( $17 \pm 5$  and  $9.5 \pm 3\%$ , respectively), SJL/J ( $10.5 \pm 3.5$  and  $21.0 \pm 2\%$ ), A/J ( $7 \pm 1$  and  $25 \pm 1.5\%$ ) and SWR/J ( $14.5 \pm 4$  and  $19 \pm 6\%$ ) strains (Arnaud et al. 1989).

The analysis of the metabolites in urine collected at 24-h showed with unchanged paraxanthine ( $10 \pm 4\%$  of the administered dose) the presence of 1MX ( $15 \pm 4\%$ ), 1MU ( $17 \pm 2\%$ ), 1,7DMU ( $15 \pm 3\%$ ), 1,7DAU ( $4 \pm 2\%$ ), 7MX ( $5 \pm 3\%$ ), AFMU ( $18 \pm 6\%$ ), and 7MU ( $4 \pm 3\%$ ) (Arnaud and Welsch 1980a; Callahan et al. 1982). The quantitative urinary excretion of paraxanthine metabolites in man and in various animal species, expressed as the percentage of the administered dose, is shown in Table 1.

## References

- Abdi F, Pollard I, Wilkinson J (1993) Placental transfer and foetal disposition of caffeine and its immediate metabolites in the 20-day pregnant rat: function of dose. *Xenobiotica* 23 (4):449–456
- Abernethy D, Todd E (1985) Impairment of caffeine clearance by chronic use of low-dose estrogen-containing oral contraceptives. *Eur J Clin Pharmacol* 28(4):425–428
- Abernethy DR, Todd EL, Schwartz JB (1985) Caffeine disposition in obesity. *Br J Clin Pharmacol* 20(1):61–66
- Aldridge A, Neims AH (1979) The effects of phenobarbital and beta-naphthoflavone on the elimination kinetics and metabolite pattern of caffeine in the beagle dog. *Drug Metab Dispos* 7(6):378–382
- Aldridge A, Neims AH (1980) Relationship between the clearance of caffeine and its 7-N-demethylation in developing beagle puppies. *Biochem Pharmacol* 29(3):1909–1914
- Amchin J, Zarycranski W, Taylor KP, Albano D, Klockowski PM (1999) Effect of venlafaxine on CYP1A2-dependent pharmacokinetics and metabolism of caffeine. *J Clin Pharmacol* 39 (3):252–259
- Amodio P, Lauro S, Rondana M, Crema G, Merkel C, Gatta A, Ruol A (1991) Theophylline pharmacokinetics and liver function indexes in chronic liver disease. *Respiration* 58 (2):106–111
- Anonymous (1991) International Agency for Research on Cancer (IARC) monographs on the evaluation of carcinogenic risks to humans, WHO, vol 51. Coffee, tea, maté, methylxanthines and methylglyoxal. WHO, Geneva, p 513
- Aranda JV, Louridas AT, Vitullo BB, Thom P, Aldridge A, Haber R (1979) Metabolism of theophylline to caffeine in human fetal liver. *Science* 206(4424):1319–1321
- Arimori K, Nakano M (1988) Dose-dependency in the exsorption of theophylline and the intestinal dialysis of theophylline by oral activated charcoal in rats. *J Pharm Pharmacol* 40(2):101–105
- Arnaud MJ (1976) Identification, kinetic and quantitative study of [2-<sup>14</sup>C] and [1-Me-<sup>14</sup>C] caffeine metabolites in rat's urine by chromatographic separations. *Biochem Med* 16(1):67–76
- Arnaud MJ (1980) Metabolism of labeled paraxanthine in the rat: comparison of rat and human metabolic pathway. *Nutr Rev* 38(5):196–200
- Arnaud MJ (1983) The effect of dietary factors on the bioavailability of methylxanthines. *Proceedings of the toxicology forum*, Aspen, Colorado, pp 234–243
- Arnaud MJ (1984) Products of metabolism of caffeine. In: Dewes PB (ed) caffeine. Springer, Berlin, pp 3–38
- Arnaud MJ (1985) Comparative metabolic disposition of [1-Me<sup>14</sup>C] caffeine in rats, mice, and Chinese hamsters. *Drug Metab Dispos* 13(4):471–478
- Arnaud MJ (1987) The pharmacology of caffeine. *Prog Drug Res* 31:273–313
- Arnaud MJ (1988) The metabolism of coffee constituents in coffee, vol 3. In: Clarke RJ, Macrae R (eds) *Physiology*. Elsevier, London, pp 33–55
- Arnaud MJ (1993a) Metabolism of caffeine and other components of coffee. In: Garattini S (ed) *Caffeine, coffee and health*. Raven, New York, pp 43–95
- Arnaud MJ (1993b) Caffeine, vol 1. In: Macrae R, Robinson RK, Sadler MJ (eds) *Encyclopaedia of food science, food technology and nutrition*. Academic, London, pp 566–571
- Arnaud MJ (1998) Pharmacokinetics and metabolism of caffeine. In: Snel J, Lorist MM (eds) *Nicotine, caffeine and social drink, behaviour and brain function*. Harwood, Amsterdam, pp 153–165
- Arnaud MJ, Ben Zvi Z, Yaari A, Gorodischer R (1986a) 1,3,8-Trimethylallantoin: a major caffeine metabolite formed by rat liver. *Res Commun Chem Pathol Pharmacol* 52(3):407–410
- Arnaud MJ, Bracco I (1981) Distribution and metabolism of theophylline in the pregnant rat: presence of a blood brain barrier. *Experientia* 37(6):665

- Arnaud MJ, Bracco I, Getaz F (1989) Synthesis of ring labelled caffeine for the study of metabolic and pharmacokinetics mouse interstrain differences in relation to pharmacologic and toxic effects. In: Baillie TA, Jones JR (eds) Synthesis and applications of isotopically labelled compounds. Elsevier, Amsterdam, pp 645–648
- Arnaud MJ, Bracco I, Sauvageat JL, Clerc MF (1983) Placental transfer of the major caffeine metabolite in the rat using 6-amino-5[N-formylmethylamino], 3[Me-<sup>14</sup>C]-dimethyluracil administered orally or intravenously to the pregnant rat. *Toxicol Lett* 16(3–4):271–279
- Arnaud MJ, Bracco I, Welsch C (1982a) Metabolism and distribution of labeled theophylline in the pregnant rat. Impairment of theophylline metabolism by pregnancy and absence of a blood-brain barrier in the fetus. *Pediatr Res* 16(3):167–171
- Arnaud MJ, Enslin M (1992) The role of paraxanthine in mediating physiological effects of caffeine. In: 14th international conference in coffee science, San Francisco, 14–19 July 1991. Proceedings ASIC, Paris, pp 71–79
- Arnaud MJ, Gétaz F (1982) Postnatal establishment of a blood-brain barrier for theobromine in the rat. *Experientia* 38(6):752
- Arnaud MJ, Gétaz F (1983) Theobromine distribution in the pregnant rat and the fetus and the impairment of its metabolism due to pregnancy. *Experientia* 39(6):678
- Arnaud MJ, Richli U, Philipposian G (1986b) Isolation and identification of paraxanthine glucuronide as the major caffeine metabolite in mice. *Experientia* 42(6):696
- Arnaud MJ, Thelin-Doerner A, Ravussin E, Acheson KJ (1980) Study of the demethylation of [1,3,7-Me-<sup>13</sup>C] caffeine in man using respiratory exchange measurements. *Biomed Mass Spectrom* 7(11–12):521–524
- Arnaud MJ, Welsch C (1979a) Metabolic pathway of theobromine in the rat and identification of two new metabolites in human urine. *J Agric Food Chem* 27(3):524–527
- Arnaud MJ, Welsch C (1979b) Metabolism of [1-Me-<sup>14</sup>C] paraxanthine in the rat: identification of a new metabolite. *Experientia* 35(7):946
- Arnaud MJ, Welsch C (1980a) Caffeine metabolism in human subjects. In: Proceedings of the ninth international colloquium on science and technology of coffee. Association Scientifique Internationale du Cafe, London, pp 385–395
- Arnaud MJ, Welsch C (1980b) Quantitative analysis of theophylline metabolites by HPLC, after oral or i.v. administration. *Experientia* 36(6):704
- Arnaud MJ, Welsch C (1982) Theophylline and caffeine metabolism in man. In: Woodcock BG, Staib AH, Rietbrock N (eds) *Methods in clinical pharmacology*, 3: theophylline and other methylxanthines. Vieweg, Brunswick, pp 135–148
- Arnaud MJ, Wietholtz H, Voegelin M, Bircher J, Preisig R (1982b) Assessment of the cytochrome P-448 dependent liver enzyme system by a caffeine breath test. In: Sato R (ed) *Microsomes drug oxidation and drug toxicity*. Wiley, New York, pp 443–444
- Arold G, Donath F, Maurer A, Diefenbach K, Bauer S, Henneicke-von Zepelin HH, Friede M, Roots I (2005) No relevant interaction with alprazolam, caffeine, tolbutamide, and digoxin by treatment with a low-hyperforin St John's wort extract. *Planta Med* 71(4):331–337
- Azcona O, Barbanj MJ, Torrent J, Jané F (1995) Evaluation of the central effects of alcohol and caffeine interaction. *Br J Clin Pharmacol* 40(4):393–400
- Backman JT, Karjalainen MJ, Neuvonen M, Laitila J, Neuvonen PJ (2006) Rofecoxib is a potent inhibitor of cytochrome P450 1A2: studies with tizanidine and caffeine in healthy subjects. *Br J Clin Pharmacol* 62(3):345–357
- Bapiro TE, Sayi J, Hasler JA, Jande M, Rimoy G, Masselle A, Masimirembwa CM (2005) Artemisinin and thiabendazole are potent inhibitors of cytochrome P450 1A2 (CYP1A2) activity in humans. *Eur J Clin Pharmacol* 61(10):755–761
- Bayar C, Ozer I (1997) A study on the route of 1-methylurate formation in theophylline metabolism. *Eur J Drug Metab Pharmacokinet* 22(4):415–419
- Bchir F, Dogui M, Ben Fradj R, Arnaud MJ, Saguem S (2006) Differences in pharmacokinetic and electroencephalographic responses to caffeine in sleep-sensitive and non-sensitive subjects. *C R Biol* 329(7):512–519

- Beach CA, Mays DC, Sterman BM, Gerber N (1985) Metabolism, distribution, seminal excretion and pharmacokinetics of caffeine in the rabbit. *J Pharmacol Exp Ther* 233(1):18–23
- Becker AB, Simons KJ, Gillespie CA, Simons FE (1984) The bronchodilator effects and pharmacokinetics of caffeine in asthma. *N Engl J Med* 310(12):743–746
- Begas E, Kouvaras E, Tsakalof A, Papakosta S, Asproдини EK (2007) In vivo evaluation of CYP1A2, CYP2A6, NAT-2 and xanthine oxidase activities in a Greek population sample by the RP-HPLC monitoring of caffeine metabolic ratios. *Biomed Chromatogr* 21(2):190–200
- Benowitz NL, Jacob P 3rd, Mayan H, Denaro C (1995) Sympathomimetic effects of paraxanthine and caffeine in humans. *Clin Pharmacol Ther* 58(6):684–691
- Berdel D, Süverkrüp R, Heimann G, von Berg A, Liappis N, Stühmer A (1987) Total theophylline clearance in childhood: the influence of age-dependent changes in metabolism and elimination. *Eur J Pediatr* 146(1):41–43
- Bienvu T, Pons G, Rey E, Richard MO, d'Athis P, Arnaud MJ, Olive G (1990) Effect of growth hormone on caffeine metabolism in hypophysectomized rats. *Drug Metab Dispos* 18(3):327–330
- Birkett DJ, Miners JO, Attwood J (1983) Secondary metabolism of theophylline biotransformation products in man—route of formation of 1-methyluric acid. *Br J Clin Pharmacol* 15(1):117–119
- Birkett DJ, Dahlqvist R, Miners JO, Lelo A, Billing B (1985) Comparison of theophylline and theobromine metabolism in man. *Drug Metab Dispos* 13(6):725–728
- Birkett DJ, Miners JO (1991) Caffeine renal clearance and urine caffeine concentrations during steady state dosing Implications for monitoring caffeine intake during sport events. *Br J Clin Pharmacol* 31(4):405–408
- Blake MJ, Abdel-Rahman SM, Pearce RE, Leeder JS, Kearns GL (2006) Effect of diet on the development of drug metabolism by cytochrome P-450 enzymes in healthy infants. *Pediatr Res* 60(6):717–723
- Blanchard J, Sawers SJA (1983a) The absolute bioavailability of caffeine in man. *Eur J Clin Pharmacol* 24(1):93–98
- Blanchard J, Sawers SJA (1983b) Comparative pharmacokinetics of caffeine in young and elderly men. *J Pharmacokinet Biopharm* 11(2):109–126
- Blanchard J, Sawers SJ (1983c) Relationship between urine flow rate and renal clearance of caffeine in man. *J Clin Pharmacol* 23(4):134–138
- Blanchard J, Sawers SJA, Jonkman JHG, Tang-Liu D-S (1985) Comparison of the urinary metabolite profile of caffeine in young and elderly males. *Br J Clin Pharmacol* 19(2):225–232
- Blanchard J, Harvey S, Morgan WJ (1992) Relationship between serum and saliva theophylline levels in patients with cystic fibrosis. *Ther Drug Monit* 14(1):48–54
- Bologa M, Tang B, Klein J, Tesoro A, Koren G (1991) Pregnancy-induced changes in drug metabolism in epileptic women. *J Pharmacol Exp Ther* 257(2):735–740
- Bonati M, Latini R, Galletti F, Young JF, Tognoni G, Garattini S (1982) Caffeine disposition after oral doses. *Clin Pharmacol Ther* 32(1):98–106
- Bonati M, Latini R, Sadurska B, Riva E, Galletti F, Borzelleca JF, Tarka SM, Arnaud MJ, Garattini S (1984) Kinetics and metabolism of theobromine in male rats. *Toxicology* 30(4):327–341
- Bortolotti A, Jiritano L, Bonati M (1985) Pharmacokinetics of paraxanthine, one of the primary metabolites of caffeine, in the rat. *Drug Metab Dispos* 13(2):227–231
- Bory C, Baltassat P, Porthault M, Bethenod M, Frederich A, Aranda JV (1979) Metabolism of theophylline to caffeine in premature newborn infants. *J Pediatr* 94(6):988–993
- Boutroy MJ, Vert P, Monin P, Royer RJ, Royer-Morrot MJ (1979) Methylation of theophylline to caffeine in premature infants. *Lancet* 14(8120):830
- Bracco D, Ferrara J-M, Arnaud MJ, Jequier E, Schutz Y (1995) Effects of caffeine on energy metabolism, heart rate, and methylxanthine metabolism in lean and obese women. *Am J Physiol* 269(4 Pt 1):E671–E678
- Brachtel D, Richter E (1988) Effect of altered gastric emptying on caffeine absorption. *Z Gastroenterol* 26(5):245–251

- Brandstetter Y, Kaplanski J, van Creveld C, Ben-Zvi Z (1986) Theophylline pharmacokinetics in pregnant and lactating rats. *Res Commun Chem Pathol Pharmacol* 53(2):269–272
- Brazier JL, Descotes J, Lery N, Ollagnier M, Evreux JC (1980a) Inhibition by idrocilamide of the disposition of caffeine. *Eur J Clin Pharmacol* 17(1):37–43
- Brazier JL, Ribon B, Desage M, Salle B, Salle B (1980b) Study of theophylline metabolism in premature human newborns using stable isotope labelling. *Biomed Mass Spectrom* 7(5):189–192
- Brösen K, Skjelbo E, Rasmussen BB, Poulsen HE, Loft S (1993) Fluvoxamine is a potent inhibitor of cytochrome P4501A2. *Biochem Pharmacol* 45(6):1211–1214
- Brouwers J, Ingels F, Tack J, Augustijns P (2005) Determination of intraluminal theophylline concentrations after oral intake of an immediate- and a slow-release dosage form. *J Pharm Pharmacol* 57(8):987–996
- Brown CR, Benowitz NL (1989) Caffeine and cigarette smoking: behavioral, cardiovascular, and metabolic interactions. *Pharmacol Biochem Behav* 34(3):565–570
- Brown CR, Jacob P 3rd, Wilson M, Benowitz NL (1988) Changes in rate and pattern of caffeine metabolism after cigarette abstinence. *Clin Pharmacol Ther* 43(5):488–491
- Bruguerolle B (1987) [Changes in the pharmacokinetics of theophylline during estrus in rats]. *Pathol Biol* 35(2):181–183
- Butler MA, Lang NP, Young JF, Caporaso NE, Vineis P, Hayes RB, Teitel CH, Massengill JP, Lawsen MF, Kadlubar FF (1992) Determination of CYP1A2 and NAT2 phenotypes in human populations by analysis of caffeine urinary metabolites. *Pharmacogenetics* 2(3):116–127
- Callahan MM, Robertson RS, Arnaud MJ, Branfman AR, McComish MF, Yesair DW (1982) Human metabolism of [1-methyl-<sup>14</sup>C]- and [2-<sup>14</sup>C]caffeine after oral administration. *Drug Metab Dispos* 10(4):417–423
- Campbell ME, Grant DM, Inaba T, Kalow W (1987a) Biotransformation of caffeine, paraxanthine, theophylline, and theobromine by polycyclic aromatic hydrocarbon-inducible cytochrome(s) P-450 in human liver microsomes. *Drug Metab Dispos* 15(2):237–249
- Campbell ME, Spielberg SP, Kalow W (1987b) A urinary metabolic ratio that reflects systemic caffeine clearance. *Clin Pharmacol Ther* 42(2):157–165
- Caraco Y, Zylber-Katz E, Berry EM, Levy M (1995) Caffeine pharmacokinetics in obesity and following significant weight reduction. *Int J Obes* 19(4):234–239
- Carrillo JA, Christensen M, Ramos SI, Alm C, Dahl ML, Benitez J, Bertilsson L (2000) Evaluation of caffeine as an in vivo probe for CYP1A2 using measurements in plasma, saliva, and urine. *Ther Drug Monit* 22(4):409–417
- Chen Y, Tu JH, He YJ, Zhang W, Wang G, Tan ZR, Zhou G, Fan L, Zhou HH (2009a) Effect of sodium tanshinone II A sulfonate on the activity of CYP1A2 in healthy volunteers. *Xenobiotica* 39(7):508–513
- Chen Y, Xiao P, Ou-Yang DS, Fan L, Guo D, Wang YN, Han Y, Tu JH, Zhou G, Huang YF, Zhou HH (2009b) Simultaneous action of the flavonoid quercetin on cytochrome P450 (CYP) 1A2, CYP2A6, N-Acetyltransferase and xanthine oxidase activity in healthy volunteers. *Clin Exp Pharmacol Physiol* 36(8):828–833
- Cheng WS, Murphy TL, Smith MT, Cooksley WG, Halliday JW, Powell LW (1990) Dose-dependent pharmacokinetics of caffeine in humans: relevance as a test of quantitative liver function. *Clin Pharmacol Ther* 47(4):516–524
- Christensen M, Andersson K, Dalén P, Mirghani RA, Muirhead GJ, Nordmark A, Tybring G, Wahlberg A, Yaşar U, Bertilsson L (2003) The Karolinska cocktail for phenotyping of five human cytochrome P450 enzymes. *Clin Pharmacol Ther* 73(6):517–528
- Chung WG, Kang JH, Park CS, Cho MH, Cha YN (2000) Effect of age and smoking on in vivo CYP1A2, flavin-containing monooxygenase, and xanthine oxidase activities in Koreans: determination by caffeine metabolism. *Clin Pharmacol Ther* 67(3):258–266
- Chvasta TE, Cooke AR (1971) Emptying and absorption of caffeine from the human stomach. *Gastroenterology* 61(6):838–843

- Collomp K, Anselme F, Audran M, Gay JP, Chanal JL, Prefaut C (1991) Effects of moderate exercise on the pharmacokinetics of caffeine. *Eur J Clin Pharmacol* 40(3):279–282
- Conner DP, Millora E, Zamani K, Nix D, Almirez RG, Rhyne-Kirsch P, Peck CC (1991) Transcutaneous chemical collection of caffeine in normal subjects: relationship to area under the plasma concentration-time curve and sweat production. *J Invest Dermatol* 96:186–190
- Cooling DS (1993) Theophylline toxicity. *J Emerg Med* 11(4):415–425
- Cornish HH, Christman AA (1957) A study of the metabolism of theobromine, theophylline, and caffeine in man. *J Biol Chem* 228(1):315–323
- Crowley JJ, Cusack BJ, Jue SG, Koup JR, Park BK, Vestal RE (1988) Aging and drug interactions. II. Effect of phenytoin and smoking on the oxidation of theophylline and cortisol in healthy men. *J Pharmacol Exp Ther* 245(2):513–523
- Cysneiros RM, Farkas D, Harmatz JS, von Moltke LL, Greenblatt DJ (2007) Pharmacokinetic and pharmacodynamic interactions between zolpidem and caffeine. *Clin Pharmacol Ther* 82(1):54–62
- Danielson TJ, Golsteyn LR (1996) Systemic clearance and demethylation of caffeine in sheep and cattle. *Drug Metab Dispos* 24(10):1058–1061
- Darwish M, Kirby M, Robertson P Jr, Hellriegel ET (2008) Interaction profile of armodafinil with medications metabolized by P enzymes 1A2, 3A4 and 2C19 in healthy subjects. *Clin Pharmacokinet* 47(1):61–74
- DeGraves FJ, Ruffin DC, Duran SH, Spano JS, Whatley EM, Schumacher J, Riddell MG (1995) Pharmacokinetics of caffeine in lactating dairy cows. *Am J Vet Res* 56(5):619–622
- Delahunty T, Schoendorfer D (1998) Caffeine demethylation monitoring using a transdermal sweat patch. *J Anal Toxicol* 22(7):596–600
- Denaro CP, Brown CR, Wilson M, Jacob P, Benowitz NL (1990) Dose-dependency of caffeine metabolism with repeated dosing. *Clin Pharmacol Ther* 48(3):277–285
- Derby KS, Cuthrell K, Caberto C, Carmella SG, Franke AA, Hecht SS, Murphy SE, Le Marchand L (2008) Nicotine metabolism in three ethnic/racial groups with different risks of lung cancer. *Cancer Epidemiol Biomarkers Prev* 17(12):3526–3535
- Derkenne S, Curran CP, Shertzer HG, Dalton TP, Dragin N, Nebert DW (2005) Theophylline pharmacokinetics: comparison of Cyp1a1(-/-) and Cyp1a2(-/-) knockout mice, humanized hCYP1A1\_1A2 knock-in mice lacking either the mouse Cyp1a1 or Cyp1a2 gene, and Cyp1(+/-) wild-type mice. *Pharmacogenet Genomics* 15(7):503–511
- Desmond PV, Patwardhan RV, Johnson RF, Schenker S (1980) Impaired elimination of caffeine in cirrhosis. *Dig Dis Sci* 25(3):193–197
- Devoe LD, Murray C, Youssif A, Arnaud M (1993) Maternal caffeine consumption and fetal behavior in normal third-trimester pregnancy. *Am J Obstet Gynecol* 168(4):1105–1111 (discussion 1111–1112)
- Djordjevic N, Ghotbi R, Bertilsson L, Jankovic S, Aklillu E (2008) Induction of CYP1A2 by heavy coffee consumption in Serbs and Swedes. *Eur J Clin Pharmacol* 64(4):381–385
- Dorrbecker SH, Ferraina RA, Dorrbecker BR, Kramer PA (1987) Caffeine and paraxanthine pharmacokinetics in the rabbit: concentration and product inhibition effects. *J Pharmacokinet Biopharm* 15(2):117–132
- Dorrbecker SH, Kramer PA, Dorrbecker BR, Raye JR (1988a) Caffeine disposition in the pregnant rabbit. II. Fetal distribution of caffeine and paraxanthine during chronic maternal caffeine administration. *Dev Pharmacol Ther* 11(2):118–124
- Dothey CI, Tserng KY, Kaw S, King KC (1989) Maturational changes of theophylline pharmacokinetics in preterm infants. *Clin Pharmacol Ther* 45(5):461–468
- Doude van Troostwijk LJ, Koopmans RP, Vermeulen HD, Guchelaar HJ (2003a) CYP1A2 activity is an important determinant of clozapine dosage in schizophrenic patients. *Eur J Pharm Sci* 20(4–5):451–457
- Doude van Troostwijk LJ, Koopmans RP, Guchelaar HJ (2003b) Two novel methods for the determination of CYP1A2 activity using the paraxanthine/caffeine ratio. *Fundam Clin Pharmacol* 17(3):355–362



- Driscoll MS, Ludden TM, Casto DT, Littlefield LC (1989) Evaluation of theophylline pharmacokinetics in a pediatric population using mixed effects models. *J Pharmacokinet Biopharm* 17 (2):141–168
- Drouillard DD, Vesell ES, Dvorchik BH (1978) Studies on theobromine disposition in normal subjects. Alterations induced by dietary abstention from or exposure to methylxanthines. *Clin Pharmacol Ther* 23(3):296–302
- du Preez MJ, Botha JH, McFadyen ML, Holford NH (1999) The pharmacokinetics of theophylline in premature neonates during the first few days after birth. *Ther Drug Monit* 21(6):598–603
- Ellis EF, Koysooko R, Levy G (1976) Pharmacokinetics of theophylline in children with asthma. *Pediatrics* 58(4):542–547
- Faber MS, Fuhr U (2004) Time response of cytochrome P450 1A2 activity on cessation of heavy smoking. *Clin Pharmacol Ther* 76(2):178–184
- Fleetham JA, Bird CE, Nakatsu K, Wigle RD, Munt PW (1981) Dose-dependency of theophylline clearance and protein binding. *Thorax* 36(5):382–386
- Fontana RJ, deVries TM, Woolf TF, Knapp MJ, Brown AS, Kaminsky LS, Tang BK, Foster NL, Brown RR, Watkins PB (1998) Caffeine based measures of CYP1A2 activity correlate with oral clearance of tacrine in patients with Alzheimer's disease. *Br J Clin Pharmacol* 46(3):221–228
- Fuhr U, Anders EM, Mahr G, Sörgel F, Staib AH (1992) Inhibitory potency of quinolone antibacterial agents against cytochrome P4501A2 activity in vivo and in vitro. *Antimicrob Agents Chemother* 36(5):942–948
- Fuhr U, Klittich K, Staib AH (1993) Inhibitory effect of grapefruit juice and its bitter principal, naringenin, on CYP1A2 dependent metabolism of caffeine in man. *Br J Clin Pharmacol* 35 (4):431–436
- Fuhr U, Maier A, Keller A, Steinijans VW, Sauter R, Staib AH (1995) Lacking effect of grapefruit juice on theophylline pharmacokinetics. *Int J Clin Pharmacol Ther* 33(6):311–314
- Fuhr U, Jetter A, Kirchheiner J (2007) Appropriate phenotyping procedures for drug metabolizing enzymes and transporters in humans and their simultaneous use in the “cocktail” approach. *Clin Pharmacol Ther* 81(2):270–83
- Gal P, Jusko WJ, Yurchak AM, Franklin BA (1978) Theophylline disposition in obesity. *Clin Pharmacol Ther* 23(4):438–444
- Gardner MJ, Jusko WJ (1982) Effect of age and sex on theophylline clearance in young subjects. *Pediatr Pharmacol* 2(2):157–169
- Gardner MJ, Tornatore KM, Jusko WJ, Kanarkowski R (1983) Effects of tobacco smoking and oral contraceptive use on theophylline disposition. *Br J Clin Pharmacol* 16(3):271–280
- Gardner MJ, Schatz M, Cousins L, Zeiger R, Middleton E, Jusko WJ (1987) Longitudinal effects of pregnancy on the pharmacokinetics of theophylline. *Eur J Clin Pharmacol* 32(3):289–295
- Gates S, Miners JO (1999) Cytochrome P450 isoform selectivity in human hepatic theobromine metabolism. *Br J Clin Pharmacol* 47(3):299–305
- George J, Murphy T, Roberts R, Cooksley WG, Halliday JW, Powell LW (1986) Influence of alcohol and caffeine consumption on caffeine elimination. *Clin Exp Pharmacol Physiol* 13(10):731–736
- Ghotbi R, Christensen M, Roh HK, Ingelman-Sundberg M, Aklillu E, Bertilsson L (2007) Comparisons of CYP1A2 genetic polymorphisms, enzyme activity and the genotype-phenotype relationship in Swedes and Koreans. *Eur J Clin Pharmacol* 63(6):537–546
- Gilbert SG, Stavric B, Klassen RD, Rice DC (1985) The fate of chronically consumed caffeine in the monkey (*Macaca fascicularis*). *Fundam Appl Toxicol* 5(3):578–587
- Gorodischer R, Zmora E, Ben-Zvi Z, Warszawski D, Yaari A, Sofer S, Arnaud MJ (1986a) Urinary metabolites of caffeine in the premature infant. *Eur J Clin Pharmacol* 31(4):497–499
- Gorodischer R, Yaari A, Margalith M, Warszawski D, Ben-Zvi Z (1986b) Changes in theophylline metabolism during postnatal development in rat liver slices. *Biochem Pharmacol* 35(18): 3077–3081
- Granfors MT, Backman JT, Neuvonen M, Neuvonen PJ (2004) Ciprofloxacin greatly increases concentrations and hypotensive effect of tizanidine by inhibiting its cytochrome P450 1A2-mediated presystemic metabolism. *Clin Pharmacol Ther* 76(6):598–606

- Granfors MT, Backman JT, Laitila J, Neuvonen PJ (2005) Oral contraceptives containing ethinyl estradiol and gestodene markedly increase plasma concentrations and effects of tizanidine by inhibiting cytochrome P450 1A2. *Clin Pharmacol Ther* 78(4):400–411
- Grant DM, Tang BK, Kalow W (1983) Polymorphic N-acetylation of a caffeine metabolite. *Clin Pharmacol Ther* 33(3):355–359
- Grant DM, Tang BK, Kalow W (1984) A simple test for acetylation phenotype using caffeine. *Br J Clin Pharmacol* 17(4):459–464
- Grant DM, Tang BK, Campbell ME, Kalow W (1986) Effect of allopurinol on caffeine disposition in man. *Br J Clin Pharmacol* 21(4):454–458
- Groen K, Horan MA, Roberts NA, Gulati RS, Miljkovic B, Jansen EJ, Paramsothy V, Breimer DD, van Bezooijen CF (1993) The relationship between phenazone (antipyrene) metabolite formation and theophylline metabolism in healthy and frail elderly women. *Clin Pharmacokinet* 25(2):136–144
- Grosso LM, Bracken MB (2005) Caffeine metabolism, genetics, and perinatal outcomes: a review of exposure assessment considerations during pregnancy. *Ann Epidemiol* 15(6):460–466
- Grosso LM, Triche E, Benowitz NL, Bracken MB (2008) Prenatal caffeine assessment: fetal and maternal biomarkers or self-reported intake? *Ann Epidemiol* 18(3):172–178
- Grygiel JJ, Birkett DJ (1980) Effect of age on patterns of theophylline metabolism. *Clin Pharmacol Ther* 28(4):456–462
- Grygiel JJ, Birkett DJ (1981) Cigarette smoking and theophylline clearance and metabolism. *Clin Pharmacol Ther* 30(4):491–496
- Gu L, Gonzalez FJ, Kalow W, Tang BK (1992) Biotransformation of caffeine, paraxanthine, theobromine and theophylline by cDNA-expressed human CYP1A2 and CYP2E1. *Pharmacogenetics* 2(2):73–77
- Gurley BJ, Gardner SF, Hubbard MA, Williams DK, Gentry WB, Cui Y, Ang CY (2002) Cytochrome P450 phenotypic ratios for predicting herb-drug interactions in humans. *Clin Pharmacol Ther* 72(3):276–287
- Ha HR, Chen J, Freiburghaus AU, Follath F (1995) Metabolism of theophylline by cDNA-expressed human s P. *Br J Clin Pharmacol* 39(3):321–326
- Ha HR, Chen J, Krahenbuhl S, Follath F (1996) Biotransformation of caffeine by cDNA-expressed human s P. *Eur J Clin Pharmacol* 49(4):309–315
- Haller CA, Jacob P 3rd, Benowitz NL (2002) Pharmacology of ephedra alkaloids and caffeine after single-dose dietary supplement use. *Clin Pharmacol Ther* 71(6):421–432
- Hamon-Vilcot B, Simon T, Becquemont L, Poirier JM, Piette F, Jaillon P (2004) Effects of malnutrition on cytochrome P450 1A2 activity in elderly patients. *Therapie* 59(2):247–251
- Hashiguchi M, Fujimura A, Ohashi K, Ebihara A (1992) Diurnal effect on caffeine clearance. *J Clin Pharmacol* 32(2):184–187
- Hendeles L, Weinberger M, Bighley L (1977) Absolute bioavailability of oral theophylline. *Am J Hosp Pharm* 34(5):525–527
- Hendeles L, Massanari MJ, Weinberger M (1986) Theophylline. In: Evans WE, Schentag JJ, Jusko WJ (eds) *Applied pharmacokinetics*, 2nd edn. Applied Therapeutics, San Francisco, pp 1105–1188
- Holstege A, Staiger M, Haag K, Gerok W (1989) Correlation of caffeine elimination and Child's classification in liver cirrhosis. *Klin Wochenschr* 67(1):6–15
- Igarashi T, Iwakawa S (2009) Effect of gender on theophylline clearance in the asthmatic acute phase in Japanese pediatric patients. *Biol Pharm Bull* 32(2):304–307
- Ingvast-Larsson C, Appelgren LE, Nyman G (1992) Distribution studies of theophylline: microdialysis in rat and horse and whole body autoradiography in rat. *J Vet Pharmacol Ther* 15(4):386–394
- Jackson SH, Johnston A, Woollard R, Turner P (1989) The relationship between theophylline clearance and age in adult life. *Eur J Clin Pharmacol* 36(1):29–34
- Jarboe CH, Hurst HE, Rodgers GC, Metaxas JM (1986) Toxicokinetics of caffeine elimination in an infant. *Clin Toxicol* 24(5):415–428

- Jennings TS, Nafziger AN, Davidson L, Bertino JS Jr (1993) Gender differences in hepatic induction and inhibition of theophylline pharmacokinetics and metabolism. *J Lab Clin Med* 122(2):208–216
- Jeppesen U, Loft S, Poulsen HE, Brøsen K (1996) A fluvoxamine-caffeine interaction study. *Pharmacogenetics* 6(3):213–222
- Jodynis-Liebert J, Flieger J, Matuszewska A, Juszczyk J (2004) Serum metabolite/caffeine ratios as a test for liver function. *J Clin Pharmacol* 44(4):338–347
- Jonkman JH, van der Boon WJ, Balant LP, Le Cottonnec JY (1985) Food reduces the rate but not the extent of the absorption of theophylline from an aqueous solution. *Eur J Clin Pharmacol* 28(2):225–227
- Jonkman JH, Sollie FA, Sauter R, Steinijans VW (1991) The influence of caffeine on the steady-state pharmacokinetics of theophylline. *Clin Pharmacol Ther* 49(3):248–255
- Jung D, Nanavaty M (1990) The effects of age and dietary protein restriction on the pharmacokinetics of theophylline in the rat. *Pharmacol Toxicol* 66(5):361–366
- Jusko WJ (1979) Influence of cigarette smoking on drug metabolism in man. *Drug Metab Rev* 9(2):221–236
- Jusko WJ, Gardner MJ, Mangione A, Schentag JJ, Koup JR, Vance JW (1979) Factors affecting theophylline clearances: age, tobacco, marijuana, cirrhosis, congestive heart failure, obesity, oral contraceptives, benzodiazepines, barbiturates, and ethanol. *J Pharm Sci* 68(11):1358–1366
- Kadlubar FF, Butler TG, MA TCH, Massengill JP, Lang NP (1990) Determination of carcinogenic arylamine N-oxidation phenotype in humans by analysis of caffeine urinary metabolites. *Prog Clin Biol Res* 340B:107–114
- Kall MA, Clausen J (1995) Dietary effect on mixed function P4501A2 activity assayed by estimation of caffeine metabolism in man. *Hum Exp Toxicol* 14(10):801–807
- Kalow W (1986) Ethnic differences in reactions to drugs and xenobiotics Caffeine and other drugs. *Prog Clin Biol Res* 214:331–341
- Kalow W, Tang BK (1991a) Use of caffeine metabolic ratios to explore CYP1A2 and xanthine oxidase activities. *Clin Pharmacol Ther* 50(5 Pt 1):508–519
- Kalow W, Tang BK (1991b) Caffeine as a metabolic probe: exploration of the enzyme-inducing effect of cigarette smoking. *Clin Pharmacol Ther* 49(1):44–48
- Kamei K, Matsuda M, Momose A (1975) New sulfur-containing metabolites of caffeine. *Chem Pharm Bull* 23(3):683–685
- Kamimori GH, Somani SM, Knowlton RG, Perkins RM (1987) The effects of obesity and exercise on the pharmacokinetics of caffeine in lean and obese volunteers. *Eur J Clin Pharmacol* 31(5):595–600
- Kamimori GH, Lugo SI, Penetar DM, Chamberlain AC, Brunhart GE, Brunhart AE, Eddington ND (1995) Dose-dependent caffeine pharmacokinetics during severe sleep deprivation in humans. *Int J Clin Pharmacol Ther* 33(3):182–186
- Kawai H, Kokubun S, Matsumoto T, Kojima J, Onodera K (2000) Pharmacokinetic study of theophylline in dogs after intravenous administration with and without ethylenediamine. *Methods Find Exp Clin Pharmacol* 22(3):179–184
- Kearns GL, Hill DE, Tumbelson ME (1986) Theophylline and caffeine disposition in the neonatal piglet. *Dev Pharmacol Ther* 9(6):389–401
- Kilbane AJ, Silbart LK, Manis M, Beitins IZ, Weber WW (1990) Human N-acetylation genotype determination with urinary caffeine metabolites. *Clin Pharmacol Ther* 47(4):470–477
- Kim YC, Lee AK, Lee JH, Lee I, Lee DC, Kim SH, Kim SG, Lee MG (2005) Pharmacokinetics of theophylline in diabetes mellitus rats: induction of CYP1A2 and CYP2E1 on 1,3-dimethyluric acid formation. *Eur J Pharm Sci* 26(1):114–123
- Kimmel CA, Kimmel GL, White CG, Grafton TF, Young JF, Nelson CJ (1984) Blood flow changes and conceptual development in pregnant rats in response to caffeine. *Fundam Appl Toxicol* 4(2 Pt 1):240–247
- Kimura M, Yamazaki H, Fujieda M, Kiyotani K, Honda G, Saruwatari J, Nakagawa K, Ishizaki T, Kamataki T (2005) Cyp2a6 is a principal enzyme involved in hydroxylation of 1,7-dimethyl-xanthine, a main caffeine metabolite, in humans. *Drug Metab Dispos* 33(9):1361–1366

- Kinzig-Schippers M, Fuhr U, Zaigler M, Dammeyer J, Rüsing G, Labeledzki A, Bulitta J, Sörgel F (1999) Interaction of pefloxacin and enoxacin with the human cytochrome P450 enzyme CYP1A2. *Clin Pharmacol Ther* 65(3):262–274
- Kokwaro GO, Szwandt IS, Glazier AP, Ward SA, Edwards G (1993) Metabolism of caffeine and theophylline in rats with malaria and endotoxin-induced fever. *Xenobiotica* 23(12):1391–1397
- Kolski GB, Levy J, Anolik R (1987) The use of theophylline clearance in pediatric status asthmaticus. I. Interpatient and inpatient theophylline clearance variability. *Am J Dis Child* 141(3):282–287
- Korrapati MR, Vestal RE, Loi CM (1995) Theophylline metabolism in healthy nonsmokers and in patients with insulin-dependent diabetes mellitus. *Clin Pharmacol Ther* 57(4):413–418
- Kotake AN, Schoeller DA, Lambert GH, Baker AL, Schaffer DO, Josephs H (1982) The caffeine CO<sub>2</sub> breath test: dose response and route of N-demethylation in smokers and nonsmokers. *Clin Pharmacol Ther* 32(2):261–269
- Kraus DM, Fischer JH, Reitz SJ, Kecskes SA, Yeh TF, McCulloch KM, Tung EC, Cwik MJ (1993) Alterations in theophylline metabolism during the first year of life. *Clin Pharmacol Ther* 54(4):351–359
- Kuh HJ, Shim CK (1994) Nonlinear renal excretion of theophylline and its metabolites, 1-methyluric acid and 1,3-dimethyluric acid, in rats. *Arch Pharm Res* 17(2):124–130
- Labeledzki A, Buters J, Jabrane W, Fuhr U (2002) Differences in caffeine and paraxanthine metabolism between human and murine CYP1A2. *Biochem Pharmacol* 63(12):2159–67
- Landi MT, Sinha R, Lang NP, Kadlubar FF (1999) Human cytochrome P4501A2. *IARC Sci Publ* 148:173–195
- Lane JD, Steege JF, Rupp SL, Kuhn CM (1992) Menstrual cycle effects on caffeine elimination in the human female. *Eur J Clin Pharmacol* 43(5):543–546
- Latini R, Bonati M, Castelli D, Garattini S (1978) Dose-dependent kinetics of caffeine in rats. *Toxicol Lett* 2(5):267–270
- Latini R, Bonati M, Marzi E, Garattini S (1981) Urinary excretion of an uracilic metabolite from caffeine by rat, monkey and man. *Toxicol Lett* 7(3):267–272
- Latini R, Bonati M, Gaspari F, Traina GL, Jiritano L, Bortolotti A, Borzelleca JF, Tarka SM, Arnaud MJ, Garattini S (1984) Kinetics and metabolism of theobromine in male and female non-pregnant and pregnant rabbits. *Toxicology* 30(4):343–354
- Lau CE, Ma F, Falk JL (1995) Oral and IP caffeine pharmacokinetics under a chronic food-limitation condition. *Pharmacol Biochem Behav* 50(2):245–252
- Le Marchand L, Sivaraman L, Franke AA, Custer LJ, Wilkens LR, Lau AF, Cooney RV (1996) Predictors of N-acetyltransferase activity: should caffeine phenotyping and NAT2 genotyping be used interchangeably in epidemiological studies? *Cancer Epidemiol Biomarkers Prev* 5(6):449–455
- Lee TC, Charles BG, Steer PA, Flenady VJ, Grant TC (1996) Theophylline population pharmacokinetics from routine monitoring data in very premature infants with apnoea. *Br J Clin Pharmacol* 41(3):191–200
- Lelo A, Birkett DJ, Robson RA, Miners JO (1986a) Comparative pharmacokinetics of caffeine and its primary demethylated metabolites paraxanthine, theobromine and theophylline in man. *Br J Clin Pharmacol* 22(2):177–182
- Lelo A, Miners JO, Robson RA, Birkett DJ (1986b) Quantitative assessment of caffeine partial clearances in man. *Br J Clin Pharmacol* 22(2):183–186
- Lelo A, Kjellen G, Birkett DJ, Miners JO (1989) Paraxanthine metabolism in humans: determination of metabolic partial clearances and effects of allopurinol and cimetidine. *J Pharmacol Exp Ther* 248(1):315–319
- Lelo A, Birkett DJ, Miners JO (1990) Mechanism of formation of 6-amino-5-(N-methylformylamino)-1-methyluracil and 3,7-dimethyluric acid from theobromine in the rat in vitro: involvement of P- and a cellular thiol. *Xenobiotica* 20(8):823–833
- Lenz TL, Lenz NJ, Faulkner MA (2004) Potential interactions between exercise and drug therapy. *Sports Med* 34(5):293–306

- Lesko LJ, Tabor KJ, Johnson BF (1981) Theophylline serum protein binding in obstructive airways disease. *Clin Pharmacol Ther* 29(6):776–781
- Levy M, Granit L, Zylber-Katz E (1984) Chronopharmacokinetics of caffeine in healthy volunteers. *Annu Rev Chronopharmacol* 1:97–100
- Loi CM, Parker BM, Cusack BJ, Vestal RE (1997) Aging and drug interactions. III. Individual and combined effects of cimetidine and cimetidine and ciprofloxacin on theophylline metabolism in healthy male and female nonsmokers. *J Pharmacol Exp Ther* 280(2):627–637
- Lowry JA, Jarrett RV, Wasserman G, Pettett G, Kauffman RE (2001) Theophylline toxicokinetics in premature newborns. *Arch Pediatr Adolesc Med* 155(8):934–939
- Madyastha KM, Sridhar GR (1998) A novel pathway for the metabolism of caffeine by a mixed culture consortium. *Biochem Biophys Res Commun* 249(1):178–181
- Matzke GR, Frye RF, Early JJ, Straka RJ, Carson SW (2000) Evaluation of the influence of diabetes mellitus on antipyrine metabolism and CYP1A2 and CYP2D6 activity. *Pharmacotherapy* 20(2):182–190
- McLean C, Graham TE (2002) Effects of exercise and thermal stress on caffeine pharmacokinetics in men and eumenorrheic women. *J Appl Physiol* 93(4):1471–1478
- McManus ME, Miners JO, Gregor D, Stupans I, Birkett DJ (1988) Theophylline metabolism by human, rabbit and rat liver microsomes and by purified forms of cytochrome P450. *J Pharm Pharmacol* 40(6):388–391
- McNamara PJ, Burgio D, Yoo SD (1992) Pharmacokinetics of caffeine and its demethylated metabolites in lactating adult rabbits and neonatal offspring. Predictions of breast milk to serum concentration ratios. *Drug Metab Dispos* 20(2):302–308
- Miller GE, Radulovic LL, DeWit RH, Brabec MJ, Tarka SM, Cornish HH (1984) Comparative theobromine metabolism in five mammalian species. *Drug Metab Dispos* 12(2):154–160
- Miners JO, Attwood J, Birkett DJ (1982) Theobromine metabolism in man. *Drug Metab Dispos* 10(6):672–675
- Miners JO, Attwood J, Wing LM, Birkett DJ (1985) Influence of cimetidine, sulfipyrazone, and cigarette smoking on theobromine metabolism in man. *Drug Metab Dispos* 13(5):598–601
- Monks TJ, Caldwell J, Smith RL (1979) Influence of methylxanthine-containing foods on theophylline metabolism and kinetics. *Clin Pharmacol Ther* 26(4):513–524
- Monks TJ, Lawrie CA, Caldwell J (1981) The effect of increased caffeine intake on the metabolism and pharmacokinetics of theophylline in man. *Biopharm Drug Dispos* 2(1):31–37
- Moore ES, Faix RG, Banagale RC, Grasela TH (1989) The population pharmacokinetics of theophylline in neonates and young infants. *J Pharmacokinetic Biopharm* 17(1):47–66
- Morisot C, Simoens C, Trublin F, Lhermitte M, Gremillet C, Robert MH, Lequin P (1990) Efficacité de la caffeine transcutanée dans le traitement des apnées du prématuré. *Arch Fr Pediatr* 47(3):221–224
- Mose T, Kjaerstad MB, Mathiesen L, Nielsen JB, Edelfors S, Knudsen LE (2008) Placental passage of benzoic acid, caffeine, and glyphosate in an ex vivo human perfusion system. *J Toxicol Environ Health A* 71(15):984–991
- Mumford GK, Benowitz NL, Evans SM, Kaminski BJ, Preston KL, Sannerud CA, Silverman K, Griffiths RR (1996) Absorption rate of methylxanthines following capsules, cola and chocolate. *Eur J Clin Pharmacol* 51(3–4):319–325
- Murphy TL, McIvor C, Yap A, Cooksley WG, Halliday JW, Powell LW (1988) The effect of smoking on caffeine elimination: implications for its use as a semiquantitative test of liver function. *Clin Exp Pharmacol Physiol* 15(1):9–13
- Murray RD, Breech L, Ailabouni A, Zingerelli J, Nahata MC (1993) Absorption of theophylline from the small and large intestine of the neonatal piglet. *Eur J Drug Metab Pharmacokin* 18(4):375–379
- Muscat JE, Pittman B, Kleinman W, Lazarus P, Stellman SD, Richie JP Jr (2008) Comparison of CYP1A2 and NAT2 phenotypes between black and white smokers. *Biochem Pharmacol* 76(7):929–937

- Nagel RA, Dirix LY, Hayllar KM, Preisig R, Tredger JM, Williams R (1990) Use of quantitative liver function tests-caffeine clearance and galactose elimination capacity-after orthotopic liver transplantation. *J Hepatol* 10(2):149–157
- Nakazawa K, Tanaka H, Arima M (1985) The effect of caffeine ingestion on pharmacokinetics of caffeine and its metabolites after a single administration in pregnant rats. *J Pharmacobiodyn* 8(3):151–160
- Nam BH, Sohn DH, Ko G, Kim JB (1997) Effect of hepatic cirrhosis on the pharmacokinetics of theophylline in rats. *Arch Pharm Res* 20(4):318–323
- Newton R, Broughton LJ, Lind MJ, Morrisson PJ, Rogers HJ, Bradbrook ID (1981) Plasma and salivary pharmacokinetics of caffeine in man. *Eur J Clin Pharmacol* 21(1):45–52
- Obase Y, Shimoda T, Kawano T, Saeki S, Tomari S, Matsuse H, Kinoshita M, Kohno S (2003) Polymorphisms in the CYP1A2 gene and metabolism in patients with asthma. *Clin Pharmacol Ther* 73:468–474
- Ogilvie RI (1978) Clinical pharmacokinetics of theophylline. *Clin Pharmacokinet* 3(4):267–293
- Oliveto AH, Hughes JR, Terry SY, Bickel WK, Higgins ST, Pepper SL, Fenwick JW (1991) Effects of caffeine on tobacco withdrawal. *Clin Pharmacol Ther* 50(2):157–164
- Omarini D, Barzago MM, Bortolotti A, Aramayona J, Bonati M (1991) Placental transfer of theophylline during in situ perfusion in the rabbit. *J Pharmacol Methods* 25(4):263–273
- Ou-Yang DS, Huang SL, Wang W, Xie HG, Xu ZH, Shu Y, Zhou HH (2000) Phenotypic polymorphism and gender-related differences of CYP1A2 activity in a Chinese population. *Br J Clin Pharmacol* 49(2):145–151
- Pan WJ, Goldwater DR, Zhang Y, Pilmer BL, Hunt RH (2000) Lack of a pharmacokinetic interaction between lansoprazole or pantoprazole and theophylline. *Aliment Pharmacol Ther* 14(3):345–352
- Park EJ, Ko G, Kim J, Sohn DH (1999) Biotransformation of theophylline in cirrhotic rats induced by biliary obstruction. *Arch Pharm Res* 22(1):60–67
- Park GJ, Katelaris PH, Jones DB, Seow F, Le Couteur DG, Ngu MC (2003) Validity of the <sup>13</sup>C-caffeine breath test as a noninvasive, quantitative test of liver function. *Hepatology* 38(5):1227–1236
- Parra P, Limon A, Ferre S, Guix T, Jane F (1991) High-performance liquid chromatographic separation of caffeine, theophylline, theobromine and paraxanthine in rat brain and serum. *J Chromatogr* 570(1):185–190
- Parsons W, Neims A (1978) Effect of smoking on caffeine clearance. *Clin Pharmacol Ther* 24(1):40–45
- Patwardhan R, Desmond P, Johnson R, Schenker S (1980) Impaired elimination of caffeine by oral contraceptive steroid. *J Lab Clin Med* 95(4):603–608
- Perry DF, Walson PD, Blanchard J (1984) Effect of pH on theophylline transfer across the everted rat jejunum. *J Pharm Sci* 73(3):320–325
- Pons G, Blais J-C, Rey E, Plissonnier M, Richard M-O, Carrier O, D'Athis P, Moran Badoual J, Olive G (1988) Maturation of caffeine N-demethylation in infancy: a study using the <sup>13</sup>CO<sub>2</sub> breath test. *Pediatr Res* 23(6):632–636
- Powell JR, Thiercelin JF, Vozeh S, Sansom L, Riegelman S (1977) The influence of cigarette smoking and sex on theophylline disposition. *Am Rev Respir Dis* 116(1):17–23
- Rasmussen BB, Brix TH, Kyvik KO, Brøsen K (2002) The differences in the 3-demethylation of caffeine alias CYP1A2 is determined by both genetic and environmental factors. *Pharmacogenetics* 12:473–478
- Renner E, Wietholtz H, Huguenin P, Arnaud MJ, Preisig R (1984) Caffeine: a model compound for measuring liver function. *Hepatology* 4(1):38–46
- Resman BH, Blumenthal P, Jusko WJ (1977) Breast milk distribution of theobromine from chocolate. *J Pediatr* 91(3):477–480
- Rietveld EC, Broekman MM, Houben JJ, Eskes TK, van Rossum JM (1984) Rapid onset of an increase in caffeine residence time in young women due to oral contraceptive steroids. *Eur J Clin Pharmacol* 26(3):371–373

- Robson RA, Miners JO, Matthews AP, Stupans I, Meller D, McManus ME, Birkett DJ (1988) Characterisation of theophylline metabolism by human liver microsomes. Inhibition and immunochemical studies. *Biochem Pharmacol* 37(9):1651–1659
- Robson RA (1992) The effects of quinolones on xanthine pharmacokinetics. *Am J Med* 92 (4A):22S–25S
- Rodopoulos N, Norman A (1996) Assessment of dimethylxanthine formation from caffeine in healthy adults: comparison between plasma and saliva concentrations and urinary excretion of metabolites. *Scand J Clin Lab Invest* 56(3):259–268
- Rodopoulos N, Höjvall L, Norman A (1996) Elimination of theobromine metabolites in healthy adults. *Scand J Clin Lab Invest* 56(4):373–383
- Rodopoulos N, Norman A (1997) Elimination of theophylline metabolites in healthy adults. *Scand J Clin Lab Invest* 57(3):233–240
- Rosal R, Rodríguez A, Perdígón-Melón JA, Petre A, García-Calvo E, Gómez MJ, Agüera A, Fernández-Alba AR (2010) Occurrence of emerging pollutants in urban wastewater and their removal through biological treatment followed by ozonation. *Water Res* 44 (2):578–588
- Rost KL, Roots I (1994) Accelerated caffeine metabolism after omeprazole treatment is indicated by urinary metabolite ratios: coincidence with plasma clearance and breath test. *Clin Pharmacol Ther* 55(4):402–411
- Ryu JY, Song IS, Sunwoo YE, Shon JH, Liu KH, Cha JJ, Shin JG (2007) Development of the “Inje cocktail” for high-throughput evaluation of five human cytochrome P450 isoforms in vivo. *Clin Pharmacol Ther* 82(5):531–540
- Sachse KT, Jackson EK, Wisniewski SR, Gillespie DG, Puccio AM, Clark RS, Dixon CE, Kochanek PM (2008) Increases in cerebrospinal fluid caffeine concentration are associated with favorable outcome after severe traumatic brain injury in humans. *J Cereb Blood Flow Metab* 28(2):395–401
- Salomon G (1883) Über Paraxanthine und Heteroxanthin. *Ber Dtsch Chem Ges* 18:3406–3410
- Sarkar MA, Hunt C, Guzelian PS, Karnes HT (1992) Characterization of human liver cytochromes P-450 involved in theophylline metabolism. *Drug Metab Dispos* 20(1):31–37
- Sarkar MA, Jackson BJ (1994) Theophylline *N*-demethylations as probes for P4501A1 and P4501A2. *Drug Metab Dispos* 22(6):827–834
- Saruwatari J, Nakagawa K, Shindo J, Tajiri T, Fujieda M, Yamazaki H, Kamataki T, Ishizaki T (2002) A population phenotyping study of three drug-metabolizing enzymes in Kyushu, Japan, with use of the caffeine test. *Clin Pharmacol Ther* 72(2):200–208
- Sato J, Nakata H, Owada E, Kikuta T, Umetsu M, Ito K (1993) Influence of usual intake of dietary caffeine on single-dose kinetics of theophylline in healthy human subjects. *Eur J Clin Pharmacol* 44(3):295–298
- Sato S, Nakajima M, Honda A, Konishi T, Miyazaki H (2007) Pharmacokinetics of theophylline in Guinea pig tears. *Drug Metab Pharmacokinet* 22(3):169–177
- Schaad HJ, Renner EL, Wietholtz H, Arnaud MJ, Preisig R (1995) Caffeine demethylation measured by breath analysis in experimental liver injury in the rat. *J Hepatol* 22(1):82–87
- Schlaeffer F, Engelberg I, Kaplanski J, Danon A (1984) Effect of exercise and environmental heat on theophylline kinetics. *Respiration* 45(4):438–442
- Schmider J, Brockmüller J, Arold G, Bauer S, Roots I (1999) Simultaneous assessment of CYP3A4 and CYP1A2 activity in vivo with alprazolam and caffeine. *Pharmacogenetics* 9 (6):725–734
- Schrenk D (1998) Impact of dioxin-type induction of drug-metabolizing enzymes on the metabolism of endo- and xenobiotics. *Biochem Pharmacol* 5(8):1155–1162
- Scott NR, Chakraborty J, Marks V (1984) Determination of caffeine, theophylline and theobromine in serum and saliva using high-performance liquid chromatography. *Ann Clin Biochem* 21(Pt 2):120–124
- Scott NR, Chakraborty J, Marks V (1986) Urinary metabolites of caffeine in pregnant women. *Br J Clin Pharmacol* 22(4):475–478

- Scott NR, Stambuk D, Chakraborty J, Marks V, Morgan MY (1988) Caffeine clearance and biotransformation in patients with chronic liver disease. *Clin Sci* 74(4):377–384
- Scott NR, Stambuk D, Chakraborty J, Marks V, Morgan MY (1989) The pharmacokinetics of caffeine and its dimethylxanthine metabolites in patients with chronic liver disease. *Br J Clin Pharmacol* 27(2):205–213
- Shaw LM, Fields L, Mayock R (1982) Factors influencing theophylline serum protein binding. *Clin Pharmacol Ther* 32(4):490–496
- Shimada T, Yamazaki H, Mimura M, Inui Y, Guengerich FP (1994) Interindividual variations in human liver cytochrome P-450 enzymes involved in the oxidation of drugs, carcinogens and toxic chemicals: studies with liver microsomes of 30 Japanese and 30 Caucasians. *J Pharmacol Exp Ther* 270:414–423
- Shively CA, Tarka SM Jr (1983) Theobromine metabolism and pharmacokinetics in pregnant and nonpregnant Sprague-Dawley rats. *Toxicol Appl Pharmacol* 67(3):376–382
- Shively CA, Tarka SM Jr (1984) Methylxanthine composition and consumption patterns of cocoa and chocolate products. In: Spiller GA (ed) *The methylxanthine beverages in foods: chemistry, consumption, and health effects*. Alan R Liss, New York, pp 149–178
- Shively CA, Tarka SM Jr, Arnaud MJ, Dvorchik BH, Passananti GT, Vesell ES (1985) High levels of methylxanthines in chocolate do not alter theobromine disposition. *Clin Pharmacol Ther* 37(4):415–424
- Siegel IA, Ben-Aryeh H, Gozal D, Colin AA, Szargel R, Laufer D (1990) Comparison of unbound and total serum theophylline concentrations with those of stimulated and unstimulated saliva in asthmatic children. *Ther Drug Monit* 12(5):460–464
- Simons FE, Rigatto H, Simons KJ (1981) Pharmacokinetics of theophylline in neonates. *Semin Perinatol* 5(4):337–345
- Sjöberg P, Olofsson IM, Lundqvist T (1992) Validation of different microdialysis methods for the determination of unbound steady-state concentrations of theophylline in blood and brain tissue. *Pharm Res* 9(12):1592–1598
- Soyka LF, Neese AL, Main D, Main E (1981) Studies of caffeine and theophylline in the neonate. *Semin Perinatol* 5(4):332–336
- Sperber AD (1991) Toxic interaction between fluvoxamine and sustained release theophylline in an 11-year-old boy. *Drug Saf* 6(6):460–462
- Spigset O, Hägg S, Söderström E, Dahlqvist R (1999a) The paraxanthine:caffeine ratio in serum or in saliva as a measure of CYP1A2 activity: when should the sample be obtained? *Pharmacogenetics* 9(3):409–412
- Spigset O, Hägg S, Söderström E, Dahlqvist R (1999b) Lack of correlation between fluvoxamine clearance and CYP1A2 activity as measured by systemic caffeine clearance. *Eur J Clin Pharmacol* 54(12):943–946
- Stähle L (1991) Drug distribution studies with microdialysis. I. Tissue dependent difference in recovery between caffeine and theophylline. *Life Sci* 49(24):1835–1842
- Stähle L, Segersvärd S, Ungerstedt U (1991) Drug distribution studies with microdialysis. II. Caffeine and theophylline in blood, brain and other tissues in rats. *Life Sci* 49(24):1843–1852
- Statland BE, Demas TJ (1980) Serum caffeine half-lives. Healthy subjects vs. patients having alcoholic hepatic disease. *Am J Clin Pathol* 73(3):90–93
- Straka RJ, Burkhardt RT, Lang NP, Hadsall KZ, Tsai MY (2006) Discordance between N-acetyltransferase 2 phenotype and genotype in a population of Hmong subjects. *J Clin Pharmacol* 46(7):802–11
- Streetman DS, Bleakley JF, Kim JS, Nafziger AN, Leeder JS, Gaedigk A, Gotschall R, Kearns GL, Bertino JS Jr (2000) Combined phenotypic assessment of CYP1A2, CYP2C19, CYP2D6, CYP3A, N-acetyltransferase-2, and xanthine oxidase with the “Cooperstown cocktail”. *Clin Pharmacol Ther* 68(4):375–383
- Takata K, Saruwatari J, Nakada N, Nakagawa M, Fukuda K, Tanaka F, Takenaka S, Mihara S (2006) Phenotype-genotype analysis of CYP1A2 in Japanese patients receiving oral theophylline therapy. *Eur J Clin Pharmacol* 62(1):23–28



- Tanaka E, Ishikawa A, Yamamoto Y, Uchida E, Kobayashi S, Yasuhara H, Misawa S (1992a) Simplified approach for evaluation of hepatic drug-oxidizing capacity with a simultaneous measurement of caffeine and its primary demethylated metabolites in carbon tetrachloride-intoxicated rats. *Xenobiotica* 22(5):535–541
- Tanaka E, Ishikawa A, Yamamoto Y, Osada A, Tsuji K, Fukao K, Misawa S, Iwasaki Y (1992b) A simple useful method for the determination of hepatic function in patients with liver cirrhosis using caffeine and its three major dimethylmetabolites. *Int J Clin Pharmacol Ther Toxicol* 30(9):336–341
- Tanaka E, Ishikawa A, Yamamoto Y, Osada A, Tsuji K, Fukao K, Iwasaki Y (1993) Comparison of hepatic drug-oxidizing activity after simultaneous administration of two probe drugs, caffeine and trimethadione, to human subjects. *Pharmacol Toxicol* 72(1):31–33
- Tanaka E, Ishikawa A, Misawa S (1995) Changes in the metabolism of three model substrates catalysed by different P isozymes when administered as a cocktail to the carbon tetrachloride-intoxicated rat. *Xenobiotica* 25(10):1111–1118
- Tang BK, Grant DM, Kalow W (1983) Isolation and identification of 5-acetylamino-6-formylamino-3-methyluracil as a major metabolite of caffeine in man. *Drug Metab Dispos* 11(3):218–220
- Tang BK, Zubovits T, Kalow W (1986) Determination of acetylated caffeine by high-performance exclusion chromatography. *J Chromatogr* 375(1):170–173
- Tang BK, Zhou Y, Kadar D, Kalow W (1994) Caffeine as a probe for CYP1A2 activity: potential influence of renal factors on urinary phenotypic trait measurements. *Pharmacogenetics* 4(3):117–224
- Tang-Liu DD, Riegelman S (1981) Metabolism of theophylline to caffeine in adults. *Res Commun Chem Pathol Pharmacol* 34(3):371–380
- Tang-Liu DDS, Williams RL, Reigelman S (1983) Disposition of caffeine and its metabolites in man. *J Pharmacol Exp Ther* 224(1):180–185
- Tarka SM Jr (1982) The toxicology of cocoa and methylxanthines: a review of the literature. *Crit Rev Toxicol* 9(4):275–312
- Tarka SM Jr, Arnaud MJ, Dvorchik BH, Vesell ES (1983) Theobromine kinetics and metabolic disposition. *Clin Pharmacol Ther* 34(4):546–555
- Tassaneeyakul W, Birkett DJ, McManus ME, Tassaneeyakul W, Veronese ME, Andersson T, Tukey RH, Miners JO (1994) Caffeine metabolism by human hepatic cytochromes P450: contributions of 1A2, 2E1 and 3A isoforms. *Biochem Pharmacol* 47(10):1767–1776
- Teichmann AT (1990) Influence of oral contraceptives on drug therapy. *Am J Obstet Gynecol* 163(6 Pt 2):2208–2213
- Teunissen MW, Brorens IO, Geerlings JM, Breimer DD (1985) Dose-dependent elimination of theophylline in rats. *Xenobiotica* 15(2):165–171
- Tornatore KM, Kanarkowski R, McCarthy TL, Gardner MJ, Yurchak AM, Jusko WJ (1982) Effect of chronic oral contraceptive steroids on theophylline disposition. *Eur J Clin Pharmacol* 23(2):129–134
- Trang JM, Blanchard J, Conrad KA, Harrison GG (1985) Relationship between total body clearance of caffeine and urine flow rate in elderly men. *Biopharm Drug Dispos* 6(1):51–56
- Trnavská Z (1990) Theophylline protein binding. *Arzneimittelforschung* 40(2 Pt 1):166–169
- Tröger U, Meyer FP (1995) Influence of endogenous and exogenous effectors on the pharmacokinetics of theophylline focus on biotransformation. *Clin Pharmacokinet* 28(4):287–314
- Tserng KY, Takieddine FN, King KC (1983) Developmental aspects of theophylline metabolism in premature infants. *Clin Pharmacol Ther* 33(4):522–528
- Tsutsumi K, Kotegawa T, Matsuki S, Tanaka Y, Ishii Y, Kodama Y, Kuranari M, Miyakawa I, Nakano S (2001) The effect of pregnancy on cytochrome P4501A2, xanthine oxidase, and N-acetyltransferase activities in humans. *Clin Pharmacol Ther* 70(2):121–125
- Tukker JJ, Meulendijk AJ (1991) Is a diurnal rhythm in bioavailability caused by a rhythm in intestinal absorption? *Eur J Drug Metab Pharmacokinet Spec No* 3:66–70

- Upton RA (1991) Pharmacokinetic interactions between theophylline and other medication (Part I). *Clin Pharmacokinet* 20(1):66–80
- Vickroy TW, Chang SK, Chou CC (2008) Caffeine-induced hyperactivity in the horse: comparisons of drug and metabolite concentrations in blood and cerebrospinal fluid. *J Vet Pharmacol Ther* 31(2):156–166
- Vistisen K, Loft S, Poulsen HE (1991) Cytochrome P450 IA2 activity in man measured by caffeine metabolism: effect of smoking, broccoli and exercise. *Adv Exp Biol Biol* 283:407–411
- Vistisen K, Poulsen HE, Loft S (1992) Foreign compound metabolism capacity in man measured from metabolites of dietary caffeine. *Carcinogenesis* 13(9):1561–1568
- Walton K, Dorne JL, Renwick AG (2001) Uncertainty factors for chemical risk assessment: interspecies differences in the in vivo pharmacokinetics and metabolism of human CYP1A2 substrates. *Food Chem Toxicol* 39(7):667–680
- Warszawski D, Gorodischer R, Moses SW, Bark H (1977) Caffeine pharmacokinetics in young and adult dogs. *Biol Neonate* 32(3–4):138–142
- Warszawski D, Ben-Zvi Z, Gorodischer R, Arnaud MJ, Bracco I (1982) Urinary metabolites of caffeine in young dogs. *Drug Metab Dispos* 10(4):424–428
- Weinberger M, Ginchansky E (1977) Dose-dependent kinetics of theophylline disposition in asthmatic children. *J Pediatr* 91(5):820–824
- Wietholtz H, Voegelin M, Arnaud MJ, Bircher J, Preisig R (1981) Assessment of cytochrome P-448 dependent liver enzyme system by a caffeine breath test. *Eur J Clin Pharmacol* 21(1):53–59
- Wijnands GJ, Cornel JH, Martea M, Vree TB (1990) The effect of multiple-dose oral lomefloxacin on theophylline metabolism in man. *Chest* 98(6):1440–1444
- Wilkinson JM, Pollard I (1993) Accumulation of theophylline, theobromine and paraxanthine in the fetal rat brain following a single oral dose of caffeine. *Brain Res Dev Brain Res* 75(2):193–199
- Wu C, Witter JD, Spongberg AL, Czajkowski KP (2009) Occurrence of selected pharmaceuticals in an agricultural landscape, western Lake Erie basin. *Water Res* 43(14):3407–416
- Xu H, Rajesan R, Harper P, Kim RB, Lonnerdal B, Yang M, Uematsu S, Hutson J, Watson-MacDonell J, Ito S (2005) Induction of P 1A by cow milk-based formula: a comparative study between human milk and formula. *Br J Pharmacol* 146(2):296–305
- Yang KH, Lee JH, Lee MG (2008) Effects of CYP inducers and inhibitors on the pharmacokinetics of intravenous theophylline in rats: involvement of CYP1A1/2 in the formation of 1,3-DMU. *J Pharm Pharmacol* 60(1):45–53
- Yano I, Tanigawara Y, Yasuhara M, Mikawa H, Hori R (1993) Population pharmacokinetics of theophylline. I: Intravenous infusion to children in the acute episode of asthma. *Biol Pharm Bull* 16(1):59–62
- Yesair DW, Branfman AR, Callahan MM (1984) Human disposition and some biochemical aspects of methylxanthines. *Prog Clin Biol Res* 158:215–233
- Zandvliet AS, Huitema AD, de Jonge ME, den Hoed R, Sparidans RW, Hendriks VM, van den Brink W, van Ree JM, Beijnen JH (2005) Population pharmacokinetics of caffeine and its metabolites theobromine, paraxanthine and theophylline after inhalation in combination with diacetylmorphine. *Basic Clin Pharmacol Toxicol* 96(1):71–79
- Zevin S, Benowitz NL (1999) Drug interactions with tobacco smoking. An update. *Clin Pharmacokinet* 36(6):425–438
- Zhang Z-Y, Kaminsky LS (1995) Characterization of human cytochromes P450 involved in theophylline 8-hydroxylation. *Biochem Pharmacol* 50(2):205–211
- Zhu B, Ou-Yang DS, Chen XP, Huang SL, Tan ZR, He N, Zhou HH (2001) Assessment of cytochrome P450 activity by a five-drug cocktail approach. *Clin Pharmacol Ther* 70(5):455–461
- Zyset T, Wietholtz H (1991) Pharmacokinetics of caffeine in patients with decompensated type I and type II diabetes mellitus. *Eur J Clin Pharmacol* 41(5):449–452

# Inhibition of Cyclic Nucleotide Phosphodiesterases by Methylxanthines and Related Compounds

Sharron H. Francis, Konjeti R. Sekhar, Hengming Ke, and Jackie D. Corbin

## Contents

1	Identification of Biologically Active Alkylxanthines .....	94
1.1	History of Methylxanthines as Biological Stimulants and Inhibitors of Cyclic Nucleotide Phosphodiesterases .....	94
1.2	Superfamily of Mammalian PDEs .....	96
1.3	Development of Potent and Selective Alkylxanthine-Related Compounds for Inhibition of cN PDEs .....	99
1.4	Mode of Action of Methylxanthine-Related Compounds in Beverages, Foods, and Medications .....	100
2	Inhibition of cN PDEs by Alkylxanthine-Related Compounds .....	106
2.1	Mechanism of Action for Inhibition of PDEs by Derivatized Alkylxanthines .....	106
2.2	Molecular Basis for High Potency of Vardenafil over Sildenafil, Substrate, and Weak Inhibitors of PDE5: An Example .....	107
2.3	Concerns for Specificity of Derivatized Alkylxanthines as Inhibitors of PDEs .....	107
3	Design of Potent Xanthine-Based PDE5 Inhibitors That Recapitulate Structural Features of the Entire cGMP Molecule .....	108
3.1	Cyclic GMP and Cyclic AMP in Solution Exist in Equilibrium Between Two Conformations .....	109
3.2	Use of cN Analogs To Map cN-Binding Sites of cN-Dependent Protein Kinases .....	111
3.3	Use of cN Analogs To Map the Allosteric cGMP-Binding Sites and Catalytic Sites of PDEs .....	112
3.4	Design and Use of 8-Substituted IBMX Analogs for Potent Inhibition of PDE .....	114

---

S.H. Francis (✉) and J.D. Corbin

Department of Molecular Physiology and Biophysics, Vanderbilt University School of Medicine, Light Hall Room 702, Nashville, TN 37232-0615, USA

e-mail: sharron.francis@vanderbilt.edu

K.R. Sekhar

Department of Radiation Biology, Vanderbilt University School of Medicine, DD-1205 MCN, Nashville, TN 37232, USA

H. Ke

Department of Biochemistry and Biophysics, University of North Carolina at Chapel Hill, 306 Mary Ellen Jones Building, Chapel Hill, NC 27599-7260, USA

4	Insights into Interaction of PDEs with Alkylxanthine-Related Compounds .....	116
4.1	Interactions of IBMX and Other Xanthine-Related PDE Inhibitors with PDE Catalytic Sites as Determined by X-ray Crystallography .....	117
4.2	Use of Site-Directed Mutagenesis To Quantify the Impact of Amino Acid Contacts with Substrates and Xanthine-Related PDE Inhibitors .....	122
4.3	Use of cN Analogs to Probe Structural Elements; Comparison of Findings Using X-ray Crystallography .....	122
5	Concluding Remarks .....	123
	References .....	123

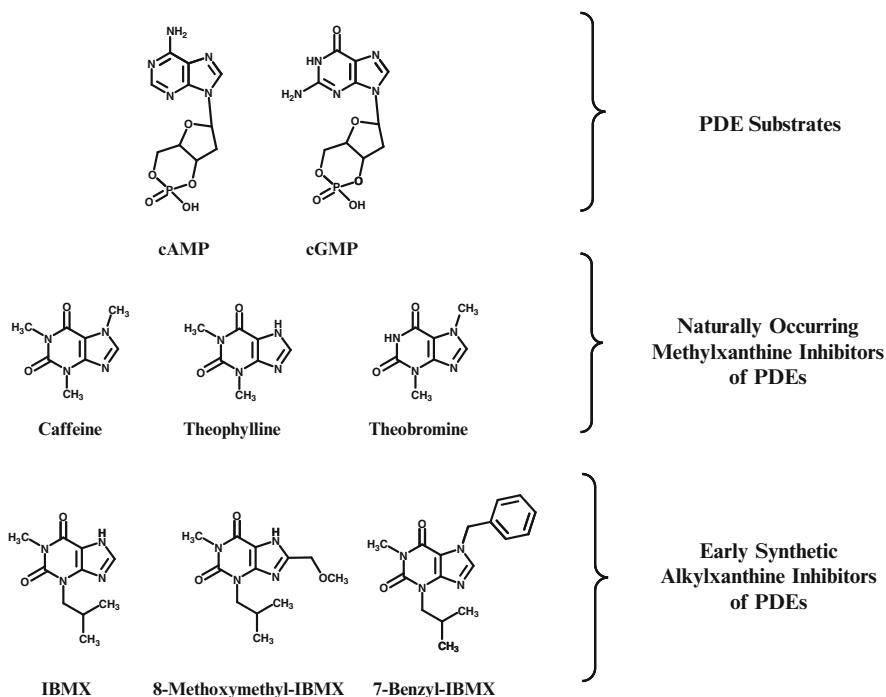
**Abstract** Naturally occurring methylxanthines were the first inhibitors of cyclic nucleotide (cN) phosphodiesterases (PDEs) to be discovered. To improve potency and specificity for inhibition of various PDEs in research and for treatment of diseases, thousands of compounds with related structures have now been synthesized. All known PDE inhibitors contain one or more rings that mimic the purine in the cN substrate and directly compete with cN for access to the catalytic site; this review focuses on inhibitors that contain a nucleus that is closely related to the xanthine ring of theophylline and caffeine and the purine ring of cNs. The specificity and potency of these compounds for blocking PDE action have been improved by appending groups at positions on the rings as well as by modification of the number and distribution of nitrogens and carbons in those rings. Several of these inhibitors are highly selective for particular PDEs; potent and largely selective PDE5 inhibitors are used clinically for treatment of erectile dysfunction [sildenafil (Viagra™), tadalafil (Cialis™) and vardenafil (Levitra™)] and pulmonary hypertension [sildenafil (Revatio™) and tadalafil (Adenocirca)]. Related compounds target other PDEs and show therapeutic promise for a number of maladies.

**Keywords** Phosphodiesterases · Cyclic AMP · Cyclic GMP · Theophylline · Xanthine · Caffeine · Phosphodiesterase inhibitors · Caffeine in beverages and foods

## 1 Identification of Biologically Active Alkylxanthines

### 1.1 *History of Methylxanthines as Biological Stimulants and Inhibitors of Cyclic Nucleotide Phosphodiesterases*

In 1958–1960 Earl Sutherland, winner of the Nobel Prize in Physiology or Medicine in 1971 for discovery of cyclic AMP (cAMP; Fig. 1), and his colleague Ted Rall identified caffeine (1,3,7-trimethylxanthine) (Fig. 1), a plant-derived alkaloid, as an inhibitor of cAMP breakdown by crude preparations of cyclic nucleotide (cN) phosphodiesterases (PDEs) (Sutherland and Rall 1958); this was the first PDE inhibitor to be identified and the first enzymatic effect of caffeine to be defined.



**Fig. 1** Molecular structures of cyclic nucleotides (cNs) and those of naturally occurring methylxanthine inhibitors of phosphodiesterases (PDEs) and representative early synthetic alkylxanthine PDE inhibitors

In their classical studies on the phosphorylation and activation of purified phosphorylase or phosphorylase in dilute liver homogenates, Wosilait and Sutherland (1956) discovered that dephosphorylation of phosphophosphorylase by an “inactivating enzyme” (later known as the LP-phosphatase activity) was increased in the presence of caffeine or theobromine; binding of the purine to an allosteric site on phosphorylase caused a conformational change that facilitated dephosphorylation. Consequently, caffeine was included in their protocols in order to promote dephosphorylation of phosphorylase, thereby lowering the blank that was due to the active phosphophosphorylase in tissue extracts (Berthet et al. 1957). However, in studies of epinephrine or glucagon action using liver homogenates, they found surprisingly that this effect of caffeine was observed only with low concentrations of caffeine and was quite modest; at higher caffeine concentrations, the amount of phosphophosphorylase actually increased. Moreover, they found that the effect of caffeine was greatest when low concentrations of glucagon or epinephrine were employed and that inclusion of caffeine in these experiments increased sensitivity to the hormones (Berthet et al. 1957). Having already established that the effect of glucagon or epinephrine was due to a heat-stable factor (cAMP), they concluded that there must be an enzyme (now known to be PDE) that destroyed the factor and

that this enzyme was inhibited by the caffeine in the reaction mixtures. Indeed, in unpublished experiments involving crude preparations from liver, brain, or heart, they found that caffeine enhanced accumulation of cAMP in these tissue extracts by blocking the PDE activity that breaks down cAMP. Thus, at low glucagon or epinephrine concentrations, caffeine acted to synergistically increase the cAMP level, thereby increasing the activities cAMP-dependent protein kinase (PKA) and phosphorylase kinase activities, resulting in increased amounts of active phosphophosphorylase (Berthet et al. 1957). However, the exact details describing the insights that led Sutherland and Rall to the realization that caffeine might act to block PDE action and foster cAMP accumulation are not known (we are grateful to Bill Butcher for his historical insights). In 1962, Butcher and Sutherland (1962) demonstrated that theophylline (1,3-dimethylxanthine) and theobromine (Fig. 1) also inhibit PDE action; in that instance, the inhibitory potencies of caffeine and theobromine were comparable, whereas theophylline was approximately six-fold more potent (Butcher and Sutherland 1962).

The stimulatory effects of plant extracts containing caffeine and theophylline had long been appreciated, but their mode of action was not understood (Fredholm 2010). Historical accounts suggest that about 3,000 years ago Ethiopian herders observed that goats that foraged on beans of coffee plants in the daytime were restless and wakeful in the evening. They subsequently found that consumption of extracts from these beans increased their own wakefulness; that was the discovery of coffee, and the pharmacological action that they experienced was due to the stimulant effect of caffeine in the beverage. Tea, which contains caffeine and theophylline, both of which are stimulants, is thought to have been discovered in China in more distant antiquity. Caffeine along with theobromine (3,7-dimethylxanthine) is also found in cocoa beans and is an additive in many foodstuffs; paraxanthine (1,7-dimethylxanthine), theophylline, and theobromine, natural breakdown products of caffeine (Arnaud 2010), are present in the body after caffeine consumption and elicit a variety of biological effects (Fig. 1) (Guerreiro et al. 2010; Smit 2010; Müller and Jacobson 2010).

## 1.2 Superfamily of Mammalian PDEs

Investigators conducting early studies on the effects of caffeine to inhibit PDEs suspected that there were multiple types of PDEs in a given tissue extract, and that was soon proven to be the case (Sutherland and Rall 1958). It is now known that there are 21 genes for PDEs in the human genome (Bender and Beavo 2006). The protein products of these genes comprise the superfamily of PDEs that has been subdivided into 11 families (PDEs 1–11) (Table 1); some families are encoded by a single gene, whereas others are products of multiple genes. Further complexity results from extensive alternative splicing of the messenger RNA to produce a vast array of PDEs with different regulatory features, catalytic characteristics, tissue

**Table 1** Substrate specificities and kinetic characteristics of mammalian phosphodiesterase (PDE) families

Isoenzyme	Substrate specificity	$K_m$ ( $\mu\text{M}$ )		$V_{\text{max}}$ ( $\mu\text{mol}/\text{min}/\text{mg}$ )		References
		cGMP	cAMP	cGMP	cAMP	
PDE1A	cAMP < cGMP	3–4	73–120	50–300	70–450	Hansen et al. (1988), Sharma et al. (1984), Snyder et al. (1999), Sonnenburg et al. (1995)
PDE1B	cAMP < cGMP	1–6	10–24	30	10	Bender et al. (2005), Sharma and Wang (1986)
PDE1C	cAMP = cGMP	1–2	0.3–1	?	?	Loughney et al. (1996), Yan et al. (1996)
PDE2A	cAMP = cGMP	10	30	123	120	Martins et al. (1982), Rosman et al. (1997)
PDE3A	cAMP > cGMP	0.02–0.2	0.2	0.3	3–6	Grant and Colman (1984), Harrison et al. (1986)
PDE3B	cAMP > cGMP	0.3	0.4	2	9	Degerman et al. (1987)
PDE4A	cAMP >> cGMP	?	3–10	?	0.6	Rena et al. (2001), Salanova et al. (1998), Wallace et al. (2005), Wang et al. (1997)
PDE4B	cAMP >> cGMP	?	2–5	?	0.1	Huston et al. (1997), Salanova et al. (1998), Wang et al. (1997)
PDE4C	cAMP >> cGMP	?	2	?	0.3	Wang et al. (1997)
PDE4D	cAMP >> cGMP	?	1–6	?	0.03–2	Salanova et al. (1998), Wang et al. (1997)
PDE5A	cGMP >> cAMP	1–6	90	1–3	1–3	Loughney et al. (1998), Thomas et al. (1990), Turko et al. (1998), Zoraghi et al. (2006)
PDE6A/B	cGMP >> cAMP	15	700	2,300	?	Gillespie and Beavo (1988), Zhang et al. (2005)
PDE6C	cGMP >> cAMP	17	610	1,400	?	Gillespie and Beavo (1988), Zhang et al. (2005)
PDE7A	cAMP >> cGMP	?	0.1–0.2	?	?	Han et al. (1997), Michaeli et al. (1993)
PDE7B	cAMP >> cGMP	?	0.03–0.1	?	?	Hetman et al. (2000), Sasaki et al. (2000, 2002)
PDE8A	cAMP >> cGMP	?	0.1	?	?	Fisher et al. (1998)
PDE8B	cAMP >> cGMP	?	0.1	?	?	Gamanuma et al. (2003)
PDE9A	cGMP >> cAMP	0.2–0.7	230	?	?	Fisher et al. (1998), Soderling et al. (1998)

*(continued)*

**Table 1** (continued)

Isoenzyme	Substrate specificity	$K_m$ ( $\mu\text{M}$ )		$V_{\text{max}}$ ( $\mu\text{mol}/\text{min}/\text{mg}$ )		References
		cGMP	cAMP	cGMP	cAMP	
PDE10A	cAMP > cGMP	13	0.2–1	?	?	Fujishige et al. (1999), Kotera et al. (1999)
PDE11A	cAMP = cGMP	0.4–2	0.5–3	?	?	Fawcett et al. (2000), Weeks et al. (2007), Yuasa et al. (2001)

The results are compiled from listed references and reproduced with modifications and permission (Bender and Beavo 2006). The range of concentrations indicates differences among different laboratories. These are likely to be due to different assay conditions and different preparations of the respective enzymes. *Question marks* indicate no reliable data are currently available  
*cAMP* cyclic AMP, *cGMP* cyclic GMP

distribution, and subcellular localizations. At present there are estimated to be about 100 PDEs derived from the 21 PDE genes and additional forms are being identified with some regularity (Bender and Beavo 2006; Conti and Beavo 2007).

All mammalian PDEs share a common catalytic domain that is located toward the carboxyl-terminal portion of the proteins and comprises about 270 amino acids; despite the fact that the identity of sequence among catalytic domains of PDEs varies from 24–50%, all X-ray crystal structures of these domains reveal a very similar overall structure that is composed primarily of  $\alpha$ -helices and a similar catalytic pocket (Ke and Wang 2007). The catalytic pocket where the cNs or inhibitors bind occupies about  $330 \text{ \AA}^3$  and contains two divalent cations, which is typically a tightly bound zinc and another more loosely bound (likely a magnesium or manganese) metal that are required for hydrolysis of the cyclic phosphate bond. Some PDEs are highly specific for hydrolysis of either cAMP (PDEs 4, 7, and 8) or cyclic GMP (cGMP) (PDEs 5, 6, and 9), whereas others readily hydrolyze both nucleotides (PDEs 1–3, 10, and 11).

While all known PDE catalytic domains interact to some extent with methylxanthines and related compounds, there is a very wide range of affinities (Table 2); methylxanthine-related compounds that are profoundly potent inhibitors for certain PDEs are ineffective for inhibiting other PDEs (Beavo et al. 2006). Moreover, members of the PDE8 and PDE9 families are not significantly inhibited by methylxanthine inhibitors such as 3-isobutyl-1-methylxanthine (IBMX) that are commonly described as “nonspecific” and are used in many tissue studies in an effort to block PDE action (Lavan et al. 1989). The affinities of the various PDEs for IBMX vary by more than 100-fold, for zaprinast by more than 1,000-fold, for sildenafil by more than 7,000-fold, and for vardenafil by more than 300,000-fold (Table 2). Thus, despite the overall similarities in the catalytic pockets of these enzymes, there are elements that strongly discriminate among these compounds.

Although PDEs in general are in low abundance in cells, almost all cells contain multiple PDEs, all of which contribute important regulatory control of cNs in that



**Table 2** Comparison of inhibitory potencies ( $IC_{50}$ ) of various methylxanthine-related compounds for mammalian PDE families

Isoenzyme	IBMX ( $\mu$ M)	Zaprinast ( $\mu$ M)	Sildenafil ( $\mu$ M)	Vardenafil ( $\mu$ M)
PDE1	3–10	6	0.28	0.07
PDE2	6–50	NA	>30	6.2
PDE3	2–10	NA	16	>1
PDE4	5–20	NA	7.7	6.1
PDE5	2–10	0.13–0.8	0.004	0.0001–0.0004
PDE6 <sup>a</sup>	1–5	0.03	0.005–0.01	0.0003–0.0007
PDE7	2	NA	21	>30
PDE8	>100	NA	30	>30
PDE9	>100	35	2.6	0.6
PDE10	3	11–22	~1	3.0
PDE11	25–80	11–33	2.7	0.16

The range of values in many instances reflects measurements from different laboratories using different assay conditions, substrate concentrations, etc. Values for sildenafil and vardenafil inhibition of PDEs 1–5 and 7–11 (Ballard et al. 1998; Corbin and Francis 2002; Gibson 2001) *IBMX* 3-isobutyl-1-methylxanthine, *NA* not available

Values for IBMX potency of inhibition for PDEs 1–5 (Dent and Rabe 1996). Values for zaprinast inhibition of PDEs 9–11 (Gibson 2001; Nakamizo et al. 2003)

<sup>a</sup>Values for PDE6 (Zhang et al. 2005) and the range of concentrations reflect the potency of inhibition of rod PDE6 and cone PDE6

particular cell. These individual contributions result from the substrate selectivity and affinity for a given cN, regulatory features of the PDEs (e.g., effects of phosphorylation, allosteric cN binding, calcium/calmodulin), and subcellular localization. Representatives of particular PDE families (PDEs 1–4), are widespread and relatively abundant in mammalian tissues, whereas others (PDEs 5–11) occur in lower abundance and have a more confined distribution. Nevertheless, particular PDEs that are low in overall abundance in a particular tissue may occur in high abundance in particular regions of a cell and can thereby still significantly impact physiological and pharmacological responses. Some PDEs (PDEs 2, 5, 6, 10, and 11) contain subdomains within their regulatory domains that can bind either cGMP (PDEs 2, 5, 6, and 11) or cAMP (PDE 10), and direct regulation of enzyme function by these allosteric sites has been demonstrated for PDEs 2, 5, and 6 (Bender and Beavo 2006; Conti and Beavo 2007). These sites are evolutionarily distinct from the PDE catalytic sites and do not interact appreciably with caffeine, theophylline, or any of the other known PDE inhibitors.

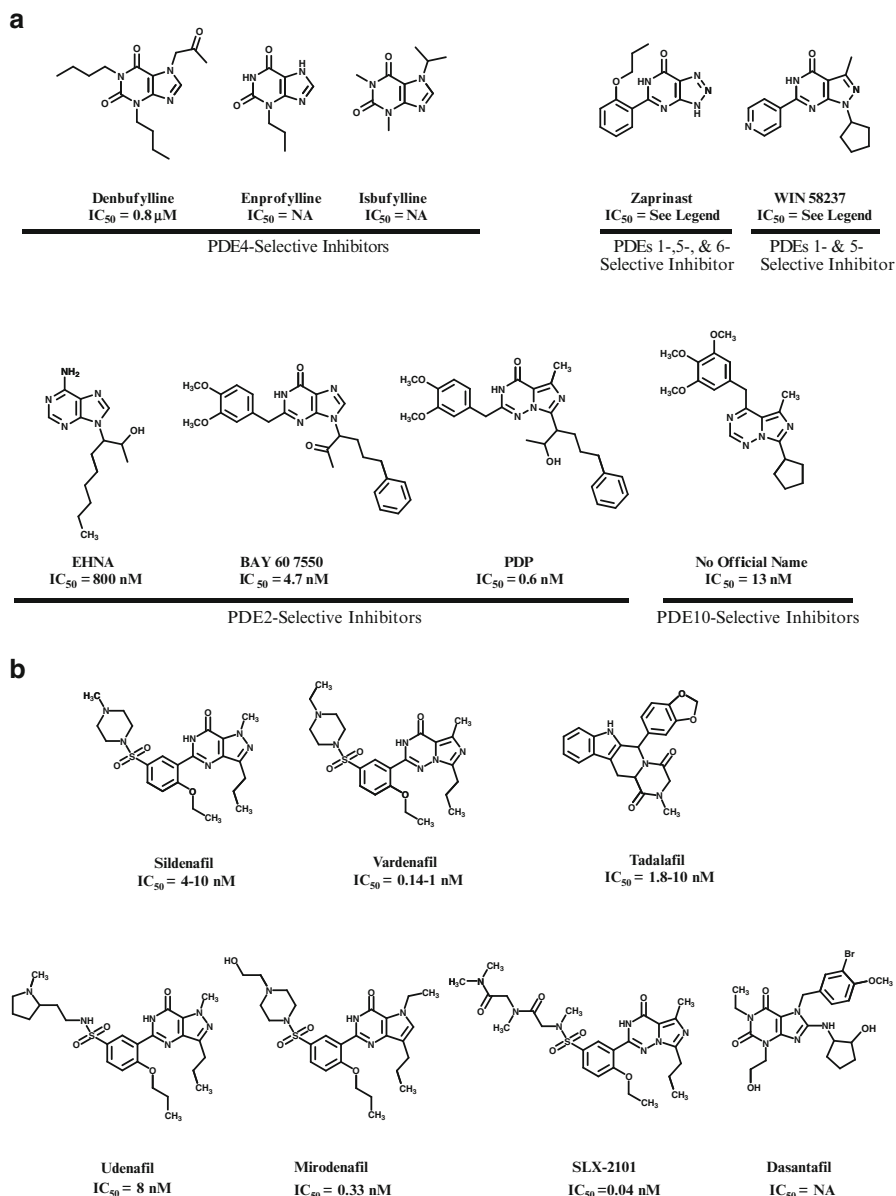
### 1.3 Development of Potent and Selective Alkylxanthine-Related Compounds for Inhibition of cN PDEs

Until the early 1970s, naturally occurring methylxanthines constituted the majority of compounds that were available for use as PDE inhibitors in research focused on cN signaling. Caffeine and theophylline weakly inhibit most cN PDEs, with  $IC_{50}$

values ranging from 100 to 1,000  $\mu\text{M}$  (Beavo et al. 2006; Choi et al. 1988; Dent and Rabe 1996; Smellie et al. 1979), and show little selectivity among PDE families (PDEs 1–11). However, it was shown early on that very modest (less than 20%) inhibition of crude PDE activity by alkylxanthines resulted in a significant increase in lipolysis (Beavo et al. 1970). To improve potency and selectivity of these inhibitors, chemists synthesized a collection of alkylxanthines; IBMX was one of the first produced, and a number of IBMX analogs with substituents appended at N7 and C8 quickly ensued (Fig. 1). These were valuable tools for studying cN signaling and for defining characteristics of different PDEs (Fig. 1) (Garst et al. 1976; Mushlin et al. 1981; Wells and Kramer 1981). Chemists in both academia and in the pharmaceutical industry eventually synthesized thousands of compounds that were in the same general family as caffeine and theophylline, i.e., either alkylxanthines or closely related compounds (Fig. 2). (Boolell et al. 1996; Boyle et al. 2005; Buckle et al. 1994; Corbin and Francis 1999; Kaplan et al. 1995; McKenna and Muller 2006; Miyamoto et al. 1993, 1994; Sekhar et al. 1996; Silver et al. 1994; Wang et al. 2002). These efforts produced compounds with greatly improved inhibitory potency for PDEs and, in certain instances, greatly improved selectivity among PDEs; this collection continues to evolve and includes pentoxifylline (a nonselective inhibitor) (Tjon and Riemann 2001), enprofylline, isbufylline, denbufylline (PDE4 inhibitors), 8-substituted-IBMX analogs (primarily PDE1 and PDE5 inhibitors), zaprinast (a PDE1, PDE5, and PDE6 inhibitor), WIN 58237 (a PDE1/PDE5 inhibitor), *erythro*-9-(2-hydroxyl-3-nonyl)-adenine (EHNA; a PDE2 inhibitor) (Fig. 2a), and a group of potent PDE5 inhibitors including sildenafil and vardenafil [both of which are also potent for PDE6 (about threefold to tenfold lower potency than for PDE5)], tadalafil, udenafil, mirodenafil, SLX-2101, and dasantafil. (Beavo et al. 2006; Francis et al. 2009) (Fig. 2b). As interest in therapeutic use of PDE inhibitors increased, attention was also directed to modifications that improved the pharmacokinetics of these compounds, including considerations of the rate of absorption, stability in the gastrointestinal tract, metabolism, and clearance.

#### ***1.4 Mode of Action of Methylxanthine-Related Compounds in Beverages, Foods, and Medications***

Despite wide use of theophylline and caffeine in beverages, foods, and medications, the exact mode and site of action of these compounds and their metabolites are still not fully understood. In the 1960s, PDE inhibition was thought to mediate the pharmacological effects of caffeine or theophylline, and evidence suggests that a number of the effects of these compounds are indeed mediated via this process (Mascali et al. 1996; Mushlin et al. 1981; Rabe et al. 1995; Sullivan et al. 1994a, b; Torphy et al. 1992). However, it is now appreciated that there are numerous modes of actions of these compounds and their metabolites. Many actions of theophylline



**Fig. 2** Molecular structures of xanthine-related PDE inhibitors. **a** Xanthine-related inhibitors that are selective for particular PDE families; **b** Xanthine-related compounds that are highly potent and selective for PDE1. Approximate  $IC_{50}$  of zaprinast for PDE1 (6  $\mu M$ ) (Beavo et al. 2006), PDE5 (0.33  $\mu M$ ) (Zhang et al. 2005), and PDE6 (0.03  $\mu M$ ) (Zhang et al. 2005) and  $IC_{50}$  of WIN 58237 for PDE1 (1.5  $\mu M$ ) and PDE5 (0.17  $\mu M$ ) (Silver et al. 1994). *NA* not available

and caffeine are mediated by antagonism of adenosine effects on adenosine receptors ( $A_1$ ,  $A_{2A}$ , and  $A_{2B}$ ), where they have modest affinity (Sawynok and Yaksh 1993; Yang et al. 2009a, b; Müller and Jacobson 2010). Blockade of the  $A_1$  receptor by these compounds accounts for their well-known natriuretic and diuretic effects (Rieg et al. 2005) (see Osswald and Schnermann 2010) and certain neurological effects such as wakefulness and increased alertness may be due, at least in part, to effects on one of the adenosine receptor isoforms (Boutrel and Koob 2004; Huang et al. 2005; Solinas et al. 2002; Yu et al. 2009) (see Porkka-Heiskanen 2010). Paraxanthine, the primary metabolite of caffeine, acts through the ryanodine receptor to elevate intracellular calcium levels and increases viability of neuronal cells in culture (Guerreiro et al. 2008, 2010). Therapeutic concentrations of theophylline also increase histone deacetylase activity, apparently via an indirect mechanism, thereby reducing inflammatory processes (Haskó and Cronstein 2010; Ohta and Sitkovsky 2010) (Barnes 2003a; Ito et al. 2002; Marwick et al. 2008).

#### 1.4.1 Role of PDE Inhibition in Effects of Caffeine Derived from Beverages and Foods

Surprisingly, caffeine is found in many beverages and foods; in some instances, this caffeine is derived from the source itself, whereas in other instances, it is added. The possibility that caffeine may act in part through inhibition of PDEs is increased following consumption of caffeine-rich “energy drinks,” foods, and gum alone, in combination, or in conjunction with caffeine-containing medications. Energy drinks are commonly ingested over a short period and can contain 6–14 times the amount of caffeine in a cup of coffee based on the number of milligrams per ounce (Table 3). Following consumption of beverages containing caffeine (about five or six cups of coffee daily) or therapeutic doses of caffeine, the plasma caffeine concentration is usually 10–50  $\mu\text{M}$  (Benowitz 1990) although the level and persistence of caffeine and its metabolites in plasma vary widely owing to many factors. Caffeine is quickly converted to paraxanthine (84%), theobromine (12%), and theophylline (4%). Generally, caffeine inhibition of PDEs requires higher levels (100–1,000  $\mu\text{M}$ ) than for interaction with adenosine receptors (10–100  $\mu\text{M}$ ), but the potencies of its metabolites, paraxanthine and theobromine, for inhibition of most known PDEs have not been studied (Butcher and Sutherland 1962; Sattin 1971; Sattin and Rall 1970). Depending on the serving size, the caffeine content of beverages and/or food, and the variation in clearance times, it is entirely plausible that plasma and cellular caffeine levels could be within the range for pharmacological action on PDEs (Chou and Bell 2007). It is also well established that the intracellular concentration/action of a PDE inhibitor cannot be confidently predicted strictly on the basis of its extracellular concentration (Thompson 1991).

The caffeine contents of a number of common foods, certain gums, and mints are shown in Table 4. In most instances, individuals are unaware of their daily caffeine

**Table 3** Compilation of caffeine content in popular beverages

Beverage	Weight (oz)	Total caffeine (mg)	Ratio (mg/oz)
Fixx Extreme	0.17	400	2,352
Ammo	1	171	171
Redline Power Rush	2.5	350	140
Mana Energy Potion	1.4	160	119
Extreme Energy 6-Hour Shot	2	220	110
Jolt Endurance Shot	2	200	100
Powershot	1	100	100
Charge! Super Shot	2	200	100
Fuel Cell	2	180	90
Stok Black Coffee Shots	0.4	40	91
Upshot	2.5	200	80
NOS Powershot	2	125	63
925 Energy Shot	2	120	60
SLAM Energy Drink	2	107	54
SPIKE shooter	8.4	300	36
Cocaine Energy Drink	8.4	280	33
Redline Princess	8	250	31
Starbucks Short Coffee	8	180	23
Wired X344	16	344	22
Dark Chocolate	1	20	20
Brewed Coffee	8	108	13
Bookoo Energy	24	360	15
Vamp	16	240	15
Rockstar Roasted	15	225	15
Dopamine Energy Drink	8.4	120	14
Arizona Caution Energy Drink	16	200	13
No Fear	16	174	11
Red Bull	8	80	10
Monster	16	160	10
Rockstar	16	160	10
Brewed tea	8	60	8
Bawls	10	67	7
Afri-Cola	12	89	7
Diet Pepsi Max	12	69	6
Mountain Dew	12	54	5
Diet Coke	12	45	4
Dr. Pepper	12	41	3
Coca Cola Classic	12	35	3
Pepsi-Cola	12	38	3

Values in this table were compiled from Energyfiend.com (Energyfiend.com 2005a)

consumption and the amount of caffeine in their beverages or foods. Use of caffeine-containing energy drinks, in their beverages or foods coffee, tea, or caffeine-rich foods in combination with medications containing caffeine or theophylline can significantly increase the risk of side effects due to higher plasma levels of these compounds. Certain “buzz beers” contain 50–60 mg of caffeine per 250 mL and combine the effects of alcohol, a depressant, and caffeine, a stimulant, both of which promote diuresis and potential dehydration (O’Brien et al. 2008).

**Table 4** Compilation of caffeine content in representative foods

Food	Caffeine (mg)	Serving
Crackheads 2	600	Per box
Dark Chocolate Coated Coffee Beans	311	200 cal
Milk Chocolate Coated Coffee Beans	291	200 cal
Cocoa, Dry Powder, Unsweetened	202	200 cal
NRG Potato Chips	175	1.75-oz bag
Engobi	140	Per ounce
Alien Energy Jerky	110	Per pack
Foosh Energy Mints Powershot	100	Per mint
Buzz Bites Chocolate Chews	100	Per chew
Go Fast Energy Gum	100	Per piece
Jet Alert	100	Per tablet
Edy's Grand Espresso Chip	90	Per cup (8 oz)
Ben & Jerry's Coffee Heath Bar Crunch	84	Per cup (8 oz)
Butterfinger Buzz	80	Per Pkg <sup>-1</sup>
Ben & Jerry's Fair Trade Coffee Ice Cream	70	Per cup (8 oz)
Ben & Jerry's coffee flavored ice cream	68	Per cup (8 oz)
Morning Spark Energy Instant Oatmeal	60	Per packet
Blitz Energy Gum	55	Per piece
Jelly Belly Extreme Sports Beans	50	Per 1-oz bag
Starbucks Coffee Ice Cream	50–60	Per cup (8 oz)
Haagen-Dazs coffee ice cream	48	Per cup (8 oz)
Hershey's Special Dark Chocolate Bar	31	1.45 oz
Breyer's All Natural Coffee	30	Per cup (8 oz)
Semisweet Chocolate	26	200 cal
Chocolate-Coated Graham Crackers	19	200 cal
Milky Way Midnight Bar	14	200 cal
Quaker Cocoa Blasts	11	200 cal
Chocolate Flavored Puddings	10	200 cal
Chocolate Flavored Lite Syrup	8	200 cal
Frozen Chocolate Yogurt	4–6	200 cal
Tootsie Roll	4	200 cal
Snickers Marathon Energy Bar	3	200 cal

Values in this table were compiled from Energyfiend.com (Energyfiend.com 2005b)

#### 1.4.2 Role of PDE Inhibition by Theophylline and Caffeine in Medications

In particular, caffeine is included in many prescription and over-the-counter medicines; common over-the-counter drugs that contain caffeine include NoDoz<sup>®</sup> (200 mg/tablet), Vivarin<sup>®</sup> (200 mg/tablet), Excedrin Extra Strength<sup>®</sup> (65 mg/tablet), Anacin Maximum Strength<sup>®</sup> (32 mg/tablet), and Midol Menstrual Complete<sup>®</sup> (60 mg/tablet) (Interest 2007).

Theophylline either alone or in combination with other medications has been used clinically in the Western world for more than 60 years for treatment of bronchospasm associated with asthma or chronic obstructive pulmonary disease (COPD) (Barnes 2003a, b, 2005; Barnes and Stockley 2005; Tilley 2010), and

the anti-inflammatory effects of theophylline in airways are well established (Barnes 2003a; Dent et al. 1994; Dent and Rabe 1996). Much of the credit for use of methylxanthines in treatment of asthma has been given to Herrmann and Greene (Greene et al. 1937; Herrmann et al. 1937), who studied the effectiveness of theophylline in treating this malady. However, a treatise in the *Edinburgh Medical Journal* in 1859 by a Scottish physician, H. Salter, who was both a medical expert on asthma and himself an asthmatic stated that “One of the commonest and best-reputed remedies of asthma, one that is almost sure to have been tried in any case that may come under our observation, and one that in many cases is more efficacious than any other is strong coffee” (Salter 1859). Salter “prescribed” the use of this approach by instructing patients to ingest several cups of very strong black coffee preferably in the morning on an empty stomach. The relief Salter experienced was most likely attributable to the pharmacological action of the caffeine in the coffee, but whether this effect was mediated by inhibition of PDEs is unclear.

Systematic studies of methylxanthines found that relatively high plasma concentrations of these compounds are required to achieve significant bronchodilation; in the case of theophylline, 10–20 mg/L, which translates to about 50–100  $\mu\text{M}$ , is required (Barnes 2006). On the basis of  $\text{IC}_{50}$  values of theophylline for a number of PDEs, these levels could significantly diminish PDE activity and cause elevation of the levels of cNs (Butcher and Sutherland 1962; Dent and Rabe 1996). Evidence suggests that in airway smooth muscle the effects of theophylline are mediated through inhibition of PDEs 3–5 to cause increases in the levels of cAMP and cGMP and activation of signaling pathways for these cNs (Rabe et al. 1995). In industrialized countries, theophylline, either alone or in combination with other medications, is still in limited use in a subset of asthma patients, but more effective medications with fewer adverse side effects are preferred. Despite its narrow therapeutic window and problems in determining due to appropriate dosages for treatment of different patients, theophylline is still widely used in developing countries for treatment of asthma due to its low cost (Barnes 2003a, b). The effects of theophylline to blunt airway inflammation in COPD occur below 10 mg/mL plasma (Hirano et al. 2006; Kobayashi et al. 2004) and are therefore unlikely to act through PDE inhibition.

### 1.4.3 Role of PDE Inhibition in Effects of Derivatized Alkylxanthine Medications

Pentoxifylline (Trental<sup>TM</sup>, Sanofi-Aventis), a derivatized alkylxanthine inhibitor of PDEs 1, 2, and 4 (Fig. 2a), entered the market in the 1970s for treatment of symptoms related to intermittent claudication, which is associated with chronic occlusive arterial disease of the limbs (Tjon and Riemann 2001). It reportedly improves peripheral perfusion by decreasing blood viscosity and increasing erythrocyte deformability, but its mode of action and therapeutic efficacy are debated (Aviado and Porter 1984; Regensteiner and Hiatt 2002). Sildenafil (Viagra<sup>TM</sup>),

a pyrazolopyrimidinone, and vardenafil (Levitra™), an imidazotriazinone, are highly selective and potent inhibitors of PDEs 5 and 6, both of which specifically hydrolyze cGMP (Cote 2006; Francis et al. 2006). The heterocyclic ring structures of these inhibitors closely mimic those of theophylline and caffeine, and both are highly successful in treatment of erectile dysfunction (Carson and Lue 2005; Corbin and Francis 1999; Francis and Corbin 2003, 2005a, b; Stief et al. 2004). Sildenafil (marketed as Revatio™) is also used for treatment of some forms of pulmonary hypertension (Galie and Branzi 2005; Ghofrani et al. 2006); each of the approved PDE5 inhibitors, as well as newly available PDE5 inhibitors (Fig. 2b), are continually being tested for use in treatment of other maladies (Al-Ameri et al. 2009; Black et al. 2008; Burnett et al. 2006; Gotshall et al. 2009; Jeong et al. 2008; Kumar et al. 2009; Levien 2006; Lubamba et al. 2008; Medeiros et al. 2008; Puzzo et al. 2008; Rutten et al. 2009; Salloum et al. 2006; Stief et al. 2008). To date, most of the known biological effects of these inhibitors are mediated through inhibition of cGMP breakdown at the PDE5 catalytic site; visual perturbations in some patients who take sildenafil or vardenafil are attributed to inhibition of PDE6 in photoreceptor cells (Francis and Corbin 2003). Both sildenafil and vardenafil have outstanding safety records, with reports of only minor and transient side effects. A number of closely related compounds (udenafil, mirodenafil, SLX-2101, dasantafil, and avanafil) have either entered or are poised to enter the market (Doh et al. 2002; Hatzimouratidis 2008).

## 2 Inhibition of cN PDEs by Alkylxanthine-Related Compounds

### 2.1 *Mechanism of Action for Inhibition of PDEs by Derivatized Alkylxanthines*

The general structures for cNs and the derivatized alkylxanthine inhibitors, which are competitive inhibitors, are similar (Figs. 1, 2). The heterocyclic ring of these inhibitors, which mimics the purine of cNs, comprises a six-membered pyrimidine ring conjoined with a five-membered ring containing two or more nitrogens. The affinity of a particular PDE for the various inhibitors is dictated by (1) the chemical characteristics of the particular heterocyclic ring, (2) groups appended to the ring, (3) differences in the distribution of electrons within the ring, and (4) structural restrictions of PDE catalytic sites for entry and binding of derivatized alkylxanthines with particular molecular structures. Substituents appended to the ring can impact affinity for the various PDE catalytic sites by forming new contacts with regions in and around the catalytic sites of PDEs or by interference (steric or chemical) with these regions. The distribution and number of carbons and nitrogens within the ring system can alter the electron distribution, thereby impacting the strength of the interactions (Corbin et al. 2006; Erneux et al. 1984).



## ***2.2 Molecular Basis for High Potency of Vardenafil over Sildenafil, Substrate, and Weak Inhibitors of PDE5: An Example***

Despite vardenafil and sildenafil appearing to have very similar chemical structures (Fig. 2b), the potency of vardenafil for inhibition of the PDE5 catalytic site ( $K_i \sim 0.1\text{--}0.4$  nM) exceeds that of sildenafil ( $K_i \sim 4$  nM) by 10–40-fold; the affinity of the PDE5 catalytic site for cGMP is 6,000–24,000 times weaker ( $K_m \sim 2,500$  nM) than that for vardenafil. The potency of caffeine or theophylline for the PDE5 catalytic site is approximately 10-million to 40-million-fold and 600,000–2,400,000 times weaker, respectively, than that of vardenafil (Francis and Corbin 2003). The strong preference of the PDE5 catalytic site for vardenafil, a potent inhibitor, compared with that for weak inhibitors such as caffeine and theophylline is determined by unique structural features of both the inhibitor and the PDE5 catalytic site (Blount et al. 2006; Corbin et al. 2004, 2006; Sung et al. 2003; Wang et al. 2008b). The 10 to 40-fold difference in potency of vardenafil and sildenafil is largely due to features of the respective heterocyclic rings (Corbin et al. 2004, 2006), which differ in having either a carbon or a nitrogen at two positions in the five-membered ring. This difference substantially alters the electron distribution in the rings and the electronegativity of individual nitrogens (Corbin et al. 2006). Moreover, higher affinity of the PDE5 catalytic site for vardenafil requires structural features in the regulatory domain (Blount et al. 2006) since in the absence of this domain the potencies of vardenafil and sildenafil are the same. Vardenafil, like sildenafil, is also quite selective for inhibition of PDE5 compared with other PDEs (Francis and Corbin 2003) owing to the fact that their novel structures exploit unique features in PDE5 (Blount et al. 2006).

## ***2.3 Concerns for Specificity of Derivatized Alkylxanthines as Inhibitors of PDEs***

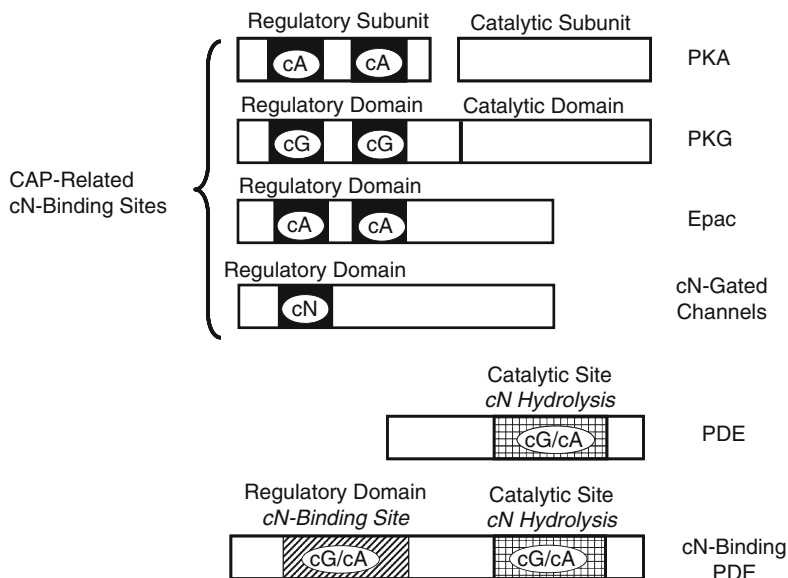
Given the abundance of cell proteins that bind or utilize purine-containing ligands (adenosine, adenine nucleotides, guanine nucleotides, etc.), the potential for interaction of alkylxanthine-related PDE inhibitors with proteins other than PDEs is always a concern. EHNA, a selective and weak inhibitor of PDE2, also inhibits adenosine deaminase (Mery et al. 1995), and zaprinast, which was historically considered to be a relatively selective inhibitor for PDEs 1 and 5, is a high-affinity agonist for GPR-35, an orphan receptor that is involved in modulating calcium homeostasis (Taniguchi et al. 2006). Some notable differences in the effects of vardenafil and sildenafil in *in vivo* studies have been reported (Toque et al. 2008), but whether this reflects action through non-PDE5 targets remains to be determined. Importantly, most of the alkylxanthine-based PDE inhibitors that have been studied to date do not appreciably interact with cN-binding sites on other proteins, including cN-gated channels, cN-dependent protein kinases, cAMP-modulated guanine-nucleotide exchange factors, or cN-binding allosteric sites on PDEs 2, 5,

6, 10, and 11. Use of radiolabeled inhibitors has not yet detected non-PDE targets of these compounds; however, this technique is only successful when inhibitor-binding proteins are sufficiently abundant for detection of the radioactive signal (Corbin et al. 2005).

### **3 Design of Potent Xanthine-Based PDE5 Inhibitors That Recapitulate Structural Features of the Entire cGMP Molecule**

Early use of cN analogs such as 2'-*O*-monobutyryl cAMP, N<sup>2</sup>, 2'-*O*-dibutyryl cAMP, and 8-Br-cAMP as agonists for activation of PKA led to the realization that these analogs could also bind to other cN-binding proteins such as cGMP-dependent protein kinase (PKG), Epacs, cN-gated channels, and PDEs. In many instances, the cN acted as a PDE inhibitor and thereby fostered accumulation of naturally occurring cN in cells or tissue extracts. An added advantage was that these analogs could traverse the cell membrane, whereas unmodified cAMP or cGMP is typically excluded from entering the cell. The cN-binding sites on PKA, PKG, Epacs, and cN-gated channels are homologous and belong to the catabolite gene activator protein (CAP) family of cN-binding sites (Fig. 3); as a result they share many similarities in preference for particular cN analogs, although those selective for each have been identified. In contrast, the catalytic sites and allosteric cN-binding sites in PDEs are evolutionarily unrelated to the CAP family of cN-binding sites, as well as to each other (McAllister-Lucas et al. 1993; Takio et al. 1984). As a result, the analog specificities of these sites are quite different from those of the CAP-related family of cN-binding sites.

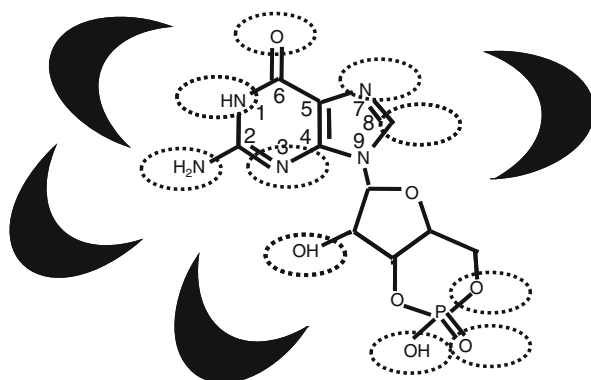
Although it was recognized early on that cN analogs acted as PDE inhibitors, the efficacy of these compounds in this capacity was often limited by their breakdown by PDEs. For these and other reasons, design of PDE inhibitors primarily mimicked only the purine of cAMP or cGMP. Increased availability of a broader spectrum of cN analogs with a range of chemical modification (Fig. 4) led to advances in understanding structural features that impact interaction of cNs and cN analogs with target proteins (kinases, cN-gated channels, Epacs, and PDEs). This, in turn, led to the development of analogs that were more potent and specific as agonists for the respective target proteins and more resistant to breakdown by PDEs, thus making them more effective PDE inhibitors. Moreover, cN analogs could be used to more rigorously define characteristics of cN-binding sites in various proteins, including PDEs (Beltman et al. 1995; Butt et al. 1995; Corbin and Doskeland 1983; Dao et al. 2006; Doskeland et al. 1983; Erneux et al. 1984, 1985; Erneux and Miot 1988; Francis et al. 1990; Rehmann et al. 2003; Thomas et al. 1992). Information that can be derived from cN analogs will be discussed in this context; cGMP analogs will be used for illustration.



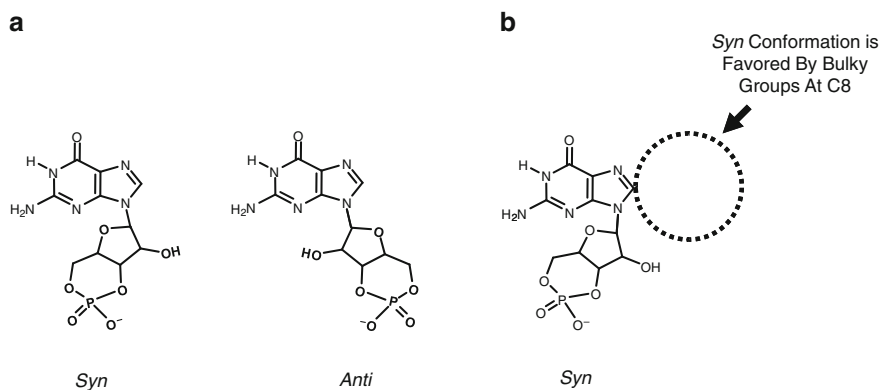
**Fig. 3** cN-binding proteins in mammalian tissues. cN-binding sites in mammals fall into three categories: **a** those whose amino acid sequences are related to the bacterial catabolite gene activator protein, **b** allosteric sites on certain PDEs (PDEs 2,5,6,10, and 11), and **c** catalytic sites of PDEs. Differences in the evolutionary origin and molecular structures are indicated by different fill patterns in the blocks representing the sites. Sites in cyclic AMP (cAMP)-dependent protein kinase and cyclic GMP (cGMP)-dependent protein kinase are selective for cAMP and cGMP, respectively, but at high concentrations either nucleotide will bind to these sites. Epacs have very high specificity for cAMP and contain either one or two cN-binding sites. Different cN-gated channels have selectivity for either cAMP or cGMP. Allosteric sites in PDEs most commonly bind cGMP with high selectivity, but PDE10 binds cAMP. Catalytic sites of PDEs can either be highly selective for cN or hydrolyze both cNs with equal efficacy

### 3.1 Cyclic GMP and Cyclic AMP in Solution Exist in Equilibrium Between Two Conformations

In solution, cGMP and cAMP are in equilibrium between two conformations (*syn* and *anti*) that differ in the orientation of the ribose phosphate moiety around the glycosyl bond at N9; the example for cGMP is shown in Fig. 5a. These conformations in gaseous and aqueous phases were predicted using the computational method self-consistent reaction field. The *anti* conformation is more stable in aqueous solution according to predictions from both the Onsager model and the isodensity polarized continuum method (Salter et al. 2003). However, appending groups at various positions in the purine can introduce steric constraints that perturb the equilibrium between the conformers so that the majority of the compound is primarily in the *anti* or the *syn* conformation (Fig. 5b). As a result, the favored cGMP conformation for binding of a particular cN analog to a target protein may be



**Fig. 4** Modifications of cN structure for development of analogs. cGMP is used as the example to indicate points in the structure that have been altered for development of analogs for use in biochemical studies. Positions on the purine ring are numbered. *Dotted ovals* indicate atoms that have been either removed or substituted by other atoms. *Black arcs* indicate regions that have had various types of groups appended to test for the effects of bulk and/or chemical characteristics



**Fig. 5** cNs in solution are in equilibrium between two conformations. **a** cGMP, shown as an example, in solution is in equilibrium between the *syn* and *anti* conformations owing to free rotation of the ribose–cyclic phosphate moiety around the N9 glycosyl bond. **b** Introduction of a bulky group at C8 sterically interferes with rotation of the ribose–cyclic phosphate moiety into the *anti* conformation and therefore increases the amount of cN in the *syn* conformation

more abundant at a particular concentration of the compound; for instance, the bulk of a large group, such as Br or a phenyl appended at C8 (Fig. 5b), physically interferes with rotation of the ribose phosphate moiety around the glycosyl bond, thereby strongly favoring the *syn* conformation. Moreover, groups appended to the purine can alter other features of the molecule, including the electron distribution and the dipole moment across the ring system (Corbin et al. 2006; Erneux et al. 1984). Insights derived from predictions of the favored conformations of cN analogs and the interactions of these compounds with target proteins were used to

model the cN-binding sites in proteins. Some proteins such as PKA and PKG preferentially bind cNs in the *syn* conformation (Corbin and Doskeland 1983; Sekhar et al. 1992; Wolfe et al. 1989); thus, cN analogs with large groups appended at C8, which foster the *syn* conformation, had high affinity for the cGMP-binding sites on PKG. Preference for either the *syn* or the *anti* conformer of cGMP for interaction with PDE catalytic sites varied (described later) (Beltman et al. 1995; Butt et al. 1995; Francis et al. 1990; Thomas et al. 1992). In the cN-signaling pathways, cN analogs can have dual and synergistic roles as activators of cN-dependent protein kinases and inhibitors of PDEs, whereas most alkylxanthine inhibitors of PDEs (e.g., IBMX and sildenafil) interact only with catalytic sites of PDEs but not with allosteric cN-binding sites in PKA, PKG, Epacs, cN-gated channels, or cN-binding PDEs.

### ***3.2 Use of cN Analogs To Map cN-Binding Sites of cN-Dependent Protein Kinases***

As more cN analogs with distinct modifications became available, they were used to define important points of contact with the cN-binding sites on PKA and PKG and to assess the potency and selectivity of analogs for activation of the respective phosphotransferase activities (Corbin and Doskeland 1983; Doskeland et al. 1983). These seminal studies provided a mechanistic basis for use of a catalog of cN analogs that continues to expand. This approach continues to be used today to better define the role of particular cN-binding proteins in mediating the effects of cN signaling in myriad biological processes (Beebe et al. 1984, 1985, 1988; Dremier et al. 2007; Francis et al. 1988; Petersen et al. 2008; Poppe et al. 2008; Strassmaier and Karpen 2007). To better understand the molecular basis of cN-mediated relaxation of smooth muscle, Francis et al. (1988) correlated known potencies of cGMP and cAMP analogs for activation of PKG or PKA with their potencies for relaxation of smooth muscle. The results convincingly showed that PKGI $\alpha$  plays a role in smooth muscle relaxation but did not support significant PKA involvement. During the course of this study, PKGI $\beta$ , which had a different pattern of cN analog selectivity, was discovered (Wolfe et al. 1989); this discovery required the design of new analogs to better define the roles of the PKGI isoenzymes.

Many new cGMP analogs were synthesized and studied for potency in activating PKGs and for blocking PDE activity (Corbin and Doskeland 1983; Sekhar et al. 1992; Wolfe et al. 1989). Analogs of cGMP with bulky groups at C8, e.g., a phenylthio group, which would strongly favor the *syn* conformer of cGMP, preferentially activated PKGI $\alpha$  over PKGI $\beta$  and potently relaxed precontracted pig coronary arteries. These studies strengthened the correlation between PKGI $\alpha$  activation and smooth muscle relaxation (Francis et al. 1988; Sekhar et al. 1992). The potency of cGMP analogs containing an 8-phenylthio group was further enhanced by appending an electron-donating substituent (e.g., hydroxyl, methoxy, amino) rather than an electron-withdrawing substituent (e.g., nitro) on the 8-phenylthio

**Table 5** Comparison of selectivity of allosteric cGMP-binding sites of cGMP-dependent protein kinase (PKG) I isoenzymes for cGMP analogs modified at C8

	$K'_1$ PKGI $\alpha$		$K'_1$ PKGI $\beta$	
	Site a	Site b	Site a	Site b
8-Br-cGMP	2.5	0.6	3.5	0.59
8- <i>p</i> -OH-Ph- <i>S</i> -cGMP	6.6	1.7	5.7	2.1
8-Di-OH-Ph- <i>S</i> -cGMP	10.4	0.7	14.4	1.2
8- <i>p</i> -NH <sub>2</sub> -Ph- <i>S</i> -cGMP	2.7	16.4	5.3	6.7
8- <i>p</i> -OMe-Ph- <i>S</i> -cGMP	0.9	2.8	2.1	1.9
8- <i>o</i> -Br-Ph- <i>S</i> -cGMP	1.6	0.9	ND	ND
8-Br-PET-cGMP	14.3	17.3	93.1	20.4

The value is the ratio of the affinity of the indicated site for a particular analog compared with that for cGMP. A high number indicates greater affinity for the analog than for cGMP

ND not determined

moiety (Sekhar et al. 1992). All of the analogs thus modified were poor activators of PKGII (Gamm et al. 1995) (unpublished data). Analogs of cGMP that were modified at N1 and C2, e.g., 1-*N*<sup>2</sup>-phenyletheno-cGMP (PET-cGMP) and 8-Br-PET-cGMP, activated PKGI $\alpha$  and PKGI $\beta$  with nearly equal potency (Sekhar et al. 1992; Wolfe et al. 1989).

The amino acid sequences of the cGMP-binding region of PKGI $\alpha$  and PKGI $\beta$  are identical and each PKG monomer contains two homologous allosteric cGMP-binding sites (site a and site b) that differ in amino acid sequence and kinetic characteristics. Remarkably, the affinities of these sites for cGMP in the two PKGs and their specificities for cGMP analogs differ (Table 5), but each site is involved in activation of the phosphotransferase activity (Corbin and Doskeland 1983; Sekhar et al. 1992; Wernet et al. 1989; Wolfe et al. 1989). The  $K'_1$  values in Table 5 represent affinities of a group of cN analogs for sites a and b in PKGI $\alpha$  and PKGI $\beta$  relative to the affinity for cGMP; a value of 10 indicates tenfold greater affinity than that for cGMP. This information was used to design potent and specific cN activators of PKGI $\alpha$  and PKGI $\beta$  and further study the roles of these PKGs in smooth muscle relaxation. 8-(4-Hydroxyphenyl-*S*)-PET-cGMP, which is modified at N1, C2, and C8 of cGMP, was very potent in activating both PKGs and in relaxing smooth muscle (Sekhar et al. 1992). The results suggested that these substitutions are favorable for the cGMP-binding sites in both PKGs.

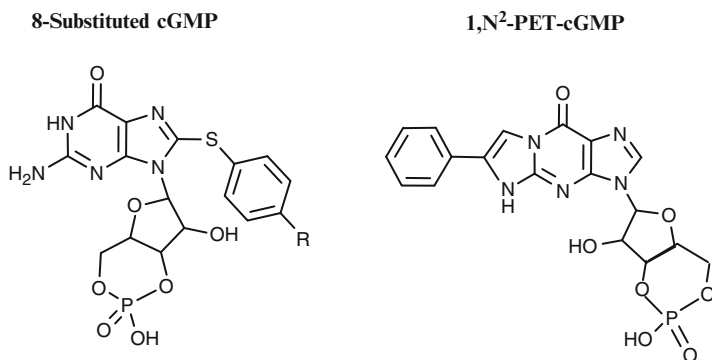
### 3.3 Use of cN Analogs To Map the Allosteric cGMP-Binding Sites and Catalytic Sites of PDEs

Cyclic nucleotide (cN) analogs were also used in studies to map the allosteric cGMP-binding sites and catalytic sites of several PDEs (Beltman et al. 1995; Butt et al. 1995; Francis et al. 1990; Thomas et al. 1992). The catalytic domains of PDEs are conserved and vary in amino acid sequence by 22–50% (Bender and Beavo 2006; Conti and Beavo 2007). Unlike cN-dependent protein kinases that strongly

prefer cNs in the *syn* conformation, the preference of PDE catalytic sites for the *anti* versus the *syn* conformation varies (Beltman et al. 1995; Butt et al. 1995; Sekhar et al. 1996; Thomas et al. 1992). Analogs of cGMP substituted at C8 had low affinity for the PDE5 catalytic site, whereas analogs with modifications at N1 and/or C2 were well tolerated. This predicted that cGMP binds to the PDE5 catalytic site in the *anti* conformation (Fig. 5a). The allosteric cGMP-binding site of PDE5 is highly specific for cGMP and has low tolerance for most cGMP analogs or alkylxanthines (Francis et al. 1980, 1990; Thomas et al. 1992).

The large number of new cGMP analogs that were synthesized by Sekhar et al. (1996) allowed for better characterization of the sites on PDE5; in addition, the resistance of these analogs to breakdown by PDE5 was determined to better predict their usefulness as investigational tools and to understand the effects of substitutions on the hydrolytic process. Analogs of cGMP substituted at C8 such as 8-(4-aminophenylthio)-cGMP were very resistant to degradation by PDE5 compared with cGMP (Sekhar et al. 1996). Structural modeling suggested that a large group appended at C8 sterically forced the ribose cyclic phosphate into the *syn* conformation, thus displacing that moiety from the catalytic machinery of PDE5 and providing strong resistance to degradation.

Analogs modified at the 1,2-position of cGMP, e.g., PET-cGMP (Fig. 6), were less resistant than the 8-substituted analogs to hydrolysis by PDE5 (Sekhar et al. 1996). This increased susceptibility to hydrolysis could be explained by greater freedom of the ribose phosphate in PET-cGMP to assume either the *syn* or the *anti* conformation by unimpeded rotation around the N9 glycosyl bond and therefore a greater tendency to assume the *anti* conformation. This interpretation was supported by comparing the extent of hydrolysis of PET-cGMP, 8-Br-PET-cGMP, and 8-I-PET-cGMP by purified PDE5; the extent of hydrolysis was inversely correlated to the bulkiness (I > Br > H) of the C8 substitution (Sekhar et al. 1996). Space-filling models revealed that steric constraints at C8 cause the ribose phosphate group in 8-I-PET-cGMP to be largely fixed in the *syn* conformation. On the basis of results obtained with cN analogs and visualization of the conformations using



**Fig. 6** The steric effect introduced by a bulky group appended to cGMP at C8 versus one appended to N1, C2

space-filling models, it was predicted that the PDE5 catalytic site binds cGMP in the *anti* conformation (Sekhar et al. 1996; Thomas et al. 1992).

### 3.4 Design and Use of 8-Substituted IBMX Analogs for Potent Inhibition of PDE

#### 3.4.1 Early Insights into the Mode of Interaction of Xanthine-Based Compounds with cN PDEs

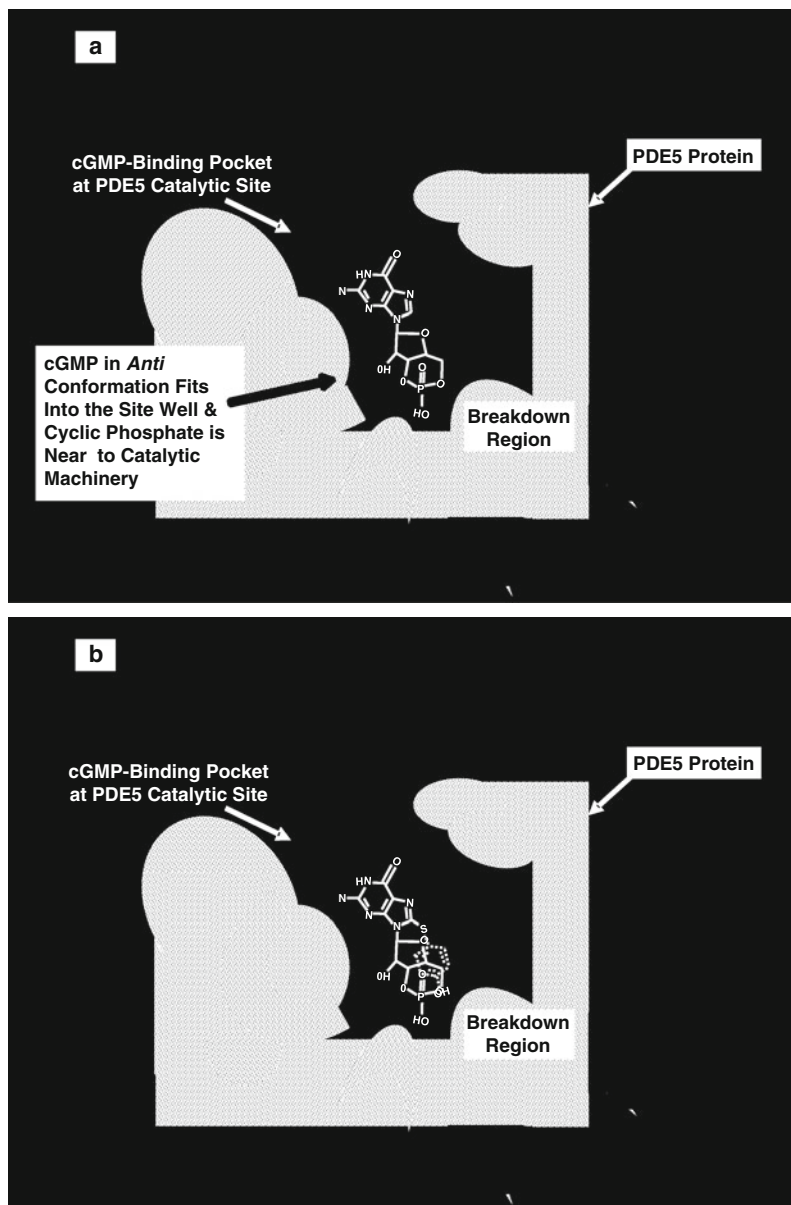
Wells et al. (1981) studied the inhibitory potency of xanthine-based compounds substituted at C1, N3, and C8 on partially purified PDEs: calmodulin-sensitive PDE and cAMP-specific PDE. On the basis of the results, they proposed that the catalytic site of calmodulin-sensitive PDE tolerates bulky groups appended at C8 of cGMP or xanthine analogs and contains a lipophilic area with which the ribose cyclic phosphate group interacts. With some xanthine analogs, this area is partially occupied by derivatized alkyl substitutions at N3, e.g., the isobutyl of IBMX. These early studies provided crucial information about PDE catalytic sites and led to the interpretation that cNs bind to certain PDEs in the *anti* conformation. It was later shown that catalytic sites of several other PDEs prefer the *syn* conformation (Beltman et al. 1995; Butt et al. 1995).

#### 3.4.2 Design of Potent and Novel Derivatized Alkylxanthines as PDE5 Inhibitors

With use of insights derived from studies of the interaction of cGMP analogs with the PDE5 catalytic site and the hydrolytic resistance of analogs, a design for potent PDE5 inhibitors was conceptualized and achieved long before the discovery of sildenafil (Sekhar et al. 1996). It was proposed that potent IBMX analogs would be achieved by mimicking the entire cGMP molecule and that the moiety which mimicked the ribose phosphate should occupy a space simulating an *anti* conformation, which is the conformation of cGMP bound in the PDE5 catalytic site (Thomas et al. 1992). At that time, despite known differences in analog specificities, it was thought that sites of interaction of cGMP on PKG and PDE5 could be homologous. Therefore, on the basis of results obtained from cN studies with PKG, it was proposed that a hydrophobic group, e.g., a phenyl, with electron-donating substitutions would foster affinity of analogs for PDE5 and thereby increase inhibitor potency. Prior to this, most PDE5 inhibitors contained a mimic of guanine but not of the ribose cyclic phosphate.

Using this strategy, Sekhar et al. (1996) synthesized a collection of IBMX analogs with a phenylthio substitution at C8, which were 10–30-fold more potent inhibitors for PDE5 than any known inhibitor. The cartoon in Fig. 7a depicts cGMP bound to the PDE5 catalytic site in the *anti* conformation with the ribose phosphate





**Fig. 7** The cN alignment in the PDE5 catalytic site. **a** cGMP is shown in the *anti* conformation in the PDE5 catalytic site; this orientation places the cyclic phosphate ring near the breakdown region containing the divalent cations that provide for hydrolysis. **b** The *dotted structure* depicts the fact that the 4-hydroxyphenylthio group appended at C8 in 3-isobutyl-1-methylxanthine (IBMX) occupies the space normally occupied by the cyclic phosphate moiety of cGMP, i.e. the two moieties are shown to be overlapping in this depiction

moiety located near the hydrolytic region of the site. To mimic cGMP in this *anti* conformation, related compounds were modeled so that the C8-substitution could occupy the space of the ribose cyclic phosphate group of cGMP when bound to the PDE5 catalytic site (Fig. 7b). This design avoided steric hindrance posed by substitutions at N9. IBMX analogs synthesized based on this reasoning produced highly potent PDE5 inhibitors that also had improved selectivity for PDE5 over other PDEs (Sekhar et al. 1996). The most potent compound [8-(norborylmethyl)-IBMX] was about 7,000-fold more potent for PDE5 ( $IC_{50} = 0.0015 \mu\text{M}$ ) than IBMX, the parent compound ( $IC_{50} = 10 \mu\text{M}$ ); the potency of this compound for PDE1 ( $IC_{50} = 0.03 \mu\text{M}$ ) was more than 200-fold greater than that of the parent IBMX ( $IC_{50} = 7 \mu\text{M}$ ). This simple modification of the nonspecific and weak inhibitor IBMX dramatically improved both the potency and specificity of this compound for PDE1 or PDE5 compared with PDEs 2–4. The  $IC_{50}$  values reported for these compounds for PDE5 underestimated the true potency because for technical reasons assays were conducted with saturating cGMP substrate. The  $K_i$  of 8-(norborylmethyl)-IBMX calculated from this  $IC_{50}$  value using the Cheng–Prusoff equation –  $K_i = IC_{50}/(1 + [S]/K_m)$  – is 0.17 nM, which is similar to that of vardenafil (0.1–0.4 nM), the most potent known inhibitor for PDE5 (Blount et al. 2004; Cheng and Prusoff 1973).

Information gained from investigations of the characteristics of the cN-binding sites of PKA and PKG was a critical antecedent to subsequent studies of the sites on PDEs (Corbin and Dosekand 1983; Dosekand et al. 1983; Sekhar et al. 1992; Wolfe et al. 1989). Synthesis and studies of IBMX analogs with restricted orientation of substitutions clearly demonstrated the preference of an *anti* conformation at the catalytic site of PDE5, a conclusion that had already been drawn on the basis of cN analog studies (Thomas et al. 1992). The *cis* isomer of 8-(4-methoxystyryl)-IBMX ( $IC_{50} = 0.016 \mu\text{M}$ ) was 23-fold more potent than its *trans* isomer ( $IC_{50} > 10 \mu\text{M}$ ). These results complemented the original design strategy and emphasized the importance of studies of substrate analogs in mapping sites that interact with cNs, and in designing effective inhibitors.

## 4 Insights into Interaction of PDEs with Alkylxanthine-Related Compounds

To advance understanding of the interaction of cNs and alkylxanthine-related inhibitors with the catalytic sites of PDEs, a number of X-ray crystal structures of isolated C domains of PDEs in complex with either the product of hydrolysis (5'-AMP or 5'-GMP) or inhibitors have been determined. To date, there is no structure for a co-crystal of a PDE with wild-type sequence containing either cAMP or cGMP in the catalytic site since the cN (even “nonhydrolyzable” analogs) is invariably hydrolyzed during the determinations. Despite that, the structures of inactive PDE4D and PDE10A in complex with cAMP and cGMP, respectively, the structures of other PDEs in complex with inhibitors and catalytic products,

site-directed mutagenesis studies, and cN analog studies have provided considerable insight into the interaction of cN with PDE catalytic sites (Corbin et al. 2006; Huai et al. 2004a; Ke and Wang 2007; Turko et al. 1998, 1999; Wang et al. 2007a, 2007b, 2008a; Zoraghi et al. 2006, 2007).

#### ***4.1 Interactions of IBMX and Other Xanthine-Related PDE Inhibitors with PDE Catalytic Sites as Determined by X-ray Crystallography***

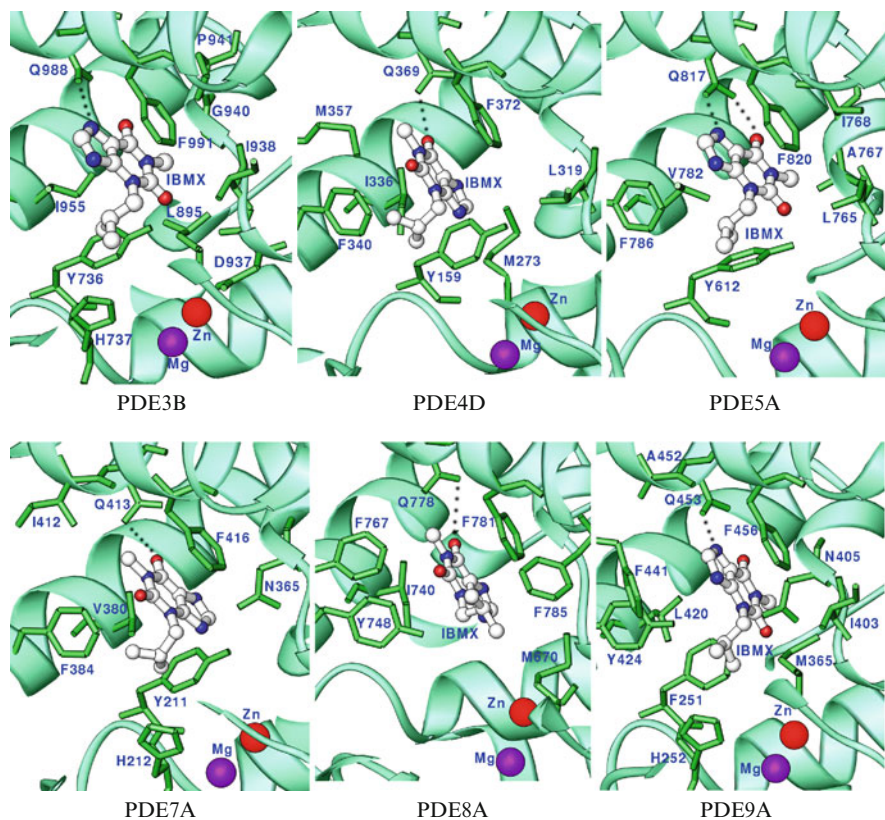
##### **4.1.1 Identification of Direct Contacts Between PDE Catalytic Sites and Derivatized Alkylxanthine Inhibitors**

Six crystal structures of isolated C domains of human PDEs in complex with IBMX (which occupies the catalytic site) have been reported; these include PDE3B (Protein Data Bank entry code 1SOJ), PDE4D (1ZKN), PDE5A (1RKP), PDE7A (1ZKL), PDE8A (3ECN), and PDE9A (2HD1) (Huai et al. 2004a, b; Ke and Wang 2007; Scapin et al. 2004; Wang et al. 2005, 2006, 2008a). In addition, X-ray crystal structures of PDE5 in complex with the derivatized alkylxanthine inhibitors sildenafil, vardenafil, and tadalafil have also been reported (Sung et al. 2003; Wang et al. 2006, 2008b).

The planar xanthine-like ring of these inhibitors mimics the purine of cN and is bound in a hydrophobic pocket of the catalytic site. In these crystal structures, the heterocyclic ring is bound by a “hydrophobic clamp” that is formed by the side chains of two amino acids [an alkyl-like group (e.g., valine or leucine) and a phenyl group of a conserved phenylalanine] (Fig. 8) (Ke and Wang 2006; Sung et al. 2003; Xu et al. 2000; Zhang 2006). IBMX binding to PDE catalytic sites shows two common features: (1) hydrophobic stacking with a conserved phenylalanine, and (2) hydrogen bonding with an invariant glutamine. For IBMX, the xanthine ring stacks against the hydrophobic side chain of Phe991 in PDE3B, Phe372 in PDE4D2, Phe820 in PDE5A1, Phe416 in PDE7A1, Phe781 in PDE8A1, and Phe456 in PDE9A2 (Fig. 8). The five-membered ring in IBMX typically forms a  $\pi$ - $\pi$  electron-stacking interaction with the phenylalanine side chain.

Hydrophobic stacking with the phenylalanine and hydrogen bonding with the glutamine appear to be basic elements for binding of PDE inhibitors since they have been shown to be common for binding of most known PDE inhibitors (Ke and Wang 2007).

While these contacts contribute importantly to high-affinity binding in certain instances, e.g., binding of IBMX, sildenafil, vardenafil, and tadalafil to PDE5A (Zoraghi et al. 2007), they alone are not sufficient for high-affinity interaction. This is evident from the very weak interaction of IBMX with PDE8A (Wang et al. 2008a) as well as the contributions of other amino acids to high affinity of PDE5 for substrates and inhibitors (Zoraghi et al., 2007). In all PDE catalytic sites studied, IBMX forms van der Waals contacts with residues other than the phenylalanine and



**Fig. 8** IBMX binding to the active sites of PDEs. The *dotted lines* represent hydrogen bonds. The PDE binuclear metal site where breakdown of the cyclic phosphate moiety of cN occurs is shown containing zinc (*large red ball*), which is bound to PDEs with high affinity and has clearly been identified in the respective X-ray crystal structures, and magnesium (*large purple ball*), which is presumed to occupy the second metal site

glutamine (Table 6); among these, some residues that are involved in IBMX binding, e.g., Ile336 of PDE4D2 or their homologs in other PDEs, are conserved within families, while different amino acids are found uniquely in particular PDE families. Despite the availability of numerous X-ray crystal structures of PDEs in co-complex with alkylxanthine-related inhibitors, there is still much that is not understood about factors that provide for high potency and selectivity for the various PDEs (Blount et al. 2006; Ke and Wang 2006; Zhang 2006).

#### 4.1.2 Difference in Orientation of IBMX in the Active Sites of PDEs

In spite of the shared pattern of interactions, IBMX binding shows significant differences among PDEs in both its orientation and position within the respective catalytic sites and specific interactions that are formed. The most significant

**Table 6** PDE catalytic domain amino acids that interact with IBMX

PDE	Y736	H737	L895	D937	I938	G940	P941	I955	F340	M357	Q988	F991
PDE3B	Y736	H737	L895	D937	I938	G940	P941	I955	F340	M357	Q988	F991
PDE4D2	Y159		M273		L319			I336			Q369	F372
PDE5A1	<b>Y612</b>					<u>L765</u>	A767	I768	<u>V782</u>	<u>F786</u>	<b>Q817</b>	<b>F820</b>
PDE7A1	Y211	H212				N365		V380	F384		Q413	F416
PDE8A1			M670					I744	Y748	F767	Q778	F781
PDE9A2	F251	H252	M365		I403	N405		L420	Y424	F441	Q453	F456

Summary of amino acids in homologous positions in the catalytic sites of PDEs that make contact with IBMX. Site-directed mutagenesis of Tyr612, Gln817, or Phe820 (*bolded and underlined*) in PDE5 holoenzyme caused a 40–90-fold loss of affinity for IBMX. Point mutations of Leu765, Val782, or Phe786 (*italicized and underlined*) caused a tenfold or lower change in affinity for IBMX

difference is the orientation of IBMX in these structures. The xanthine ring of IBMX is aligned in the same direction in the structures of PDEs 4D, 7A, and 8A, but in PDEs 3B, 5A, and 9A, the alignment is the opposite (Fig. 8). This difference in orientation results in a different pattern of hydrogen bonding. The N7 atom of IBMX forms a hydrogen bond with the O $\epsilon$  from the side chain of the invariant glutamine of PDEs 3B, 5A, and 9A; the carbonyl oxygen at C6 forms a hydrogen bond with the N $\epsilon$  of the glutamine side chain in the structures of PDEs 4D, 5A, 7A, and 8A (Fig. 8) (Huai et al. 2004a; Ke and Wang 2007). In addition, IBMX shows positional changes of several angstroms among the atoms. On the basis of the results of different positions and orientations, individual PDE families show unique interactions such as the interaction between N3 of IBMX and OH of Tyr424 in the PDE9A structure (Huai et al. 2004b).

Differences in the interactions of IBMX in these structures may be due to the fact that IBMX has a much smaller molecular volume than do the PDE catalytic site pockets (about 330 Å<sup>3</sup>) (Xu et al. 2000). As a result, IBMX can assume different orientations and exploit potential interactions, while retaining hydrophobic stacking with the conserved phenylalanine and hydrogen-bond formation with the glutamine. These differences in the IBMX binding should be useful for design of PDE-family selective inhibitors.

### 4.1.3 Extended Effects of Derivatized Alkylxanthines on the Structure of the PDE5 Catalytic Site

#### Movement of the H-Loop of PDE5

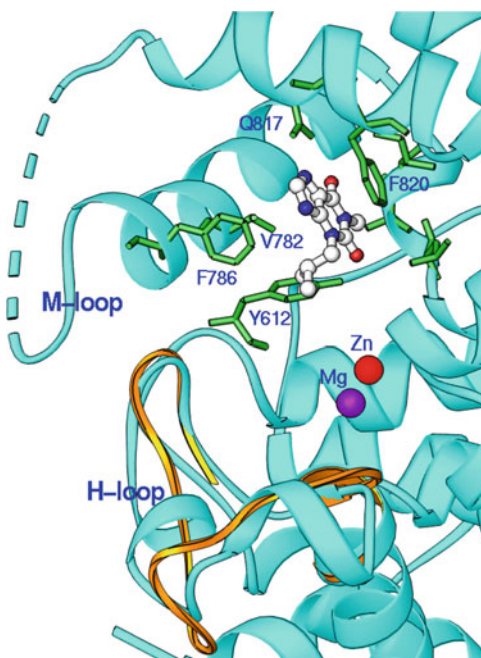
In most PDE families, the conformations of the C domain appear to be rigid. The structure of the PDE5 C domain appears to be unique among PDE families (Ke and Wang 2007) since the H-loop in the C domain can assume at least five different conformations (Chen et al. 2008; Wang et al. 2006). While some inhibitors such as sildenafil and icarisid II directly interact with the H-loop and cause 10–20-Å shifts, IBMX does not directly contact the H-loop and causes up to 7-Å movement of the H-loop (Fig. 9). The H-loop of the PDE5A–IBMX complex contains two short  $\alpha$ -helices that are comparable within most other PDE families. However, in unliganded PDE5 C domain, the H-loop is more like a random coil. The explanation for the H-loop movement is not straightforward. Apparently, IBMX binding to the pocket of the active site imposes an allosteric effect on H-loop movement (Wang et al. 2006), but the impact of the binding of other inhibitors could be due to both direct interaction and an allosteric effect.

#### Inhibitor-Induced Structural Changes in the PDE5 Catalytic Site

Another characteristic of binding of alkylxanthine-related compounds to PDE5 involves an inhibitor-induced change in affinity for the inhibitor. The molecular



**Fig. 9** Comparison of the position of PDE5 H-loop in the unliganded PDE5A versus its position in the IBMX-bound form. Ribbon structure of the PDE5 catalytic domain in the unliganded state and with IBMX bound is shown. The location of IBMX in the catalytic site is shown by the stick model and amino acids surrounding IBMX are indicated. The image emphasizes the different positions and structural features of the PDE5 H-loop in the unliganded state (*blue*) and the IBMX-bound form (*golden ribbon*)



basis for these kinetic effects has not been resolved. Binding of certain inhibitors (sildenafil, vardenafil, and tadalafil) to either PDE5 holoenzyme or isolated C domain exhibited two rates, a high-affinity component and a low-affinity component (Blount et al. 2007). However, prolonged incubation of PDE5 holoenzyme with these inhibitors converted the low-affinity component to the high-affinity component, indicating that there is a time-dependent conformational change that optimizes contacts between the compound and PDE5 catalytic site. This shift in affinity required a portion of the regulatory domain of PDE5, i.e., the GAF B subdomain, and did not occur in the isolated C domain. Since heterogeneity of the binding pattern was not evident in the X-ray crystallographic structure of the isolated C domain, there is no straightforward explanation of this phenomenon. However, it is worth noting that binding of rolipram, which is not an alkylxanthine derivative, to PDE4 occurs with low- and high-affinity states that apparently reflect binding activity of a single site on PDE4, i.e., the catalytic site (McKenna and Muller 2006). Whether the PDE4 catalytic site converts between the two kinetic states is unknown. However, these types of kinetic differences in the catalytic sites of PDEs may be more common than appreciated and add another degree of complexity to understanding ligand interactions with PDE catalytic sites.

## **4.2 Use of Site-Directed Mutagenesis To Quantify the Impact of Amino Acid Contacts with Substrates and Xanthine-Related PDE Inhibitors**

Results of site-directed mutagenesis in PDE5 holoenzyme have biochemically quantified and verified the importance of the contact(s) observed in the X-ray crystallographic studies of the isolated C domain (Zoraghi et al. 2007); contacts observed in crystal structures of other PDEs have not yet been studied in this manner. In PDE5 holoenzyme, conversion of the conserved phenylalanine (Phe820) to alanine caused a dramatic loss (60–450-fold) of affinity for substrate as well as inhibitors such as vardenafil, sildenafil, tadalafil, and IBMX. Mutation of the side chain of the invariant glutamine (Gln869) also caused a major loss (60–500-fold) of affinity for substrate as well as inhibitors (Ke and Wang 2006, 2007; Zoraghi et al. 2007); mutation of this same glutamine in PDE11 also caused a major loss of affinity for substrates (cGMP and cAMP) as well as tadalafil, a reasonably potent inhibitor (Weeks et al. 2009). Substitution of alanine for the invariant tyrosine in PDE5 (Tyr612), which also makes contacts with certain inhibitors in PDE5 catalytic site, also caused a major loss of affinity (14–120-fold) for substrate and inhibitors. Replacement of this tyrosine by phenylalanine increased the affinity for some inhibitors, consistent with a positive contribution of the tyrosine side chain to the hydrophobicity of the catalytic pocket (Corbin et al. 2006; Huai et al. 2004a; Sung et al. 2003). Most of the mutations of other amino acids that had been shown by X-ray crystallography to be in contact with inhibitors had more modest effects. Thorough mutagenesis studies have not been conducted in most other PDE families.

## **4.3 Use of cN Analogs to Probe Structural Elements; Comparison of Findings Using X-ray Crystallography**

Studies with cN analogs to characterize the catalytic sites of PDEs have provided an important adjunct to interpretations arrived at through the use of X-ray crystallographic structures and site-directed mutagenesis. Studies with cN analogs as well as those using site-directed mutagenesis are particularly important because both approaches can employ PDE holoenzymes to assess the importance of various contacts between the PDE and the substrate and/or inhibitors. Studies with cN analogs have shown major differences in the catalytic sites of PDEs, so it is difficult to make general comments. Studies with cN analogs have shown that (1) some PDEs prefer cN in the *anti* conformation, while others prefer the *syn* conformation, and these results have subsequently been validated by X-ray crystallographic studies (Wang et al. 2007a, b), (2) contact with the 2'-hydroxyl is not particularly important for any PDE tested, (3) interaction with N1 in either cAMP or cGMP is



important in many, but not all PDEs, (4) the importance of interaction with N7 varies, and (5) interaction with the substituent at C6 is in some cases important, but is not universal (Beltman et al. 1995; Butt et al. 1995; Francis et al. 1990; Thomas et al. 1992; Weeks et al., unpublished results).

## 5 Concluding Remarks

Shortly after the discovery of cAMP, the methylxanthines and related compounds became critical tools for research in cN signaling pathways. They inspired synthesis of thousands of compounds that might act as more specific and potent PDE inhibitors. Many of these compounds have become highly useful in clinical treatment of a number of disease processes, and potential treatment of many other diseases is being investigated. However, despite great advances in understanding the mode of interaction of these compounds with PDEs, the pharmacological action of these agents are not completely understood.

**Acknowledgements** This work was supported by NIH grants DK 40029 (J.D.C) and GM 059791 (H.K.)

## References

- Al-Ameri H, Thomas ML, Yoon A, Mayeda GS, Burstein S, Kloner RA, Shavelle DM (2009) Complication rate of diagnostic carotid angiography performed by interventional cardiologists. *Catheter Cardiovasc Interv* 73:661–665
- Arnaud M (2010) Pharmacokinetics and metabolism of natural methylxanthines in animal and man. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Aviado DM, Porter JM (1984) Pentoxifylline: a new drug for the treatment of intermittent claudication. Mechanism of action, pharmacokinetics, clinical efficacy and adverse effects. *Pharmacotherapy* 4:297–307
- Ballard SA, Gingell CJ, Tang K, Turner LA, Price ME, Naylor AM (1998) Effects of sildenafil on the relaxation of human corpus cavernosum tissue in vitro and on the activities of cyclic nucleotide phosphodiesterase isozymes. *J Urol* 159:2164–2171
- Barnes PJ (2003a) Theophylline: new perspectives for an old drug. *Am J Respir Crit Care Med* 167:813–818
- Barnes PJ (2003b) Therapy of chronic obstructive pulmonary disease. *Pharmacol Ther* 97:87–94
- Barnes PJ (2005) Theophylline in chronic obstructive pulmonary disease: new horizons. *Proc Am Thorac Soc* 2:334–339
- Barnes P (2006) Theophylline for COPD. *Thorax* 61:742–744
- Barnes PJ, Stockley RA (2005) COPD: current therapeutic interventions and future approaches. *Eur Respir J* 25:1084–1106
- Beavo JA, Rogers NL, Crofford OB, Hardman JG, Sutherland EW, Newman EV (1970) Effects of xanthine derivatives on lipolysis and on adenosine 3',5'-monophosphate phosphodiesterase activity. *Mol Pharmacol* 6:597–603

- Beavo J, Houslay MD, Francis SH (2006) Cyclic nucleotide phosphodiesterase superfamily. In: Beavo J, Francis SH, Houslay MD (eds) Cyclic nucleotide phosphodiesterases in health and disease. CRC, Boca Raton, pp 3–17
- Beebe SJ, Holloway R, Rannels SR, Corbin JD (1984) Two classes of cAMP analogs which are selective for the two different cAMP-binding sites of type II protein kinase demonstrate synergism when added together to intact adipocytes. *J Biol Chem* 259:3539–3547
- Beebe SJ, Redmon JB, Blackmore PF, Corbin JD (1985) Discriminative insulin antagonism of stimulatory effects of various cAMP analogs on adipocyte lipolysis and hepatocyte glycolysis. *J Biol Chem* 260:15781–15788
- Beebe SJ, Blackmore PF, Chrisman TD, Corbin JD (1988) Use of synergistic pairs of site-selective cAMP analogs in intact cells. *Methods Enzymol* 159:118–139
- Beltman J, Becker DE, Butt E, Jensen GS, Rybalkin SD, Jastorff B, Beavo JA (1995) Characterization of cyclic nucleotide phosphodiesterases with cyclic GMP analogs: topology of the catalytic domains. *Mol Pharmacol* 47:330–339
- Bender AT, Beavo JA (2006) Cyclic nucleotide phosphodiesterases: molecular regulation to clinical use. *Pharmacol Rev* 58:488–520
- Bender AT, Ostenson CL, Wang EH, Beavo JA (2005) Selective up-regulation of PDE1B2 upon monocyte-to-macrophage differentiation. *Proc Natl Acad Sci USA* 102:497–502
- Benowitz NL (1990) Clinical pharmacology of caffeine. *Annu Rev Med* 41:277–288
- Berthet J, Sutherland EW, Rall TW (1957) The assay of glucagon and epinephrine with use of liver homogenates. *J Biol Chem* 229:351–361
- Black KL, Yin D, Ong JM, Hu J, Konda BM, Wang X, Ko MK, Bayan JA, Sacapano MR, Espinoza A, Irvin DK, Shu Y (2008) PDE5 inhibitors enhance tumor permeability and efficacy of chemotherapy in a rat brain tumor model. *Brain Res* 1230:290–302
- Blount MA, Beasley A, Zoraghi R, Sekhar KR, Bessay EP, Francis SH, Corbin JD (2004) Binding of tritiated sildenafil, tadalafil, or vardenafil to the phosphodiesterase-5 catalytic site displays potency, specificity, heterogeneity, and cGMP stimulation. *Mol Pharmacol* 66:144–152
- Blount MA, Zoraghi R, Ke H, Bessay EP, Corbin JD, Francis SH (2006) A 46-amino acid segment in phosphodiesterase-5 GAF-B domain provides for high vardenafil potency over sildenafil and tadalafil and is involved in phosphodiesterase-5 dimerization. *Mol Pharmacol* 70:1822–1831
- Blount MA, Zoraghi R, Bessay EP, Beasley A, Francis SH, Corbin JD (2007) Conversion of phosphodiesterase-5 (PDE5) catalytic site to higher affinity by PDE5 inhibitors. *J Pharmacol Exp Ther* 323:730–737
- Boolell M, Allen MJ, Ballard SA, Gepi-Attee S, Muirhead GJ, Naylor AM, Osterloh IH, Gingell C (1996) Sildenafil: an orally active type 5 cyclic GMP-specific phosphodiesterase inhibitor for the treatment of penile erectile dysfunction. *Int J Impot Res* 8:47–52
- Boutrel B, Koob GF (2004) What keeps us awake: the neuropharmacology of stimulants and wakefulness-promoting medications. *Sleep* 27:1181–1194
- Boyle CD, Xu R, Asberom T, Chackalamanni S, Clader JW, Greenlee WJ, Guzik H, Hu Y, Hu Z, Lankin CM, Pissamitski DA, Stamford AW, Wang Y, Skell J, Kurowski S, Vemulapalli S, Palamanda J, Chintala M, Wu P, Myers J, Wang P (2005) Optimization of purine based PDE1/PDE5 inhibitors to a potent and selective PDE5 inhibitor for the treatment of male ED. *Bioorg Med Chem Lett* 15:2365–2369
- Buckle DR, Arch JR, Connolly BJ, Fenwick AE, Foster KA, Murray KJ, Readshaw SA, Smallridge M, Smith DG (1994) Inhibition of cyclic nucleotide phosphodiesterase by derivatives of 1,3-bis(cyclopropylmethyl)xanthine. *J Med Chem* 37:476–485
- Burnett AL, Bivalacqua TJ, Champion HC, Musicki B (2006) Long-term oral phosphodiesterase 5 inhibitor therapy alleviates recurrent priapism. *Urology* 67:1043–1048
- Butcher RW, Sutherland EW (1962) Adenosine 3',5'-phosphate in biological materials. I. Purification and properties of cyclic 3',5'-nucleotide phosphodiesterase and use of this enzyme to characterize adenosine 3',5'-phosphate in human urine. *J Biol Chem* 237:1244–1250
- Butt E, Beltman J, Becker DE, Jensen GS, Rybalkin SD, Jastorff B, Beavo JA (1995) Characterization of cyclic nucleotide phosphodiesterases with cyclic AMP analogs: topology of the

- catalytic sites and comparison with other cyclic AMP-binding proteins. *Mol Pharmacol* 47:340–347
- Carson CC, Lue TF (2005) Phosphodiesterase type 5 inhibitors for erectile dysfunction. *BJU Int* 96:257–280
- Chen G, Wang H, Robinson H, Cai J, Wan Y, Ke H (2008) An insight into the pharmacophores of phosphodiesterase-5 inhibitors from synthetic and crystal structural studies. *Biochem Pharmacol* 75:1717–1728
- Cheng Y, Prusoff WH (1973) Relationship between the inhibition constant ( $K_1$ ) and the concentration of inhibitor which causes 50 per cent inhibition ( $I_{50}$ ) of an enzymatic reaction. *Biochem Pharmacol* 22:3099–3108
- Choi OH, Shamim MT, Padgett WL, Daly JW (1988) Caffeine and theophylline analogues: correlation of behavioral effects with activity as adenosine receptor antagonists and as phosphodiesterase inhibitors. *Life Sci* 43:387–398
- Chou K, Bell L (2007) Caffeine content of prepackaged national-brand and private-label carbonated beverages. *J Food Sci* 72:C337–C442
- Conti M, Beavo J (2007) Biochemistry and physiology of cyclic nucleotide phosphodiesterases: essential components in cyclic nucleotide signaling. *Annu Rev Biochem* 76:481–511
- Corbin JD, Dorskland SO (1983) Studies of two different intrachain cGMP-binding sites of cGMP-dependent protein kinase. *J Biol Chem* 258:11391–11397
- Corbin JD, Francis SH (1999) Cyclic GMP phosphodiesterase-5: target of sildenafil. *J Biol Chem* 274:13729–13732
- Corbin JD, Francis SH (2002) Pharmacology of phosphodiesterase-5 inhibitors. *Int J Clin Pract* 56:453–459
- Corbin JD, Beasley A, Blount MA, Francis SH (2004) Vardenafil: structural basis for higher potency over sildenafil in inhibiting cGMP-specific phosphodiesterase-5 (PDE5). *Neurochem Int* 45:859–863
- Corbin JD, Beasley A, Blount MA, Francis SH (2005) High lung PDE5: a strong basis for treating pulmonary hypertension with PDE5 inhibitors. *Biochem Biophys Res Commun* 334:930–938
- Corbin J, Francis S, Zoraghi R (2006) Tyrosine-612 in PDE5 contributes to higher affinity for vardenafil over sildenafil. *Int J Impot Res* 18:251–257
- Cote RH (2006) Photoreceptor phosphodiesterase (PDE6): a G-protein-activated PDE regulating visual excitation in rod and cone photoreceptor cells. In: Beavo J, Francis SH, Houslay MD (eds) *Cyclic nucleotide phosphodiesterases in health and disease*. CRC, Boca Raton, pp 165–193
- Dao KK, Teigen K, Kopperud R, Hodneland E, Schwede F, Christensen AE, Martinez A, Dorskland SO (2006) Epac1 and cAMP-dependent protein kinase holoenzyme have similar cAMP affinity, but their cAMP domains have distinct structural features and cyclic nucleotide recognition. *J Biol Chem* 281:21500–21511
- Degerman E, Belfrage P, Newman AH, Rice KC, Manganiello VC (1987) Purification of the putative hormone-sensitive cyclic AMP phosphodiesterase from rat adipose tissue using a derivative of cilostamide as a novel affinity ligand. *J Biol Chem* 262:5797–5807
- Dent G, Rabe K (1996) Effects of theophylline and non-selective xanthine derivatives on PDE isoenzymes and cellular function. In: Schudt C, Dent G, Rabe KF (eds) *Handbook of immunopharmacology: phosphodiesterase inhibitors*. Academic, San Diego, pp 41–64
- Dent G, Giembycz MA, Rabe KF, Wolf B, Barnes PJ, Magnussen H (1994) Theophylline suppresses human alveolar macrophage respiratory burst through phosphodiesterase inhibition. *Am J Respir Cell Mol Biol* 10:565–572
- Doh H, Shin CY, Son M, Ko JI, Yoo M, Kim SH, Kim WB (2002) Mechanism of erectogenic effect of the selective phosphodiesterase type 5 inhibitor, DA-8159. *Arch Pharm Res* 25:873–878
- Dorskland SO, Ogreid D, Ekanger R, Sturm PA, Miller JP, Suva RH (1983) Mapping of the two intrachain cyclic nucleotide binding sites of adenosine cyclic 3',5'-phosphate dependent protein kinase I. *Biochemistry* 22:1094–1101

- Dremier S, Milenkovic M, Blancquaert S, Dumont JE, Doskeland SO, Maenhaut C, Roger PP (2007) Cyclic adenosine 3',5'-monophosphate (cAMP)-dependent protein kinases, but not exchange proteins directly activated by cAMP (Epac), mediate thyrotropin/cAMP-dependent regulation of thyroid cells. *Endocrinology* 148:4612–4622
- [Energyfiend.com](#) (2005a) Caffeine content of drinks. Energy Fiend
- [Energyfiend.com](#) (2005b) Caffeine in food
- Erneux C, Miot F (1988) Cyclic nucleotide analogs used to study phosphodiesterase catalytic and allosteric sites. *Methods Enzymol* 159:520–530
- Erneux C, Couchie D, Dumont JE, Jastorff B (1984) Cyclic nucleotide derivatives as probes of phosphodiesterase catalytic and regulatory sites. *Adv Cyclic Nucleotide Protein Phosphor Res* 16:107–118
- Erneux C, Miot F, Van Haastert PJ, Jastorff B (1985) The binding of cyclic nucleotide analogs to a purified cyclic GMP-stimulated phosphodiesterase from bovine adrenal tissue. *J Cyclic Nucleotide Protein Phosphor Res* 10:463–472
- Fawcett L, Baxendale R, Stacey P, McGruther C, Harrow I, Soderling S, Hetman J, Beavo JA, Phillips SC (2000) Molecular cloning and characterization of a distinct human phosphodiesterase gene family: PDE11A. *Proc Natl Acad Sci USA* 97:3702–3707
- Fisher DA, Smith JF, Pillar JS, St Denis SH, Cheng JB (1998) Isolation and characterization of PDE9A, a novel human cGMP-specific phosphodiesterase. *J Biol Chem* 273:15559–15564
- Francis SH, Corbin JD (2003) Molecular mechanisms and pharmacokinetics of phosphodiesterase-5 antagonists. *Curr Urol Rep* 4:457–465
- Francis SH, Corbin JD (2005a) Phosphodiesterase-5 inhibition: the molecular biology of erectile function and dysfunction. *Urol Clin N Am* 32:419–429, vi
- Francis SH, Corbin JD (2005b) Sildenafil: efficacy, safety, tolerability and mechanism of action in treating erectile dysfunction. *Expert Opin Drug Metab Toxicol* 1:283–293
- Francis SH, Lincoln TM, Corbin JD (1980) Characterization of a novel cGMP binding protein from rat lung. *J Biol Chem* 255:620–626
- Francis SH, Noblett BD, Todd BW, Wells JN, Corbin JD (1988) Relaxation of vascular and tracheal smooth muscle by cyclic nucleotide analogs that preferentially activate purified cGMP-dependent protein kinase. *Mol Pharmacol* 34:506–517
- Francis SH, Thomas MK, Corbin JD (1990) Cyclic GMP-binding cyclic GMP-specific phosphodiesterase from lung. In: Beavo J, Houslay MD (eds) *Cyclic nucleotide phosphodiesterases: structure, regulation and drug action*, vol 2. Wiley, New York, p 358
- Francis SH, Zoraghi R, Kotera J, Ke H, Bessay EP, Blount MA, Corbin JD (2006) Phosphodiesterase-5: molecular characteristics relating to structure, function, and regulation. In: Beavo SHF JA, Houslay MD (eds) *Cyclic nucleotide phosphodiesterases in health and disease*. CRC, Boca Raton, pp 131–164
- Francis SH, Corbin JD, Bischoff E (2009) Cyclic GMP-hydrolyzing phosphodiesterases. *Handb Exp Pharmacol* (191): 367–408
- Fredholm BB (2010) Notes on the history of caffeine use. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Fujishige K, Kotera J, Michibata H, Yuasa K, Takebayashi S, Okumura K, Omori K (1999) Cloning and characterization of a novel human phosphodiesterase that hydrolyzes both cAMP and cGMP (PDE10A). *J Biol Chem* 274:18438–18445
- Galie N, Branzi A (2005) Pulmonary arterial hypertension: therapeutic algorithm. *Ital Heart J* 6:856–860
- Gamanuma M, Yuasa K, Sasaki T, Sakurai N, Kotera J, Omori K (2003) Comparison of enzymatic characterization and gene organization of cyclic nucleotide phosphodiesterase 8 family in humans. *Cell Signal* 15:565–574
- Gamm DM, Francis SH, Angelotti TP, Corbin JD, Uhler MD (1995) The type II isoform of cGMP-dependent protein kinase is dimeric and possesses regulatory and catalytic properties distinct from the type I isoforms. *J Biol Chem* 270:27380–27388

- Garst JE, Kramer GL, Wu YJ, Wells JN (1976) Inhibition of separated forms of phosphodiesterases from pig coronary arteries by uracils and by 7-substituted derivatives of 1-methyl-3-isobutylxanthine. *J Med Chem* 19:499–503
- Ghofrani HA, Voswinckel R, Reichenberger F, Weissmann N, Schermuly RT, Seeger W, Grimminger F (2006) Hypoxia- and non-hypoxia-related pulmonary hypertension - established and new therapies. *Cardiovasc Res* 72:30–40
- Gibson A (2001) Phosphodiesterase 5 inhibitors and nitregeric transmission-from zaprinast to sildenafil. *Eur J Pharmacol* 411:1–10
- Gillespie PG, Beavo JA (1988) Characterization of a bovine cone photoreceptor phosphodiesterase purified by cyclic GMP-sepharose chromatography. *J Biol Chem* 263:8133–8141
- Gotshall RW, Hamilton KL, Foreman B, van Patot MC, Irwin DC (2009) Glutaraldehyde-polymerized bovine hemoglobin and phosphodiesterase-5 inhibition. *Crit Care Med* 37:1988–1993
- Grant PG, Colman RW (1984) Purification and characterization of a human platelet cyclic nucleotide phosphodiesterase. *Biochemistry* 23:1801–1807
- Greene J, Paul W, Faller A (1937) The action of theophylline with ethylene diamine on intrathecal and venous pressures in cardiac failure and on bronchial obstruction in cardiac failure and in bronchial asthma. *JAMA* 109:1712–1715
- Guerreiro S, Toulorge D, Hirsch E, Marien M, Sokoloff P, Michel PP (2008) Paraxanthine, the primary metabolite of caffeine, provides protection against dopaminergic cell death via stimulation of ryanodine receptor channels. *Mol Pharmacol* 74:980–989
- Guerreiro S, Marien M, Michel PP (2010) Methylxanthines and ryanodine receptor channels. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Han P, Zhu X, Michaeli T (1997) Alternative splicing of the high affinity cAMP-specific phosphodiesterase (PDE7A) mRNA in human skeletal muscle and heart. *J Biol Chem* 272:16152–16157
- Hansen RS, Charbonneau H, Beavo JA (1988) Purification of calmodulin-stimulated cyclic nucleotide phosphodiesterase by monoclonal antibody affinity chromatography. *Methods Enzymol* 159:543–557
- Harrison SA, Reifsnnyder DH, Gallis B, Cadd GG, Beavo JA (1986) Isolation and characterization of bovine cardiac muscle cGMP-inhibited phosphodiesterase: a receptor for new cardiotonic drugs. *Mol Pharmacol* 29:506–514
- Haskó G, Cronstein B (2010) Methylxanthines and inflammatory cells. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Hatzimouratidis K (2008) Words of wisdom. Re: the efficacy and safety of udenafil, a new selective phosphodiesterase type 5 inhibitor, in patients with erectile dysfunction. Paick J-S, Kim SW, Yang DY, Kim JJ, Lee SW, Ahn TY, Choi HK, Suh J-K, Kim SC. *Eur Urol* 54:946–947
- Herrmann G, Aynesworth M, Martin J (1937) Successful treatment of persistent extreme dyspnea, “status asthmaticus”-use of theophylline enthlene diamine (aminophylline USP) intravenously. *Lab Clin Med* 23:135–148
- Hetman JM, Soderling SH, Glavas NA, Beavo JA (2000) Cloning and characterization of PDE7B, a cAMP-specific phosphodiesterase. *Proc Natl Acad Sci USA* 97:472–476
- Hirano T, Yamagata T, Gohda M, Yamagata Y, Ichikawa T, Yanagisawa S, Ueshima K, Akamatsu K, Nakanishi M, Matsunaga K, Minakata Y, Ichinose M (2006) Inhibition of reactive nitrogen species production in COPD airways: comparison between inhaled corticosteroid and oral theophylline. *Thorax* 61:761–766
- Huai Q, Liu Y, Francis SH, Corbin JD, Ke H (2004a) Crystal structures of phosphodiesterases 4 and 5 in complex with inhibitor 3-isobutyl-1-methylxanthine suggest a conformation determinant of inhibitor selectivity. *J Biol Chem* 279:13095–13101
- Huai Q, Wang H, Zhang W, Colman RW, Robinson H, Ke H (2004b) Crystal structure of phosphodiesterase 9 shows orientation variation of inhibitor 3-isobutyl-1-methylxanthine binding. *Proc Natl Acad Sci USA* 101:9624–9629

- Huang ZL, Qu WM, Eguchi N, Chen JF, Schwarzschild MA, Fredholm BB, Urade Y, Hayaishi O (2005) Adenosine A2A, but not A1, receptors mediate the arousal effect of caffeine. *Nat Neurosci* 8:858–859
- Huston E, Lumb S, Russell A, Catterall C, Ross AH, Steele MR, Bolger GB, Perry MJ, Owens RJ, Houslay MD (1997) Molecular cloning and transient expression in COS7 cells of a novel human PDE4B cAMP-specific phosphodiesterase, HSPDE4B3. *Biochem J* 328(Pt 2):549–558
- Interest CfSitP (2007) Caffeine content of food and drugs
- Ito K, Lim S, Caramori G, Cosio B, Chung KF, Adcock IM, Barnes PJ (2002) A molecular mechanism of action of theophylline: induction of histone deacetylase activity to decrease inflammatory gene expression. *Proc Natl Acad Sci USA* 99:8921–8926
- Jeong KH, Lee TW, Ihm CG, Lee SH, Moon JY, Lim SJ (2008) Effects of sildenafil on oxidative and inflammatory injuries of the kidney in streptozotocin-induced diabetic rats. *Am J Nephrol* 29:274–282
- Kaplan JM, Herzyk DJ, Ruggieri EV, Bartus JO, Esser KM, Bugelski PJ (1995) Effect of TNF alpha production inhibitors BRL 61063 and pentoxifylline on the response of rats to poly I:C. *Toxicology* 95:187–196
- Ke H, Wang H (2006) Structure, catalytic mechanism, and inhibitor selectivity of cyclic nucleotide phosphodiesterases. In: Beavo J, Francis SH, Houslay MD (eds) *Cyclic nucleotide phosphodiesterases in health and disease*. CRC, Boca Raton, pp 607–625
- Ke H, Wang H (2007) Crystal structures of phosphodiesterases and implications on substrate specificity and inhibitor selectivity. *Curr Top Med Chem* 7:391–403
- Kobayashi M, Nasuhara Y, Betsuyaku T, Shibuya E, Tanino Y, Tanino M, Takamura K, Nagai K, Hosokawa T, Nishimura M (2004) Effect of low-dose theophylline on airway inflammation in COPD. *Respirology* 9:249–254
- Kotera J, Fujishige K, Yuasa K, Omori K (1999) Characterization and phosphorylation of PDE10A2, a novel alternative splice variant of human phosphodiesterase that hydrolyzes cAMP and cGMP. *Biochem Biophys Res Commun* 261:551–557
- Kumar P, Francis GS, Wilson Tang WH (2009) Phosphodiesterase 5 inhibition in heart failure: mechanisms and clinical implications. *Nat Rev Cardiol* 6:349–355
- Lavan BE, Lakey T, Houslay MD (1989) Resolution of soluble cyclic nucleotide phosphodiesterase isoenzymes, from liver and hepatocytes, identifies a novel IBMX-insensitive form. *Biochem Pharmacol* 38:4123–4136
- Levien TL (2006) Phosphodiesterase inhibitors in Raynaud's phenomenon. *Ann Pharmacother* 40:1388–1393
- Loughney K, Martins TJ, Harris EA, Sadhu K, Hicks JB, Sonnenburg WK, Beavo JA, Ferguson K (1996) Isolation and characterization of cDNAs corresponding to two human calcium, calmodulin-regulated, 3',5'-cyclic nucleotide phosphodiesterases. *J Biol Chem* 271:796–806
- Loughney K, Hill TR, Florio VA, Uher L, Rosman GJ, Wolda SL, Jones BA, Howard ML, McAllister-Lucas LM, Sonnenburg WK, Francis SH, Corbin JD, Beavo JA, Ferguson K (1998) Isolation and characterization of cDNAs encoding PDE5A, a human cGMP-binding, cGMP-specific 3',5'-cyclic nucleotide phosphodiesterase. *Gene* 216:139–147
- Lubamba B, Lecourt H, Lebacq J, Lebecque P, De Jonge H, Wallemacq P, Leal T (2008) Preclinical evidence that sildenafil and vardenafil activate chloride transport in cystic fibrosis. *Am J Respir Crit Care Med* 177:506–515
- Martins TJ, Mumby MC, Beavo JA (1982) Purification and characterization of a cyclic GMP-stimulated cyclic nucleotide phosphodiesterase from bovine tissues. *J Biol Chem* 257:1973–1979
- Marwick JA, Wallis G, Meja K, Kuster B, Bouwmeester T, Chakravarty P, Fletcher D, Whittaker PA, Barnes PJ, Ito K, Adcock IM, Kirkham PA (2008) Oxidative stress modulates theophylline effects on steroid responsiveness. *Biochem Biophys Res Commun* 377:797–802
- Mascali JJ, Cvietusa P, Negri J, Borish L (1996) Anti-inflammatory effects of theophylline: modulation of cytokine production. *Ann Allergy Asthma Immunol* 77:34–38

- McAllister-Lucas LM, Sonnenburg WK, Kadlecak A, Seger D, LeTrong H, Colbran JL, Thomas MK, Walsh KA, Francis SH, Corbin JD, Beavo JA (1993) The structure of a bovine lung cGMP-binding, cGMP-specific phosphodiesterase deduced from a cDNA clone. *J Biol Chem* 268:22863–22873
- McKenna J, Muller G (2006) Medicinal chemistry of PDE4 inhibitors. In: Beavo SHF JA, Houslay MD (eds) *Cyclic nucleotide phosphodiesterases in health and disease*. CRC, Boca Raton, pp 667–699
- Medeiros JV, Gadelha GG, Lima SJ, Garcia JA, Soares PM, Santos AA, Brito GA, Ribeiro RA, Souza MH (2008) Role of the NO/cGMP/K(ATP) pathway in the protective effects of sildenafil against ethanol-induced gastric damage in rats. *Br J Pharmacol* 153:721–727
- Mery PF, Pavoine C, Pecker F, Fischmeister R (1995) Erythro-9-(2-hydroxy-3-nonyl)adenine inhibits cyclic GMP-stimulated phosphodiesterase in isolated cardiac myocytes. *Mol Pharmacol* 48:121–130
- Michaeli T, Bloom TJ, Martins T, Loughney K, Ferguson K, Riggs M, Rodgers L, Beavo JA, Wigler M (1993) Isolation and characterization of a previously undetected human cAMP phosphodiesterase by complementation of cAMP phosphodiesterase-deficient *Saccharomyces cerevisiae*. *J Biol Chem* 268:12925–12932
- Miyamoto K, Yamamoto Y, Kurita M, Sakai R, Konno K, Sanae F, Ohshima T, Takagi K, Hasegawa T, Iwasaki N et al (1993) Bronchodilator activity of xanthine derivatives substituted with functional groups at the 1- or 7-position. *J Med Chem* 36:1380–1386
- Miyamoto K, Kurita M, Sakai R, Sanae F, Wakusawa S, Takagi K (1994) Cyclic nucleotide phosphodiesterase isoenzymes in guinea-pig tracheal muscle and bronchorelaxation by alkylxanthines. *Biochem Pharmacol* 48:1219–1223
- Müller C, Jacobson KA (2010) Xanthines as adenosine receptor antagonists. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Mushlin P, Boerth RC, Wells JN (1981) Selective phosphodiesterase inhibition and alterations of cardiac function by alkylated xanthines. *Mol Pharmacol* 20:179–189
- Nakamizo T, Kawamata J, Yoshida K, Kawai Y, Kanki R, Sawada H, Kihara T, Yamashita H, Shibasaki H, Akaike A, Shimohama S (2003) Phosphodiesterase inhibitors are neuroprotective to cultured spinal motor neurons. *J Neurosci Res* 71:485–495
- O'Brien M, McCoy T, Rhodes S, Wagoner A, Wolfson M (2008) Caffeinated cocktails: energy drink consumption, high-risk drinking, and alcohol-related consequences among college students. *Acad Emerg Med* 15:453–460
- Ohta A, Sitkovsky M (2010) Methylxanthines, inflammation and cancer: Fundamental mechanisms. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Osswald H, Schnermann J (2010) Methylxanthines and the kidney. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Petersen RK, Madsen L, Pedersen LM, Hallenborg P, Hagland H, Viste K, Doskeland SO, Kristiansen K (2008) Cyclic AMP (cAMP)-mediated stimulation of adipocyte differentiation requires the synergistic action of Epac- and cAMP-dependent protein kinase-dependent processes. *Mol Cell Biol* 28:3804–3816
- Poppe H, Rybalkin SD, Rehmann H, Hinds TR, Tang XB, Christensen AE, Schwede F, Genieser HG, Bos JL, Doskeland SO, Beavo JA, Butt E (2008) Cyclic nucleotide analogs as probes of signaling pathways. *Nat Methods* 5:277–278
- Porkka-Heiskanen T (2010) Methylxanthines and sleep. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Puzzo D, Sapienza S, Arancio O, Palmeri A (2008) Role of phosphodiesterase 5 in synaptic plasticity and memory. *Neuropsychiatr Dis Treat* 4:371–387
- Rabe KF, Magnussen H, Dent G (1995) Theophylline and selective PDE inhibitors as bronchodilators and smooth muscle relaxants. *Eur Respir J* 8:637–642
- Regensteiner JG, Hiatt WR (2002) Treatment of peripheral arterial disease. *Clin Cornerstone* 4:26–40

- Rehmann H, Schwede F, Doskeland SO, Wittinghofer A, Bos JL (2003) Ligand-mediated activation of the cAMP-responsive guanine nucleotide exchange factor Epac. *J Biol Chem* 278:38548–38556
- Rena G, Begg F, Ross A, MacKenzie C, McPhee I, Campbell L, Huston E, Sullivan M, Houslay MD (2001) Molecular cloning, genomic positioning, promoter identification, and characterization of the novel cyclic amp-specific phosphodiesterase PDE4A10. *Mol Pharmacol* 59:996–1011
- Rieg T, Steigle H, Schnermann J, Richter K, Osswald H, Vallon V (2005) Requirement of intact adenosine A<sub>1</sub> receptors for the diuretic and natriuretic of the methylxanthines theophylline and caffeine. *J Pharmacol Exp Ther* 313:403–419
- Rosman GJ, Martins TJ, Sonnenburg WK, Beavo JA, Ferguson K, Loughney K (1997) Isolation and characterization of human cDNAs encoding a cGMP-stimulated 3',5'-cyclic nucleotide phosphodiesterase. *Gene* 191:89–95
- Rutten K, Van Donkelaar EL, Ferrington L, Blokland A, Bollen E, Steinbusch HW, Kelly PA, Prickaerts JH (2009) Phosphodiesterase inhibitors enhance object memory independent of cerebral blood flow and glucose utilization in rats. *Neuropsychopharmacology* 34:1914–1925
- Salanova M, Jin SC, Conti M (1998) Heterologous expression and purification of recombinant rolipram-sensitive cyclic AMP-specific phosphodiesterases. *Methods* 14:55–64
- Salloum FN, Takenoshita Y, Ockaili RA, Daoud VP, Chou E, Yoshida KI, Kukreja RC (2006) Sildenafil and vardenafil but not nitroglycerin limit myocardial infarction through opening of mitochondrial K(ATP) channels when administered at reperfusion following ischemia in rabbits. *J Mol Cell Cardiol* 42:453–458
- Salter H (1859) On some points in the treatment and clinical history of asthma. *Edinb Med J* 4:1109–1115
- Salter E, Wierzbicki A, Sperl G, Thompson WJ (2003) Quantum mechanical study of the *syn* and *anti* conformations of solvated cyclic GMP. *Struct Chem* 14:527–533
- Sasaki T, Kotera J, Yuasa K, Omori K (2000) Identification of human PDE7B, a cAMP-specific phosphodiesterase. *Biochem Biophys Res Commun* 271:575–583
- Sasaki T, Kotera J, Omori K (2002) Novel alternative splice variants of rat phosphodiesterase 7B showing unique tissue-specific expression and phosphorylation. *Biochem J* 361:211–220
- Sattin A (1971) Increase in the content of adenosine 3',5'-monophosphate in mouse forebrain during seizures and prevention of the increase by methylxanthines. *J Neurochem* 18:1087–1096
- Sattin A, Rall TW (1970) The effect of adenosine and adenine nucleotides on the cyclic adenosine 3',5'-phosphate content of guinea pig cerebral cortex slices. *Mol Pharmacol* 6:13–23
- Sawynok J, Yaksh TL (1993) Caffeine as an analgesic adjuvant: a review of pharmacology and mechanisms of action. *Pharmacol Rev* 45:43–85
- Scapin G, Patel SB, Chung C, Varnerin JP, Edmondson SD, Mastracchio A, Parmee ER, Singh SB, Becker JW, Van der Ploeg LH, Tota MR (2004) Crystal structure of human phosphodiesterase 3B: atomic basis for substrate and inhibitor specificity. *Biochemistry* 43:6091–6100
- Sekhar KR, Hatchett RJ, Shabb JB, Wolfe L, Francis SH, Wells JN, Jastorff B, Butt E, Chakinala MM, Corbin JD (1992) Relaxation of pig coronary arteries by new and potent cGMP analogs that selectively activate type I alpha, compared with type I beta, cGMP-dependent protein kinase. *Mol Pharmacol* 42:103–108
- Sekhar KR, Grondin P, Francis SH, Corbin JD (1996) Design and synthesis of xanthines and cyclic GMP analogues as potent inhibitors of PDE5. In: Schudt C, Dent G, Rabe KF (eds) *Phosphodiesterase inhibitors*. Academic, New York, pp 135–146
- Sharma RK, Wang JH (1986) Calmodulin and Ca<sup>2+</sup>-dependent phosphorylation and dephosphorylation of 63-kDa subunit-containing bovine brain calmodulin-stimulated cyclic nucleotide phosphodiesterase isozyme. *J Biol Chem* 261:1322–1328
- Sharma RK, Adachi AM, Adachi K, Wang JH (1984) Demonstration of bovine brain calmodulin-dependent cyclic nucleotide phosphodiesterase isozymes by monoclonal antibodies. *J Biol Chem* 259:9248–9254



- Silver PJ, Dundore RL, Bode DC, de Garavilla L, Buchholz RA, van Aller G, Hamel LT, Bacon E, Singh B, Leshner GY et al (1994) Cyclic GMP potentiation by WIN 58237, a novel cyclic nucleotide phosphodiesterase inhibitor. *J Pharmacol Exp Ther* 271:1143–1149
- Smellie FW, Davis CW, Daly JW, Wells JN (1979) Alkylxanthines: inhibition of adenosine-elicited accumulation of cyclic AMP in brain slices and of brain phosphodiesterase activity. *Life Sci* 24:2475–2482
- Smit HJ (2010) Theobromine and the pharmacology of cocoa. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Snyder PB, Florio VA, Ferguson K, Loughney K (1999) Isolation, expression and analysis of splice variants of a human  $Ca^{2+}$ /calmodulin-stimulated phosphodiesterase (PDE1A). *Cell Signal* 11:535–544
- Soderling SH, Bayuga SJ, Beavo JA (1998) Identification and characterization of a novel family of cyclic nucleotide phosphodiesterases. *J Biol Chem* 273:15553–15558
- Solinas M, Ferre S, You ZB, Karcz-Kubicha M, Popoli P, Goldberg SR (2002) Caffeine induces dopamine and glutamate release in the shell of the nucleus accumbens. *J Neurosci* 22:6321–6324
- Sonnenburg WK, Seger D, Kwak KS, Huang J, Charbonneau H, Beavo JA (1995) Identification of inhibitory and calmodulin-binding domains of the PDE1A1 and PDE1A2 calmodulin-stimulated cyclic nucleotide phosphodiesterases. *J Biol Chem* 270:30989–31000
- Stief C, Porst H, Saenz De Tejada I, Ulbrich E, Beneke M (2004) Sustained efficacy and tolerability with vardenafil over 2 years of treatment in men with erectile dysfunction. *Int J Clin Pract* 58:230–239
- Stief CG, Porst H, Neuser D, Beneke M, Ulbrich E (2008) A randomised, placebo-controlled study to assess the efficacy of twice-daily vardenafil in the treatment of lower urinary tract symptoms secondary to benign prostatic hyperplasia. *Eur Urol* 53:1236–1244
- Strassmaier T, Karpen JW (2007) Novel N7- and N1-substituted cGMP derivatives are potent activators of cyclic nucleotide-gated channels. *J Med Chem* 50:4186–4194
- Sullivan P, Bekir S, Jaffar Z, Page C, Jeffery P, Costello J (1994a) Anti-inflammatory effects of low-dose oral theophylline in atopic asthma. *Lancet* 343:1006–1008
- Sullivan S, Page CP, Costello JF (1994b) *Xanthines*. Raven, New York
- Sung BJ, Hwang KY, Jeon YH, Lee JI, Heo YS, Kim JH, Moon J, Yoon JM, Hyun YL, Kim E, Eum SJ, Park SY, Lee JO, Lee TG, Ro S, Cho JM (2003) Structure of the catalytic domain of human phosphodiesterase 5 with bound drug molecules. *Nature* 425:98–102
- Sutherland EW, Rall TW (1958) Fractionation and characterization of a cyclic adenosine ribonucleotide formed by tissue particles. *J Biol Chem* 232:1077–1091
- Takio K, Wade RD, Smith SB, Krebs EG, Walsh KA, Titani K (1984) Guanosine cyclic 3',5'-phosphate dependent protein kinase, a chimeric protein homologous with two separate protein families. *Biochemistry* 23:4207–4218
- Taniguchi Y, Tonai-Kachi H, Shinjo K (2006) Zaprinast, a well-known cyclic guanosine monophosphate-specific phosphodiesterase inhibitor, is an agonist for GPR35. *FEBS Lett* 580:5003–5008
- Thomas MK, Francis SH, Corbin JD (1990) Characterization of a purified bovine lung cGMP-binding cGMP phosphodiesterase. *J Biol Chem* 265:14964–14970
- Thomas MK, Francis SH, Beebe SJ, Gettys TW, Corbin JD (1992) Partial mapping of cyclic nucleotide sites and studies of regulatory mechanisms of phosphodiesterases using cyclic nucleotide analogues. *Adv Second Messenger Phosphoprotein Res* 25:45–53
- Thompson WJ (1991) Cyclic nucleotide phosphodiesterases: pharmacology, biochemistry and function. [Review] [195 refs]. *Pharmacol Ther* 51:13–33
- Tilley SL (2010) Methylxanthines in asthma. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Tjon JA, Riemann LE (2001) Treatment of intermittent claudication with pentoxifylline and cilostazol. *Am J Health Syst Pharm* 58:485–496

- Toque HA, Teixeira CE, Priviero FB, Morganti RP, Antunes E, De Nucci G (2008) Vardenafil, but not sildenafil or tadalafil, has calcium-channel blocking activity in rabbit isolated pulmonary artery and human washed platelets. *Br J Pharmacol* 154:787–796
- Torphy TJ, Livi GP, Balcarek JM, White JR, Chilton FH, Udem BJ (1992) Therapeutic potential of isozyme-selective phosphodiesterase inhibitors in the treatment of asthma. *Adv Second Messenger Phosphoprotein Res* 25:289–305
- Turko IV, Francis SH, Corbin JD (1998) Potential roles of conserved amino acids in the catalytic domain of the cGMP-binding cGMP-specific phosphodiesterase. *J Biol Chem* 273:6460–6466
- Turko IV, Ballard SA, Francis SH, Corbin JD (1999) Inhibition of cyclic GMP-binding cyclic GMP-specific phosphodiesterase (type 5) by sildenafil and related compounds. *Mol Pharmacol* 56:124–130
- Wallace DA, Johnston LA, Huston E, MacMaster D, Houslay TM, Cheung YF, Campbell L, Millen JE, Smith RA, Gall I, Knowles RG, Sullivan M, Houslay MD (2005) Identification and characterization of PDE4A11, a novel, widely expressed long isoform encoded by the human PDE4A cAMP phosphodiesterase gene. *Mol Pharmacol* 67:1920–1934
- Wang P, Myers JG, Wu P, Cheewatrakoolpong B, Egan RW, Billah MM (1997) Expression, purification, and characterization of human cAMP-specific phosphodiesterase (PDE4) subtypes A, B, C, and D. *Biochem Biophys Res Commun* 234:320–324
- Wang Y, Chackalamannil S, Hu Z, Boyle CD, Lankin CM, Xia Y, Xu R, Asberom T, Pissarnitski D, Stamford AW, Greenlee WJ, Skell J, Kurowski S, Vemulapalli S, Palamanda J, Chintala M, Wu P, Myers J, Wang P (2002) Design and synthesis of xanthine analogues as potent and selective PDE5 inhibitors. *Bioorg Med Chem Lett* 12:3149–3152
- Wang H, Liu Y, Chen Y, Robinson H, Ke H (2005) Multiple elements jointly determine inhibitor selectivity of cyclic nucleotide phosphodiesterases 4 and 7. *J Biol Chem* 280:30949–30955
- Wang H, Liu Y, Huai Q, Cai J, Zoraghi R, Francis SH, Corbin JD, Robinson H, Xin Z, Lin G, Ke H (2006) Multiple conformations of phosphodiesterase-5: implications for enzyme function and drug development. *J Biol Chem* 281:21469–21479
- Wang H, Liu Y, Hou J, Zheng M, Robinson H, Ke H (2007a) Structural insight into substrate specificity of phosphodiesterase 10. *Proc Natl Acad Sci USA* 104:5782–5787
- Wang H, Robinson H, Ke H (2007b) The molecular basis for different recognition of substrates by phosphodiesterase families 4 and 10. *J Mol Biol* 371:302–307
- Wang H, Yan Z, Yang S, Cai J, Robinson H, Ke H (2008a) Kinetic and structural studies of phosphodiesterase-8A and implication on the inhibitor selectivity. *Biochemistry* 47:12760–12768
- Wang H, Ye M, Robinson H, Francis SH, Ke H (2008b) Conformational variations of both phosphodiesterase-5 and inhibitors provide the structural basis for the physiological effects of vardenafil and sildenafil. *Mol Pharmacol* 73:104–110
- Weeks JL 2nd, Zoraghi R, Francis SH, Corbin JD (2007) N-Terminal domain of phosphodiesterase-11A4 (PDE11A4) decreases affinity of the catalytic site for substrates and tadalafil, and is involved in oligomerization. *Biochemistry* 46:10353–10364
- Wells JN, Kramer GL (1981) Phosphodiesterase inhibitors as tools in cyclic nucleotide research: a precautionary comment. *Mol Cell Endocrinol* 23:1–9
- Wells JN, Garst JE, Kramer GL (1981) Inhibition of separated forms of cyclic nucleotide phosphodiesterase from pig coronary arteries by 1, 3-disubstituted and 1, 3, 8-trisubstituted xanthines. *J Med Chem* 24:954–958
- Wernet W, Flockerzi V, Hofmann F (1989) The cDNA of the two isoforms of bovine cGMP-dependent protein kinase. *FEBS Lett* 251:191–196
- Wolfe L, Corbin JD, Francis SH (1989) Characterization of a novel isozyme of cGMP-dependent protein kinase from bovine aorta. *J Biol Chem* 264:7734–7741
- Wosilait WD, Sutherland EW (1956) The relationship of epinephrine and glucagon to liver phosphorylase. II. Enzymatic inactivation of liver phosphorylase. *J Biol Chem* 218:469–481

- Xu RX, Hassell AM, Vanderwall D, Lambert MH, Holmes WD, Luther MA, Rocque WJ, Milburn MV, Zhao Y, Ke H, Nolte RT (2000) Atomic structure of PDE4: insights into phosphodiesterase mechanism and specificity. *Science* 288:1822–1825
- Yan C, Zhao AZ, Bentley JK, Beavo JA (1996) The calmodulin-dependent phosphodiesterase gene PDE1C encodes several functionally different splice variants in a tissue-specific manner. *J Biol Chem* 271:25699–25706
- Yang JN, Bjorklund O, Lindstrom-Tornqvist K, Lindgren E, Eriksson TM, Kahlstrom J, Chen JF, Schwarzschild MA, Tobler I, Fredholm BB (2009a) Mice heterozygous for both A1 and A2A adenosine receptor genes show similarities to mice given long-term caffeine. *J Appl Physiol* 106:631–639
- Yang JN, Chen JF, Fredholm BB (2009b) Physiological roles of A1 and A2A adenosine receptors in regulating heart rate, body temperature, and locomotion as revealed using knockout mice and caffeine. *Am J Physiol Heart Circ Physiol* 296:H1141–H1149
- Yu L, Coelho JE, Zhang X, Fu Y, Tillman A, Karaoz U, Fredholm BB, Weng Z, Chen JF (2009) Uncovering multiple molecular targets for caffeine using a drug target validation strategy combining A2A receptor knockout mice with microarray profiling. *Physiol Genomics* 37:199–210
- Yuasa K, Ohgaru T, Asahina M, Omori K (2001) Identification of rat cyclic nucleotide phosphodiesterase 11A (PDE11A): comparison of rat and human PDE11A splicing variants. *Eur J Biochem* 268:4440–4448
- Zhang KYJ (2006) Crystal structure of phosphodiesterase families and the potential for rational drug design. In: Beavo J, Francis SH, Houslay MD (eds) *Cyclic nucleotide phosphodiesterase in health and disease*. CRC, Boca Raton, pp 583–605
- Zhang X, Feng Q, Cote RH (2005) Efficacy and selectivity of phosphodiesterase-targeted drugs in inhibiting photoreceptor phosphodiesterase (PDE6) in retinal photoreceptors. *Invest Ophthalmol Vis Sci* 46:3060–3066
- Zoraghi R, Corbin JD, Francis SH (2006) Phosphodiesterase-5 Gln817 is critical for cGMP, vardenafil, or sildenafil affinity: its orientation impacts cGMP but not cAMP affinity. *J Biol Chem* 281:5553–5558
- Zoraghi R, Francis SH, Corbin JD (2007) Critical amino acids in phosphodiesterase-5 catalytic site that provide for high-affinity interaction with cGMP and inhibitors. *Biochemistry* 46:13554–13563

# Methylxanthines and Ryanodine Receptor Channels

Serge Guerreiro, Marc Marien, and Patrick P. Michel

## Contents

1	RyR Channels: Regulation and Function .....	137
2	Methylxanthines and RyR Channels in Muscle Cells .....	138
2.1	Effects of Caffeine on Muscle Cell Contractility .....	138
2.2	Effects of Other Naturally Occurring Methylxanthines .....	140
2.3	Effects of Synthetic Methylxanthines .....	141
3	Methylxanthines and RyRs in Neuronal Cells .....	141
3.1	Stimulation of ER Calcium Release .....	141
3.2	Impact on Neurotransmitter Release .....	142
3.3	Impact on Neuronal Survival .....	143
4	Are the Effects of Methylxanthines In Vitro Physiologically Relevant In Vivo? .....	145
	References .....	147

**Abstract** Methylxanthines of either natural or synthetic origin have a number of interesting pharmacological features. Proposed mechanisms of methylxanthine-induced pharmacological effects include competitive antagonism of G-coupled adenosine A<sub>1</sub> and A<sub>2A</sub> receptors and inhibition of phosphodiesterases. A number

---

S. Guerreiro

Université Pierre et Marie Curie-Paris 6, Centre de Recherche de l'Institut du Cerveau et de la Moelle Epinière, UMR-S975 Paris, France

Institut National de la Santé et de la Recherche Médicale, U-975 Paris, France

Centre National de la Recherche Scientifique, UMR 7225 Paris, France

M. Marien

Institut de Recherche Pierre Fabre, Castres, France

P.P. Michel (✉)

Université Pierre et Marie Curie-Paris 6, Centre de Recherche de l'Institut du Cerveau et de la Moelle Epinière, UMR-S975 Paris, France

Institut National de la Santé et de la Recherche Médicale, U-975 Paris, France

Centre National de la Recherche Scientifique, UMR 7225 Paris, France

Institut de Recherche Pierre Fabre, Castres, France

e-mail: patrick-pierre.michel@upmc.fr

of studies have indicated that methylxanthines also exert effects through alternative mechanisms, in particular via activation of sarcoplasmic reticulum or endoplasmic reticulum ryanodine receptor (RyR) channels. More specifically, RyR channel activation by methylxanthines was reported (1) to stimulate the process of excitation coupling in muscle cells, (2) to augment the excitability of neurons and thus their capacity to release neurotransmitters, and also (3) to improve their survival. Here, we address the mechanisms by which methylxanthines control RyR activation and we consider the pharmacological consequences of this activation, in muscle and neuronal cells.

**Keywords** Calcium · Methylxanthines · Muscle cell contraction · Neuroprotection · Neurotransmitter release · Ryanodine receptors

## Abbreviations

$\text{Ca}_{\text{cyt}}^{2+}$	Cytoplasmic $\text{Ca}^{2+}$
ER	Endoplasmic reticulum
RyR	Ryanodine receptor
SR	Sarcoplasmic reticulum

Methylxanthines are purine derivatives that have in common a xanthine core molecule with methyl groups attached in various combinations to nitrogens, at positions 1, 3, 7, or 9. Methylxanthines are generally from natural origin but a number of synthetic congeners have also been designed. The most common methylxanthines are caffeine (1,3,7-trimethylxanthine) and its hypomethylated analogues theophylline (1,3-dimethylxanthine), theobromine (3,7-dimethylxanthine), and paraxanthine (1,7-dimethylxanthine). Caffeine, theophylline, and theobromine are produced by different plant species, whereas paraxanthine occurs essentially as a caffeine metabolite in biological fluids (Magkos and Kavouras 2005)<sup>1</sup>.

Proposed mechanisms of methylxanthine-induced pharmacological actions include competitive antagonism of G-coupled adenosine  $A_1$  and  $A_{2A}$  receptors and also inhibition of phosphodiesterases (Jacobson and Gao 2006; Daly 2007)<sup>2</sup>. A number of studies have shown, however, that methylxanthines can exert effects through alternative mechanisms, in particular via the activation of sarcoplasmic reticulum (SR) or endoplasmic reticulum (ER) ryanodine receptor (RyR) channels. RyR channel activation by methylxanthines was reported originally to stimulate

---

<sup>1</sup>Botanical sources and bioconversion pathways of methylxanthines are described in detail in Ashihara et al. (2010) and Arnaud (2010), respectively.

<sup>2</sup>See Francis et al. (2010) for details on the pharmacological effects of methylxanthines that are mediated by adenosine receptor blockade or phosphodiesterase inhibition.

excitation coupling in muscle cells, a process that connects myocyte membrane depolarization and muscle contraction (Fill and Copello 2002; Zalk et al. 2007). Other studies have shown that methylxanthines can augment the excitability of neurons and thus their capacity to release neurotransmitters (Pessah et al. 1987) and may improve their survival as well (Guerreiro et al. 2008). Note that methylxanthines were also found to activate RyRs in other cell types, including pancreatic  $\beta$  cells (Bruton et al. 2003) and T cells (Ritter et al. 2001), but work on the subject is relatively limited.

The aim of this chapter is to provide an overview of the mechanisms by which methylxanthines trigger RyR channel activation in muscle and neuronal cells and to report on the pharmacological consequences of this activation. As a first step, it appears pertinent, however, to begin with a brief description of the mechanisms involved in the release of calcium ( $\text{Ca}^{2+}$ ) via RyR channels.

## 1 RyR Channels: Regulation and Function

RyR channels are intracellular  $\text{Ca}^{2+}$  channels that share some structural and functional similarities with inositol 1,4,5-trisphosphate receptors (Fill and Copello 2002). They are large tetrameric channel proteins sharing a rather peculiar four-leaf-clover-like structure when observed by electron microscopy (Fill and Copello 2002; Zalk et al. 2007). The large cytoplasmic domain of RyRs serves as a scaffold for proteins that bind to the channel and modulate its function. The RyR family comprises three major receptor subtypes. The three channel subtypes are encoded by three distinct genes that share approximately 70% sequence homology. The predominant isoform in skeletal muscle is RyR<sub>1</sub>, and in cardiac muscle it is RyR<sub>2</sub>. In neurons, RyR<sub>1</sub>, RyR<sub>2</sub>, and RyR<sub>3</sub> are all present. All three RyRs can be activated by  $\text{Ca}^{2+}$  from the cytosolic side. In addition, RyR channels are modulated by numerous factors, including a number of physiological agents (e.g., ATP, adenosine, cyclic ADP ribose, and  $\text{Mg}^{2+}$ ) and various cellular processes (e.g., redox state and energy charge phosphorylation) (Sitsapesan et al. 1995; Zalk et al. 1997).

RyR channels directly control intracellular  $\text{Ca}^{2+}$  release in skeletal and cardiac muscle cells, activating muscle contraction during excitation–contraction coupling. In neurons RyR channels can modulate neurotransmitter release. Two mechanisms account for RyR-mediated  $\text{Ca}^{2+}$  release (Verkhatsky 2005; Zalk et al. 2007).  $\text{Ca}^{2+}$  release through RyRs can result from a direct physical interaction between plasmalemmal voltage-gated  $\text{Ca}^{2+}$  channels located on the transverse tubule membrane of striated muscle cells and RyR<sub>1</sub> channels on the SR (Fill and Copello 2002; Magkos and Kavouras 2005). Alternatively, in cardiac cells and also in neurons,  $\text{Ca}^{2+}$  release is triggered through a process known as  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release (Fabiato 1983). In that case, the activation of voltage-gated channels triggers a small elevation in the level of cytoplasmic  $\text{Ca}^{2+}$  ( $\text{Ca}_{\text{cyt}}^{2+}$ ), which in turn serves to activate  $\text{Ca}^{2+}$  release through RyR activation. Ligand-gated  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release makes the kinetics of intracellular  $\text{Ca}^{2+}$  release

slower when compared with protein-mediated activation of RyR<sub>1</sub> in skeletal muscle (Näbauer et al. 1989).

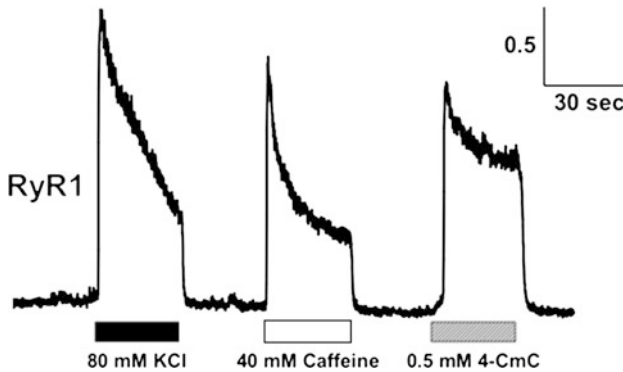
The RyR channel binds specifically the plant alkaloid ryanodine, which is the reason for its name. Ryanodine binds the channel with high affinity in a Ca<sup>2+</sup>-dependent manner (Sutko et al. 1997). The two ryanodine-binding sites, a high-affinity and a low-affinity binding site, which have been described at the C-terminus of the receptor may account for the fact that low micromolar concentrations of ryanodine cause the stimulation of RyR channels, while higher concentrations have the opposite effect and can even provoke their complete closure (Buck et al. 1992). Whereas ryanodine operates as preferential ligand, methylxanthines are also potent activators of RyR channels. The mechanisms by which methylxanthines cause the activation of RyR and the pharmacological consequences of this activation are described hereafter.

## 2 Methylxanthines and RyR Channels in Muscle Cells

### 2.1 *Effects of Caffeine on Muscle Cell Contractility*

The first evidence that caffeine could increase the contractility of isolated skeletal muscles dates back to experiments performed by Veley and Waller (1910). Subsequent studies using muscle preparations demonstrated that contractures induced by caffeine can occur in the absence of external Ca<sup>2+</sup> and without any change in resting membrane action potential (Conway and Sakai 1960; reviewed in Magkos and Kavouras 2005). These early studies showed that caffeine's effects were dose-dependent, and that threshold effective concentrations for activation were on the order of 1–2 mM, with a peak activation occurring at 5–10 mM.

It again took a number of years before several groups could demonstrate that caffeine operates by releasing Ca<sup>2+</sup> from ER stores (Endo 1977; Fabiato 1983; Kirino and Shimizu 1982). The site of caffeine's action was ascribed to RyR channels using preparations in which the channel proteins from striatal or cardiac muscles were incorporated into an artificial planar lipid bilayer (Pessah et al. 1987; Hymel et al. 1988; Meissner et al. 1988; Xu and Meissner 1988). This explains why cultured myotubes isolated from a dyspedic (IB5) mouse lacking RyR<sub>1</sub> channels were insensitive to caffeine, while the response to caffeine could be restored in this preparation by transfection with a complementary DNA encoding RyR<sub>1</sub> (Fessenden et al. 2006) (Fig. 1). A number of studies demonstrated that the activation site for caffeine was within the pore region of RyRs channels, i.e., in the C-terminal portion of the protein (Du and MacLennan 1999). In particular, several mutations in the C-terminal portion of RyR<sub>1</sub> have been reported to modulate the response of RyR<sub>1</sub> channels to caffeine in central core disease, an autosomal dominant, slow progressive or nonprogressive congenital myopathy (Du et al. 2004). Furthermore,



**Fig. 1** Caffeine-induced elevation of intracellular calcium levels in cultured 1B5 myotubes transfected with the ryanodine receptor isoform RyR<sub>1</sub>. Caffeine stimulates the release of sarcoplasmic reticulum calcium by directly activating RyR<sub>1</sub> channels. An elevated K<sup>+</sup> level was initially applied to the preparation to test for depolarization-induced calcium release, e.g., calcium release occurring via RyR<sub>1</sub>, as the result of the activation of plasmalemmal voltage-gated calcium channels. Note that untransfected 1B5 myotubes cells are insensitive to both caffeine and depolarization induced by high K<sup>+</sup> concentration as they derive from RyR null/dyspedic mice. The phenol-based compound 4-chloro-*m*-cresol, which is unrelated structurally to methylxanthines, can also operate as a potent activator of RyRs. The figure depicts changes in Fluo-4 fluorescence (F) normalized to the average resting fluorescence (F<sub>0</sub>) of the cell. Calibration bar = 0.5 F/F<sub>0</sub> arbitrary units versus 30 s. (Data from Fessenden et al. 2006 adapted with permission)

site-directed mutagenesis of the same region in RyR<sub>2</sub> channels generated mutants that were insensitive to caffeine (Wang et al. 2004).

Rousseau et al. (1988) demonstrated that caffeine increases the channel mean open time and open probability of both skeletal (RyR<sub>1</sub>) and cardiac (RyR<sub>2</sub>) SR Ca<sup>2+</sup> release channels but not their conductance. The response to caffeine was like that of ryanodine, in that it was suppressed by Mg<sup>2+</sup> and ruthenium red (Zucchi and Ronca-Testoni 1997). The regulatory effect of caffeine on excitation–contraction coupling appeared to be bimodal with (1) a Ca<sup>2+</sup>-dependent activation with threshold concentrations greater than 100 μM caffeine making RyRs (RyR<sub>1</sub> and RyR<sub>2</sub>) more sensitive to endogenous activators (e.g., Ca<sup>2+</sup>, ATP, ADP, AMP, adenosine) (Pessah et al. 1987; Magkos and Kavouras 2005) and (2) a Ca<sup>2+</sup>-independent activation observed at high (millimolar) caffeine concentrations (Sitsapesan and Williams 1990; Herrmann-Frank et al. 1999). When applied to SR vesicles, caffeine caused a significant increase in the efflux rate of Ca<sup>2+</sup> when Ca<sup>2+</sup> in the release medium was in the range of nanomolar concentrations. However, when the external Ca<sup>2+</sup> concentration was set at micromolar concentrations or above, caffeine had no stimulatory effect on Ca<sup>2+</sup> mobilization (Meissner and Henderson 1987; Moutin and Dupont 1988). It is worth noting that caffeine-mediated Ca<sup>2+</sup> release via RyRs was more sensitive to cytosolic Ca<sup>2+</sup> in cardiac preparations than in skeletal SR preparations (Meissner and Henderson 1987).



In most cases, the effect of caffeine reproduced that of adenine nucleotides. However, several lines of evidence suggested that caffeine and adenine nucleotides acted on different but possibly interacting sites: (1) caffeine and adenine nucleotides had synergistic effects on channel gating; (2) adenine and adenosine inhibited the response to ATP analogues possibly by competition, yet they stimulated the response to caffeine (McGarry and Williams 1994). The phenol-based compound 4-chloro-*m*-cresol, which is unrelated structurally to methylxanthines, also reproduced some of the effects of caffeine on RyRs (Fessenden et al. 2006). Similar to caffeine, 4-chloro-*m*-cresol activated RyR<sub>1</sub> by increasing the affinity of Ca<sup>2+</sup> binding to its activator site. Yet, these two compounds differed in that 4-chloro-*m*-cresol was approximately 25 times more potent than caffeine in activating RyR. The activation profiles of caffeine and 4-chloro-*m*-cresol were also different: whereas 4-chloro-*m*-cresol was a much more effective activator of RyR<sub>1</sub> and RyR<sub>2</sub> compared with RyR<sub>3</sub> (Fessenden et al. 2006), caffeine was much more potent in activating RyR<sub>2</sub> and RyR<sub>3</sub> compared with RyR<sub>1</sub> (Du and MacLennan 1999).

Finally, unlike many receptors that are activated only transiently and undergo a rapid process of desensitization (Kelly et al. 2008), skeletal and cardiac RyR channels did not undergo conventional inactivation upon caffeine exposure. Indeed, multiple additions of submaximal concentrations of caffeine could each induce a partial and transient Ca<sup>2+</sup> release from intracellular Ca<sup>2+</sup> stores in cells expressing RyRs, a phenomenon known as “quantal” Ca<sup>2+</sup> release (Dettbarn and Palade 1997; Kong et al. 2008). These observations can be explained by assuming either that different Ca<sup>2+</sup> release channels have different sensitivities to caffeine or that the SR is composed of several compartments, which are discharged by different concentrations of caffeine (Verkhatsky 2005).

## 2.2 *Effects of Other Naturally Occurring Methylxanthines*

Theophylline, theobromine, and paraxanthine, which occurs essentially as a metabolite of caffeine, shared several of the properties of caffeine toward RyRs. They all stimulated the release of <sup>45</sup>Ca<sup>2+</sup> from heavy SR vesicles and they also increased [<sup>3</sup>H]ryanodine binding to skeletal muscle SR vesicles (Liu and Meissner 1997). Rousseau et al. (1988) reported that paraxanthine, theobromine, and theophylline were more effective than caffeine in stimulating Ca<sup>2+</sup> release. In the same preparation, 1,3-dimethyluracil was ineffective, suggesting that the integrity of the imidazole ring (methylxanthines are formed from a pyrimidine-dione ring and an imidazole ring fused together) was required for this process. Hawke et al. (2000) also demonstrated that paraxanthine, theophylline, and theobromine were all able to transiently increase Ca<sub>cyt</sub><sup>2+</sup> concentrations to subcontracture levels in resting mammalian skeletal muscles, in a concentration-dependent fashion. The ability of procaine to inhibit this effect confirmed that the SR was the primary source for the increased Ca<sub>cyt</sub><sup>2+</sup> concentrations.

### 2.3 *Effects of Synthetic Methylxanthines*

Two methylxanthines which are structurally related to theophylline and paraxanthine, 3-isobutyl-1-methylxanthine and 7-isobutyl-1-methylxanthine, respectively, retained the ability of their parent compounds to stimulate  $\text{Ca}^{2+}$  release through RyR channels in SR vesicles (Rousseau et al. 1988; Wyskovsky 1994). Rousseau et al. (1988) also reported that among methylxanthines having an *N*-methyl substitution at position 9, 3,9-dimethylxanthine was as potent as caffeine in release experiments, whereas 1,9-dimethylxanthine was minimally effective. Using a preparation of sea urchin egg homogenates, Cavallaro et al. (1999) also compared the potential of a number of synthetic caffeine derivatives to potentiate the effects of cyclic ADP ribose, a metabolite of  $\text{NAD}^+$ , which is also a physiological activator of RyRs (Galione et al. 1991; Lee 1993). These compounds had their methyl substituent at positions 1 or 7 replaced with alkyl chains containing different functional groups. Two of these compounds, 1,3-dimethyl-7-(7-hydroxyoctyl)xanthine and 3-methyl-7-(7-oxooctyl)-1-propargylxanthine, were more potent than caffeine itself in potentiating cyclic ADP ribose induced  $\text{Ca}^{2+}$  release, while 1,3-dimethyl-7-(5-ethylcarboxypentyl)xanthine was more efficacious (Cavallaro et al. 1999).

## 3 Methylxanthines and RyRs in Neuronal Cells

### 3.1 *Stimulation of ER Calcium Release*

A number of studies have also shown that methylxanthines, including caffeine, theophylline, and 3-isobutyl-1-methylxanthine, can cause elevations of  $\text{Ca}_{\text{cyt}}^{2+}$  concentrations in ganglionic neuronal cells via activation of RyRs (McPherson et al. 1991; Usachev et al. 1993; Usachev and Verkhatsky 1995; Walz et al. 1995; Smith and Cunnane 1996). The contribution of RyR channels to this effect was established by showing that  $\text{Ca}_{\text{cyt}}^{2+}$  transients evoked by methylxanthines were blocked by high concentrations of ryanodine, by procaine, and dantrolene as well. As expected, the response to methylxanthines persisted in  $\text{Ca}^{2+}$ -free extracellular solutions. Similar to what was observed in muscle cells, caffeine and other methylxanthines appeared to sensitize RyR channels to  $\text{Ca}_{\text{cyt}}^{2+}$  ions so that  $\text{Ca}^{2+}$  release could develop even at resting  $\text{Ca}_{\text{cyt}}^{2+}$  concentrations.

Of importance, methylxanthines also caused the activation of RyRs in brain neurons, in particular hippocampal neurons, midbrain dopamine neurons, and Purkinje cells (Kano et al. 1995; Garaschuk et al. 1997; Sharma and Vijayaraghavan 2003; Patel et al. 2009). Caffeine was generally sufficient to trigger  $\text{Ca}^{2+}$  elevations at resting  $\text{Ca}_{\text{cyt}}^{2+}$  concentrations (Sharma and Vijayaraghavan 2003; Patel et al. 2009). However, in Purkinje cells from rat cerebellar slices, application of 20 mM caffeine to the resting cells rarely resulted in  $\text{Ca}^{2+}$  release. Yet, when

the  $\text{Ca}_{\text{cyt}}^{2+}$  concentration was slightly elevated by moderate depolarization (experiments were done under voltage clamp) caffeine produced robust  $\text{Ca}_{\text{cyt}}^{2+}$  concentration elevations (Kano et al. 1995). Similarly, in  $\text{CA}_1$  hippocampal neurons, a small elevation of  $\text{Ca}_{\text{cyt}}^{2+}$  concentration prior to caffeine treatment led to a several-fold increase of the subsequent caffeine-induced  $\text{Ca}_{\text{cyt}}^{2+}$  response (Garaschuk et al. 1997).

### 3.2 *Impact on Neurotransmitter Release*

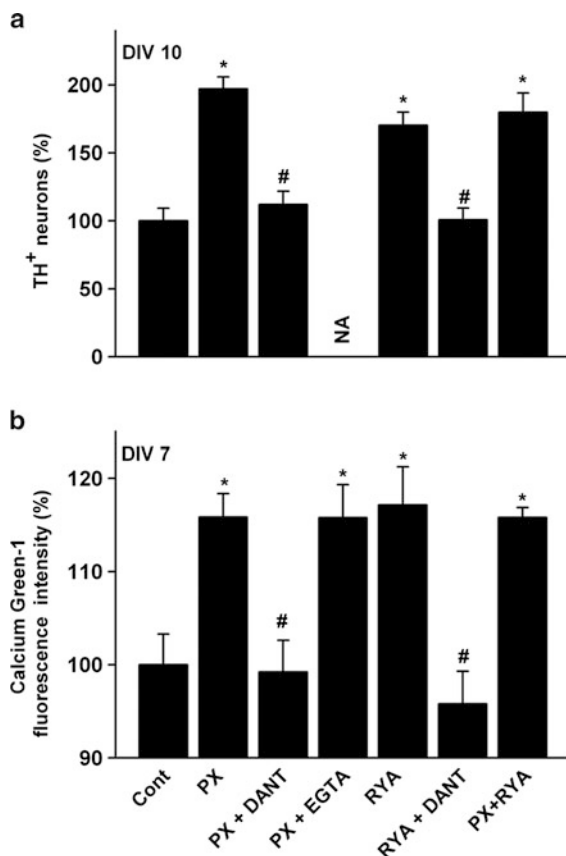
The transient increase in  $\text{Ca}^{2+}$  which triggers exocytosis after invasion of nerve terminals by the action potential is thought to arise principally through the activation of voltage-dependent  $\text{Ca}^{2+}$  channels located close to the sites of neurotransmitter release. However, the demonstration that methylxanthines can cause elevations of  $\text{Ca}_{\text{cyt}}^{2+}$  concentrations in neuronal cells indicated that RyR channels might also possibly intervene in neurotransmitter release (Galante and Marty 2003). Consistent with this view, the application of caffeine onto mossy fiber nerve terminals in hippocampal slices led to a burst of high-frequency and large-amplitude glutamatergic miniature excitatory postsynaptic currents in  $\text{CA}_3$  pyramidal neurons (Sharma and Vijayaraghavan 2003). A similar effect was also observed when nicotine was substituted for caffeine to activate presynaptic nicotinic receptors, indicating that glutamate release occurred via a mechanism that was independent of incoming action potentials. Demonstrating an obligatory requirement for stored  $\text{Ca}^{2+}$  release via RyR channels in this process, caffeine and nicotine responses were absent after depleting intracellular  $\text{Ca}^{2+}$  stores with thapsigargin, and the effect of nicotine was abolished by a high concentration of ryanodine (Sharma and Vijayaraghavan 2003). It is worth noting that caffeine also exerted a facilitatory effect on the release of glutamate evoked by 4-aminopyridine, a potassium channel blocker that depolarizes nerve terminals *in vitro*, in a manner corresponding to *in vivo* depolarization. Yet, this effect required presynaptic adenosine  $\text{A}_1$  receptor blockade by caffeine and was independent of RyR- $\text{Ca}^{2+}$  stores (Wang 2007).

Caffeine was also reported to stimulate somatodendritic release of dopamine when applied to dopamine cell bodies of the substantia nigra pars compacta in midbrain slices (Patel et al. 2009). This effect was prevented either by the RyR blocker dantrolene or by the inhibitor of the ER  $\text{Ca}^{2+}$ -ATPase cyclopiazonic acid that causes depletion of  $\text{Ca}^{2+}$  stores, suggesting that caffeine operated via activation of RyR channels (Patel et al. 2009). As expected, the effect of caffeine was accompanied by a concomitant rise in  $\text{Ca}_{\text{cyt}}^{2+}$  concentration in dopamine neurons. This finding is also in agreement with reports showing that somatodendritic release of dopamine in the substantia nigra pars compacta shows limited dependence on extracellular  $\text{Ca}^{2+}$  concentration, at variance with what is observed for axonal release of dopamine in the striatum (Chen et al. 2006). Likewise, caffeine was also reported to stimulate somatodendritic release of serotonin in leech Retzius neurons (Trueta et al. 2004).

### 3.3 *Impact on Neuronal Survival*

The mechanisms by which methylxanthines cause RyR channel activation and calcium release have been essentially studied with respect to skeletal and cardiac muscle contraction and neurotransmitter release. However, intracellular  $\text{Ca}^{2+}$  signaling also intervenes crucially in the control of neuronal survival during development and also in the adult brain (Michel et al. 2007). Therefore, one may assume that RyR channels might play a role in neurodegenerative conditions of aging and that methylxanthines may also have an impact in these disorders. This may be particularly true in the case of Parkinson's disease: (1) the CD157 gene which encodes an ectoenzyme that catalyses the production of the RyR channel endogenous agonist cyclic ADP-ribose, is a susceptibility gene for this disorder (Satake et al. 2009); (2) results of case-control and prospective studies have indicated that consumption of caffeine in coffee, tea, and caffeinated beverages may significantly reduce the risk of developing the disease after accounting for smoking and other potentially confounding factors such as estrogen replacement therapy (Ascherio et al. 2001; Elbaz and Moisan 2008; Sääksjärvi et al. 2008).

Caffeine and its primary metabolite paraxanthine produced through liver enzymatic biotransformation were shown to exert protective effects for dopamine neurons in mouse models of Parkinson's disease, but these effects were attributed to the antagonistic action of these compounds for  $\text{A}_{2\text{A}}$  adenosine receptors (Xu et al. 2010; Xu et al. 2006; Jacobson and Gao 2006). Using a model system of spontaneous dopaminergic cell death, we have shown, however, that paraxanthine was protective by a mechanism that was unrelated to adenosine receptor blockade (Guerreiro et al. 2008). We also excluded a possible mechanism involving the action of glial-cell-line-derived neurotrophic factor (Guerreiro et al. 2008), a trophic peptide which exerts neurotrophic effects for dopamine neurons in the developing and aging brain (Love et al. 2005; Kramer et al. 2007). In fact, a number of arguments suggested that survival promotion by paraxanthine was mediated by RyR channel activation (Fig. 2): (1) ryanodine mimicked the rescuing effects of paraxanthine; (2) survival-promoting effects of paraxanthine or ryanodine were abolished by blockade of RyRs with dantrolene; (3) the rescuing effects of both paraxanthine and ryanodine caused a moderate rise in  $\text{Ca}_{\text{cyt}}^{2+}$  concentration which was also prevented by dantrolene; (4) the  $\text{Ca}^{2+}$  rise persisted in the absence of extracellular  $\text{Ca}^{2+}$ . Unlike paraxanthine, caffeine provided only limited neuroprotection to dopamine neurons in this preparation. Of interest, the activation of RyR channels by paraxanthine also provided protection in an unrelated paradigm of dopamine cell death in which neuronal demise was triggered by the mitochondrial complex I inhibitor 1-methyl-4-phenylpyridinium (Guerreiro et al. 2008), the active metabolite of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine, a toxin which can cause irreversible parkinsonian symptoms in man when self-administrated accidentally (Langston et al. 1999). Interestingly, ryanodine was also neuroprotective in the same paradigm (Guerreiro et al. 2008). Altogether, these data suggest that some of



**Fig. 2** Role of RyR calcium release channels in the neuroprotective effect of paraxanthine on dopamine neurons in culture. **(a)** Number of dopamine (tyrosine hydroxylase immunopositive) neurons at 10 days in vitro in midbrain cultures exposed chronically to optimal concentrations of paraxanthine (PX) or ryanodine (RYA) in the presence or absence of dantrolene (DANT), an antagonist of RyR channels. **(b)** Calcium Green-1 fluorescence levels in 7 day in vitro cultures exposed for a short time to the same treatments as in **a**. Some measurements were performed in the presence of 3 mM ethylene glycol bis(2-aminoethyl ether)tetraacetic acid to eliminate extracellular calcium from the culture medium. Note that the effects of paraxanthine and ryanodine were not additive. Paraxanthine was used at 800  $\mu$ M, ryanodine at 10  $\mu$ M, and dantrolene at 30  $\mu$ M. \* $P < 0.05$ , compared with corresponding control (cont) cultures; # $P < 0.05$ , significant inhibition of the effect of the test compound (One-way ANOVA followed by Student Newman Keuls post-hoc test). NA not applicable, since widespread degeneration occurs when low-calcium conditions are maintained for prolonged periods of time in our culture model. (Data reproduced from Guerreiro et al. 2008 with permission)

the protective effects of caffeine reported earlier in epidemiology studies on Parkinson's disease might be due in part to paraxanthine acting through the activation of RyR channels. This hypothesis needs, however, to be further substantiated by in vivo studies.

Additional studies established that the N7- and N1-demethylated metabolites of paraxanthine, 1-methylxanthine, and 7-methylxanthine, were either less potent than paraxanthine itself or totally inactive, respectively. Incidentally, these results and the observation that 3-methylxanthine and xanthine itself failed to protect dopamine neurons, also indicated that N-substitution at position 1 was required for survival promotion by methylxanthine derivatives and that this effect was largely modulated by the presence of other N-substituents. Of interest, the extent of neuroprotection afforded by methylxanthines, including caffeine, was strictly correlated to their efficacy to elevate  $\text{Ca}_{\text{cyt}}^{2+}$  levels via RyR activation. It is worth noting that the  $\text{Ca}^{2+}$  level elevation elicited by optimal concentrations of paraxanthine (about 15% above control levels) was relatively modest and much lower than that produced by concentrations of glutamate causing excitotoxic stress (Guerreiro et al. 2008). Overall, these results are consistent with the idea that paraxanthine helped to maintain  $\text{Ca}_{\text{cyt}}^{2+}$  within levels that are optimal for the survival of dopamine neurons (Douhou et al. 2001; Salthun-Lassalle et al. 2004). The effects of methylxanthines on RyR channels reported in this paper are summarized in Table 1.

#### 4 Are the Effects of Methylxanthines In Vitro Physiologically Relevant In Vivo?

In summary, the effects of methylxanthines on RyR channels are fairly well documented. In particular, methylxanthines are useful probes to explore the effects of RyR channel activation in striated and cardiac muscle cells and in neuronal cells. The possibility that RyR modulation may also play a role in the physiological or therapeutical response to methylxanthines remains, however, speculative. Indeed, the concentrations of methylxanthines required to elicit RyR receptor activation in vitro, which are generally in the range 0.3–20 mM, are several-fold higher than corresponding plasma and brain concentrations (Hawke et al. 2000; Guerreiro et al. 2008). Nevertheless, a variety of endogenous modulators, such as fatty acyl-CoA esters, cyclic ADP ribose, and ATP have been reported to sensitize RyRs to exogenous agonists, suggesting that the effects of methylxanthines on these receptors may manifest themselves in vivo at lower concentrations than those determined from in vitro studies (Magkos and Kavouras 2005). Consistent with this view, paraxanthine was reported to increase  $\text{Ca}_{\text{cyt}}^{2+}$  concentrations in intact skeletal muscle preparations at 10  $\mu\text{M}$ , a concentration that is physiologically relevant after systemic administration of its parent compound caffeine (Ferré et al. 1990), whereas concentrations 50-fold higher were required to produce the same effect on single isolated muscle cells (Hawke et al. 2000). In addition, twitch potentiation has also been documented in isolated rat soleus muscle after exposure to 10–100  $\mu\text{M}$  caffeine (Connett et al. 1983), and in frog semitendinous muscles in the presence of 50  $\mu\text{M}$  of the theophylline dimer aminophylline (Block et al. 1992).

**Table 1** Comparative effects of methylxanthines on ryanodine receptor (*RyR*) channels

Common names of methylxanthines	<i>N</i> -Methyl substituents	Other substituents	RyR activation	References <sup>a</sup>
<b>From plants</b>				
Caffeine	1,3,7	none	+++	Rousseau et al. (1988), Fill and Copello (2002), Magkos and Kavouras (2005), Zalk et al. (2007)
Theophylline	1,3	none	++++	Rousseau et al. (1988)
Theobromine	3,7	none	++++	Rousseau et al. (1988)
1-Methylxanthine	1	none	++	Guerreiro et al. (2008)
3-Methylxanthine	3	none	0	Guerreiro et al. (2008)
7-Methylxanthine	7	none	0	Guerreiro et al. (2008)
<b>Produced essentially by bioconversion</b>				
Paraxanthine	1,7	none	++++	Hawke et al. (2000), Guerreiro et al. (2008), Rousseau et al. (1988)
<b>Produced by synthesis</b>				
3-Isobutyl-1-methylxanthine	1	3-Isobutyl	+++	Rousseau et al. (1988), Usachev and Verkhratsky (1995)
–	1	7-Isobutyl	+++	Rousseau et al. (1988), Islam et al. (1998), Wyskovsky (1994)
–	3,9	none	+++	Rousseau et al. (1988)
–	1,9	none	0	Rousseau et al. (1988)
–	1,3	7-(5 Ethylcarboxypentyl)	++++	Cavallaro et al. (1999)
–	1,3	7-(7-Hydroxyoctyl)	++++	Cavallaro et al. (1999)
–	3	7-(7-Oxoocetyl)-1-propargylxanthine	++++	Cavallaro et al. (1999)

Paraxanthine, the main metabolite of caffeine in man (Magkos and Kavouras 2005), is only produced in low amounts in plants. 1-Methylxanthine is also a secondary metabolite of paraxanthine and a metabolite of theophylline (Birkett et al. 1985; Magkos and Kavouras 2005). 3-Methylxanthine is a metabolite of caffeine, theophylline, and theobromine (Birkett et al. 1985; Magkos and Kavouras 2005). 7-Methylxanthine is the main metabolite of theobromine (Birkett et al. 1985)

<sup>a</sup>Limited to those references cited in the present article.

**Acknowledgements** The work by the authors mentioned in this paper received support from Institut de Recherche Pierre Fabre. We gratefully acknowledge support from Institut National de la Santé et de la Recherche Médicale (INSERM) and Université Pierre et Marie Curie-Paris 6.

## References

- Arnaud M (2010) Pharmacokinetics and metabolism of natural methylxanthines in animal and man. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Ascherio A, Zhang SM, Hernan MA, Kawachi I, Colditz GA, Speizer FE, Willett WC (2001) Prospective study of caffeine consumption and risk of Parkinson's disease in men and women. *Ann Neurol* 50:56–63
- Ashihara H, Kato M, Crozier A (2010) Distribution, biosynthesis and catabolism of methylxanthines in plants. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Birkett DJ, Dahlqvist R, Miners JO, Lelo A, Billing B (1985) Comparison of theophylline and theobromine metabolism in man. *Drug Metab Dispos* 13:725–728
- Block BM, Barry SR, Faulkner JA (1992) Aminophylline increases submaximum power but not intrinsic velocity of shortening of frog muscle. *J Appl Physiol* 73:71–74
- Bruton JD, Lemmens R, Shi CL, Persson-Sjögren S, Westerblad H, Ahmed M, Pyne NJ, Frame M, Furman BL, Islam MS (2003) Ryanodine receptors of pancreatic  $\beta$ -cells mediate a distinct context-dependent signal for insulin secretion. *FASEB J* 17:301–303
- Buck E, Zimanyi I, Abramson JJ, Pessah IN (1992) Ryanodine stabilizes multiple conformational states of the skeletal muscle  $\text{Ca}^{2+}$  release channel. *J Biol Chem* 267:23560–23567
- Cavallaro RA, Filocamo L, Galuppi A, Galione A, Brufani M, Genazzani AA (1999) Potentiation of cADPR-induced  $\text{Ca}^{2+}$ -release by methylxanthine analogues. *J Med Chem* 42:2527–2534
- Chen BT, Moran KA, Avshalumov MV, Rice ME (2006) Limited regulation of somatodendritic dopamine release by voltage-sensitive  $\text{Ca}^{2+}$  channels contrasted with strong regulation of axonal dopamine release. *J Neurochem* 96:645–655
- Connett RJ, Ugol LM, Hammack MJ, Hays ET (1983) Twitch potentiation and caffeine contractures in isolated rat soleus muscle. *Comp Biochem Physiol C* 74:349–354
- Conway D, Sakai T (1960) Caffeine contracture. *Proc Natl Acad Sci USA* 46:897–903
- Daly JW (2007) Caffeine analogs: biomedical impact. *Cell Mol Life Sci* 64:2153–2169
- Detbarn C, Palade P (1997)  $\text{Ca}^{2+}$  feedback on “quantal”  $\text{Ca}^{2+}$  release involving ryanodine receptors. *Mol Pharmacol* 52:1124–1130
- Douhou A, Troadec JD, Ruberg M, Raisman-Vozari R, Michel PP (2001) Survival promotion of mesencephalic dopaminergic neurons by depolarizing concentrations of  $\text{K}^{+}$  requires concurrent inactivation of NMDA or AMPA/kainate receptors. *J Neurochem* 78:163–174
- Du GG, MacLennan DH (1999)  $\text{Ca}^{2+}$  inactivation sites are located in the COOH-terminal quarter of recombinant rabbit skeletal muscle  $\text{Ca}^{2+}$  release channels (ryanodine receptors). *J Biol Chem* 274:26120–26126
- Du GG, Khanna VK, Guo X, MacLennan DH (2004) Central core disease mutations R4892W, I4897T and G4898E in the ryanodine receptor isoform 1 reduce the  $\text{Ca}^{2+}$  sensitivity and amplitude of  $\text{Ca}^{2+}$ -dependent  $\text{Ca}^{2+}$  release. *Biochem J* 382(Pt 2):557–564
- Elbaz A, Moisan F (2008) Update in the epidemiology of Parkinson's disease. *Curr Opin Neurol* 21:454–460
- Endo M (1977) Calcium release from the sarcoplasmic reticulum. *Physiol Rev* 57:71–108
- Fabiato A (1983) Calcium-induced release of calcium from the cardiac sarcoplasmic reticulum. *Am J Physiol Cell Physiol* 245:C1–C14
- Ferré S, Guix T, Sallés J, Badia A, Parra P, Jané F, Herrera-Marschitz M, Ungerstedt U, Casas M (1990) Paraxanthine displaces the binding of [ $^3\text{H}$ ]SCH 23390 from rat striatal membranes. *Eur J Pharmacol* 179:295–299



- Fessenden JD, Feng W, Pessah IN, Allen PD (2006) Amino acid residues Gln4020 and Lys4021 of the ryanodine receptor type 1 are required for activation by 4-chloro-m-cresol. *J Biol Chem* 281:21022–21031
- Fill M, Copello JA (2002) Ryanodine receptor calcium release channels. *Physiol Rev* 82: 893–922
- Francis SH, Sekhar KR, Ke H, Corbin JD (2010) Inhibition of cyclic nucleotide phosphodiesterases by methylxanthines and related compounds. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Galante M, Marty A (2003) Presynaptic ryanodine-sensitive calcium stores contribute to evoked neurotransmitter release at the basket cell-Purkinje cell synapse. *J Neurosci* 23:11229–11234
- Galione A, Lee HC, Busa WB (1991)  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release in sea urchin egg homogenates: modulation by cyclic ADP-ribose. *Science* 253:1143–1146
- Garaschuk O, Yaari Y, Konnerth A (1997) Release and sequestration of calcium by ryanodine-sensitive stores in rat hippocampal neurones. *J Physiol* 502:13–30
- Guerreiro S, Toulorge D, Hirsch E, Marien M, Sokoloff P, Michel PP (2008) Paraxanthine, the primary metabolite of caffeine, provides protection against dopaminergic cell death via stimulation of ryanodine receptor channels. *Mol Pharmacol* 74:980–989
- Hawke TJ, Allen DG, Lindinger MI (2000) Paraxanthine, a caffeine metabolite, dose dependently increases  $[\text{Ca}^{2+}]_i$  in skeletal muscle. *J Appl Physiol* 89:2312–2317
- Herrmann-Frank A, Lüttgau HC, Stephenson DG (1999) Caffeine and excitation-contraction coupling in skeletal muscle: a stimulating story. *J Muscle Res Cell Motil* 20:223–237
- Hymel L, Inui M, Fleischer S, Schindler H (1988) Purified ryanodine receptor of skeletal muscle sarcoplasmic reticulum forms  $\text{Ca}^{2+}$ -activated oligomeric  $\text{Ca}^{2+}$  channels in planar bilayers. *Proc Natl Acad Sci USA* 85:441–445
- Islam MS, Leibiger I, Leibiger B, Rossi D, Sorrentino V, Ekström TJ, Westerblad H, Andrade FH, Berggren PO (1998) In situ activation of the type 2 ryanodine receptor in pancreatic beta cells requires cAMP-dependent phosphorylation. *Proc Natl Acad Sci USA* 95:6145–6150
- Jacobson KA, Gao ZG (2006) Adenosine receptors as therapeutic targets. *Nat Rev Drug Discov* 5:247–264
- Kano M, Garaschuk O, Verkhratsky A, Konnerth A (1995) Ryanodine receptor-mediated intracellular calcium release in rat cerebellar Purkinje neurones. *J Physiol* 487:1–16
- Kelly E, Bailey CP, Henderson G (2008) Agonist-selective mechanisms of GPCR desensitization. *Br J Pharmacol* 153(Suppl 1):S379–S388
- Kirino Y, Shimizu H (1982)  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release from fragmented sarcoplasmic reticulum: a comparison with skinned muscle fiber studies. *J Biochem* 92:1287–1296
- Kong H, Jones PP, Koop A, Zhang L, Duff HJ, Chen SR (2008) Caffeine induces  $\text{Ca}^{2+}$  release by reducing the threshold for luminal  $\text{Ca}^{2+}$  activation of the ryanodine receptor. *Biochem J* 414:441–452
- Kramer ER, Aron L, Ramakers GM, Seitz S, Zhuang X, Beyer K, Smidt MP, Klein R (2007) Absence of Ret signaling in mice causes progressive and late degeneration of the nigrostriatal system. *PLoS Biol* 5:e39
- Langston JW, Forno LS, Tetrud J, Reeves AG, Kaplan JA, Karluk D (1999) Evidence of active nerve cell degeneration in the substantia nigra of humans years after 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine exposure. *Ann Neurol* 46:598–605
- Lee HC (1993) Potentiation of calcium- and caffeine-induced calcium release by cyclic ADP-ribose. *J Biol Chem* 268:293–299
- Liu W, Meissner G (1997) Structure-activity relationship of xanthines and skeletal muscle ryanodine receptor/ $\text{Ca}^{2+}$  release channel. *Pharmacology* 54:135–143
- Love S, Plaha P, Patel NK, Hotton GR, Brooks DJ, Gill SS (2005) Glial cell line-derived neurotrophic factor induces neuronal sprouting in human brain. *Nat Med* 11:703–704
- Magkos F, Kavouras SA (2005) Caffeine use in sports, pharmacokinetics in man, and cellular mechanisms of action. *Crit Rev Food Sci Nutr* 45:535–562

- McGarry SJ, Williams AJ (1994) Adenosine discriminates between the caffeine and adenine nucleotide sites on the sheep cardiac sarcoplasmic reticulum calcium-release channel. *J Membr Biol* 137:169–177
- McPherson PS, Kim YK, Valdivia H, Knudson CM, Takekura H, Franzini-Armstrong C, Coronado R, Campbell KP (1991) The brain ryanodine receptor: a caffeine-sensitive calcium release channel. *Neuron* 7:17–25
- Meissner G, Henderson JS (1987) Rapid calcium release from cardiac sarcoplasmic reticulum vesicles is dependent on  $\text{Ca}^{2+}$  and is modulated by  $\text{Mg}^{2+}$ , adenine nucleotide, and calmodulin. *J Biol Chem* 262:3065–3073
- Meissner G, Rousseau E, Lai FA, Liu QY, Anderson KA (1988) Biochemical characterization of the  $\text{Ca}^{2+}$  release channel of skeletal and cardiac sarcoplasmic reticulum. *Mol Cell Biochem* 82:59–65
- Michel PP, Alvarez-Fischer D, Guerreiro S, Hild A, Hartmann A, Hirsch EC (2007) Role of activity-dependent mechanisms in the control of dopaminergic neuron survival. *J Neurochem* 101:289–297
- Moutin MJ, Dupont Y (1988) Rapid filtration studies of  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release from skeletal sarcoplasmic reticulum. Role of monovalent ions. *J Biol Chem* 263:4228–4235
- Näbauer M, Callewaert G, Cleemann L, Morad M (1989) Regulation of calcium release is gated by calcium current, not gating charge, in cardiac myocytes. *Science* 246:1640
- Patel JC, Witkovsky P, Avshalumov MV, Rice ME (2009) Mobilization of calcium from intracellular stores facilitates somatodendritic dopamine release. *J Neurosci* 29:6568–6579
- Pessah IN, Stambuk RA, Casida JE (1987)  $\text{Ca}^{2+}$ -activated ryanodine binding: mechanisms of sensitivity and intensity modulation by  $\text{Mg}^{2+}$ , caffeine, and adenine nucleotides. *Mol Pharmacol* 31:232–238
- Ritter M, Menon S, Zhao L, Xu S, Shelby J, Barry WH (2001) Functional importance and caffeine sensitivity of ryanodine receptors in primary lymphocytes. *Int Immunopharmacol* 1:339–347
- Rousseau E, Ladine J, Liu QY, Meissner G (1988) Activation of the  $\text{Ca}^{2+}$  release channel of skeletal muscle sarcoplasmic reticulum by caffeine and related compounds. *Arch Biochem Biophys* 267:75–86
- Sääksjärvi K, Knekt P, Rissanen H, Laaksonen MA, Reunanen A, Männistö S (2008) Prospective study of coffee consumption and risk of Parkinson's disease. *Eur J Clin Nutr* 62:908–915
- Salthun-Lassalle B, Hirsch EC, Wolfart J, Ruberg M, Michel PP (2004) Rescue of mesencephalic dopaminergic neurons in culture by low-level stimulation of voltage-gated sodium channels. *J Neurosci* 24:5922–5930
- Satake W, Nakabayashi Y, Mizuta I, Hirota Y, Ito C, Kubo M, Kawaguchi T, Tsunoda T, Watanabe M, Takeda A, Tomiyama H, Nakashima K, Hasegawa K, Obata F, Yoshikawa T, Kawakami H, Sakoda S, Yamamoto M, Hattori N, Murata M, Nakamura Y, Toda T (2009) Genome-wide association study identifies common variants at four loci as genetic risk factors for Parkinson's disease. *Nat Genet* 41:1303–1307
- Sharma G, Vijayaraghavan S (2003) Modulation of presynaptic store calcium induces release of glutamate and postsynaptic firing. *Neuron* 38:929–939
- Sitsapesan R, Williams AJ (1990) Mechanisms of caffeine activation of single calcium-release channels of sheep cardiac sarcoplasmic reticulum. *J Physiol* 423:425–439
- Sitsapesan R, McGarry SJ, Williams AJ (1995) Cyclic ADP-ribose, the ryanodine receptor and  $\text{Ca}^{2+}$  release. *Trends Pharmacol Sci* 16:386–391
- Smith AB, Cunnane TC (1996) Ryanodine-sensitive calcium stores involved in neurotransmitter release from sympathetic nerve terminals of the guinea-pig. *J Physiol* 497(Pt 3):657–664
- Sutko JL, Airey JA, Welch W, Ruest L (1997) The pharmacology of ryanodine and related compounds. *Pharmacol Rev* 49:53–98
- Trueta C, Sánchez-Armass S, Morales MA, De-Miguel FF (2004) Calcium-induced calcium release contributes to somatic secretion of serotonin in leech Retzius neurons. *J Neurobiol* 61:309–316

- Usachev Y, Verkhratsky A (1995) IBMX induces calcium release from intracellular stores in rat sensory neurones. *Cell Calcium* 17:197–206
- Usachev Y, Shmigol A, Pronchuk N, Kostyuk P, Verkhratsky A (1993) Caffeine-induced calcium release from internal stores in cultured rat sensory neurons. *Neuroscience* 57:845–859
- Veley VH, Waller AD (1910) On the comparative toxicity of theobromine and caffeine, as measured by their direct effect upon the contractility of isolated muscle. *Proc R Soc Lond B Biol Sci* 82:568–574
- Verkhratsky A (2005) Physiology and pathophysiology of the calcium store in the endoplasmic reticulum of neurons. *Physiol Rev* 85:201–279
- Walz B, Baumann O, Zimmermann B, Ciriacy-Wantrup EV (1995) Caffeine- and ryanodine-sensitive  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release from the endoplasmic reticulum in honeybee photoreceptors. *J Gen Physiol* 105:537–567
- Wang R, Bolstad J, Kong H, Zhang L, Brown C, Chen SR (2004) The predicted TM10 transmembrane sequence of the cardiac  $\text{Ca}^{2+}$  release channel (ryanodine receptor) is crucial for channel activation and gating. *J Biol Chem* 279:3635–42
- Wang SJ (2007) Caffeine facilitation of glutamate release from rat cerebral cortex nerve terminals (synaptosomes) through activation protein kinase C pathway: an interaction with presynaptic adenosine  $\text{A}_1$  receptors. *Synapse* 61:401–411
- Wyskovsky W (1994) Caffeine-induced calcium oscillations in heavy-sarcoplasmic-reticulum vesicles from rabbit skeletal muscle. *Eur J Biochem* 221:317–325
- Xu L, Meissner G (1988) Regulation of cardiac muscle  $\text{Ca}^{2+}$  release channel by sarcoplasmic reticulum lumenal  $\text{Ca}^{2+}$ . *Biophys J* 75:2302–2312
- Xu K, Xu YH, Chen JF, Schwarzschild MA (2010) Neuroprotection by caffeine: time course and role of its metabolites in the MPTP model of Parkinson's disease. *Neuroscience* 167:475–478
- Xu K, Xu Y, Brown-Jermyn D, Chen JF, Ascherio A, Dluzen DE, Schwarzschild MA (2006) Estrogen prevents neuroprotection by caffeine in the mouse 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine model of Parkinson's disease. *J Neurosci* 26:535–541
- Zalk R, Lehnart SE, Marks AR (2007) Modulation of the ryanodine receptor and intracellular calcium. *Annu Rev Biochem* 76:367–385
- Zucchi R, Ronca-Testoni S (1997) The sarcoplasmic reticulum  $\text{Ca}^{2+}$  channel/ryanodine receptor: modulation by endogenous effectors, drugs and disease states. *Pharmacol Rev* 49:1–51

# Xanthines as Adenosine Receptor Antagonists

Christa E. Müller and Kenneth A. Jacobson

## Contents

1	Caffeine and Theophylline: Historical Aspects and Early Structural Modification	152
1.1	Naturally Occurring Xanthines	152
1.2	Early Modification of 8-Unsubstituted Xanthine Derivatives	153
1.3	Progression to Xanthines with Subtype Selectivity	156
2	A <sub>1</sub> Adenosine Receptor Antagonists	158
2.1	8-Aryl- and 8-Arylalkyl-Substituted Xanthines	158
2.2	8-Cycloalkylxanthines	163
2.3	Species Differences	169
2.4	Deazaxanthines and Azaxanthines	169
2.5	Tricyclic Xanthine Derivatives	170
3	A <sub>2A</sub> Adenosine Receptor Antagonists	172
3.1	8-Styrylxanthines	175
3.2	Configurationally Stable Analogues of 8-Styrylxanthines	176
3.3	A <sub>2A</sub> -Adenosine-Receptor-Selective Radiolabelled Xanthine Derivatives	177
3.4	Heterocyclic Compounds Related to Xanthines	177
4	A <sub>2B</sub> Adenosine Receptor Antagonists	177
4.1	Aryl-Substituted 1,3-Dialkylxanthines	177
4.2	1,8-Disubstituted Xanthines	178
4.3	9-Deazaxanthines	179
4.4	8-Furylmethyl-Substituted Xanthines	179
5	A <sub>3</sub> Adenosine Receptor Antagonists	180
5.1	8-Aryl-Substituted Xanthine Derivatives	180
5.2	Tricyclic Xanthine and Deazaxanthine Derivatives	180

---

C.E. Müller (✉)

PharmaCenter Bonn, Pharmaceutical Sciences Bonn (PSB), University of Bonn, Pharmaceutical Institute, An der Immenburg 4, 53121 Bonn, Germany  
e-mail: christa.mueller@uni-bonn.de

K.A. Jacobson

Molecular Recognition Section, Laboratory of Bioorganic Chemistry, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Building. 8A, Room B1A-19, 8 center Dr. 2089-0810 NIH, NIDDK, LBC, Bethesda, MD, USA

6	Xanthine Derivatives Used as Molecular Probes .....	181
6.1	Irreversible Ligand Probes .....	181
6.2	Spectroscopic Probes: Spin-Labelled and Fluorescent Probes .....	184
6.3	Specialized Radioligand Probes Based on Conjugation .....	185
6.4	Xanthine Radioligand Probes for Positron Emission Tomography .....	185
6.5	Conjugated Ligand Probes and Bivalent Ligands .....	186
7	Conclusions .....	187
	References .....	187

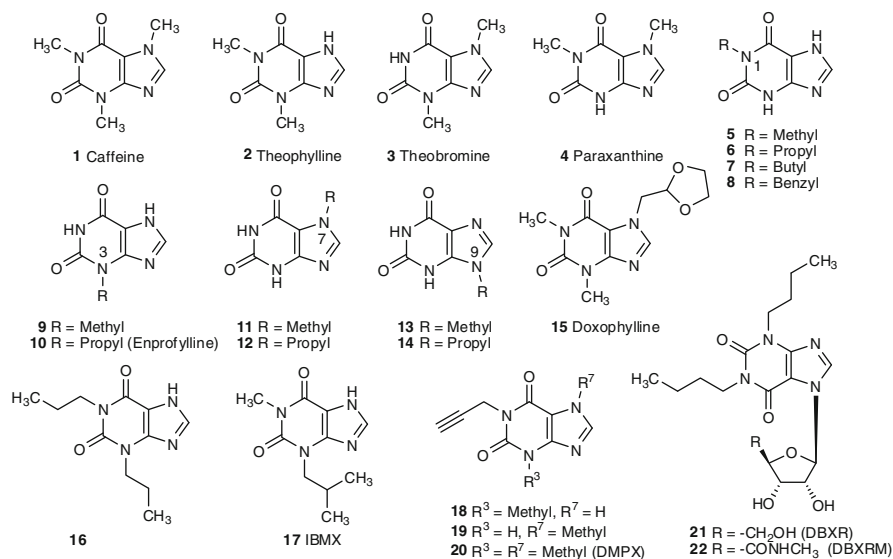
**Abstract** The natural plant alkaloids caffeine and theophylline were the first adenosine receptor (AR) antagonists described in the literature. They exhibit micromolar affinities and are non-selective. A large number of derivatives and analogues were subsequently synthesized and evaluated as AR antagonists. Very potent antagonists have thus been developed with selectivity for each of the four AR subtypes.

**Keywords** Adenosine receptors ·  $A_1$  receptor antagonists ·  $A_{2A}$  receptor antagonists ·  $A_{2B}$  receptor antagonists ·  $A_3$  receptor antagonists · Caffeine · Deazaxanthines · Molecular probes · Paraxanthine · Theobromine · Theophylline · Tricyclic xanthine derivatives · Xanthines

## 1 Caffeine and Theophylline: Historical Aspects and Early Structural Modification

### 1.1 Naturally Occurring Xanthines

The earliest adenosine receptor (AR) antagonists identified were the naturally occurring alkylxanthines, most notably among these being caffeine (1,3,7-trimethylxanthine, **1**) and theophylline (1,3-dimethylxanthine, **2**) (see Fig. 1) (Daly 1982; Fredholm et al. 1999; Stefanovich 1989). Another simple natural xanthine, theobromine (**3**), was shown to have only weak activity as an AR antagonist (Müller et al. 1993). The major caffeine metabolites in humans, paraxanthine (**4**) and 1-methylxanthine (**5**) (Krämer and Testa 2008), the latter also being the major metabolite of theophylline, are as potent as caffeine and theophylline and may contribute to their activity (Daly et al. 1986a; Müller et al. 1993). These simple alkylxanthines are of micromolar affinity, at best, at the ARs, and variation of affinity between species has been documented (see Table 1). This affinity range was later shown to apply generally to all human AR subtypes,  $A_1$ ,  $A_{2A}$ ,  $A_{2B}$ , and  $A_3$ , but only to three of the AR subtypes of the rat ( $A_1$ ,  $A_{2A}$ , and  $A_{2B}$ ). At the rat  $A_3$  AR, the



**Fig. 1** 8-Unsubstituted xanthine derivatives

simple alkylxanthines were shown to have much higher  $K_i$  values of approximately  $10^{-4}$  M or higher (Fredholm et al. 1994; van Galen et al. 1994).

## 1.2 Early Modification of 8-Unsubstituted Xanthine Derivatives

The first xanthine analogues with enhanced affinity at the ARs were modified from caffeine and theophylline (Daly 2000, 2007; Müller and Scior 1993; Fredholm and Jacobson 2009). In the early 1980s, one particular type of modification of the xanthine structure proved to be especially useful in enhancing affinity: the elongation of the 1,3-dimethyl groups to propyl or larger alkyl groups (Bruns 1981; Ukena et al. 1986b). For example, substitution of the 1,3-dimethyl groups with 1,3-dipropyl groups in **16** increased affinity at the rat A<sub>1</sub> AR by approximately 20-fold (Table 1). Substitutions at the 1-, 3-, or 7-positions, particularly small hydrophobic groups, were generally much better tolerated in AR binding than substitution at the 9-position (Daly 1982; Müller and Scior 1993). Evaluation of a series of monosubstituted xanthines (**5–14**) showed that substitution at N1 was most important for all AR subtypes (Müller et al. 1993). While a 1-propyl residue was best for A<sub>2B</sub> and A<sub>3</sub> ARs, a 1-benzyl residue was optimal for the A<sub>1</sub> and A<sub>2A</sub> AR subtypes (see Table 1). 1-Propylxanthine (**6**) shows the highest affinity of the small, simple xanthine derivatives for the human A<sub>2B</sub>AR ( $K_i$  360 nM), along with some selectivity (at least sevenfold vs. the other subtypes) (Kim

**Table 1** Adenosine receptor affinities of 8-unsubstituted xanthine derivatives

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
<b>Natural xanthine derivatives</b>				
<b>1</b> Caffeine (1,3,7-trimethylxanthine)	10,700 (h) <sup>d</sup>	23,400 (h) <sup>e</sup>	33,800 (h) <sup>j</sup>	13,300 (h) <sup>d</sup>
	44,900 (h) <sup>e</sup>	9,560 (h) <sup>d</sup>	10,400 (h) <sup>k</sup>	>100,000 (r) <sup>n</sup>
	41,000 (r) <sup>f</sup>	45,000 (r) <sup>g</sup>	20,500 (h) <sup>l</sup>	
	44,000 (r) <sup>g</sup>	32,500 (r) <sup>i</sup>	30,000 (r) <sup>m</sup>	
	47,000 (gp) <sup>h</sup>	48,000 (r) <sup>d</sup>	13,000 (m) <sup>m</sup>	
	44,000 (c) <sup>h</sup>			
<b>2</b> Theophylline (1,3-dimethylxanthine)	6,770 (h) <sup>o</sup>	1,710 (h) <sup>o</sup>	9,070 (h) <sup>k</sup>	22,300 (h) <sup>d</sup>
	14,000 (r) <sup>p</sup>	6,700 (h) <sup>d</sup>	74,000 (h) <sup>l</sup>	86,400 (h) <sup>o</sup>
	8,740 (r) <sup>d</sup>	22,000 (r) <sup>p</sup>	15,100 (r) <sup>k</sup>	>100,000 (r) <sup>n</sup>
	7,060 (gp) <sup>q</sup>	25,300 (r) <sup>d</sup>	5,630 (m) <sup>f</sup>	85,000 (r) <sup>t</sup>
	4,710 (rb) <sup>d</sup>		11,000 (gp) <sup>s</sup>	>100,000 (d) <sup>u</sup>
	9,050 (s) <sup>d</sup>		17,700 (rb) <sup>f</sup>	
	6,330 (c) <sup>q</sup>		38,700 (d) <sup>f</sup>	
<b>3</b> Theobromine (3,7-dimethylxanthine)	105,000 (r) <sup>p</sup>	>250,000 (r) <sup>p</sup>	130,000 (h) <sup>v</sup>	>100,000 (r) <sup>n</sup>
	83,400 (r) <sup>v</sup>	187,000 (r) <sup>v</sup>		
<b>4</b> Paraxanthine (1,7-dimethylxanthine)	21,000 (r) <sup>p</sup>	32,000 (r) <sup>p</sup>	4,500 (h) <sup>w</sup>	>100,000 (r) <sup>n</sup>
<b>Monosubstituted xanthine derivatives</b>				
<b>5</b> 1-Methylxanthine	36,000 (r) <sup>p</sup>	47,000 (r) <sup>p</sup>	6,600 (h) <sup>v</sup>	>100,000 (r) <sup>n</sup>
	11,400 (r) <sup>v</sup>	36,200 (r) <sup>v</sup>		
<b>6</b> 1-Propylxanthine	13,000 (r) <sup>p</sup>	33,000 (r) <sup>p</sup>	360 (h) <sup>k</sup>	2,370 (h) <sup>k</sup>
			1,880 (r) <sup>k</sup>	
<b>7</b> 1-Butylxanthine	9,000 (r) <sup>p</sup>	61,000 (r) <sup>p</sup>	421 (h) <sup>k</sup>	4,610 (h) <sup>k</sup>
<b>8</b> 1-Benzylxanthine	2,800 (r) <sup>p</sup>	22,000 (r) <sup>p</sup>	ND	ND
<b>9</b> 3-Methylxanthine	>100,000 (r) <sup>p</sup>	59,000 (r) <sup>p</sup>	87,000 (h) <sup>w</sup>	>100,000 (r) <sup>n</sup>
	35,000 (r) <sup>x</sup>			
<b>10</b> Enprofylline (3-propylxanthine)	156,000 (h) <sup>y</sup>	32,000 (h) <sup>y</sup>	7,000 (h) <sup>y</sup>	92,600 (h) <sup>d</sup>
	42,000 (h) <sup>d</sup>	81,300 (h) <sup>d</sup>	4,730 (h) <sup>k</sup>	65,000 (h) <sup>y</sup>
	32,000 (r) <sup>p</sup>	137,000 (r) <sup>p</sup>	19,800 (h) <sup>z</sup>	93,000 (r) <sup>d</sup>
	29,100 (r) <sup>v</sup>	103,000 (r) <sup>v</sup>	26,000 (r) <sup>aa</sup>	>100,000 (d) <sup>u</sup>
	>100,000 (d) <sup>u</sup>		5,630 (m) <sup>f</sup>	
		5,840 (rb) <sup>f</sup>		
		6,960 (d) <sup>f</sup>		
<b>11</b> 7-Methylxanthine	33,000 (r) <sup>p</sup>	59,000 (r) <sup>p</sup>	97,000 (h) <sup>w</sup>	>100,000 (r) <sup>n</sup>
<b>12</b> 7-Propylxanthine	18,000 (r) <sup>p</sup>	>200,000 (r) <sup>p</sup>	ND	ND
<b>13</b> 9-Methylxanthine	>250,000 (r) <sup>p</sup>	>250,000 (r) <sup>p</sup>	>1,000,000 (h) <sup>w</sup>	>100,000 (r) <sup>n</sup>
<b>14</b> 9-Propylxanthine	>250,000 (r) <sup>p</sup>	>250,000 (r) <sup>p</sup>	ND	ND
<b>Di- and trisubstituted xanthine derivatives</b>				
<b>15</b> Doxofylline	~100,000 <sup>ab</sup>	~100,000 <sup>ab</sup>	ND	ND
<b>16</b> 1,3-Dipropylxanthine	700 (r) <sup>p</sup>	6,600 (r) <sup>p</sup>	1,110 (h) <sup>k</sup>	1,940 (h) <sup>d</sup>
	450 (r) <sup>v</sup>	5,160 (r) <sup>v</sup>	680 (h) <sup>w</sup>	
	1,310 (gp) <sup>h</sup>			
	340 (c) <sup>h</sup>			
<b>17</b> 3-Isobutyl-1-methylxanthine	7,000 (r) <sup>p</sup>	16,000 (r) <sup>p</sup>	3,500 (h) <sup>v</sup>	ND
	2,460 (r) <sup>v</sup>	13,800 (r) <sup>v</sup>		
	8,600 (gp) <sup>h</sup>			
	4,400 (c) <sup>h</sup>			
<b>18</b> 3-Methyl-1-propargylxanthine	820 (r) <sup>p</sup>	4,800 (r) <sup>p</sup>	511 (h) <sup>k</sup>	10,900 (h) <sup>k</sup>
	5,830 (r) <sup>k</sup>	33,600 (r) <sup>k</sup>	2,150 (r) <sup>k</sup>	
<b>19</b> 7-Methyl-1-propargylxanthine	22,000 (r) <sup>p</sup>	16,000 (r) <sup>p</sup>	ND	ND
<b>20</b> 3,7-Dimethyl-1-propargylxanthine	45,000 (r) <sup>g</sup>	16,000 (r) <sup>g</sup>	4,130 (h) <sup>k</sup>	>10,000 (r) <sup>ac</sup>
	11,000 (r) <sup>h</sup>	5,600 (r) <sup>i</sup>		

(continued)

**Table 1** (continued)

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
	25,800 (gp) <sup>h</sup> 16,400 (c) <sup>h</sup>			
<b>21</b> DBXR	4,190 (r) <sup>n</sup>	19,500 (r) <sup>n</sup>	ND	6,030 (r) <sup>n,b</sup>
<b>22</b> DBXRM	37,300 (r) <sup>ad</sup>	>100,000 (r) <sup>27</sup>	ND	229 (r) <sup>ad,c</sup>

*DBXRM* *N*-methyl-1,3-dibutylxanthine-7-β-D-ribofuronamide

<sup>a</sup>*h* human, *c* cow, *d* dog, *gp* guinea pig, *m* mouse, *r* rat, *rb* rabbit, a few A<sub>2B</sub> data are from functional (cyclic AMP) studies; *ND* no data available

<sup>b</sup>Partial agonist

<sup>c</sup>Full agonist

<sup>d</sup>Jacobson et al. (1999)

<sup>e</sup>Abo-Salem et al. (2004)

<sup>f</sup>Grahner et al. (1994)

<sup>g</sup>Daly et al. (1991)

<sup>h</sup>Ukena et al. (1986b)

<sup>i</sup>Müller et al. (2000)

<sup>j</sup>Borrmann et al. (2009)

<sup>k</sup>Kim et al. (2002)

<sup>l</sup>Bertarelli et al. (2006)

<sup>m</sup>Brackett and Daly (1994)

<sup>n</sup>van Galen et al. (1994)

<sup>o</sup>Klotz et al. (1998)

<sup>p</sup>Müller et al. (1993b)

<sup>q</sup>Klotz et al. (1991)

<sup>r</sup>Auchampach et al. (2009)

<sup>s</sup>Fozard et al. (2003)

<sup>t</sup>Jacobson et al. (1995)

<sup>u</sup>Auchampach et al. (1997)

<sup>v</sup>Bruns et al. (1986)

<sup>w</sup>Bruns (1981)

<sup>x</sup>Shamim et al. (1989)

<sup>y</sup>Robeva et al. (1996)

<sup>z</sup>Ji et al. (2001)

<sup>aa</sup>Alexander et al. (1996)

<sup>ab</sup>Cirillo et al. (1988)

<sup>ac</sup>Müller and Ferré (2007)

<sup>ad</sup>Kim et al. (1994b)

et al. 2002), while 3-propylxanthine (enprofylline, **10**) is less potent ( $K_i$  human A<sub>2B</sub> AR 4,730 nM), but even more selective (at least 14-fold).

3,7-Dimethyl-1-propargylxanthine (DMPX, **20**) was the first A<sub>2</sub>-AR-selective antagonist described in the literature (Ukena et al. 1986b; Seale et al. 1988). It is similarly potent at A<sub>2A</sub> and A<sub>2B</sub> ARs, but the degree of selectivity versus A<sub>1</sub> ARs is low (Jacobson et al. 1992a). A comparison with the 7-unmethylated derivative 3-methyl-1-propargylxanthine (**18**) indicated that a 7-methyl group led to a large decrease in A<sub>1</sub> AR affinity and thus increased selectivity for A<sub>2A</sub> or A<sub>2B</sub> AR (Müller et al. 1993; Kim et al. 2002). The theophylline derivative doxofylline (**15**), bearing a 1,3-dioxolan-2-ylmethyl residue in the 7-position, is virtually inactive at A<sub>1</sub> and



A<sub>2A</sub>ARs and is believed to exert its antiasthmatic activity via inhibition of phosphodiesterases (Cirillo et al. 1988; Shukla et al. 2009).

The branched analogue 3-isobutyl-1-methylxanthine (IBMX, **17**) shows potency as a phosphodiesterase inhibitor in the same concentration range as is required to block ARs (Ukena et al. 1993).

1,3-Dibutylxanthine-7-ribosides (**21**, **22**) were found to bind effectively at A<sub>3</sub>ARs, indicating that the ribose group – as in adenosine – can act as a secondary anchor or recognition moiety in the receptor binding site (van Galen et al. 1994; Kim et al. 1994b; Park et al. 1998). This series of xanthine-7-ribosides also provided an early indication of modes of ligand binding at ARs, i.e. the overlay of receptor-bound positions of xanthine and adenine moieties. The 5'-uronamide modification of the CH<sub>2</sub>OH group of the ribose moiety greatly enhanced A<sub>3</sub>AR affinity as was shown for adenosine agonists, such that *N*-methyl-1,3-dibutylxanthine-7-β-D-ribofuronamide (DBXRM, **22**) was 143-fold selective (Kim et al. 1994b). DBXRM was found to be a full agonist at the A<sub>3</sub>AR, unlike other xanthine derivatives. It is proposed that the ribose moiety contains the essential structure and required flexibility to effect the conformational change of the receptor needed for activation (Gao et al. 2002).

### 1.3 Progression to Xanthines with Subtype Selectivity

In addition to substitution of the 1,3-dimethyl groups with larger 1,3-dialkyl groups, a means of increasing affinity at the rat A<sub>1</sub>AR was found to be the introduction of 8-aryl substituents (Fig. 2, Table 2) (Bruns 1981; Daly et al. 1986b; Jacobson et al. 1988, 1992a; Müller and Stein 1996). For example, placement of a phenyl group at the 8-position, generally increased A<sub>1</sub>AR affinity by at least an order of magnitude (Table 2). The first analogue having both 1,3-dialkyl and 8-phenyl modifications to be studied in detail was 1,3-diethyl-8-phenylxanthine (DPX, **25**), which displayed a *K<sub>i</sub>* value of 44 nM at the rat A<sub>1</sub>AR and was beginning to show selectivity for that subtype (Bruns et al. 1987a). [<sup>3</sup>H]DPX was demonstrated as the first AR antagonist radioligand, but its hydrophobicity limited its use (Bruns et al. 1980). Homologation to the 1,3-propyl groups in the 8-aryl analogue (**26**) provided a desired boost in affinity; however, the unintended consequence of rapidly diminishing aqueous solubility made this series unable to be used in typical pharmacology studies (Ukena et al. 1986b; Bruns and Fergus 1989). The 8-*p*-hydroxyphenyl-substituted derivative (**33**) (NPC-205) was slightly more potent than **26** (Shamim et al. 1988). The low aqueous solubility, which is both a function of the lipophilic groups present on the xanthine and the tendency of 8-arylxanthine derivatives to form highly stable crystal lattices, resulted in low bioavailability of **26** and similar compounds (Müller and Stein 1996; Frédéricck et al. 2005).

Several approaches were taken to increase the water solubility. A sulfonate group was introduced at the *para* position of the 8-phenyl ring, which greatly increased the solubility (Daly et al. 1985; Shamim et al. 1989). However, this

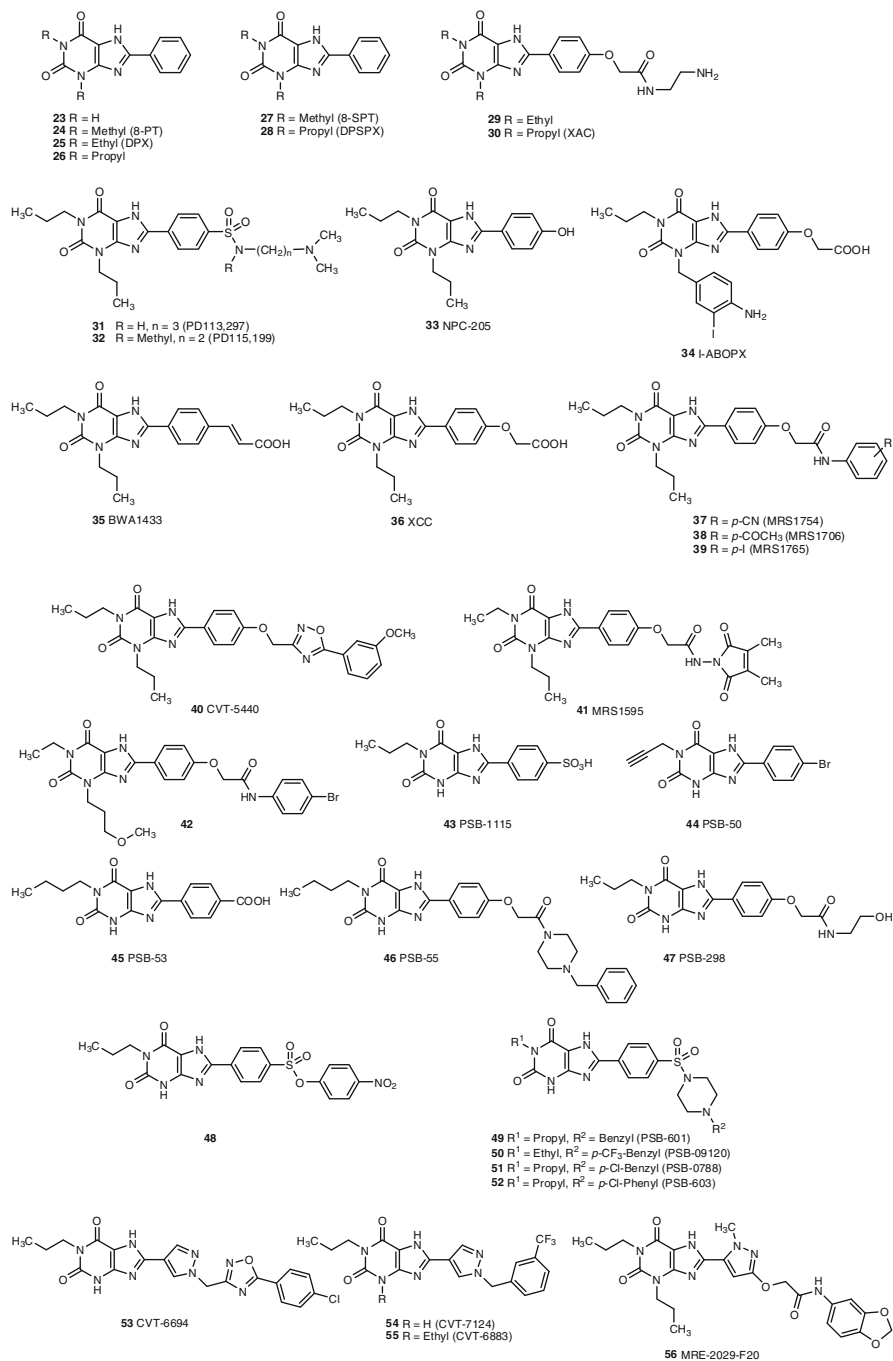
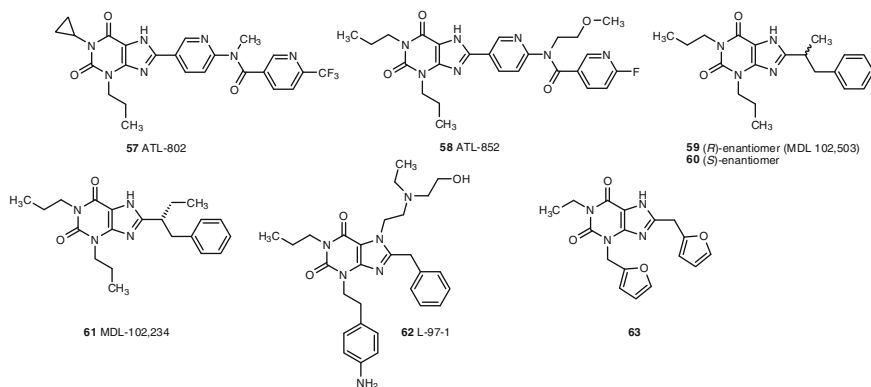


Fig. 2 (Continued)



**Fig. 2** 8-Phenyl- and 8-phenylalkyl-substituted xanthines and heteroaromatically substituted derivatives

modification tended to decrease both the affinity and the selectivity in comparison with the uncharged 8-phenyl analogues. Thus, SPT (**27**) and DPSPX (**28**), the latter of which is somewhat more potent, were both useful in pharmacological experiments where a blockade of all AR subtypes is required. It has to be kept in mind that these compounds do not block rat  $A_3$  ARs but are active in other species, such as human and sheep (Table 2). SPT (**27**) was shown not to penetrate into the brain owing to its high polarity (Baumgold et al. 1992).

## 2 $A_1$ Adenosine Receptor Antagonists

### 2.1 8-Aryl- and 8-Arylalkyl-Substituted Xanthines

An alternative approach to the introduction of charged groups for increasing water solubility resulted in the synthesis of the 8-aryl derivatives in which the charged group was separated from the phenyl ring by a spacer group. Various substitutions of an 8-phenyl ring indicated that an electron-donating group provided a favourable AR affinity (Jacobson et al. 1985b; Shamim et al. 1988). Thus, a methoxy substituent was replaced by a carboxymethoxy group, resulting in the carboxylic congener XCC (**36**) and the amine congener 8-[4-[[[(2-aminoethyl)amino]carbonyl]methyl]oxy]phenyl]-1,3-dipropylxanthine (XAC, **30**) (Fig. 2, Table 2) (Jacobson et al. 1985b, 1999). By placement of the charged group at a distance from the 8-aryl ring, it was possible to maintain and even enhance the high affinity seen with neutral, but poorly soluble analogues. Thus, XAC was found to have a  $K_i$  value of 1.2 nM at the rat  $A_1$  AR and approximately 50-fold selectivity in comparison with the rat  $A_2$  AR. The initial measure of  $A_2$  AR affinity used in Jacobson et al. (1985b)

**Table 2** Adenosine receptor affinities of 8-phenyl- and 8-phenylalkyl-substituted xanthines and heteroaromatically substituted derivatives

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
First-generation 8-phenylxanthine derivatives				
23 8-Phenylxanthine	2,500 (r) <sup>c</sup>	21,000 (r) <sup>c</sup>	810 (h) <sup>d</sup>	ND
24 8-Phenyltheophylline	1,340 (h) <sup>e</sup>	454 (h) <sup>e</sup>	415 (h) <sup>i</sup>	1,250 (h) <sup>e</sup>
	115 (h) <sup>f</sup>	850 (r) <sup>g</sup>	436 (m) <sup>j</sup>	>100,000 (r) <sup>k</sup>
	86 (r) <sup>g</sup>		249 (rb) <sup>j</sup>	
	76 (r) <sup>h</sup>		371 (d) <sup>j</sup>	
	1,540 (gp) <sup>h</sup>			
25 1,3-Diethyl-8-phenylxanthine	7.6 (c) <sup>h</sup>	860 (r) <sup>l</sup>	62.0 (h) <sup>i</sup>	ND
	44 (r) <sup>l</sup>	190 (h) <sup>m</sup>		
26 1,3-Dipropyl-8-phenylxanthine	10 (r) <sup>h</sup>	180 (r) <sup>n</sup>	18.9 (h) <sup>i</sup>	ND
	0.22 (c) <sup>h</sup>	2100 (h) <sup>o</sup>		
27 SPT	20.9 (gp) <sup>h</sup>			
	1,000 (h) <sup>e</sup>	7,050 (h) <sup>e</sup>	1,330 (h) <sup>i</sup>	5,890 (h) <sup>f</sup>
	4,500 (r) <sup>g</sup>	14,000 (r) <sup>g</sup>	1,590 (r) <sup>q</sup>	11,000 (h) <sup>s</sup>
	1,000 (r) <sup>h</sup>		4,990 (m) <sup>j</sup>	>>10,000 (r) <sup>r</sup>
	10,100 (gp) <sup>h</sup>		2,190 (gp) <sup>q</sup>	25,300 (d) <sup>p</sup>
28 DPSPX	6,460 (d) <sup>p</sup>		2,370 (rb) <sup>j</sup>	
	300 (c) <sup>h</sup>		7,240 (d) <sup>j</sup>	
			224 (d) <sup>q</sup>	
	210 (r) <sup>g</sup>	1,400 (r) <sup>g</sup>	568 (m) <sup>j</sup>	183 (s) <sup>l</sup>
	140 (r) <sup>k</sup>	790 (r) <sup>k</sup>	200 (rb) <sup>j</sup>	>100,000 (r) <sup>k</sup>
29			721 (d) <sup>j</sup>	22,500 (rb) <sup>u</sup>
				ND
30 XAC	12 (r) <sup>v</sup>	83 (r) <sup>v</sup>	7.8 (h) <sup>y</sup>	91.9 (h) <sup>x</sup>
	6.8 (h) <sup>w</sup>	18 (h) <sup>w</sup>	7.8 (h) <sup>y</sup>	26 (h) <sup>w</sup>
	29.1 (h) <sup>x</sup>	1.00 (h) <sup>x</sup>	16.0 (h) <sup>i</sup>	71 (h) <sup>p</sup>
	1.2 (r) <sup>y</sup>	63 (r) <sup>y</sup>	42.7 (r) <sup>q</sup>	29,000 (r) <sup>p</sup>
	0.49 (r) <sup>u</sup>		4.51 (m) <sup>j</sup>	106 (rb) <sup>p</sup>
	5.49 (gp) <sup>u</sup>		17.8 (gp) <sup>q</sup>	180 (s) <sup>t,z</sup>
	0.45 (rb) <sup>u</sup>		4.47 (rb) <sup>j</sup>	138 (d) <sup>p</sup>
	0.09 (s) <sup>u</sup>		29.8 (d) <sup>j</sup>	
	0.03 (c) <sup>u</sup>		3.55 (d) <sup>q</sup>	
159 (d) <sup>u</sup>				
31 PD113,297	5.59 (r) <sup>n</sup>	70.0 (r) <sup>n</sup>	ND	ND
32 PD115,199	14 (r) <sup>aa</sup>	16 (r) <sup>aa</sup>	160 (m) <sup>ac</sup>	ND
	4.05 (r) <sup>v</sup>	3.86 (rb) <sup>ab</sup>		
33 NPC-205	3.5 (r) <sup>ad</sup>	48 (h) <sup>ad</sup>	50 (gp) <sup>ae</sup>	ND
34 I-ABOPX (BW-A522)	70 (h) <sup>au</sup>	95 (h) <sup>au</sup>	30 (h) <sup>au</sup>	18 (h) <sup>z</sup>
	37 (r) <sup>af</sup>	700 (r) <sup>af</sup>		1,170 (r) <sup>ag</sup>
	601 (d) <sup>p</sup>			1,500 (r) <sup>p</sup>
35 BWA1433		nd	15.6 (h) <sup>au</sup>	179 (rb) <sup>p</sup>
	20 (r) <sup>af</sup>			37.5 (d) <sup>p</sup>
	132 (d) <sup>p</sup>			54 (h) <sup>z</sup>
				15,000 (r) <sup>p</sup>
			384 (rb) <sup>p</sup>	
			1,880 (d) <sup>p</sup>	

(continued)

**Table 2** (continued)

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
<b>36</b> XCC	175 (h) <sup>w</sup> 58 (r) <sup>y</sup>	2200 (h) <sup>w</sup> 595 (h) <sup>w</sup>	13.6 (h) <sup>y</sup> 40 (h) <sup>i</sup> 2,200 (r) <sup>y</sup>	3,910 (h) <sup>w</sup> 75,700 (r) <sup>w</sup>
A <sub>2B</sub> -selective 8-phenylxanthine derivatives and heteroaromatically substituted derivatives				
<b>37</b> MRS1754	403 (h) <sup>w</sup> 16.8 (r) <sup>w</sup>	503 (h) <sup>w</sup> 612 (r) <sup>w</sup>	1.97 (h) <sup>w</sup> 12.8 (r) <sup>w</sup> 16.6 (r) <sup>q</sup> 3.39 (m) <sup>j</sup> 9.12 (gp) <sup>q</sup> 1.79 (rb) <sup>j</sup> 12.8 (d) <sup>j</sup> 12.3 (d) <sup>q</sup>	570 (h) <sup>w</sup>
<b>38</b> MRS1706	157 (h) <sup>w</sup> 38 (r) <sup>w</sup>	112 (h) <sup>w</sup> 548 (r) <sup>w</sup>	1.4 (h) <sup>w</sup>	230 (h) <sup>w</sup>
<b>39</b> MRS1765	152 (h) <sup>w</sup> 15.7 (r) <sup>w</sup>	293 (h) <sup>w</sup> 1640 (r) <sup>w</sup>	2.13 (h) <sup>w</sup>	1270 (h) <sup>w</sup>
<b>40</b> CVT-5440	>10,000 (h) <sup>ah</sup>	>10,000 (h) <sup>ah</sup>	50 (h) <sup>ah</sup>	>10,000 (h) <sup>ah</sup>
<b>41</b> MRS1595	3,030 (h) <sup>w</sup> 11.1 (r) <sup>w</sup>	1,970 (h) <sup>w</sup> 126 (r) <sup>w</sup>	26.6 (h) <sup>w</sup>	670 (h) <sup>w</sup>
<b>42</b>	100 (r) <sup>ai</sup>	97.7 (h) <sup>ai</sup>	2.88 (h) <sup>ai</sup>	1,290 (h) <sup>ai</sup>
<b>43</b> PSB-1115	>10,000 (h) <sup>d</sup> 2,200 (r) <sup>c</sup>	24,000 (r) <sup>c</sup>	53.4 (h) <sup>d</sup>	>10,000 (h) <sup>d</sup>
<b>44</b> PSB-50	60 (r) <sup>d</sup>	199 (r) <sup>d</sup>	6.8 (h) <sup>d</sup>	477 (h) <sup>d</sup>
<b>45</b> PSB-53	1,181 (h) <sup>d</sup> 481 (r) <sup>d</sup>	~10,000 (h) <sup>d</sup> 3,800 (r) <sup>d</sup>	24 (h) <sup>d</sup>	4,622 (h) <sup>d</sup>
<b>46</b> PSB-55	122 (h) <sup>d</sup> 37 (r) <sup>d</sup>	~10,000 (r) <sup>d</sup> 550 (r) <sup>d</sup>	1.3 (h) <sup>d</sup>	475 (h) <sup>d</sup>
<b>47</b> PSB-298	68 (h) <sup>d</sup> 35 (r) <sup>d</sup>	2,139 (r) <sup>d</sup>	1.2 (h) <sup>d</sup> 60 (h) <sup>aj</sup>	422 (h) <sup>d</sup>
<b>48</b>	3.6 (r) <sup>ak</sup>	74 (r) <sup>ak</sup>	5.4 (h) <sup>ak</sup>	≥10,000 (h) <sup>ak</sup>
<b>49</b> PSB-601	2,067 (h) <sup>al</sup> 260 (r) <sup>al</sup>	484 (h) <sup>al</sup> 93.7 (r) <sup>al</sup>	3.6 (h) <sup>al</sup>	>1,000 (h) <sup>al</sup>
<b>50</b> PSB-09120	>10,000 (h) <sup>am</sup> >1,000 (r) <sup>am</sup>	22.7 (h) <sup>am</sup> 122 (r) <sup>am</sup>	0.157 (h) <sup>am</sup>	>10,000 (h) <sup>am</sup>
<b>51</b> PSB-0788	2,240 (h) <sup>am</sup> 386 (r) <sup>am</sup>	333 (h) <sup>am</sup> 1,730 (r) <sup>am</sup>	0.393 (h) <sup>am</sup>	>1,000 (h) <sup>am</sup>
<b>52</b> PSB-603	>10,000 (h) <sup>am</sup> >10,000 (r) <sup>am</sup>	>10,000 (h) <sup>am</sup> >10,000 (r) <sup>am</sup>	0.553 (h) <sup>am</sup> $K_D$ 0.403 (h) <sup>am</sup> $K_D$ 0.351 (m) <sup>am</sup>	>10,000 (h) <sup>am</sup>
<b>53</b> CVT-6694	>6,000 (h) <sup>an</sup>	>5,000 (h) <sup>an</sup>	7 (h) <sup>an</sup>	>9,000 (h) <sup>an</sup>
<b>54</b> CVT-7124	>6,000 (h) <sup>an</sup>	>5,000 (h) <sup>an</sup>	6 (h) <sup>an</sup>	>9,000 (h) <sup>an</sup>
<b>55</b> CVT-6883	1,940 (h) <sup>ao</sup>	3,280 (h) <sup>ao</sup>	22 (h) <sup>ao</sup>	1,070 (h) <sup>ao</sup>
<b>56</b> MRE-2029-F20	200 (h) <sup>ap</sup>	>1,000 (h) <sup>ap</sup>	5.5 (h) <sup>ap</sup>	>1,000 (h) <sup>ap</sup>
<b>57</b> ATL 802	369 (h) <sup>aq</sup> 9,583 (m) <sup>aq</sup>	654 (h) <sup>aq</sup> 8,393 (m) <sup>aq</sup>	2.36 (h) <sup>aq</sup> 8.58 (m) <sup>aq</sup>	>1,000 (h) <sup>aq</sup> >10,000 (m) <sup>aq</sup>
<b>58</b> ATL 852	ND	ND	28.5 (h) <sup>b</sup>	ND
8-Phenylalkyl-substituted xanthines				
<b>59</b> MDL 102,503	6.9 (r) <sup>ar</sup>	157 (r) <sup>ar</sup>	ND	ND
<b>60</b>	60.7 (r) <sup>ar</sup>	848 (r) <sup>ar</sup>	ND	ND

(continued)

**Table 2** (continued)

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
<b>61</b> MDL 102,234	23.2 (r) <sup>ar</sup>	3,510 (r) <sup>ar</sup>	ND	ND
<b>62</b> L-97-1	580 (h) <sup>as</sup>	>100,000 (h) <sup>as</sup>	>100,000 (h) <sup>as</sup>	ND
<b>63</b>	102 (r) <sup>at</sup>	83.2 (h) <sup>at</sup>	7.41 (h) <sup>at</sup>	10,000 (h) <sup>at</sup>

XAC 8-[4-[[[(2-aminoethyl)amino]carbonyl]methyl]oxy]phenyl]-1,3-dipropylxanthine

<sup>a</sup>*h* human, *c* cow, *d* dog, *gp* guinea pig, *m* mouse, *r* rat, *rb* rabbit, a few A<sub>2B</sub> data are from functional (cyclic AMP) studies, *ND* no data available

<sup>b</sup>Personal communication (J. Linden), see also Cagnina et al. (2009)

<sup>c</sup>Müller et al. (1993b)

<sup>d</sup>Hayallah et al. (2002)

<sup>e</sup>Kim et al. (1999)

<sup>f</sup>Ferkany et al. (1986)

<sup>g</sup>Daly (1991)

<sup>h</sup>Ukena et al. (1986b)

<sup>i</sup>Kim et al. (2002)

<sup>j</sup>Auchampach et al. (2009)

<sup>k</sup>van Galen et al. (1994)

<sup>l</sup>Bruns et al. (1987a)

<sup>m</sup>Ukena et al. (1986a)

<sup>n</sup>Bruns et al. (1986)

<sup>o</sup>Shamim et al. (1989)

<sup>p</sup>Auchampach et al. (1997)

<sup>q</sup>Fozard et al. (2003)

<sup>r</sup>Abo-Salem et al. (2004)

<sup>s</sup>Martin et al. (1996)

<sup>t</sup>Linden et al. (1993)

<sup>u</sup>Klotz et al. (1991)

<sup>v</sup>Jacobson et al. (1988)

<sup>w</sup>Kim et al. (2000)

<sup>x</sup>Klotz et al. (1998)

<sup>y</sup>Jacobson et al. (1999)

<sup>z</sup>Salvatore et al. (1993)

<sup>aa</sup>Bruns et al. (1987b)

<sup>ab</sup>Ji et al. (1991)

<sup>ac</sup>Brackett and Daly (1994)

<sup>ad</sup>Shamim et al. (1988)

<sup>ae</sup>Daly et al. (1986b)

<sup>af</sup>Linden (1994)

<sup>ag</sup>Kim et al. (1994a)

<sup>ah</sup>Zablocki et al. (2005)

<sup>ai</sup>Nieto et al. (2009)

<sup>aj</sup>Bertarelli et al. (2006)

<sup>ak</sup>Yan and Müller (2004)

<sup>al</sup>Yan et al. (2006)

<sup>am</sup>Borrmann et al. (2009)

<sup>an</sup>Kalla et al. (2008)

<sup>ao</sup>Elzein et al. (2008)

<sup>ap</sup>Baraldi et al. (2004)

<sup>aq</sup>Cagnina et al. (2009)

<sup>ar</sup>Peet et al. (1993)

<sup>as</sup>Obiefuna et al. (2005)

<sup>at</sup>Balo et al. (2009)

<sup>au</sup>Linden et al. (1999)

was the inhibition of cyclic AMP accumulation in rat brain slices, which corresponds more closely to the  $A_{2B}$  AR, rather than the  $A_{2A}$  AR. However, subsequent tests at the rat  $A_{2A}$  AR confirmed that there was still a margin of selectivity of XAC in binding to the  $A_1$  AR in rat (Ukena et al, 1986c). The substitution of the 1,3-dipropyl groups of XAC with 1,3-diethyl increased the affinity at the  $A_{2A}$  AR while decreasing it at the  $A_1$  AR (Jacobson et al. 1987a). The aqueous solubility of XAC was found to be 25  $\mu$ M, which was an improvement over the uncharged 8-aryl derivatives. Therefore, XAC was suitable for use in pharmacological experiments as a general AR antagonist and was the first such antagonist to display moderate  $A_1$  AR selectivity, at least in rat.

Given the promise of a relatively water soluble and somewhat selective xanthine derivative, this amine-functionalized derivative of 1,3-dipropyl-8-phenylxanthine was specifically radiolabelled on the 1,3-dipropyl groups by catalytic reduction of a 1,3-diallyl precursor. The resulting [ $^3$ H]XAC was useful as a radiotracer in binding experiments at rat cerebral cortical  $A_1$  ARs, with a  $K_D$  value of approximately 1 nM, and was thus the first generally useful antagonist radioligand for study of this receptor (Jacobson et al. 1986a). [ $^3$ H]XCC (**36**) was also introduced as a high-affinity radioligand for the  $A_1$  AR (Jarvis et al. 1987).

Another rationale for the design of XAC with a primary amino group was the “functionalized congener approach” to drug design (Jacobson et al. 1986b; Jacobson 2009). By this approach, a chemically functionalized chain is incorporated at an insensitive site on the xanthine pharmacophore and can be extended to enable a conjugation strategy. Such high-affinity conjugates are useful for AR characterization and can be coupled to radioactive or spectroscopic reporter groups without them losing their ability to bind to the receptor (Jacobson et al. 1987b). XAC was also used as an immobilized high-affinity ligand for the purpose of affinity chromatography leading to the isolation of both  $A_1$  and  $A_{2A}$  ARs and their purification to homogeneity (Olah et al. 1989; Weiss and Grisshammer 2002). While in XAC the polar, basic residue was connected to the 8-phenyl ring via ether and amide linkages, in another series, sulfonamide-linked derivatives were investigated (**31**, **32**, Fig. 2). Compound **31** and its congeners were potent  $A_1$  AR antagonists, but showed only a moderate degree of selectivity (Table 2) (Bruns et al. 1986, 1987a).

Besides 8-phenylxanthine derivatives, 8-benzyl-substituted xanthines (**62**) and 8-phenylethyl-substituted xanthines (**59–61**) have also been investigated and optimized with respect to  $A_1$  AR affinity (Peet et al. 1993). Among 8-(arylalkyl)-xanthine derivatives, 3-[2-(4-aminophenyl)ethyl]-8-benzyl-7-{2-ethyl(2-hydroxyethyl)amino}ethyl]-1-propyl-3,7-dihydropurine-2,6-dione (L-97-1, **62**) is a weaker binder than typical 8-aryl xanthine probes, but is a water-soluble  $A_1$  AR antagonist bearing a basic substituent at N7 that has been proposed for the treatment of asthma (Obiefuna et al. 2005). 1,3-Dipropyl-8-phenylethylxanthine derivatives with a methyl (**59**, **60**) or ethyl (**61**) substituent at the  $\alpha$ -carbon atom adjacent to the xanthine C8 position showed particularly high affinity and selectivity for the  $A_1$  AR, with a configurational preference for the *R* enantiomer over the *S* enantiomer (Peet et al. 1993).

## 2.2 8-Cycloalkylxanthines

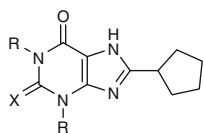
The introduction of 8-cycloalkyl groups instead of 8-aryl groups proved to be beneficial for affinity at the A<sub>1</sub> AR, and also allowed sufficient aqueous solubility for broad pharmacological application (Fig. 3, Table 3). The 8-cycloalkylxanthine derivative that is most widely used as a pharmacological tool is 1,3-dipropyl-8-cyclopentylxanthine (DPCPX) (**65**, also known as CPX), which is approximately 500-fold selective for the rat A<sub>1</sub> AR in comparison with the A<sub>2A</sub> AR (Bruns et al. 1987a). Among the human ARs, the A<sub>1</sub> AR selectivity is less than in the rat (Table 3). The corresponding cyclohexyl analogue has a similar pharmacological profile (Shamim et al. 1989). Curiously, DPCPX was in clinical trials for the treatment of cystic fibrosis through a non-AR related mechanism. It was found to act on the cystic-fibrosis-related chloride transporter to enhance the level of chloride in cell systems, an action that is unrelated to its AR antagonism (Cohen et al. 1997; Sorbera et al. 2000).

Bulkier cycloalkyl substituents in the xanthine 8-position, such as 3-noradamantyl [e.g. rolofylline (KW3902, **69**) and **77**], (substituted) norbornyl [naxifylline (BG-9719, CVT124, **80**), and the lactone **81**], dicyclopropylmethyl (MPDX, **83**, and KF15372, **84**), and bicyclo[2.2.2]octyl [toponafylline (BG-9928, **82**)] yielded very potent and selective A<sub>1</sub> AR antagonists (Fig. 3, Table 3).

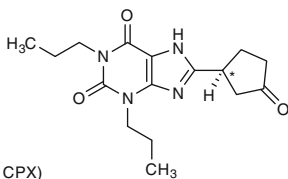
The 2-thio analogue of DPCPX (**65**), 2-thio-DPCPX (**66**), was as potent and selective as **65**, showing that in the 2-position a hydrogen-bond acceptor (such as a keto group) was not required (Jacobson et al. 1989b). Replacement of the 3-propyl residue in DPCPX by a phenyl (**73**), a benzyl (**72**), or a chiral methylbenzyl residue (**70**, **71**) was well tolerated (Weyler et al. 2006). However, the affinity for the human A<sub>3</sub> AR was increased by the lipophilic, aromatic residues, and the compounds lost A<sub>1</sub> AR selectivity in comparison with the A<sub>3</sub> AR affinity. The introduction of polar hydroxy groups at the 3-position was well tolerated by the A<sub>1</sub> AR, but not by the A<sub>3</sub> AR, leading to very potent and highly selective A<sub>1</sub> AR antagonists (**74**, **75**, **77–79**) (Weyler et al. 2006; Massip et al. 2006). In fact, 1-butyl-3-(3-hydroxypropyl)8-(3-noradamantyl)xanthine (PSB-36, **77**) is one of the most potent A<sub>1</sub> AR antagonists described to date, showing K<sub>i</sub> values of 0.124 nM (rat) and 0.7 nM (human). The hydroxylated DPCPX derivative **74** (PSB-16) was converted to its phosphoric acid ester disodium salt, yielding a highly water soluble A<sub>1</sub> AR antagonist prodrug suitable for parenteral application without the need for detergents and organic solvents (Weyler et al. 2006).

Several other polar derivatives and analogues of DPCPX were developed in order to further improve water solubility and bioavailability. Apaxifylline (**67**), with a keto group at C3 of the cyclopentyl ring, was clinically evaluated as a memory-enhancing drug for the treatment of dementia (Schingnitz et al. 1991). An amino-substituted DPCPX derivative, midaxifylline (**68**), has also been investigated (Ceccarelli et al. 1995). A promising second-generation compound currently undergoing clinical trials for the treatment of chronic heart failure is toponafylline (**82**), which contains a carboxylate function (Doggrell, 2005). Roloxylline (**69**), an A<sub>1</sub> AR antagonist of the first generation, had shown promising results in a pilot phase III study in patients with acute heart failure, but in a recently published larger

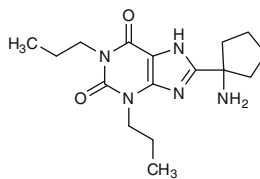




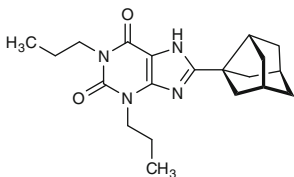
**64** R = Methyl, X = O (CPT)  
**65** R = Propyl, X = O (DPCPX, CPX)  
**66** R = Propyl, X = S



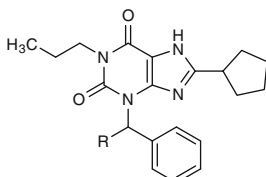
**67** Apaxifylline  
 ((*S*)-enantiomer, BIP20)  
 (Racemate: KFM19)



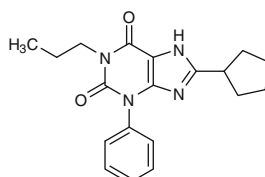
**68** Midaxifylline  
 (IRF1117)



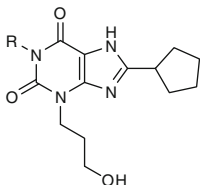
**69** Rolofylline (KW3902, NAX)



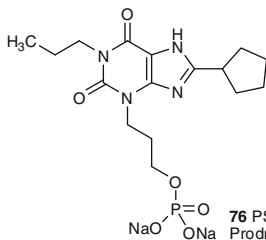
**70** R = Methyl, (*S*)-enantiomer  
**71** R = Methyl, (*R*)-enantiomer  
**72** R = H



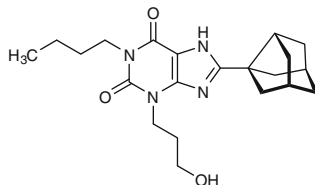
**73**



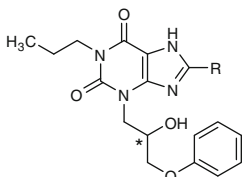
**74** R = Propyl (PSB-16)  
**75** R = Butyl



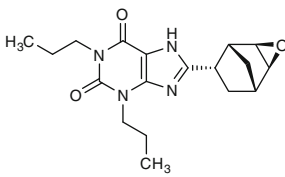
**76** PSB-16P  
 Prodrug of **74**



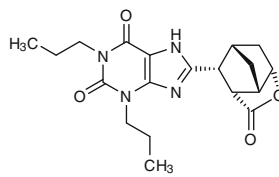
**77** PSB-36



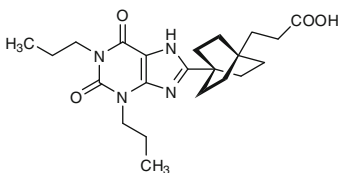
**78** R = Cyclopentyl  
**79** R = 3-Noradamantyl



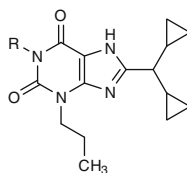
**80** Naxifylline  
 (BG-9719, CVT-124)



**81**



**82** Toponafylline (BG-9928)



**83** R = Methyl: MPDX  
**84** = Propyl: KF15372

**Fig. 3** 8-Cycloalkylxanthines

**Table 3** Adenosine receptor affinities of 8-cycloalkylxanthines

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
<b>64</b> 8-Cyclopentyltheophylline	24 (r) <sup>c</sup>	1,400 (r) <sup>c</sup>	710 (h) <sup>d</sup>	~100,000 (h) <sup>c</sup>
	6.3 (r) <sup>e</sup>	3,170 (r) <sup>ac</sup>	902 (h) <sup>ac</sup>	>10,000 (r) <sup>ac</sup>
	26.1 (gp) <sup>e</sup>			
	6.4 (rb) <sup>e</sup>			
	2.9 (s) <sup>e</sup>			
	1.4 (c) <sup>e</sup>			
<b>65</b> DPCPX	3.0 (h) <sup>f</sup>	129 (h) <sup>j</sup>	51 (h) <sup>f</sup>	795 (h) <sup>o</sup>
	0.50 (r) <sup>f</sup>	60 (h) <sup>k</sup>	63.8 (h) <sup>g</sup>	243 (h) <sup>f</sup>
	1.0 (r) <sup>g</sup>	157 (r) <sup>l</sup>	186 (r) <sup>g</sup>	509 (h) <sup>i</sup>
	0.18 (r) <sup>c</sup>	500 (r) <sup>g</sup>	200 (r) <sup>m</sup>	3,960 (h) <sup>j</sup>
	1.06 (gp) <sup>e</sup>		86.2 (m) <sup>n</sup>	>10,000 (r) <sup>k</sup>
	3.9 (gp) <sup>h</sup>		145 (gp) <sup>m</sup>	43,000 (r) <sup>i</sup>
	0.21 (rb) <sup>e</sup>		96.0 (rb) <sup>n</sup>	708 (rb) <sup>i</sup>
	0.10 (s) <sup>e</sup>		147 (d) <sup>n</sup>	115 (d) <sup>i</sup>
	0.05 (c) <sup>e</sup>		132 (d) <sup>m</sup>	
	0.29 (c) <sup>h</sup>			
	11.4 (d) <sup>i</sup>			
<b>66</b> 2-Thio-DPCPX	0.655 (r) <sup>p</sup>	314 (r) <sup>p</sup>	2800 (h) <sup>q</sup>	nd
<b>67</b> Apaxifylline [S(-)-configured enantiomer] [KFM19 (racemate), BIIP- 20 (S(-))]	10.5 (mk) <sup>r,b</sup>	1,512 (mk) <sup>r,b</sup>	nd	nd
	3 (r) <sup>s</sup>	2,640 (r) <sup>s</sup>		
<b>68</b> Midaxifylline [8-(1-aminocyclopentyl)-1,3- dipropylxanthine, IRFI117]	26 <sup>t</sup>	54,600 <sup>t</sup>	nd	nd
<b>69</b> Rolofylline [KW3902, NAX, 1,3-dipropyl-8-(3- noradamantyl)xanthine]	0.72 (h) <sup>u</sup>	108 (h) <sup>u</sup>	296 (h) <sup>v</sup>	4,390 (h) <sup>v</sup>
	8.0 (h) <sup>v</sup>	673 (h) <sup>v</sup>		
	0.19 (r) <sup>v</sup>	380 (r) <sup>w</sup>		
	12.6 (r) <sup>u</sup>	510 (r) <sup>u</sup>		
<b>70</b> 1-Propyl-3-(S)-1- methylbenzyl-8- cyclopentylxanthine	10.1 (r) <sup>k</sup>	3,500 (r) <sup>k</sup>	8,000 (h) <sup>k</sup>	85 (h) <sup>k</sup> >10,000 (r) <sup>k</sup>
<b>71</b> 1-Propyl-3-(R)-1- methylbenzyl-8- cyclopentylxanthine	23.8 (r) <sup>k</sup>	2,400 (r) <sup>k</sup>	2,960 (h) <sup>k</sup>	370 (h) <sup>k</sup>
<b>72</b> 1-Propyl-3-benzyl-8- cyclopentylxanthine	24.3 (h) <sup>k</sup>	511 (r) <sup>k</sup>	8,000 (h) <sup>k</sup>	54.6 (h) <sup>k</sup>
	8.70 (r) <sup>k</sup>			
<b>73</b> 1-Propyl-3-phenyl-8- cyclopentylxanthine	7.1 (h) <sup>k</sup>	1,200 (h) <sup>k</sup>	625 (h) <sup>k</sup>	395 (h) <sup>k</sup>
	1.01 (r) <sup>k</sup>	492 (r) <sup>k</sup>		
<b>74</b> 1-Propyl-3-(3-hydroxypropyl)- 8-cyclopentylxanthine (PSB-16)	5.74 (h) <sup>k</sup>	664 (r) <sup>k</sup>	194 (h) <sup>k</sup>	3,100 (h) <sup>k</sup>
	0.57 (r) <sup>k</sup>			
<b>75</b> 1-Butyl-3-(3-hydroxypropyl)- 8-cyclopentylxanthine	0.45 (r) <sup>k</sup>	582 (r) <sup>k</sup>	ND	1,190 (h) <sup>k</sup>
<b>77</b> 1-Butyl-3-(3-hydroxypropyl)- 8-(3-noradamantyl) xanthine (PSB-36)	0.7 (h) <sup>k</sup>	980 (h) <sup>k</sup>	187 (h) <sup>k</sup>	2,300 (h) <sup>k</sup>
	0.124 (r) <sup>k</sup>	552 (r) <sup>k</sup>		6,500 (r) <sup>k</sup>
<b>78</b>	49 (h) <sup>x</sup>	>10,000 (h) <sup>x</sup>	ND	3,550 (h) <sup>x</sup>
	55 (r) <sup>x</sup>	>10,000 (r) <sup>x</sup>		

(continued)

**Table 3** (continued)

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
<b>79</b>	29 (h) <sup>x</sup> 21 (r) <sup>x</sup>	>10,000 (h) <sup>x</sup> >10,000 (r) <sup>x</sup>	ND	>10,000 (h) <sup>x</sup>
<b>80</b> Naxifylline (BG-9719, CVT-124)	0.45 (h) <sup>u</sup> 12 (h) <sup>w</sup> 0.67 (r) <sup>u</sup>	1,100 (h) <sup>u</sup> 1,660 (h) <sup>w</sup> 1,250 (r) <sup>u</sup>	611 (h) <sup>w</sup> 1,010 (m) <sup>n</sup> 470 (rb) <sup>n</sup> 742 (d) <sup>n</sup>	4,810 (h) <sup>w</sup>
<b>81</b>	18 (h) <sup>w</sup> 3.0 (r) <sup>w</sup>	657 (h) <sup>w</sup> 264 (r) <sup>w</sup>	802 (h) <sup>w</sup>	>1,000 (h) <sup>w</sup>
<b>82</b> Toponafylline (BG-9928)	7.4 (h) <sup>y</sup> 3.9 (mk) <sup>y</sup> 1.3 (r) <sup>v</sup> 29 (d) <sup>y</sup>	6,410 (h) <sup>y</sup> 943 (mk) <sup>y</sup> 2,440 (r) <sup>v</sup> 4307 (d) <sup>y</sup>	90 (h) <sup>y</sup>	>10,000 (h) <sup>y</sup>
<b>83</b> MPDX (1-methyl analogue of KF 15372)	4.2 (r) <sup>z</sup>	>100 (r) <sup>z</sup>	ND	ND
<b>84</b> KF 15372	0.99 (r) <sup>aa</sup> 3.0 (r) <sup>ab</sup> 3.0 (gp) <sup>aa</sup>	430 (r) <sup>aa</sup>	ND	ND

*DPCPX* 1,3-dipropyl-8-cyclopentylxanthine

<sup>a</sup>*h* human, *c* cow, *d* dog, *gp* guinea pig, *m* mouse, *mk* monkey, *r* rat, *rb* rabbit, a few A<sub>2B</sub> data are from functional (cyclic AMP) studies, *ND* no data available

<sup>b</sup>Data for the racemate (KFM-19)

<sup>c</sup>van Galen et al. (1994)

<sup>d</sup>Bruns et al. (1986)

<sup>e</sup>Klotz et al. (1991)

<sup>f</sup>Bulicz et al. (2006)

<sup>g</sup>Kim et al. (2002)

<sup>h</sup>Ukena et al. (1986b)

<sup>i</sup>Auchampach et al. (1997)

<sup>j</sup>Klotz et al. (1998)

<sup>k</sup>Weyler et al. (2006)

<sup>l</sup>Müller et al. (2000)

<sup>m</sup>Fozard et al. (2003)

<sup>n</sup>Auchampach et al. (2009)

<sup>o</sup>Hayallah et al. (2002)

<sup>p</sup>Jacobson et al. (1989b)

<sup>q</sup>Jacobson et al. (1999)

<sup>r</sup>Schingnitz et al. (1991)

<sup>s</sup>Müller (1997)

<sup>t</sup>Ceccarelli et al. (1995)

<sup>u</sup>Pfister et al. (1997)

<sup>v</sup>Kiesman et al. (2006b)

<sup>w</sup>Kiesman et al. (2006a)

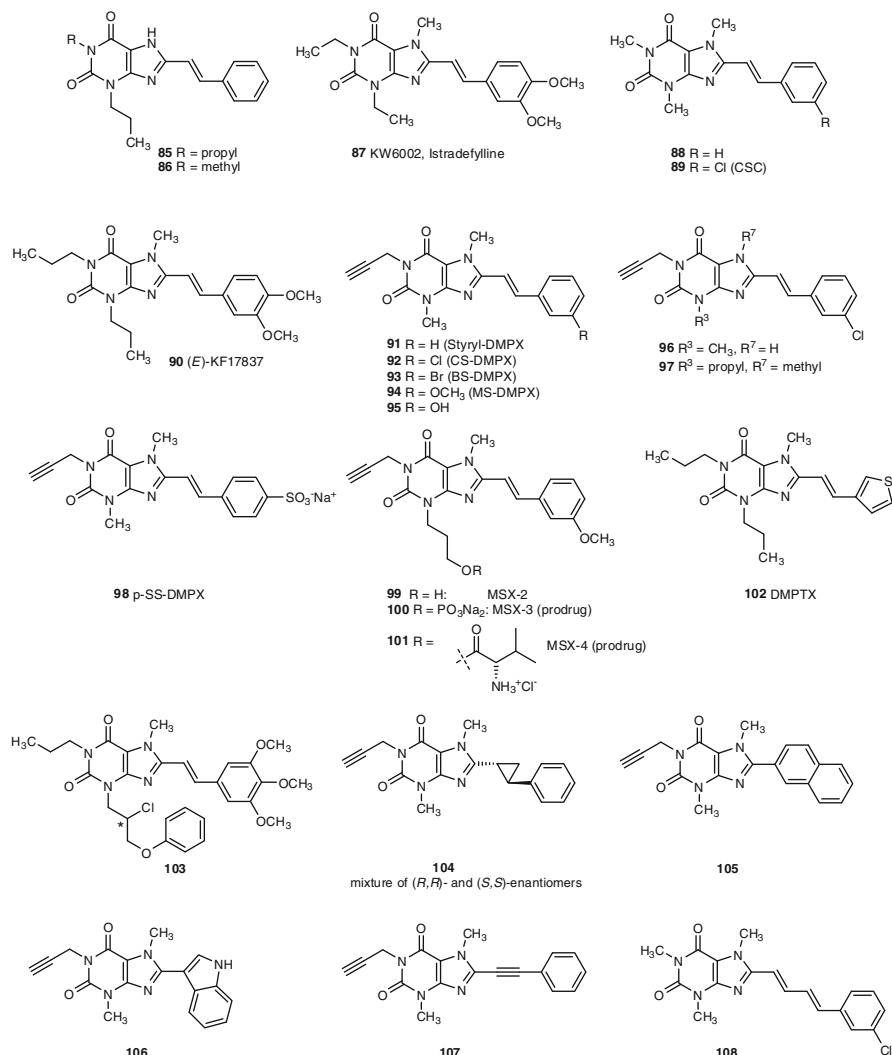
<sup>x</sup>Massip et al. (2006)

<sup>y</sup>Doggrell (2005)

<sup>z</sup>Noguchi et al. (1997)

<sup>aa</sup>Suzuki et al. (1992a)

<sup>ab</sup>Shimada et al. (1992)



**Fig. 4** 8-Styrylxanthines and configurationally stable analogues

phase III study (PROTECT) it did not exhibit significant improvement over a placebo (Slawsky and Givertz 2009). Further potential applications for A<sub>1</sub> AR antagonists include hypertension and renal diseases owing to their diuretic and kidney-protective effects. The feasibility of designing kidney-selective prodrugs of an A<sub>1</sub> AR antagonist has been demonstrated (Barone et al. 1989). Yet other applications are cardiac arrhythmia, asthma and other respiratory disorders, and the prevention of organ damage, e.g. resulting from transplantation (Jacobson et al. 1992a; Müller and Stein 1996; Müller 1997; Jacobson and Gao 2006; Moro et al. 2006; Givertz 2009).

**Table 4** Adenosine receptor affinities of 8-strylylxanthines and configurationally stable analogues

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
Styrylxanthines <sup>b</sup>				
<b>85</b> 1,3-Dipropyl-8-strylylxanthine	22.2 (r) <sup>f</sup>	85.1 (r) <sup>f</sup>	ND	ND
<b>86</b> 1-Methyl-3-propyl-8-strylylxanthine	31.1 (r) <sup>f</sup>	46.5 (r) <sup>f</sup>	ND	ND
<b>87</b> Istradefylline (KW-6002) ( $K_i$ MAO-B 28,000 nM) <sup>g</sup>	841 (h) <sup>c</sup> 230 (r) <sup>c</sup>	12 (h) <sup>h</sup> 91.2 (h) <sup>c</sup> 2.2 (r) <sup>i</sup> 4.46 (r) <sup>j</sup>	>10,000 (h) <sup>c</sup>	4,470 (h) <sup>c</sup>
<b>88</b> 8-Styrylcaffeine ( $K_i$ MAO-B 2,864 nM) <sup>k</sup>	3,890 (r) <sup>l</sup>	94 (r) <sup>l</sup>	ND	ND
<b>89</b> <i>m</i> -Chlorostyrylcaffeine ( $K_i$ MAO-B 80.6 nM) <sup>j</sup>	28,000 (r) <sup>l</sup>	54 (r) <sup>l</sup>	8,200 <sup>m</sup>	>10,000 (r) <sup>n</sup>
<b>90</b> KF17837	390 (r) <sup>o</sup>	7.9 (r) <sup>o</sup> ( <i>E/Z</i> ) 1.0 (r) <sup>o</sup> ( <i>E</i> )	1,500 (h) <sup>o</sup>	ND
<b>91</b> Styryl-DMPX	1,100 (r) <sup>p</sup>	27 (r) <sup>p</sup>	ND	ND
<b>92</b> <i>m</i> -Chlorostyryl-DMPX	1,300 (r) <sup>p</sup>	13 (r) <sup>p</sup>	ND	ND
<b>93</b> <i>m</i> -Bromostyryl-DMPX	1,200 (r) <sup>p</sup>	8.2 (r) <sup>p</sup> 10 (r) <sup>q</sup>	>10,000 (h) <sup>f</sup>	>10,000 (h) <sup>f</sup>
<b>94</b> <i>m</i> -Methoxystyryl-DMPX	1,280 (r) <sup>f</sup>	12 (r) <sup>f</sup>	ND	ND
<b>95</b> <i>m</i> -Hydroxystyryl-DMPX	940 (r) <sup>f</sup>	21 (r) <sup>f</sup>	ND	ND
<b>96</b> 7-Unsubstituted analogue of <i>m</i> -Chlorostyryl-DMPX	250 (r) <sup>p</sup>	410 (r) <sup>p</sup>	ND	ND
<b>97</b> 3-Propyl analogue of <i>m</i> -Chlorostyryl-DMPX	102 (r) <sup>p</sup>	5.1 (r) <sup>p</sup>	ND	ND
<b>98</b> <i>p</i> -Sulfostyryl-DMPX	4,900 (r) <sup>q</sup>	240 (r) <sup>q</sup>	ND	ND
<b>99</b> MSX-2	900 (r) <sup>s</sup> 2,500 (h) <sup>s</sup>	8.04 (r) <sup>r,s</sup> 5.38 (h) <sup>s,d</sup> 14.5 (h) <sup>s,e</sup>	>10,000 (h) <sup>s</sup> 2,900 (h) <sup>t</sup>	>10,000 (h) <sup>s</sup>
<b>102</b> DMPTX	561 (r) <sup>u</sup>	19 (r) <sup>u</sup>	ND	ND
<b>103</b>	44 (r) <sup>v</sup>	>10,000 (r) <sup>v</sup>	ND	ND
Analogues of styrylxanthines				
<b>104</b> Phenylcyclopropyl-DMPX ( <i>trans</i> , <i>rac</i> )	4,600 (r) <sup>w</sup>	1,700 (r) <sup>w</sup>	ND	ND
<b>105</b> β-Naphthyl-DMPX	980 (r) <sup>w</sup>	380 (r) <sup>w</sup>	ND	ND
<b>106</b> 3-Indolyl-DMPX	1,000 (r) <sup>w</sup>	300 (r) <sup>w</sup>	ND	ND
<b>107</b> Phenylethynyl-DMPX	>3,000 (r) <sup>w</sup>	314 (h) <sup>c</sup> 300 (r) <sup>w</sup>	ND	5,000 (h) <sup>c</sup>
<b>108</b> Phenylbutadienylxanthine ( $K_i$ MAO-B 42.1 nM) <sup>j</sup>	ND	104 (r) <sup>w</sup>	ND	ND

MSX-2 3-(3-hydroxypropyl)-7-methyl-8-(*m*-methoxystyryl)-1-propargylxanthine, DMPX 3,7-dimethyl-1-propargylxanthine, DMPTX [8-(3-thienylethenyl)-1,3-dipropylxanthine], MAO-B monoaminoxidase type B

<sup>a</sup>*h* human, *c* cow, *d* dog, *gp* guinea pig, *mmouse*, *mk* monkey, *r* rat, *rb* rabbit, a few A<sub>2B</sub> data are from functional (cyclic AMP) studies, ND no data available

<sup>b</sup>Most data probably represent data from mixture of *E/Z* isomers since in dilute solutions light-induced isomerization occurs very fast and is difficult to avoid under standard testing conditions

<sup>c</sup>Müller, C.E., Hockemeyer, J., Diekmann, M. unpublished data

<sup>d</sup>Recombinant receptors expressed in Chinese hamster ovary cells

<sup>e</sup>Native receptors (postmortem human brain cortex)

### 2.3 *Species Differences*

Among the human ARs, the A<sub>1</sub> AR affinity and selectivity (vs. A<sub>2A</sub> and A<sub>2B</sub> ARs) of 8-cycloalkyl and 8-aryl derivatives are typically less than in the rat (see Tables 2, 3). Several groups have studied the species dependence of the AR affinity of xanthine derivatives (Ukena et al. 1986b; Klotz et al. 1991; Müller et al. 1993; Müller 1997; Kull et al. 1999; Fozard et al. 2003; Auchampach et al. 2009). An early conclusion was that the affinity of typical 8-substituted analogues (both aryl and cycloalkyl) was greatest at the bovine A<sub>1</sub> AR, intermediate at the rat A<sub>1</sub> AR, and lowest at the porcine A<sub>1</sub> AR. Later, it was found that the human A<sub>1</sub> AR most closely resembled the porcine A<sub>1</sub> AR, in that respect. At the A<sub>2A</sub> AR and the A<sub>2B</sub> AR, the opposite is true, although the differences are moderate: 8-substituted xanthines, such as XAC and DPCPX, are more potent at the human receptor than at the rat orthologue. The largest species differences are observed for the A<sub>3</sub> AR: 8-phenylxanthines and 8-cyclopentylxanthines are typically much more potent at the human than at the rat A<sub>3</sub> AR (Linden 1994; Ji et al. 1994; Jacobson 1998; Müller 2001, 2003).

### 2.4 *Deazaxanthines and Azaxanthines*

Analogues of xanthine derivatives, such as caffeine, theophylline, and 1,3-dialkyl-8-phenylxanthine, have been synthesized which are lacking either the N7 (“7-deazaxanthines”) or the N9 (“9-deazaxanthines”) nitrogen atom in the imidazole partial structure (compounds 109–116, Fig. 5, Table 5). It was found that the nitrogen atom at the 9-position was not required for high receptor affinity, the

---

<sup>f</sup>Erickson et al. (1991)

<sup>g</sup>Petzer et al. (2003)

<sup>h</sup>Kase (2003)

<sup>i</sup>Shimada et al. (1997)

<sup>j</sup>Pretorius et al. (2008)

<sup>k</sup>Vlok et al. (2006)

<sup>l</sup>Jacobson et al. (1993a)

<sup>m</sup>Daly and Jacobson (1995)

<sup>n</sup>van Galen et al. (1994)

<sup>o</sup>Nonaka et al. (1994a)

<sup>p</sup>Müller et al. (1997a)

<sup>q</sup>Müller et al. (1998b)

<sup>r</sup>Müller et al. (2000)

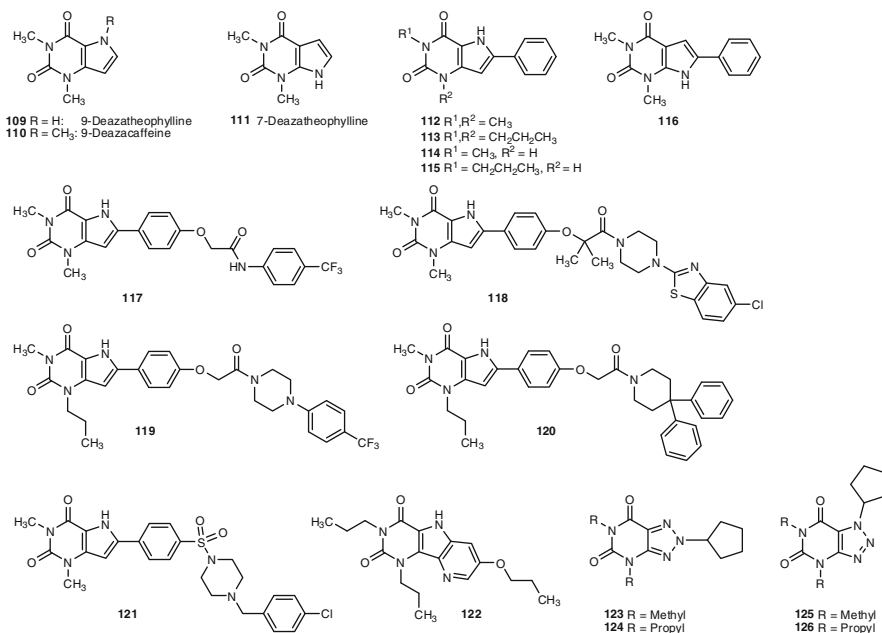
<sup>s</sup>Sauer et al. (2000)

<sup>t</sup>Solinas et al. (2005)

<sup>u</sup>Del Giudice et al. (1996)

<sup>v</sup>Massip et al. (2006)

<sup>w</sup>Müller et al. (1997a)



**Fig. 5** Deazaxanthines and azaxanthines

9-deazaxanthines being even slightly more potent at the A<sub>1</sub> AR in comparison with the corresponding xanthine derivatives (Grahner et al. 1994). In contrast, 7-deazaxanthines were much less potent, proving that the xanthines will bind as 7*H* rather than 9*H* tautomers to the receptors (Grahner et al. 1994). The addition of another nitrogen atom to the 8-position of xanthines was less successful: 8-azaxanthines (**123–126**, Fig. 5, Table 5) showed only moderate affinity for the receptors (Franchetti et al. 1994) which can be explained by the lack of the N7 hydrogen atom that is required as a hydrogen-bond donor for high-affinity binding.

## 2.5 Tricyclic Xanthine Derivatives

Several different types of tricyclic xanthine derivatives have been prepared and investigated (Fig. 6, Table 6). Cycloalkyl-substituted dihydroimidazo[2,1-*i*]purinones (**127**, **128**) showed high A<sub>1</sub> AR affinity and selectivity combined with improved water solubility owing to the presence of a basic nitrogen atom that can be protonated (Suzuki et al. 1992b; Vu et al. 2006). A new class of heterotricyclic xanthine derivatives in which the 3-alkyl substituent is tethered to the N9 atom – pyrimido[1,2,3-*cd*]purinediones (**151–153**) – was synthesized and investigated (Fig. 6) (Weyler et al. 2006). Interestingly, the cyclopentyl-substituted derivative **151**, an analogue of DPCPX, was only weakly active, probably due to the lack of

**Table 5** Adenosine receptor affinities of deazaxanthines and azaxanthines

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
Deazaxanthines				
<b>109</b> 9-Deazatheophylline	5,400 (r) <sup>b</sup>	12,000 (r) <sup>b</sup>	ND	ND
<b>110</b> 9-Deazacaffeine	32,000 (r) <sup>b</sup>	72,000 (r) <sup>b</sup>	ND	ND
<b>111</b> 7-Deazatheophylline	43,000 (r) <sup>b</sup>	>250,000 (r) <sup>b</sup>	ND	ND
<b>112</b> 1,3-Dimethyl-8-phenyl-9-deazaxanthine	47 (r) <sup>b</sup>	510 (r) <sup>b</sup>	ND	ND
<b>113</b> 1,3-Dipropyl-8-phenyl-9-deazaxanthine	13 (r) <sup>b</sup>	450 (r) <sup>b</sup>	ND	ND
<b>114</b> 1-Methyl-8-phenyl-9-deazaxanthine	97 (r) <sup>b</sup>	2,000 (r) <sup>b</sup>	520 (h) <sup>c</sup>	2,098 (h) <sup>c</sup>
<b>115</b> 1-Propyl-8-phenyl-9-deazaxanthine	45 (h) <sup>c</sup> 39 (r) <sup>b</sup>	>10,000 (h) <sup>c</sup> 1,200 (r) <sup>b</sup>	42 (h) <sup>c</sup>	380 (h) <sup>c</sup>
<b>116</b> 1,3-Dimethyl-8-phenyl-7-deazaxanthine	3,100 <sup>b</sup>	12,000 <sup>b</sup>	ND	ND
<b>117</b>	14.8 (h) <sup>d</sup>	64.6 (h) <sup>d</sup>	3.02 (h) <sup>d</sup>	>1,000 (h) <sup>d</sup>
<b>118</b>	>1,000 (h) <sup>e</sup>	10,000 (h) <sup>e</sup>	11.0 (h) <sup>e</sup>	>1,000 (h) <sup>e</sup>
<b>119</b>	89.1 (h) <sup>e</sup>	324 (h) <sup>e</sup>	2.04 (h) <sup>e</sup>	2,240 (h) <sup>e</sup>
<b>120</b>	676 (h) <sup>e</sup>	3,550 (h) <sup>e</sup>	5.26 (h) <sup>e</sup>	>1,000 (h) <sup>e</sup>
<b>121</b>	183 (h) <sup>f</sup>	ND	1 (h) <sup>f</sup>	12,260 (h) <sup>f</sup>
<b>122</b> Tricyclic 9-deazaxanthine	346 (h) <sup>g</sup>	164 (h) <sup>g</sup>	ND	3.82 (h) <sup>g</sup>
8-Azaxanthines				
<b>123</b> 1,3-Dimethyl-8-cyclopentyl-8-azaxanthine	110,000 (c) <sup>h</sup>	58,000 (c) <sup>h</sup>	ND	ND
<b>124</b> 1,3-Dipropyl-8-cyclopentyl-8-azaxanthine	1,300 (c) <sup>h</sup>	13,000 (c) <sup>h</sup>	ND	ND
<b>125</b> 1,3-Dimethyl-7-cyclopentyl-8-azaxanthine	11,000 (c) <sup>h</sup>	292,000 (c) <sup>h</sup>	ND	ND
<b>126</b> 1,3-Dipropyl-7-cyclopentyl-8-azaxanthine	340 (c) <sup>h</sup>	10,000 (c) <sup>h</sup>	ND	ND

<sup>a</sup>*n* human, *c* cow, *r* rat, a few A<sub>2B</sub> data may be from functional (cyclic AMP) studies, *ND* no data available

<sup>b</sup>Grahner et al. (1994)

<sup>c</sup>Hayallah et al. (2002)

<sup>d</sup>Carotti et al. (2006)

<sup>e</sup>Stefanachi et al. (2008)

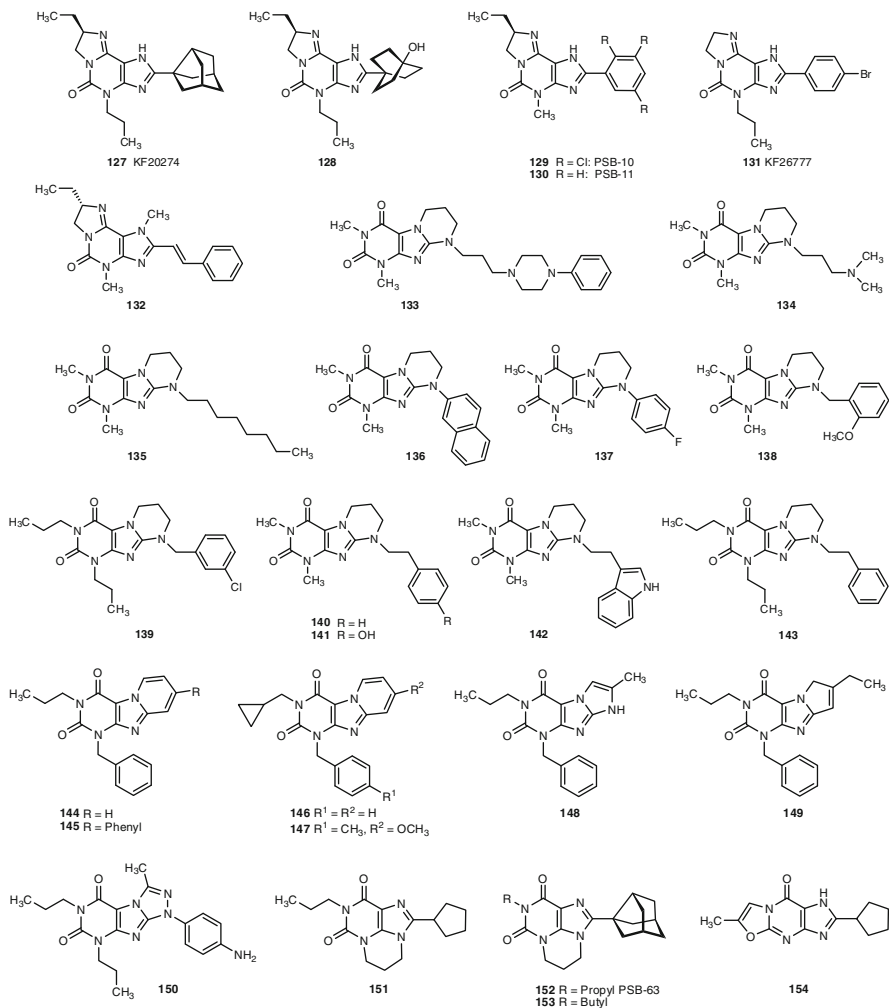
<sup>f</sup>Esteve et al. (2006)

<sup>g</sup>Ishiyama et al. (2009)

<sup>h</sup>Franchetti et al. (1994)

the N7 hydrogen atom. In contrast, the 3-noradamantyl-substituted analogues (**152**, **153**) showed relatively high A<sub>1</sub> AR affinity. While propyl derivative **152** (PSB-63) was very selective compared with the other AR subtypes, butyl derivative **153** was also quite potent at the human A<sub>3</sub> AR (Table 6). Another novel tricyclic analogue of DPCPX, the oxazo[3,2-*a*]purinone derivative **154**, showed only weak affinity for ARs (Table 6) (Müller 1994). In a series of tricyclic pyrimido[2,1-*f*]purinediones the *N,N*-dipropyl-substituted derivative **139** (Fig. 6), bearing a *m*-chlorobenzyl





**Fig. 6** Tricyclic xanthine derivatives

residue attached to the additional ring, was a relatively potent A<sub>1</sub> AR antagonist with some selectivity (Table 6) (Drabczynska et al. 2007a).

### 3 A<sub>2A</sub> Adenosine Receptor Antagonists

A<sub>2A</sub>-AR-selective antagonists of both xanthine and nonxanthine classes have been developed and some have entered clinical trials for Parkinson's disease, based on the opposing action of adenosine and dopamine in the striatal pathways in the brain

**Table 6** Adenosine receptor affinities of tricyclic xanthine derivatives

Name	$K_i$ (nM) <sup>a</sup>				
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>	
Imidazo[2,1- <i>i</i> ]purin-5-ones					
<b>127</b>	KF20274	2.7 (r) <sup>b</sup>	290 (r) <sup>b</sup>	ND	ND
<b>128</b>		22 (h) <sup>c</sup>	4,400 (h) <sup>c</sup>	580 (h) <sup>c</sup>	>10,000 (h) <sup>c</sup>
		6 (r) <sup>c</sup>	2,700 (r) <sup>c</sup>		
<b>129</b>	PSB-10	1,700 (h) <sup>d</sup>	2,700 (h) <sup>d</sup>	ND	0.441 (h) <sup>e</sup>
		805 (r) <sup>e</sup>	6,040 (r) <sup>e</sup>		
<b>130</b>	PSB-11	1,640 (h) <sup>d</sup>	1,280 (h) <sup>d</sup>	2,100 (m) <sup>e</sup>	2.34 (h) <sup>d</sup>
		440 (r) <sup>d</sup>	2,100 (r) <sup>d</sup>		$K_D$ 4.9 (h) <sup>f</sup>
<b>131</b>	KF26777	1,800 (h) <sup>g</sup>	470 (h) <sup>g</sup>	620 (h) <sup>g</sup>	0.20 (h) <sup>g</sup>
<b>132</b>		14,900 (r) <sup>e</sup>	424 (r) <sup>e</sup>	3,700 (m) <sup>e</sup>	30,600 (h) <sup>e</sup>
Pyrimido[2,1- <i>f</i> ]purinediones					
<b>133</b>		15,000 (r) <sup>h</sup>	16,000 (r) <sup>h</sup>	ND	ND
<b>134</b>		20,000 (r) <sup>h</sup>	>250,000 (r) <sup>h</sup>	ND	ND
<b>135</b>		>25,000 (r) <sup>i</sup>	998 (r) <sup>i</sup>	5,200 (h) <sup>i</sup>	12,300 (h) <sup>i</sup>
<b>136</b>		26,800 (h) <sup>j</sup>	2,870 (h) <sup>j</sup>	~10,000 (h) <sup>j</sup>	>10,000 (h) <sup>j</sup>
		≥25,000 (r) <sup>j</sup>	219 (r) <sup>j</sup>		
<b>137</b>		16,700 (h) <sup>j</sup>	1,880 (h) <sup>j</sup>	~10,000 (h) <sup>j</sup>	>10,000 (h) <sup>j</sup>
		>25,000 (r) <sup>j</sup>	147 (r) <sup>j</sup>		
<b>138</b>		>25,000 (r) <sup>k</sup>	11,300 (h) <sup>k</sup>	ND	ND
			699 (r) <sup>k</sup>		
<b>139</b>		89 (r) <sup>k</sup>	478 (r) <sup>k</sup>	ND	1,290 (h) <sup>k</sup>
<b>140</b>		>10,000 (h) <sup>l</sup>	2,890 (h) <sup>l</sup>	~10,000 (h) <sup>l</sup>	>10,000 (h) <sup>l</sup>
		>25,000 (r) <sup>l</sup>	320 (r) <sup>l</sup>		
<b>141</b>		>25,000 (h) <sup>l</sup>	630 (h) <sup>l</sup>	7,200 (h) <sup>l</sup>	>10,000 (h) <sup>l</sup>
		~25,000 (r) <sup>l</sup>	230 (r) <sup>l</sup>		
<b>142</b>		>25,000 (h) <sup>l</sup>	4,560 (h) <sup>l</sup>	~10,000 (h) <sup>l</sup>	>10,000 (h) <sup>l</sup>
		~25,000 (r) <sup>l</sup>	330 (r) <sup>l</sup>		
<b>143</b>		620(r) <sup>l</sup>	860(r) <sup>l</sup>	590 (h) <sup>l</sup>	3,660 (h) <sup>l</sup>
Pyrido[2,1- <i>f</i> ]purinediones					
<b>144</b>		50 (h) <sup>m</sup>	119 (h) <sup>m</sup>	ND	4.0 (h) <sup>m</sup>
<b>145</b>		>10,000 (h) <sup>m</sup>	>10,000 (h) <sup>m</sup>	ND	35 (h) <sup>m</sup>
<b>146</b>		>1,000 (h) <sup>n</sup>	242 (h) <sup>n</sup>	ND	4.2 (h) <sup>n</sup>
<b>147</b>		>1,000 (h) <sup>n</sup>	>1,000 (h) <sup>n</sup>	>1,000 (h) <sup>n</sup>	2.24 (h) <sup>n</sup>
Imidazopurinediones, pyrrolopurinediones, and triazolopurinediones					
<b>148</b>		>1,000 (h) <sup>o</sup>	>1,000 (h) <sup>o</sup>	>1,000 (h) <sup>o</sup>	0.8 (h) <sup>o</sup>
<b>149</b>		>1,000 (h) <sup>o</sup>	>1,000 (h) <sup>o</sup>	>1,000 (h) <sup>o</sup>	3.5 (h) <sup>o</sup>
<b>150</b>		>10,000 (h) <sup>p</sup>	2,050 (h) <sup>p</sup>	>100,000 (h) <sup>p</sup>	1,330 (h) <sup>p</sup>
4,5-Dihydro-6 <i>H</i> ,8 <i>H</i> -pyrimido[1,2,3- <i>cd</i> ]purine-8,10-diones					
<b>151</b>		1,440 (r) <sup>q</sup>	12,400 (r) <sup>q</sup>	42,600 (h) <sup>q</sup>	ND
<b>152</b>	PSB-63	16.9 (r) <sup>q</sup>	22,000 (r) <sup>q</sup>	3,190 (h) <sup>q</sup>	>10,000 (h) <sup>q</sup>
		90.6 (h) <sup>q</sup>	34,500 (h) <sup>q</sup>		
<b>153</b>		40.6 (r) <sup>q</sup>	23,400 (r) <sup>q</sup>	22,300 (h) <sup>q</sup>	188 (h) <sup>q</sup>
		13.8 (h) <sup>q</sup>	~25,000 (h) <sup>q</sup>		
Oxazolo[3,2- <i>a</i> ]purinone					
<b>154</b>		770 (r) <sup>r</sup>	20,600 (r) <sup>r</sup>	ND	ND

<sup>a</sup>*h* human, *m* mouse, *r* rat, a few A<sub>2B</sub> data may be from functional (cyclic AMP) studies, *ND* no data available

<sup>b</sup>Suzuki et al. (1992b)

(Richardson et al. 1997; Schapira et al 2006; Schwarzschild et al. 2006; Müller and Ferré 2007; Baraldi et al. 2008). The cellular mechanisms of the motor and neuroprotective effects of  $A_{2A}$  AR antagonists have been explored (Yu et al. 2008). Recently, ameliorating effects of  $A_{2A}$  AR antagonists including xanthine derivatives on animal models of Alzheimer's disease and cognitive dysfunction have been reported (Dall'Igna et al. 2007; Cunha et al. 2008; Takahashi et al. 2008). Since the early 1990s, there has been a major medicinal chemical effort to increase the  $A_{2A}$  AR selectivity of simple xanthines by structural modification.

Prior to the synthesis of truly  $A_{2A}$ -AR-selective antagonists, certain high-affinity xanthines were used in a non-selective fashion as probes of the  $A_{2A}$  AR. For example, [ $^3$ H]XAC (**30**) was useful as a radiotracer in binding experiments at the  $A_{2A}$  AR in human platelets and was therefore the first antagonist radioligand with high affinity at the  $A_{2A}$  AR (Ukena et al. 1986a). PD115,199 (**32**) was prepared in tritiated form and shown to bind with high affinity to the rat  $A_{2A}$  AR (Bruns et al. 1987b).

The first "selective"  $A_{2A}$  AR antagonist described in the literature was the caffeine analogue DMPX (**20**, Fig. 1, Table 1) (Ukena et al. 1986b). Like caffeine, the compound possesses low  $A_{2A}$  AR affinity and moderate selectivity compared with  $A_1$  ARs. Nevertheless, this compound has been widely used in in vivo studies because of its good water solubility and bioavailability (Seale et al. 1988; Thorsell et al. 2007). Later it was found that DMPX is as potent at the  $A_{2B}$  AR as at the  $A_{2A}$  AR. The species dependence of affinity at the  $A_{2A}$  AR of 1,3,7- and 1,3,8-trisubstituted xanthines has been reported (Stone et al. 1988).

An early example of a caffeine analogue that displayed selectivity for the  $A_{2A}$  AR was 8-trifluoromethylcaffeine, but the affinity was still low, with a  $K_i$  value in binding at the rat  $A_{2A}$  AR of 29  $\mu$ M (Jacobson et al. 1993b). This effect of the 8-trifluoromethyl group was not observed in the corresponding (inactive) theophylline derivative. An 8-(*trans*-2-carboxyvinyl) derivative of caffeine also proved to be similarly selective for the  $A_{2A}$  AR.

---

<sup>c</sup>Vu et al. (2006)

<sup>d</sup>Ozola et al. (2003)

<sup>e</sup>Müller et al. (2002a)

<sup>f</sup>Müller et al. (2002b)

<sup>g</sup>Saki et al. (2002)

<sup>h</sup>Geis et al. (1995)

<sup>i</sup>Drabczynska et al. (2004)

<sup>j</sup>Drabczynska et al. (2006)

<sup>k</sup>Drabczynska et al. (2007a)

<sup>l</sup>Drabczynska et al. (2007b)

<sup>m</sup>Priego et al. (2002)

<sup>n</sup>Priego et al. (2008)

<sup>o</sup>Baraldi et al. (2005)

<sup>p</sup>Pastorin et al. (2005)

<sup>q</sup>Weyler et al. (2006)

<sup>r</sup>Müller (1994)

### 3.1 8-Styrylxanthines

The observation that N7-methylation in 8-substituted xanthine derivatives was better tolerated by the A<sub>2A</sub> AR than the A<sub>1</sub> AR (Shamim et al. 1989) and that the 8-substituent had to be coplanar to achieve high A<sub>2A</sub> AR affinity (Erickson et al. 1991) led to the first highly potent and selective A<sub>2A</sub> AR antagonists: the 1,3,7-alkyl-substituted 8-styrylxanthine derivatives **87–95** and **99** (Fig. 4, Table 4).

A small alkyl group at N1 (methyl, ethyl, propyl, propargyl) proved to be optimal for high A<sub>1</sub> AR affinity and selectivity, while methylation is required at the 7-position (Jacobson et al. 1993a; Nonaka et al. 1994a; Shimada et al. 1997; Müller et al. 2000; Kase 2003). The 8-styryl residue has to be *E*-configured, and *m*-chloro or *m*-methoxy substitution improved affinity and selectivity. The *meta* position of the 8-styryl ring can be substituted with elongated chains with retention of A<sub>2A</sub> AR selectivity and enhancement of water solubility (Jacobson et al. 1993a). The phenyl ring in the 8-styryl residue can be substituted by heterocyclic rings, such as a 3-thienyl ring (**102**) (Del Giudice et al. 1996).

The most common substituents at N3 in A<sub>2A</sub>-AR-selective xanthine derivatives are small alkyl residues, such as methyl, propyl, and 3-hydroxypropyl (reviewed in Müller 2000; Cacciari et al. 2003; Vu 2005; Yuzlenko and Kiec-Kononowicz 2006; Müller and Ferré 2007; Cristalli et al. 2007, 2009). Recently, the development of a new synthetic approach allowed the preparation of a series of xanthine derivatives with more variations in the 3-position (Massip et al. 2006). It was found that the A<sub>2A</sub> AR tolerated bulky, functionalized substituents at the 3-position. For instance, N3-phenoxypopyl-substituted 8-(methoxystyryl)xanthine derivatives (e.g. **103**) are potent and selective A<sub>2A</sub> AR antagonists (Massip et al. 2006).

Some of the best A<sub>2A</sub> AR antagonists were istradefylline (KW6002, **87**), *m*-chlorostyrylcaffeine ((*E*)-8-*m*-Chlorostyrylcaffeine, **89**), *m*-bromostyryl-DMPX (**93**), and 3-(3-hydroxypropyl)-7-methyl-8-(*m*-methoxystyryl)-1-propargylxanthine (MSX-2, **99**). (*E*)-8-(3-Chlorostyryl)caffeine (**89**) is not only a potent A<sub>2A</sub> AR antagonist ( $K_i$  rat A<sub>2A</sub> AR 54 nM), but in addition, it has been reported to be a potent inhibitor of monoaminoxidase type B (MAO-B) (baboon MAO-B,  $K_i$  80.6 nM), an enzyme which metabolizes dopamine (van den Berg et al. 2007; Petzer et al. 2009). This activity may contribute to the potency of CSC in in vivo studies, e.g. in animal models of Parkinson's disease. All other styrylxanthine derivatives investigated so far, including 8-styrylcaffeine (**88**) and istradefylline (**87**), are considerably less potent as MAO-B inhibitors than CSC. Recently, a chain-extended homologue of CSC, 8-(*m*-chlorophenylbutadienyl)caffeine (**108**), has been described as showing similar dual activity as an A<sub>2A</sub> AR antagonist and an MAO-B inhibitor (Pretorius et al. 2008).

Istradefylline (KW-6002, **87**) has been intensively studied in in vitro and in a number of animal models. Until recently (Fernandez et al. 2010), it was in phase IIIb clinical trials for Parkinson's disease. In phase II clinical trials istradefylline reduced motoric dysfunction without producing dyskinesias (reviewed by Knutsen

and Weiss 2001). A  $^{11}\text{C}$ -labelled version of istradefylline has been prepared and used for positron emission tomography (PET) studies in healthy human brain (Hirani et al. 2001).

A major drawback of styrylxanthine derivatives, however, is their high lipophilicity and low water solubility. Introduction of a polar sulfonate group into 8-styryl-DMPX, resulting in compound **98**, led to an almost tenfold reduction in  $\text{A}_{2\text{A}}$  affinity, but increased water solubility (Müller et al. 1998). A more successful approach has been the preparation of water-soluble prodrugs, particularly of the 3-(3-hydroxypropyl)-substituted 1-propargyl-8-styrylxanthine derivative MSX-2 (**99**) (Müller 2009). MSX-3 (**100**) is a water-soluble phosphate prodrug of MSX-2, which is cleaved in vivo by ubiquitous phosphatases to release the  $\text{A}_{2\text{A}}$  AR antagonist MSX-2 (Sauer et al. 2000). MSX-3 has proven useful for animal studies and is widely used for studying the in vivo effects of  $\text{A}_{2\text{A}}$  AR antagonists (Hauber et al. 1998, 2001; Strömberg et al. 2000; Ferré et al. 2001, 2008; Nagel et al. 2003; Blum et al. 2003; Schindler et al. 2004, 2005; Antoniou et al. 2005; Karcz-Kubicha et al. 2003a, b; Filip et al. 2006; Fuxe et al. 2007; Ishiwari et al. 2007; Farrar et al. 2007; Carriba et al. 2007; Salamone et al. 2008a, b; Marcellino et al. 2008; Mott et al. 2009; Worden et al. 2009). Owing to its very high water solubility at the physiological pH of 7.4 (9 mg/mL), it can be directly injected into specific brain areas, but is also an effective  $\text{A}_{2\text{A}}$  AR antagonist after systemic application. Recently, an amino acid ester prodrug of MSX-2, MSX-4 (**101**), was synthesized, and was found to be very soluble in water, highly stable in artificial gastric fluid, but readily cleaved by esterases and may be a suitable prodrug for peroral administration (Vollmann et al. 2008).

Care has to be taken when using the *E*-configured styrylxanthines since they easily undergo light-induced isomerization in dilute solutions yielding mixtures of *E* and *Z* isomers, the *Z* isomers being only weakly active or inactive (Nonaka et al. 1993; Müller et al. 1998). This isomerization does not occur in concentrated solution, e.g. during synthesis of the compounds, or when the compounds are applied as solid dosage forms. However, styrylxanthines can also undergo light-induced dimerization ([2 + 2] cycloaddition reaction) in the solid state, and therefore have to be rigorously stored under the exclusion of light (Hockemeyer et al. 2004).

### 3.2 Configurationally Stable Analogues of 8-Styrylxanthines

To overcome the problem of photoisomerization, the styryl moiety has been replaced with different, more stable bioisosteric groups (e.g. replacement of the double bond for a cyclopropyl ring in **104**, a 2-naphthyl residue in **105**, a triple bond in **107** (Müller et al. 1997c), or a tricyclic constrained structure (**133–143**) (Kiec-Kononowicz et al. 2001; Drabczynska et al. 2003, 2004, 2006, 2007b; Fhid et al. 2003). In most cases, a significant loss of affinity was observed by such modifications. The most promising compounds were the pyrimido[2,1-*f*]

purinedione derivative (**141**) ( $K_i$  human  $A_{2A}$  AR 630 nM, rat  $A_{2A}$  AR 230 nM) and 8-phenylethynyl-DMPX (**107**,  $K_i$  human  $A_{2A}$  AR 314 nM, rat  $A_{2A}$  AR 300 nM), both endowed with high selectivity. The latter class of compounds has been optimized for increased  $A_{2A}$  AR affinity and the highly potent and selective  $A_{2A}$  AR antagonists obtained were described in a recent patent (Müller et al. 2008). Furthermore, a substitution of the ethenyl group with a diazo structure has been performed. The compounds obtained retained selectivity but showed only moderate affinity (Müller et al. 1997b).

### 3.3 $A_{2A}$ -Adenosine-Receptor-Selective Radiolabelled Xanthine Derivatives

The tritiated derivative of the 8-styrylxanthine KF17837S [the equilibrium mixture of (*E*)-KF17837 and (*Z*)-KF17837 isomers] was shown to bind to rat striatal membranes in a saturable and reversible way, with  $K_D$  values of low nanomolar concentration (Nonaka et al. 1994b). Another  $A_{2A}$  AR antagonist radioligand was prepared, [ $^3$ H]MSX-2. This molecule showed high affinity ( $K_D = 8.0$  nM) for rat and human  $A_{2A}$  ARs, with saturable and reversible binding, and also a  $A_{2A}$ -selectivity of at least 2 orders of magnitude compared with all other AR subtypes (Müller et al. 2000).

### 3.4 Heterocyclic Compounds Related to Xanthines

A tricyclic styryl-substituted imidazo[2,1-*i*]purin-5-one derivative (**132**, Fig. 6, Table 6) showed enhanced water solubility but reduced  $A_{2A}$  AR affinity and moderate selectivity (Müller et al. 2002a).

## 4 $A_{2B}$ Adenosine Receptor Antagonists

### 4.1 Aryl-Substituted 1,3-Dialkylxanthines

From the initial studies of Daly and coworkers using cyclic AMP studies in the brain slice, it was recognized that 1,3,7- and 1,3,8-trisubstituted xanthines have considerable affinity at the  $A_{2B}$  AR. Also, the simple xanthine enprofylline (**10**) was discovered to have slight selectivity for the  $A_{2B}$  AR, which was proposed to be responsible for its antiasthmatic action in the clinic (Stefanovich 1989; Daly 2000, 2007). Screening efforts by Bruns (1981) followed by more detailed studies by Müller, Daly, and Jacobson showed that 1-monosubstituted xanthine derivatives, such as 1-propylxanthine (**6**) and 1-butylxanthine (**7**), were about

tenfold more potent than enprofylline at  $A_{2B}$  AR and equally selective (Müller et al. 1993; Kim et al. 2002).

In fact, the unintended interaction at the  $A_{2B}$ AR of widely used xanthine antagonists of the ARs has proven to be a complication in pharmacology studies.

Many known xanthines were screened at the  $A_{2B}$ AR to identify leads for the design of novel  $A_{2B}$  AR antagonists. The first successful efforts to enhance the activity of 1,3,8-trisubstituted xanthines at the  $A_{2B}$ AR by Jacobson and colleagues resulted in one compound of intermediate selectivity at the human, but not rat  $A_{2B}$  AR, MRS1595 (**41**), which is a hydrazide derivative of XCC (Fig. 2, Table 2) (Kim et al. 2000). Then, further probing of the structure–activity relationship culminated in the introduction of MRS1754 (**37**), which was the first selective  $A_{2B}$  AR antagonist with nanomolar affinity at the human receptor (Kim et al. 2000). The degree of selectivity for the human  $A_{2B}$  AR was more than 120-fold, but selectivity for the rat  $A_{2B}$  AR was considerably less (Fig. 2, Table 2). Thus, it remained a challenge to design a rat  $A_{2B}$ -AR-selective xanthine antagonist. Another drawback in the series of anilide derivatives of XCC is the low aqueous solubility, which is partly remedied in related antagonists such as MRS1706 (**38**). Nevertheless, [ $^3$ H] MRS1754 has found application as a useful radioligand of the  $A_{2B}$  AR (Ji et al. 2001). Structurally related 8-phenylxanthine derivatives include CVT-5440 (**40**), in which additional aromatic rings were attached by an ether linkage, and **42**, with a modified 3-substituent (3-methoxypropyl), and were developed as potent and selective  $A_{2B}$  AR antagonists (Kim et al. 2000; Nieto et al. 2009). Newer derivatives in this series, which have two pyridine rings linked by an amide group, include the highly selective  $A_{2B}$  AR antagonists ATL-802 (**57**) and ATL-852 (**58**). [ $^3$ H] ATL-852 has been reported as a high-affinity radioligand at this receptor (Cagnina et al. 2009).

8-Pyrazolyl-substituted xanthines that have been developed as selective human  $A_{2B}$  AR antagonists include MRE-2029-F20 (**56**), which was also reported as a high-affinity radioligand (Baraldi et al. 2004). A different series of isomeric 8-pyrazolylxanthines yielded the highly potent  $A_{2B}$  AR antagonists CVT-6694, CVT-7124, and CVT-6883 (**53–55**) (Kalla et al. 2008; Elzein et al. 2008; Kalla and Zablocki 2009). High selectivity at human receptors has been found for all of these pyrazolylxanthines, but no data for rodent receptors have been reported. CVT-6883 (**55**) is a promising candidate for the treatment of diabetes or asthma and has entered phase I clinical trials. Pain treatment is another potential area under consideration for  $A_{2B}$  AR antagonists (Abo-Salem et al. 2004; Akkari et al. 2006; Bilkei-Gorzo et al. 2008; Michael et al. 2010).

## 4.2 1,8-Disubstituted Xanthines

The observation that 1-monosubstituted and 1,8-disubstituted xanthine derivatives showed high affinity and increased selectivity for the  $A_{2B}$  AR led to the development of a series of 1-alkyl-8-phenylxanthine derivatives (**43–52**) (Hayallah

et al. 2002; Yan and Müller 2004; Yan et al. 2006; Borrmann et al. 2009). These compounds also appeared to show reduced affinity at the rat A<sub>1</sub> AR and therefore increased A<sub>2B</sub> AR selectivity in rat. 1-Propyl-8-*p*-sulphophenylxanthine (PSB-1115, **43**) was developed as a water-soluble A<sub>2B</sub> AR antagonist, useful as a pharmacological tool for in vivo studies (Müller et al. 1993; Kirfel et al. 1997; Abo-Salem et al. 2004; Bilkei-Gorzo et al. 2008). 1-Butyl-8-(*p*-carboxyphenyl)xanthine (PSB-53, **45**) showed similar affinity and selectivity. 1-Propargyl-8-*p*-bromophenylxanthine (PSB-50, **44**) was more potent, but somewhat less selective and much less water soluble. The 8-phenylxanthine derivatives PSB-55 (**46**), a benzylpiperazine derivative, and PSB-298 (**47**), a hydroxyethylamide, were synthesized to obtain more polar compounds with high A<sub>2B</sub>AR affinity. PSB-298 was obtained in tritiated form and was found to have a low degree of non-specific binding (Bertarelli et al. 2006). However, its affinity and selectivity were not satisfactory.

Starting from the sulfonate PSB-1115 (**43**), sulfonic acid esters (e.g. **48**) and sulfonamides (e.g. **49–52**) were obtained (Yan and Müller 2004; Yan et al. 2006). Compound **48** can be envisaged as a lipophilic prodrug of the highly polar sulfonate **43**, which may show peroral bioavailability and release of **43** after absorption (Yan and Müller 2004). However, **48** has high A<sub>2B</sub>AR affinity itself and can therefore be classified as a limited prodrug, although without selectivity versus the A<sub>1</sub> AR (Fig. 2, Table 2). The most potent and selective A<sub>2B</sub>AR antagonists described to date are the sulfonamide derivatives **50–52**, whose development was based on PSB-601 (**49**), an already very potent and selective A<sub>2B</sub>AR antagonist ( $K_i$  3.6 nM). Compounds **50–52** show subnanomolar affinity for A<sub>2B</sub> ARs and very high selectivity in humans and in rodents. [<sup>3</sup>H]PSB-603 was prepared as a selective, high-affinity A<sub>2B</sub>AR antagonist radioligand with  $K_D$  values of 0.403 nM at human and 0.351 nM at mouse A<sub>2B</sub>AR (Borrmann et al. 2009).

### 4.3 9-Deazaxanthines

Several series of 9-deazaxanthine derivatives (**115**, **117–121**) were developed as A<sub>2B</sub>AR antagonists, and were structurally related to the 8-phenylxanthine derivatives described earlier (Fig. 5, Table 5). A number of compounds with low nanomolar affinity were obtained, and some were selective for the human A<sub>2B</sub>AR (Carotti et al. 2006; Esteve et al. 2006; Stefanachi et al. 2008).

### 4.4 8-Furylmethyl-Substituted Xanthines

8-(2-Furyl)methyl-substituted xanthined derivatives, e.g. **63**, have been developed as A<sub>2B</sub> AR antagonists (Fig. 2, Table 2). Some of them showed high A<sub>2B</sub> AR affinity but only moderate selectivity (Balo et al. 2009).



## 5 A<sub>3</sub> Adenosine Receptor Antagonists

In the search for A<sub>3</sub>AR antagonists, alkylxanthines were initially rejected as a suitable lead in favour of non-xanthine chemically diverse heterocycles, because of the exceptionally low affinity of alkylxanthines at the rat A<sub>3</sub>AR. For example, the classic adenosine antagonists caffeine and theophylline have  $K_i$  values of more than 100  $\mu\text{M}$  at the rat A<sub>3</sub>AR (Table 1). Initial structure–activity relationship studies at the rat A<sub>3</sub>AR were conducted using multiply substituted xanthines, many of which retained selectivity for the A<sub>3</sub>AR (van Galen et al. 1994). Only slight A<sub>3</sub>AR selectivity was observed for analogues containing 8-alkyl and 2-thio substitutions (Kim et al. 1994b). However, when other orthologues of the A<sub>3</sub>AR were cloned and studied pharmacologically, such as sheep, and human A<sub>3</sub>AR, many xanthines were found to display good affinity for those A<sub>3</sub>ARs (Linden 1994). Thus, attention returned to the xanthines as a source for A<sub>3</sub>AR antagonist leads.

One of the earliest approaches to enhancing the affinity of xanthines at the A<sub>3</sub>AR was to attach a ribose group at the 7-position (**21**, **22**). The 5'-uronamide derivative DBXRM (**22**) is 140-fold selective for the A<sub>3</sub>AR, but the presence of the uronamide function increases the efficacy such that it is an agonist at this receptor (van Galen et al. 1994; Kim et al. 1994b; Bridson et al. 1998).

### 5.1 8-Aryl-Substituted Xanthine Derivatives

8-Phenylxanthine derivatives bearing a carboxylate group attached to the phenyl ring via an ethylene (**35**) or an oxymethylene (**34**) spacer were initially found to be potent antagonists at the human A<sub>3</sub>AR, but both compounds are also very potent A<sub>1</sub> and A<sub>2B</sub>AR antagonists (Fig. 2, Table 2). [<sup>125</sup>I]-ABOPX (BW-A522, **34**) has been used as a radioligand for labelling the A<sub>1</sub>AR as well as human A<sub>3</sub> and human A<sub>2B</sub>ARs (Patel et al. 1988; Salvatore et al. 1993; Linden et al. 1999). The corresponding *p*-azido derivative was previously used for photoaffinity labelling of the A<sub>1</sub>AR. The xanthine derivative **34** is characterized by a *p*-amino-*m*-iodobenzyl residue at N3 of the xanthine core (Fig. 2) and is therefore quite lipophilic. As observed for other xanthine antagonists, **34** is much less potent (65-fold) at rat than at human A<sub>3</sub>ARs.

### 5.2 Tricyclic Xanthine and Deazaxanthine Derivatives

Cyclized derivatives of xanthines, such as (8*R*)-8-ethyl-4-methyl-2-phenyl-4,5,7,8-tetrahydro-1*H*-imidazo[2.1-*i*]purin-5-one (PSB-11, **130**), its trichlorophenyl-substituted derivative PSB-10 (**129**), and the 8-unsubstituted 8-bromophenyl derivative **131** are very potent A<sub>3</sub>-AR-selective antagonists (Fig. 6, Table 6) (Müller et al. 2002a; Saki et al. 2002; Ozola et al. 2003). PSB-11 ( $K_D$  4.9 nM) was prepared

as a radioligand by catalytic hydrogenation from the polychlorinated precursor PSB-10 (Müller et al. 2002b; Burbiel et al. 2003). Owing to its increased polarity and solubility in comparison with xanthines, [ $^3\text{H}$ ]PSB-11 shows only low non-specific binding. Further tricyclic xanthine derivatives, in which an additional ring was attached to the imidazole rather than the pyrimidine ring of the xanthine core structure (pyridopurinediones, imidazopurinediones, pyrrolopurinediones, and triazolopurinediones **144–150**), were developed as  $\text{A}_3\text{-AR}$ -selective antagonists (Priego et al. 2002; Baraldi et al. 2005; Pastorin et al. 2005; Priego et al. 2008). An aromatic residue (mostly benzyl) attached to N3 (xanthine numbering) increases  $\text{A}_3\text{AR}$  affinity in xanthine derivatives, including the tricyclic ones.

Besides tricyclic xanthine derivatives, tricyclic deaxanthines have also been obtained (Ishiyama et al. 2009). Compound **122** (Fig. 5, Table 5) was one of the most potent and selective compounds in this series.

## 6 Xanthine Derivatives Used as Molecular Probes

### 6.1 Irreversible Ligand Probes

The amine congener XAC (**30**) was coupled to a variety of bifunctional cross-linking reagents to form products containing a single chemically reactive group (Fig. 7, Table 7). For example, 1,3-phenylene and 1,4-phenylene diisothiocyanates

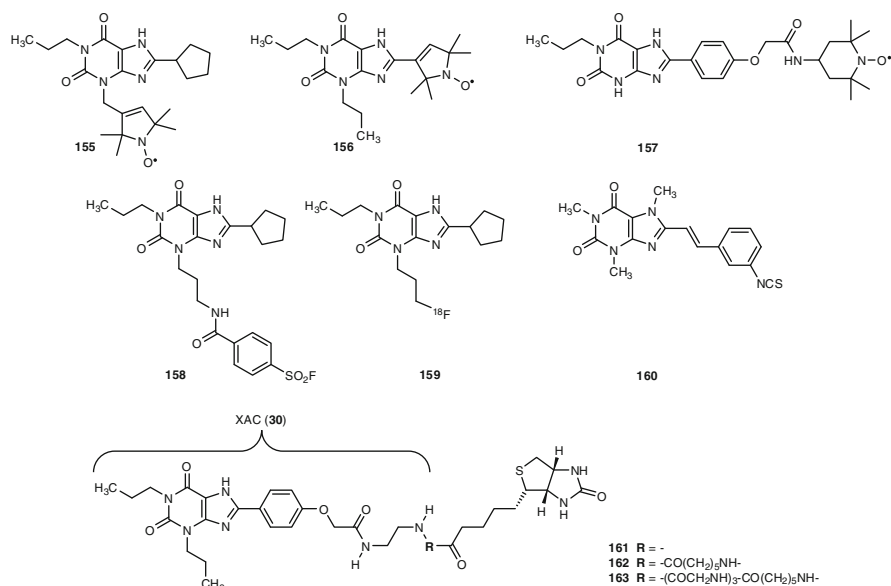
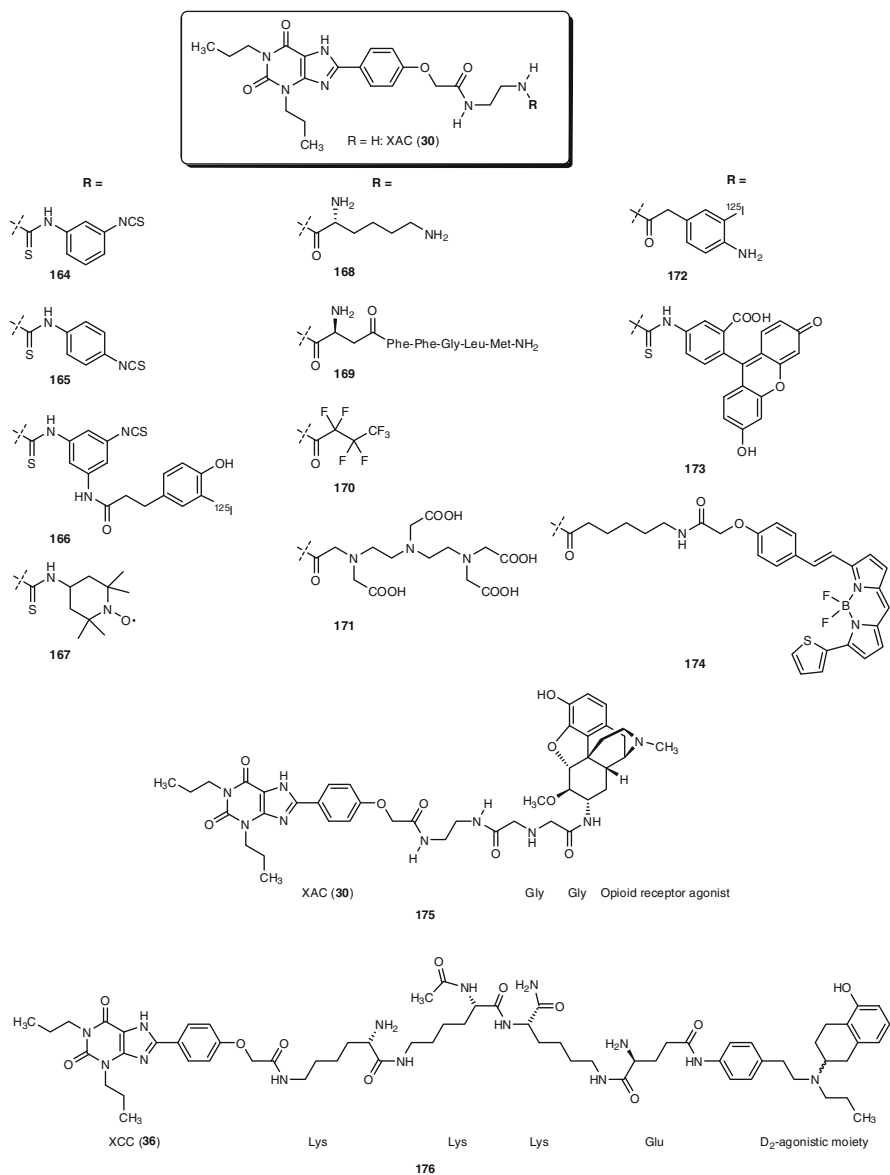


Fig. 7 (Continued)



**Fig. 7** Functionalized xanthines as molecular probes

were conjugated to XAC to form the A<sub>1</sub>-AR-selective antagonists *m*-phenylenedithiocyanate-XAC (**164**) and *p*-DITC-XAC (**165**), which were demonstrated to bind irreversibly at submicromolar concentrations to the A<sub>1</sub>AR to act as covalent affinity labels. Similarly, prosthetic reagents designed for radiolabelling and photoaffinity labelling could be coupled to XAC and similar amine congeners

**Table 7** Adenosine receptor affinities of functionalized xanthines as molecular probes

Name	$K_i$ (nM) <sup>a</sup>			
	A <sub>1</sub>	A <sub>2A</sub>	A <sub>2B</sub>	A <sub>3</sub>
Spin-labeled probes				
<b>155</b>	5.47 (r) <sup>b</sup>	8,780 (r) <sup>b</sup>	>1,000 (h) <sup>b</sup>	1,700 (h) <sup>b</sup>
<b>156</b>	8.23 (r) <sup>b</sup>	3,800 (r) <sup>b</sup>	3,100 (h) <sup>b</sup>	~10,000 (h) <sup>b</sup>
<b>157</b>	15.7 (r) <sup>b</sup>	1,270 (r) <sup>b</sup>	48 (h) <sup>b</sup>	350 (h) <sup>b</sup>
<b>167</b> TEMPO-XAC	4.9 (r) <sup>c</sup> 0.30 (c) <sup>c</sup>	ND	ND	ND
Irreversible ligands				
<b>158</b> FSCPX <sup>d,e</sup>	10 (r) <sup>d</sup>	ND	ND	ND
<b>160</b> ISC	42,600 (r) <sup>f</sup> 51,400 (gp) <sup>f</sup> 89,500 (rb) <sup>f</sup> 63,400 (c) <sup>f</sup>	146 (r) <sup>f</sup> 160 (gp) <sup>f</sup> 413 (rb) <sup>f</sup> 516 (c) <sup>f</sup>	ND	ND
<b>164</b> <i>m</i> -DITC-XAC	2.39 (r) <sup>g</sup> 52 (r) <sup>h</sup>	ND	ND	ND
<b>165</b> <i>p</i> -DITC-XAC	6.60 (r) <sup>g</sup> 27 (r) <sup>h</sup>	ND	ND	ND
Radioligands				
<b>159</b> [ <sup>18</sup> F]CFPFX	1.26 (h) <sup>i</sup> 0.63 (r) <sup>i</sup> 1.37(p) <sup>i</sup> 0.18 (c) <sup>i</sup>	940 (h) <sup>i</sup> 812 (r) <sup>i</sup>	ND	ND
<b>166</b>	40 (IC <sub>50</sub> ) (c) <sup>i</sup>	ND	ND	ND
<b>172</b> [ <sup>125</sup> I]PAPA-XAC	0.1 (c) <sup>k</sup>	ND	ND	ND
Biotin conjugates				
<b>161</b>	54 (r) <sup>l,m</sup>	ND	ND	ND
<b>162</b>	50 (r) <sup>l,m</sup>	ND	ND	ND
<b>163</b>	60 (r) <sup>m</sup>	ND	ND	ND
Various conjugates				
<b>168</b> <i>D</i> -Lys-XAC	1.74 (IC <sub>50</sub> ) (r) <sup>n</sup>	159 (IC <sub>50</sub> ) (r) <sup>n</sup>	ND	ND
<b>169</b>	35 (r) <sup>m</sup>	ND	ND	ND
<b>170</b>	8.1 (r) <sup>c</sup> 0.8 (c) <sup>c</sup>	ND	ND	ND
<b>171</b> DTPA-XAC	59.5 (r) <sup>c</sup> 3.25 (c) <sup>c</sup>	ND	ND	ND
Fluorescent ligands				
<b>173</b> FITC-XAC	125 (r) <sup>c</sup> 9.3 (c) <sup>c</sup>	ND	ND	ND
<b>174</b> XAC-BY630	151 (h) <sup>o</sup>	ND	ND	ND
Bivalent ligand conjugates				
<b>175</b> <sup>l,p</sup>	31 (r)	ND	ND	ND
<b>176</b> A <sub>2A</sub> antagonist/D <sub>2</sub> agonist for A <sub>2A</sub> /D <sub>2</sub> receptor heteromers ( $K_i$ D <sub>2</sub> (s) = 1.0 nM) <sup>q</sup>	ND	55 (s) <sup>q</sup>	ND	ND

TEMPO 2,2,6,6-tetramethylpiperidine-1-oxyl, FSCPX 8-cyclopentyl-3-*N*-[3-(3-(4-fluorosulphonyl)benzoyl)-oxy]-propyl]-1-*N*-propyl-xanthine, ISC 8-(3-isothiocyantostyryl)caffeine, DITC phenylenediisothiocyanate, PAPA *p*-phenylacetyl, DTPA diethylenediaminepentaaetic acid, FITC fluorescein isothiocyanate

(Stiles and Jacobson 1988; Jacobson et al. 1989a). The conjugate with the *p*-aminophenylacetyl moiety was readily iodinated to form the radioligand **172**, which could then be converted to the photoactivatable *p*-azide (Stiles and Jacobson 1987). This azide then served to cross-link a radiolabelled AR antagonist suitable for the A<sub>1</sub>AR protein, which could be visualized by gel electrophoresis. The sulfonyl fluoride group present in **158** was also a means of cross-linking a xanthine derivative to the A<sub>1</sub>AR (Scammels et al. 1994; van Muijlwijk-Koezen et al. 2001).

## 6.2 Spectroscopic Probes: Spin-Labelled and Fluorescent Probes

Other types of reporter groups could be similarly incorporated into xanthine-functionalized congeners with retention of moderate AR affinity, for example chelating groups capable of complexing radioactive metal ions; spin labels for electron spin resonance spectroscopy, e.g. **167**; a perfluorinated acyl prosthetic group, as in **170**, intended for use in fluorine-NMR spectroscopy; and fluorescent dyes, e.g. **173** and **174** (Jacobson et al. 1987b). The fluorescent conjugate (**174**) of XAC and BODIPY [6-(((4,4-difluoro-5-(2-thienyl)-4-bora-3a,4a-diaza-*s*-indacene-3-yl)styryloxy)acetic acid)], with an intermediate  $\epsilon$ -aminocaproyl spacer, has proven useful in fluorescence correlation spectroscopy to characterize ligand complexes of the A<sub>1</sub> AR (Briddon et al. 2004).

Spin-labelled probes that retained high A<sub>1</sub> AR affinity were obtained by inserting the spin label into the molecule as part of the pharmacophore. The most potent and A<sub>1</sub>-AR-selective compounds were the DPCPX analogues **155** (replacement of the 3-substituent by a spin label) and **156** (substitution of the cyclopentyl ring by a structurally related spin label). Both compounds showed affinity for the A<sub>1</sub> AR in the low nanomolar range combined with high selectivity compared with the other receptor subtypes (Ilas et al. 2005). The 1-propyl-8-phenyl derivative (**157**), in

---

<sup>a</sup>*h* human, *c* cow, *d* dog, *gp* guinea pig, *p* pig, *r* rat, *rb* rabbit, *s* sheep

<sup>b</sup>Ilas et al. (2005)

<sup>c</sup>Jacobson et al. (1987b)

<sup>d</sup>Scammels et al. (1994)

<sup>e</sup>van Muijlwijk-Koezen et al. (2001)

<sup>f</sup>Ji et al. (1993)

<sup>g</sup>Jacobson et al. (1989a)

<sup>h</sup>Stiles and Jacobson (1988)

<sup>i</sup>Holschbach et al. (2002)

<sup>j</sup>Jacobson et al. (1992b)

<sup>k</sup>Stiles and Jacobson (1987)

<sup>l</sup>Jacobson (2009)

<sup>m</sup>Jacobson et al. (1987c)

<sup>n</sup>Jacobson et al. (1986b)

<sup>o</sup>Briddon et al. (2004)

<sup>p</sup>Jacobson, K.A., Kirk, K.L., Daly, J.W., Lipkowski, A.W., Rice, K.C., and Jacobson, A.A., unpublished

<sup>q</sup>Soriano et al. (2009)

which the spin label was integrated into the 8-substituent, showed good affinity for A<sub>1</sub> as well as A<sub>2B</sub> ARs (Fig. 7, Table 7).

### 6.3 Specialized Radioligand Probes Based on Conjugation

Trifunctional probes derived from XAC were synthesized for the purpose of cross-linking to both a reporter group and the receptor (Boring et al. 1991). By this means, the xanthine would deliver a radioactive or spectroscopic prosthetic group to the receptor, to which it would react irreversibly by virtue of an electrophilic group such as an isothiocyanate. This approach was illustrated with a series of analogues of *m*-DITC-XAC containing a third substituent in the phenyl isothiocyanate ring. For example, in **166** the third substituent contained a 3-(4-hydroxyphenyl)propionate moiety for radioiodination (Jacobson et al. 1992b). This antagonist derivative effectively radiolabelled the bovine A<sub>1</sub>AR in a covalent manner. Similar trifunctional xanthine probes for covalent labelling of ARs that furthermore contained a cleavable disulfide linkage within the chain linked to the xanthine moiety were reported (Jacobson et al. 1995). The intended strategy was to be able to remove the label after isolation of the modified receptor in order to regenerate the binding ability of the receptor.

### 6.4 Xanthine Radioligand Probes for Positron Emission Tomography

There is a need for the development of imaging agents based on high-affinity ligands for ARs. For example, ligands for *in vivo* PET imaging of A<sub>1</sub>, A<sub>2A</sub>, and A<sub>3</sub> ARs have been developed. The high-affinity A<sub>1</sub> AR antagonist DPCPX gave rise to the high-affinity analogue in which a terminal hydrogen of the 3-propyl group has been substituted with radiofluorine: 8-cyclopentyl-1-propyl-3-(3-[<sup>18</sup>F]fluoropropyl)-xanthine ([<sup>18</sup>F]CPFPX, **159**), similar in structure to DPCPX. This tracer is being developed for PET imaging of the A<sub>1</sub>AR in the brain (Holschbach et al. 2002; Bauer and Ishiwata 2009).

PET ligands for the A<sub>2A</sub> AR in the 8-styrylxanthine series that are structurally related to KW6002 have been developed: for example [7-methyl-<sup>11</sup>C]-(*E*)-8-(3,4,5-trimethoxystyryl)-1,3,7-trimethylxanthine([<sup>11</sup>C]TMSX) (Ishiwata et al. 2000a). This compound was alternatively named [<sup>11</sup>C]KF18446 ([7-methyl-<sup>11</sup>C]-(*E*)-8-(3,4,5-trimethoxystyryl)-1,3,7-trimethylxanthine; Ishiwata et al. 2000b, 2002, 2003a, 2003b). *Ex vivo* autoradiography for this molecule showed a high striatal uptake and a high uptake ratio of the striatum in comparison with other brain regions; [<sup>11</sup>C]KF18446 was therefore proposed as a suitable radioligand for mapping the A<sub>2A</sub>AR of the brain by PET (Mishina et al. 2007). In 2001 the

synthesis and the testing of the 8-styrylxanthine derivative [ $^{11}\text{C}$ ]KW-6002 as a PET ligand was reported. This molecule showed high retention in the striatum but it bound also to extrastriatal regions, so its potential as a PET ligand appeared to require further investigation (Hirani et al. 2001; Brooks et al. 2008).

In an earlier study,  $^{11}\text{C}$ -labelled (*E*)-KF17837 was synthesized and tested, and it was proposed as a potential PET radioligand for mapping the  $A_{2A}$  AR in the heart and the brain (Ishiwata et al. 1996, 1997). Further studies on radiolabelled xanthine derivatives as  $A_{2A}$  AR radioligands were carried out by preparing and testing a  $^{11}\text{C}$ -labelled selective antagonist, (*E*)-8-(3-chlorostyryl)-1,3-dimethyl-7- $^{11}\text{C}$ -methylxanthine [ $^{11}\text{C}$ ](*E*)-8-*m*-chlorostyrylcaffeine). This molecule was shown to accumulate in the striatum, and PET studies on rabbits showed a fast brain uptake of [ $^{11}\text{C}$ ]CSC, reaching a maximum in less than 2 min (Marian et al. 1999). A few years later, iodinated and brominated styrylxanthine derivatives labelled with  $^{11}\text{C}$  were tested as *in vivo* probes (Ishiwata et al. 2000c). [7-Methyl- $^{11}\text{C}$ ]-(*E*)-3,7-dimethyl-8-(3-iodostyryl)-1-propargylxanthine ([ $^{11}\text{C}$ ]IS-DMPX) and [7-methyl- $^{11}\text{C}$ ]-(*E*)-8-(3-bromostyryl)-3,7-dimethyl-1-propargylxanthine ([ $^{11}\text{C}$ ]BS-DMPX) showed  $K_i$  affinities of 8.9 and 7.7 nM respectively, and high  $A_{2A}/A_1$  AR selectivity values. Unfortunately, biological studies proved that the two ligands were only slightly concentrated in the striatum, and that they were not suitable as *in vivo* ligands because of low selectivity for the striatal  $A_{2A}$  ARs and a high non-specific binding (Ishiwata et al. 2000c).

## 6.5 Conjugated Ligand Probes and Bivalent Ligands

Three biotin conjugates (**161–163**) of 1,3-dipropyl-8-phenylxanthine (Fig. 7) were reported as being able to bind competitively to the rat  $A_1$  AR, but in the case of **161** and **162** only in the absence of avidin. This was in contrast to similar conjugates of functionalized nucleoside agonists, which more readily bound simultaneously to both avidin and the  $A_1$ AR. The results were interpreted in terms of the possible reorientation of the ligands at the receptor binding site (Jacobson et al. 1985a; Jacobson 1990).

Two different pharmacophores, one being a xanthine AR antagonist, have been tethered with the intention to create a dual selectivity in a single functional unit. For example, XAC was coupled covalently through an L-Lys linker to a segment derived from the neurotransmitter peptide substance P to form a binary drug (**169**) (Jacobson et al. 1987c). The L-Lys linker served to increase aqueous solubility and to preserve  $A_1$  AR by virtue of a free amino group in the spacer chain. Conjugate **169** bound to the rat  $A_1$  AR with a  $K_i$  value of 35 nM and to the neurokinin type 1 receptor with a  $K_i$  value of 300 nM. Similarly, XAC was coupled to functionalized agonist ligands for opioid receptors, e.g. **175**, and for  $D_2$  dopamine receptors, e.g. **176** (Fig. 7, Table 7) (Jacobson 2009; Soriano et al. 2009). Each of these conjugates bound effectively to both relevant receptors.

## 7 Conclusions

The pharmacological activity of the natural xanthines currently used in therapy, namely theophylline (as an antiasthmatic) and caffeine (as a CNS stimulant, for the treatment of apnoea in newborn babies and as an analgesic in combination therapy e.g. for the treatment of headaches) is mainly mediated by a (non-selective) inhibition of AR subtypes. AR-subtype-selective xanthine derivatives with high potency have been developed and evaluated in animal models and clinical trials.

**Acknowledgements** CEM is grateful for support by BMBF (BioPharma - Neuroallianz), DFG, DAAD, European Commission (ERANET Neuron), and the State of North-Rhine Westfalia (NRW International Research Graduate Schools BIOTECH-PHARMA and Chemical Biology). KAJ acknowledges support from the Intramural Research Program of NIIDK, NIH.

## References

- Abo-Salem OM, Hayallah AM, Bilkei-Gorzo A, Filipek B, Zimmer A, Müller CE (2004) Antinociceptive effects of novel  $A_{2B}$  adenosine receptor antagonists. *J Pharmacol Exp Ther* 308:358–366
- Alexander SP, Cooper J, Shine J, Hill SJ (1996) Characterization of the human brain putative  $A_{2B}$  adenosine receptor expressed in Chinese hamster ovary (CHO.A2B4) cells. *Br J Pharmacol* 119:1286–1290
- Antoniou K, Daifoti-Papadopoulou Z, Hyphantis T, Papathanasiou G, Bekris E, Marselos M, Panlilio L, Müller CE, Goldberg SR, Ferré S (2005) A detailed behavioural analysis of the acute motor effects of caffeine in the rat: involvement of adenosine  $A_1$  and  $A_{2A}$  receptors. *Psychopharmacology* 183:154–162
- Akkari R, Burbiel JC, Hockemeyer J, Müller CE (2006) Recent progress in the development of adenosine receptor ligands as antiinflammatory drugs. *Curr Top Med Chem* 6:1375–1399
- Auchampach JA, Jin X, Wan TC, Caughey GH, Linden J (1997) Canine mast cell adenosine receptors: cloning and expression of the  $A_3$  receptor and evidence that degranulation is mediated by the  $A_{2B}$  receptor. *Mol Pharmacol* 52:846–860
- Auchampach JA, Kreckler LM, Wan TC, Maas JE, van der Hoeven D, Gizewski E, Narayanan J, Maas GE (2009) Characterization of the  $A_{2B}$  adenosine receptor from mouse, rabbit, and dog. *J Pharm Exp Ther* 329:2–13
- Balo MC, Brea J, Caamano O, Fernandez F, Garcia-Mera X, Lopez C, Loza MI, Nieto MI, Rodriguez-Borges JE (2009) Synthesis and pharmacological evaluation of novel 1- and 8-substituted 3-furfurylxanthines as adenosine receptor antagonists. *Bioorg Med Chem* 17:6755–6760
- Baraldi PG, Tabrizi MA, Preti D, Bovero A, Romagnoli R, Fruttarolo F, Zaid NA, Moorman AR, Varani K, Gessi S, Merighi S, Borea PA (2004) Design, synthesis, and biological evaluation of new 8-heterocyclic xanthine derivatives as highly potent and selective human  $A_{2B}$  adenosine receptor antagonists. *J Med Chem* 47:1434–1447
- Baraldi PG, Preti D, Tabrizi MA, Fruttarolo F, Romagnoli R, Zaid NA, Moorman AR, Merighi S, Varani K, Borea PA (2005) New pyrrolo[2,1-f]purine-2, 4-dione and imidazo[2,1-f]purine-2, 4-dione derivatives as potent and selective human  $A_3$  adenosine receptor antagonists. *J Med Chem* 48:4697–4701
- Baraldi PG, Tabrizi MA, Gessi S, Borea PA (2008) Adenosine receptor antagonists: translating medicinal chemistry and pharmacology into clinical utility. *Chem Rev* 108:238–263



- Barone S, Churchill PC, Jacobson KA (1989) Adenosine receptor prodrugs: towards kidney-selective dialkylxanthines. *J Pharm Exp Ther* 250:79–85
- Bauer A, Ishiwata K (2009) Adenosine receptor ligands and PET imaging of the CNS. *Handb Exp Pharmacol* 193:617–642
- Baumgold J, Nikodijevic O, Jacobson KA (1992) Penetration of adenosine antagonists into mouse brain as determined by *ex vivo* binding. *Biochem Pharmacol* 43:889–894
- Bertarelli DCG, Diekmann M, Hayallah AM, Rüsing D, Iqbal J, Preiss B, Verspohl EJ, Müller CE (2006) Characterization of human and rodent native and recombinant adenosine A<sub>2B</sub> receptors by radioligand binding studies. *Purinergic Signal* 2:559–571
- Bilkei-Gorzo A, Abo-Salem OM, Hayallah AM, Michel K, Müller CE, Zimmer A (2008) Adenosine receptor subtype-selective antagonists in inflammation and hyperalgesia. *Naunyn Schmiedebergs Arch Pharmacol* 377:65–76
- Blum D, Galas M-C, Pintor A, Brouillet E, Ledent C, Müller CE, Bantubungi K, Galluzzo M, Gall D, Cuvelier L, Rolland A-S, Popoli P, Schiffmann SN (2003) A dual role of adenosine A<sub>2A</sub> receptors in the modulation of 3-nitropropionic acid-induced striatal lesions: implications for the neuroprotective potential of A<sub>2A</sub> antagonists. *J Neurosci* 23:5361–5369
- Boring DL, Ji XD, Zimmet J, Taylor KE, Stiles GL, Jacobson KA (1991) Trifunctional agents as a design strategy for tailoring ligand properties: Irreversible inhibitors of A<sub>1</sub> adenosine receptors. *Bioconjug Chem* 2:77–88
- Borrmann T, Hinz S, Bertarelli DCG, Li W, Florin NC, Scheiff AB, Müller CE (2009) 1-Alkyl-8-(piperazine-1-sulfonyl)phenylxanthines: development and characterization of adenosine A<sub>2B</sub> receptor antagonists and a new radioligand with subnanomolar affinity and subtype specificity. *J Med Chem* 52:3994–4006
- Brackett LE, Daly JW (1994) Functional characterization of the A<sub>2b</sub> adenosine receptor in NIH 3T3 fibroblasts. *Biochem Pharmacol* 47:801–814
- Bridson SJ, Middleton RJ, Cordeaux Y, Flavin FM, Weinstein JA, George MW, Kellam B, Hill SJ (2004) Quantitative analysis of the formation and diffusion of A<sub>1</sub>-adenosine receptor-antagonist complexes in single living cells. *Proc Natl Acad Sci USA* 101:4673–4678
- Bridson PK, Lin X, Mleman N, Ji XD, Jacobson KA (1998) Synthesis and adenosine receptor affinity of 7-β-D-ribofuranosylxanthine. *Nucleosides Nucleotides* 17:759–768
- Brooks DJ, Doder M, Osman S, Luthra SK, Hirani E, Hume S, Kase H, Kilborn J, Martindill S, Mori A (2008) Positron emission tomography analysis of [<sup>11</sup>C]KW-6002 binding to human and rat adenosine A<sub>2A</sub> receptors in the brain. *Synapse* 62:671–681
- Bruns RF (1981) Adenosine antagonism by purines, pteridines and benzopteridines in human fibroblasts. *Biochem Pharmacol* 30:325–333
- Bruns RF, Daly JW, Snyder SH (1980) Adenosine receptors in brain membranes: binding of N6-cyclohexyl[3H]adenosine and 1,3-diethyl-8-[3H]phenylxanthine. *Proc Natl Acad Sci USA* 77:5547–5551
- Bruns RF, Lu GH, Pugsley TA (1986) Characterization of the A<sub>2</sub> adenosine receptor labeled by [<sup>3</sup>H]NECA in rat striatal membranes. *Mol Pharmacol* 29:331–346
- Bruns RF, Lu GH, Pugsley TA (1987a) In: Gerlach E, Becker BF (eds) *Topics and perspectives in adenosine research*. Springer, New York, pp 59–73
- Bruns RF, Fergus JH, Badger EW, Bristol JA, Santay LA, Hays SJ (1987b) PD 115, 199: an antagonist ligand for adenosine A<sub>2</sub> receptors. *Naunyn Schmiedebergs Arch Pharmacol* 335:64–69
- Bruns RF, Fergus JH (1989) Solubilities of adenosine antagonists determined by radioreceptor assay. *J Pharm Pharmacol* 41:590–594
- Bulicz J, Bertarelli DCG, Baumert D, Fülle F, Müller CE, Heber D (2006) Synthesis and pharmacology of pyrido[2,3-d]pyrimidinediones bearing polar substituents as adenosine receptor antagonists. *Bioorg Med Chem* 14:2837–2849
- Burbiel J, Thorand M, Müller CE (2003) Improved efficient synthesis for multigram-scale production of PSB-10, a potent antagonist at human A<sub>3</sub> adenosine receptors. *Heterocycles* 60:1425–1432

- Cacciari B, Pastorin G, Spalluto G (2003) Medicinal chemistry of A<sub>2A</sub> adenosine receptor antagonists. *Curr Top Med Chem* 3:403–411
- Cagnina RE, Ramos SI, Marshall MA, Wang G, Frazier CR, Linden J (2009) Adenosine A<sub>2B</sub> receptors are highly expressed on murine type II alveolar epithelial cells. *Am J Physiol Lung Cell Mol Physiol* 297:L467–L474
- Carotti A, Cadavid MI, Centeno NB, Esteve C, Loza MI, Martinez A, Nieto RE, Sanz F, Segarra V, Sotelo E, Stefanachi A, Vidal B (2006) Design, synthesis, and structure-activity relationships of 1-, 3-, 8- and 9-substituted 9-deazaxanthines at the human A<sub>2B</sub> adenosine receptor. *J Med Chem* 49:282–299
- Carriba P, Ortiz O, Patkar K, Justinova Z, Stroik J, Themann A, Müller C, Woods AS, Hope BT, Ciruela F, Casado V, Canela EI, Lluís C, Goldberg SR, Moratalla R, Franco R, Ferré S (2007) Striatal adenosine A<sub>2A</sub> and cannabinoid CB<sub>1</sub> receptors form functional heteromeric complexes that mediate the motor effects of cannabinoids. *Neuropsychopharmacology* 32:2249–2259
- Ceccarelli S, Altobelli M, D'Alessandro A, Paesano A (1995) A novel hydrophilic 8-cycloalkylxanthine derivative (IRFI 117) is a highly selective antagonist at A<sub>1</sub> adenosine receptors. *Res Commun Mol Pathol Pharmacol* 87:101–102
- Cirillo R, Barone D, Franzone JS (1988) Doxofylline, an antiasthmatic drug lacking affinity for adenosine receptors. *Arch Int Pharmacodyn Ther* 295:221–237
- Cohen BE, Lee G, Jacobson KA, Kim YC, Huang Z, Sorscher E, Pollard HB (1997) CPX (1,3-dipropyl-8-cyclopentylxanthine) and other alkyl-xanthines differentially bind to wild type and DF508 mutant first nucleotide binding fold (NBF-1) domains of the cystic fibrosis transmembrane conductance regulator. *Biochemistry* 36:6455–6461
- Cristalli G, Cacciari B, Dal Ben D, Lambertucci C, Moro S, Spalluto G, Volpini R (2007) Highlights on the development of A<sub>2A</sub> adenosine receptor agonists and antagonists. *Chem-MedChem* 2:260–281
- Cristalli G, Müller CE, Volpini G (2009) Recent development in adenosine A<sub>2A</sub> receptor ligands. In: Wilson CN, Mustafa SJ (eds) *Handbook of experimental pharmacology 193: adenosine receptors in health and disease*, Springer Verlag, Berlin, pp 59–98
- Cunha GM, Canas PM, Melo CS, Hockemeyer J, Müller CE, Oliveira CR, Cunha RA (2008) Adenosine A<sub>2A</sub> receptor blockade prevents memory dysfunction caused by beta-amyloid peptides but not by scopolamine or MK-801. *Exp Neurol* 210:776–781
- Dall'Igna OP, Fett P, Gomes MW, Souza DO, Cunha RA, Lara DR (2007) Caffeine and adenosine A<sub>2A</sub> receptor antagonists prevent beta-amyloid (25-35)-induced cognitive deficits in mice. *Exp Neurol* 203:241–245
- Daly JW (1982) Adenosine receptors: targets for future drugs. *J Med Chem* 25:197–207
- Daly JW, Padgett W, Shamim MT, Butts-Lamb P, Waters J (1985) 1, 3-Dialkyl-8-(p-sulfophenyl) xanthines: potent water-soluble antagonists for A<sub>1</sub>- and A<sub>2</sub>-adenosine receptors. *J Med Chem* 28:487–492
- Daly JW, Padgett WL, Shamim MT (1986a) Analogues of caffeine and theophylline: effect of structural alterations on affinity at adenosine receptors. *J Med Chem* 29:1305–1308
- Daly JW, Padgett WL, Shamim MT (1986b) Analogues of 1,3-dipropyl-8-phenylxanthine: enhancement of selectivity at A<sub>1</sub>-adenosine receptors by aryl substituents. *J Med Chem* 29:1520–1524
- Daly JW, Hide I, Müller CE, Shamim M (1991) Caffeine analogs: structure-activity relationships at adenosine receptors. *Pharmacology* 42:309–321
- Daly JW (1991) Analogs of caffeine and theophylline: activity as antagonists at adenosine receptors. In: Imai S, Nakazawa M (eds) *Role of adenosine and adenine nucleotides in the biological system*. Amsterdam, Elsevier, pp 119–129
- Daly JW, Jacobson KA (1995) Adenosine and adenine nucleotides: from molecular biology to integrative physiology. Kluwer, Boston, 155
- Daly JW (2000) Alkylxanthines as research tools. *J Auton Nerv Syst* 81:44–52
- Daly JW (2007) Caffeine analogs: biomedical impact. *Cell Mol Life Sci* 64:2153–2169

- Del Giudice MR, Borioni A, Mustazza C, Gatta F, Dionisotti S, Zocchi C, Ongini E (1996) (E)-1-(Heterocyclyl or cyclohexyl)-2-[1,3,7-trisubstituted(xanthin-8-yl)]ethenes as adenosine A<sub>2A</sub> receptors antagonists. *Eur J Med Chem* 31:59–63
- Doggrell SA (2005) BG-9928 (Biogen Idec). *Curr Opin Investig Drugs* 6:962–968
- Drabczynska A, Schumacher B, Müller CE, Karolak-Wojciechowska J, Michalak B, Pekala E, Kiec-Kononowicz K (2003) Impact of the aryl substituent kind and distance from pyrimido [2,1-f]purinediones on the adenosine receptor selectivity and antagonistic properties. *Eur J Med Chem* 38:397–402
- Drabczynska A, Müller CE, Schumacher B, Hinz S, Karolak-Wojciechowska J, Michalak B, Pekala E, Kiec-Kononowicz K (2004) Tricyclic oxazolo[2,3-f]purinediones: potency as adenosine receptor ligands and anticonvulsants. *Bioorg Med Chem* 12:4895–4908
- Drabczynska A, Müller CE, Lacher SK, Schumacher B, Karolak-Wojciechowska J, Nasal A, Kawczak P, Yuzlenko O, Pekala E, Kiec-Kononowicz K (2006) Synthesis and biological activity of tricyclic arylimidazo-, pyrimido-, and diazepinopurinediones. *Bioorg Med Chem* 14:7258–7281
- Drabczynska A, Müller CE, Karolak-Wojciechowska J, Schumacher B, Schiedel A, Yuzlenko O, Kiec-Kononowicz K (2007a) N<sup>9</sup>-Benzyl-substituted 1,3-dimethyl- and 1,3-dipropyl-pyrimido-[2,1-f]purinediones: synthesis and structure-activity relationships at adenosine A<sub>1</sub> and A<sub>2A</sub> receptors. *Bioorg Med Chem* 15:5003–5017
- Drabczynska A, Müller CE, Schiedel A, Schumacher B, Karolak-Wojciechowska J, Fruzinski A, Zobnina W, Yuzlenko O, Kiec-Kononowicz K (2007b) Phenylethyl-substituted pyrimido [2,1-f]purinediones and related compounds: structure-activity relationships as adenosine A<sub>1</sub> and A<sub>2A</sub> receptor ligands. *Bioorg Med Chem* 15:6956–6974
- Elzein E, Kalla RV, Li X, Perry T, Gimbel A, Zeng D, Lustig D, Leung K, Zablocki J (2008) Discovery of a novel A<sub>2B</sub> adenosine receptor antagonist as a clinical candidate for chronic inflammatory airway diseases. *J Med Chem* 51:2267–2278
- Erickson RH, Hiner RN, Feeney SW, Blake PR, Rzeszutarski WJ, Hicks RP, Costello DG, Abreu ME (1991) 1,3,8-Trisubstituted xanthines. Effects of substitution pattern upon adenosine receptor A<sub>1</sub>/A<sub>2</sub> affinity. *J Med Chem* 34:1431–1435
- Esteve C, Nueda JL, Beleta J, Cardenas A, Lozoya E, Cadavid MI, Loza MI, Ryder H, Vidal B (2006) New pyrrolopyrimidin-6-ylbenzenesulfonamides: potent A<sub>2B</sub> adenosine receptor antagonists. *Bioorg Med Chem Lett* 16:3642–3645
- Farrar AM, Pereira M, Velasco F, Hockemeyer J, Müller CE, Salamone J (2007) Adenosine A<sub>2A</sub> receptor antagonism reverses the effects of dopamine receptor antagonism on instrumental output and effort-related choice in the rat: implications for studies of psychomotor slowing. *Psychopharmacology* 191:579–586
- Ferkany JW, Valentine HL, Stone GA, Williams M (1986) Adenosine A<sub>1</sub> receptors in mammalian brain: species differences in their interactions with agonists and antagonists. *Drug Dev Res* 9:85–93
- Fernandez HH, Greeley DR, Zweig RM, Wojcieszek J, Mori A, Sussman NM. 6002-US-051 Study Group (2010) Istradefylline as monotherapy for Parkinson disease: results of the 6002-US-051 trial. *Parkinsonism Relat Disord* 16:16–20
- Ferré S, Popoli P, Giménez-Llort L, Rimondini R, Müller CE, Strömberg I, Ögren SO, Fuxe K (2001) Adenosine/dopamine interaction: implication for the treatment of Parkinson's disease. *Parkinsonism Relat Disord* 7:235–241
- Ferré S, Ciruela F, Borycz J, Solinas M, Quarta D, Antoniou K, Quiroz C, Justinova Z, Lluís C, Franco R, Goldberg SR (2008) Adenosine A<sub>1</sub>-A<sub>2A</sub> receptor heteromers: new targets for caffeine in the brain. *Front Biosci* 13:2391–2399
- Fhidi O, Pawlowski M, Jurczyk S, Müller CE, Schumacher B (2003) Pyridin-8-on[2,1-f]theophylline-9-alkylcarboxylic acid amides as A<sub>1</sub> and A<sub>2A</sub> adenosine receptor ligands. *Farmacologia* 58:439–444
- Filip M, Frankowska M, Zaniewska M, Przegalinski E, Müller CE, Agnati LF, Franco R, Roberts DCS, Fuxe K (2006) Involvement of adenosine A<sub>2A</sub> and dopamine receptors in the locomotor and sensitizing effects of cocaine. *Brain Res* 1077:67–80

- Fozard JR, Baur F, Wolber C (2003) Antagonist pharmacology of adenosine A<sub>2B</sub> receptors from rat, guinea pig and dog. *Eur J Pharmacol* 475:79–84
- Franchetti P, Messini L, Cappellacci L, Grifantini M, Lucacchini A, Martini C, Senatore G (1994) 8-Azaxanthine derivatives as antagonists of adenosine receptors. *J Med Chem* 37:2970–2975
- Frédéric R, Ooms F, Castagnoli N Jr, Petzer JP, Feng JF, Schwarzschild MA, Van der Schyf CJ, Wouters J (2005) (E)-8-(3-Chlorostyryl)-1,3,7-trimethylxanthine, a caffeine derivative acting both as antagonist of adenosine A<sub>2A</sub> receptors and as inhibitor of MAO-B. *Acta Crystallogr C* 61:o531–o532
- Fredholm BB, Abbracchio MP, Burnstock G, Daly JW, Harden KT, Jacobson KA, Leff P, Williams M (1994) Nomenclature and classification of purinoceptors: a report from the IUPHAR subcommittee. *Pharmacol Rev* 46:143–156
- Fredholm BB, Bättig K, Holmén J, Nehlig A, Zvartau EE (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fredholm BB, Jacobson KA (2009) John W. Daly and the early characterization of adenosine receptors. *Heterocycles* 79:73–83
- Fuxe K, Marcellino D, Genedani S, Agnati L (2007) Adenosine A<sub>2A</sub> receptors, dopamine D<sub>2</sub> receptors and their interactions in Parkinson's disease. *Mov Disord* 22:1990–2017
- Gao ZG, Kim SK, Biadatti T, Chen W, Lee K, Barak D, Kim SG, Johnson CR, Jacobson KA (2002) Structural determinants of A<sub>3</sub> adenosine receptor activation: nucleoside ligands at the agonist/antagonist boundary. *J Med Chem* 45:4471–4484
- Geis U, Grahner B, Pawlowski M, Drabczynska A, Gorczyca M, Müller CE (1995) Tricyclic theophylline derivatives with high water-solubility: structure-activity relationships at adenosine receptors, phosphodiesterases and benzodiazepine binding sites. *Pharmazie* 50:333–336
- Givertz MM (2009) Adenosine A<sub>1</sub> receptor antagonists at a fork in the road. *Circ Heart Fail* 2:519–522
- Grahner B, Winiwarter S, Lanzner W, Müller CE (1994) Synthesis and structure-activity relationships of deazaxanthines: analogs of potent A<sub>1</sub>- and A<sub>2</sub>-adenosine receptor antagonists. *J Med Chem* 37:1526–1534
- Hayallah AM, Sandoval-Ramírez J, Reith U, Schobert U, Preiss B, Schumacher B, Daly JW, Müller CE (2002) 1, 8-Disubstituted xanthine derivatives: synthesis of potent A<sub>2B</sub>-selective adenosine receptor antagonists. *J Med Chem* 45:1500–1510
- Hauber W, Nagel J, Sauer R, Müller CE (1998) Motor effects induced by a blockade of adenosine A<sub>2A</sub> receptors in the caudate-putamen. *Neuroreport* 9:1803–1806
- Hauber W, Neuscheler P, Nagel J, Müller CE (2001) Catalepsy induced by a blockade of dopamine D<sub>1</sub> or D<sub>2</sub> receptors was reversed by a concomitant blockade of A<sub>2A</sub> receptors in the caudate-putamen of rats. *Eur J Neurosci* 14:1287–1293
- Hirani E, Gillies J, Karasawa A, Shimada J, Kase H, Opacka-Juffry J, Osman S, Luthra SK, Hume SP, Brooks DJ (2001) Evaluation of [4-O-methyl-<sup>11</sup>C]KW-6002 as a potential PET ligand for mapping central adenosine A<sub>2A</sub> receptors in rats. *Synapse* 42:164–176
- Hockemeyer J, Burbiel JC, Müller CE (2004) Multigram-scale syntheses, stability, and photo-reactions of A<sub>2A</sub> adenosine receptor antagonists with 8-styrylxanthine structure: potential drugs for Parkinson's disease. *J Org Chem* 69:3308–3318
- Holschbach MH, Olsson RA, Bier D, Wutz W, Sihver W, Schüller M, Palm B, Coenen HH (2002) Synthesis and evaluation of no-carrier-added 8-cyclopentyl-3-(3-[<sup>18</sup>F]fluoropropyl)-1-propyl-xanthine ([<sup>18</sup>F]CPFPX): a potent and selective A<sub>1</sub>-adenosine receptor antagonist for in vivo imaging. *J Med Chem* 45:5150–5156
- Ilas J, Pekar S, Hockemeyer J, Euler H, Kirfel A, Müller CE (2005) Development of spin-labeled probes for adenosine receptors. *J Med Chem* 48:2108–2114
- Ishiwari K, Madson LJ, Farrar AM, Mingote SM, Valenta JP, DiGianvittorio MD, Frank LE, Correa M, Hockemeyer J, Müller CE, Salamone JD (2007) Injections of the selective

- adenosine A<sub>2A</sub> antagonist MSX-3 into the nucleus accumbens core attenuate the locomotor suppression induced by haloperidol in rats. *Behav Brain Res* 178:190–199
- Ishiwata K, Noguchi J, Toyama H, Sakiyama Y, Koike N, Ishii S, Oda K, Endo K, Suzuki F, Senda M (1996) Synthesis and preliminary evaluation of [<sup>11</sup>C]KF17837, a selective adenosine A<sub>2A</sub> antagonist. *Appl Radiat Isot* 47:507–511
- Ishiwata K, Sakiyama Y, Sakiyama T, Shimada J, Toyama H, Oda K, Suzuki F, Senda M (1997) Myocardial adenosine A<sub>2A</sub> receptor imaging of rabbit by PET with [<sup>11</sup>C]KF17837. *Ann Nucl Med* 11:219–225
- Ishiwata K, Noguchi J, Wakabayashi S, Shimada J, Ogi N, Nariai T, Tanaka A, Endo K, Suzuki F, Senda M (2000a) <sup>11</sup>C-labeled KF18446: a potential central nervous system adenosine A<sub>2A</sub> receptor ligand. *J Nucl Med* 41:345–354
- Ishiwata K, Ogi N, Shimada J, Nonaka H, Tanaka A, Suzuki F, Senda M (2000b) Further characterization of a CNS adenosine A<sub>2A</sub> receptor ligand [<sup>11</sup>C]KF18446 with in vitro autoradiography and in vivo tissue uptake. *Ann Nucl Med* 14:81–89
- Ishiwata K, Shimada J, Wang WF, Harakawa H, Ishii S, Kiyosawa M, Suzuki F, Senda M (2000c) Evaluation of iodinated and brominated [<sup>11</sup>C]styrylxanthine derivatives as in vivo radioligands mapping adenosine A<sub>2A</sub> receptor in the central nervous system. *Ann Nucl Med* 14:247–253
- Ishiwata K, Ogi N, Hayakawa N, Oda K, Nagaoka T, Toyama H, Suzuki F, Endo K, Tanaka A, Senda M (2002) Adenosine A<sub>2A</sub> receptor imaging with [<sup>11</sup>C]KF18446 PET in the rat brain after quinolinic acid lesion: comparison with the dopamine receptor imaging. *Ann Nucl Med* 16:467–475
- Ishiwata K, Kawamura K, Kimura Y, Oda K, Ishii K (2003a) Potential of an adenosine A<sub>2A</sub> receptor antagonist [<sup>11</sup>C]TMSX for myocardial imaging by positron emission tomography: a first human study. *Ann Nucl Med* 17:457–462
- Ishiwata K, Wang WF, Kimura Y, Kawamura K, Ishii K (2003b) Preclinical studies on [<sup>11</sup>C]TMSX for mapping adenosine A<sub>2A</sub> receptors by positron emission tomography. *Ann Nucl Med* 17:205–211
- Ishiyama H, Nakajima H, Nakata H, Kobayashi J (2009) Synthesis of hybrid analogues of caffeine and eudistomin D and their affinity for adenosine receptors. *Bioorg Med Chem* 17:4280–4284
- Jacobson KA, Kirk KL, Padgett W, Daly JW (1985a) Probing the adenosine receptor with adenosine and xanthine biotin conjugates. *FEBS Lett* 184:30–35
- Jacobson KA, Kirk KL, Padgett WL, Daly JW (1985b) Functionalized congeners of 1,3-dialkylxanthines: preparation of analogues with high affinity for adenosine receptors. *J Med Chem* 28:1334–1340
- Jacobson KA, Ukena D, Kirk KL, Daly JW (1986a) [3H]Xanthine amine congener of 1,3-dipropyl-8-phenylxanthine: an antagonist radioligand for adenosine receptors. *Proc Natl Acad Sci USA* 83:4089–4093
- Jacobson KA, Kirk KL, Padgett WL, Daly JW (1986b) A functionalized congener approach to adenosine receptor antagonists: amino acid conjugates of 1,3-dipropylxanthine. *Mol Pharmacol* 29:126–133
- Jacobson KA, Ukena D, Padgett W, Daly JW, Kirk KL (1987a) Xanthine functionalized congeners as potent ligands at A<sub>2</sub>-adenosine receptors. *J Med Chem* 30:211–214
- Jacobson KA, Ukena D, Padgett W, Kirk KL, Daly JW (1987b) Molecular probes for extracellular adenosine receptors. *Biochem Pharmacol* 36:1697–1707
- Jacobson KA, Lipowski AW, Moody TW, Padgett W, Pijl E, Kirk KL, Daly JW (1987c) Binary drugs: conjugates of purines and a peptide that bind to both adenosine and substance P receptors. *J Med Chem* 30:1529–1532
- Jacobson KA, de la Cruz R, Schulick R, Kiriasis L, Padgett W, Pfeleiderer W, Kirk KL, Neumeyer JL, Daly JW (1988) 8-Substituted xanthines as antagonists at A<sub>1</sub> and A<sub>2</sub>-adenosine receptors. *Biochem Pharmacol* 37:3653–3661
- Jacobson KA, Barone S, Kammula U, Stiles GL (1989a) Electrophilic derivatives of purines as irreversible inhibitors of A<sub>1</sub>-adenosine receptors. *J Med Chem* 32:1043–1051

- Jacobson KA, Kiriasis L, Barone S, Bradbury BJ, Kammula U, Campagne JM, Daly JW, Neumeyer JL, Pfeleiderer W (1989b) Sulfur-containing xanthine derivatives as selective antagonists at A<sub>1</sub>-adenosine receptors. *J Med Chem* 32:1873–1879
- Jacobson KA (1990) Probing adenosine receptors using biotinylated purine conjugates. *Methods Enzymol* 184:668–671
- Jacobson KA, van Galen PJM, Williams M (1992a) Perspective, adenosine receptors: pharmacology, structure activity relationships and therapeutic potential. *J Med Chem* 35:407–422
- Jacobson KA, Olah ME, Stiles GL (1992b) Trifunctional ligands: a radioiodinated high affinity acylating antagonist for the A<sub>1</sub> adenosine receptor. *Pharmacol Commun* 1:145–154
- Jacobson KA, Gallo-Rodriguez C, Melman N, Fischer B, Maillard M, van Bergen A, van Galen PJ, Karton Y (1993a) Structure-activity relationships of 8-styrylxanthines as A<sub>2</sub>-selective adenosine antagonists. *J Med Chem* 36:1333–1342
- Jacobson KA, Shi D, Gallo-Rodriguez C, Manning M Jr, Müller C, Daly JW, Neumeyer JL, Kiriasis L, Pfeleiderer W (1993b) Effect of trifluoromethyl and other substituents on activity of xanthines at adenosine receptors. *J Med Chem* 36:2639–2644
- Jacobson KA, Fischer B, Ji XD (1995) A “cleavable trifunctional” approach to receptor affinity labeling: regeneration of binding to A<sub>1</sub>-adenosine receptors. *Bioconjug Chem* 6:255–263
- Jacobson KA (1998) A<sub>3</sub> adenosine receptors: novel ligands and paradoxical effects. *Trends Pharmacol Sci* 19:184–191
- Jacobson KA, IJzerman AP, Linden J (1999) 1,3-Dialkylxanthine derivatives having high potency as antagonists at human A<sub>2B</sub> adenosine receptors. *Drug Devel Res* 47:45–53
- Jacobson KA, Gao ZG (2006) Adenosine receptors as therapeutic targets. *Nat Rev Drug Discov* 5:247–264
- Jacobson KA (2009) Functionalized congener approach to the design of ligands for G protein-coupled receptors (GPCRs). *Bioconjug Chem* 20:1816–1835
- Jarvis MF, Jacobson KA, Williams M (1987) Autoradiographic localization of adenosine A<sub>1</sub> receptors in rat brain using [3H]XCC, a functionalized congener of 1,3-dipropylxanthine. *Neurosci Lett* 81:69–74
- Ji XD, Stiles GL, Jacobson KA (1991) [<sup>3</sup>H]XAC (xanthine amine congener) is a radioligand for A<sub>2</sub>-adenosine receptors in rabbit striatum. *Neurochem Int* 18:207–213
- Ji XD, Gallo-Rodriguez C, Jacobson KA (1993) 8-(3-Isothiocyanatostyryl)caffeine is a selective irreversible inhibitor of striatal A<sub>2</sub>-adenosine receptors. *Drug Dev Res* 29:292–298
- Ji XD, von Lubitz D, Olah ME, Stiles GL, Jacobson KA (1994) Species differences in ligand affinity at central A<sub>3</sub>-adenosine receptors. *Drug Dev Res* 33:51–59
- Ji XD, Kim YC, Ahern DG, Linden J, Jacobson KA (2001) [<sup>3</sup>H]MRS 1754, a selective antagonist radioligand for A<sub>2B</sub> adenosine receptors. *Biochem Pharmacol* 61:657–663
- Kalla R, Elzein E, Perry T, Li X, Gimbel A, Yang M, Zeng D, Zablocki J (2008) Selective, high affinity A<sub>2B</sub> adenosine receptor antagonists: N-1 monosubstituted 8-(pyrazol-4-yl)xanthines. *Bioorg Med Chem Lett* 18:1397–1401
- Kalla R, Zablocki J (2009) Progress in the discovery of selective, high affinity A<sub>2B</sub> adenosine receptor antagonists as clinical candidates. *Purinergic Signal* 5:21–29
- Karcz-Kubicha M, Quarta D, Hope BT, Antoniou K, Müller CE, Morales M, Schindler CW, Goldberg SR, Ferré S (2003a) Enabling role of adenosine A<sub>1</sub> receptors in adenosine A<sub>2A</sub> receptor-mediated striatal expression of c-fos. *Eur J Neurosci* 18:296–302
- Karcz-Kubicha M, Antoniou K, Terasmaa A, Quarta D, Solinas M, Justinova Z, Pezzola A, Reggioro R, Müller CE, Fuxe K, Goldberg SR, Popoli P, Ferré S (2003b) Involvement of adenosine A<sub>1</sub> and A<sub>2A</sub> receptors in the motor effects of caffeine after its acute and chronic administration. *Neuropsychopharmacology* 28:1281–1291
- Kase H (2003) The adenosine A<sub>2A</sub> receptor selective antagonist KW6002: research toward a novel nondopaminergic therapy for Parkinson’s disease. *Neurology* 61(Suppl 6):S97–S100
- Kiec-Kononowicz K, Drabczynska A, Pekala E, Michalak B, Müller CE, Schumacher B, Karolak-Wojciechowska J, Duddeck H, Rockitt S, Wartchow R (2001) New developments in A<sub>1</sub> and A<sub>2</sub> adenosine receptor antagonists. *Pure Appl Chem* 73:1411–1420

- Kiesman WF, Zhao J, Conlon PR, Petter RC, Jin X, Smits G, Lutterodt F, Sullivan GW, Linden J (2006a) Norbornyllactone-substituted xanthines as adenosine A<sub>1</sub> receptor antagonists. *Bioorg Med Chem* 14:3654–3661
- Kiesman WF, Zhao J, Conlon PR, Dowling JE, Petter RC, Lutterodt F, Jin X, Smits G, Fure M, Jayaraj A, Kim J, Sullivan G, Linden J (2006b) Potent and orally bioavailable 8-bicyclo[2.2.2]octylxanthines as adenosine A<sub>1</sub> receptor antagonists. *J Med Chem* 49: 7119–7131
- Kim HO, Ji XD, Melman N, Olah ME, Stiles GL, Jacobson KA (1994a) Structure activity relationships of 1,3-dialkylxanthine derivatives at rat A<sub>3</sub>-adenosine receptors. *J Med Chem* 37:3373–3382
- Kim HO, Ji XD, Melman N, Olah ME, Stiles GL, Jacobson KA (1994b) Selective ligands for rat A<sub>3</sub>-adenosine receptors: structure-activity relationships of 1,3-dialkylxanthine-7-riboside derivatives. *J Med Chem* 37:4020–4030
- Kim YC, Karton Y, Ji XD, Melman N, Linden J, Jacobson KA (1999) Acyl-hydrazide derivatives of a xanthine carboxylic congener (XCC) as selective antagonists at human A<sub>2B</sub> adenosine receptors. *Drug Dev Res* 47:178–188
- Kim Y-S, Ji X, Melman N, Linden J, Jacobson KA (2000) Anilide derivatives of an 8-phenyl-xanthine carboxylic congener are highly potent and selective antagonists at human A<sub>2B</sub> adenosine receptors. *J Med Chem* 43:1165–1172
- Kim S-A, Marschall MA, Melman N, Kim HS, Müller CE, Linden J, Jacobson KA (2002) Structure-activity relationships at human and rat A<sub>2B</sub> adenosine receptors of xanthine derivatives substituted at the 1-, 3-, 7-, and 8-positions. *J Med Chem* 45:2131–2138
- Kirfel A, Schwabenländer F, Müller CE (1997) Crystal structure of 1-propyl-8-(4-sulfophenyl)-7H-imidazo[4,5-d]pyrimidin-2,6(1H,3H)-dione dehydrate, C<sub>14</sub>H<sub>14</sub>N<sub>4</sub>O<sub>5</sub>S×2 H<sub>2</sub>O. *Z Kristallogr New Cryst Struct* 3:447–448
- Klotz KN, Vogt H, Tawfik-Schlieper H (1991) Comparison of adenosine receptors in brain from different species by radioligand binding and photoaffinity labelling. *Naunyn Schmiedebergs Arch Pharmacol* 343:196–201
- Klotz KN, Hessling J, Hegler J, Owman C, Kull B, Fredholm BB, Lohse MJ (1998) Comparative pharmacology of human adenosine receptor subtypes - characterization of stably transfected receptors in CHO cells. *Naunyn Schmiedebergs Arch Pharmacol* 357:1–9
- Knutsen LJ, Weiss SM (2001) KW-6002 (Kyowa Hakko Kogyo). *Curr Opin Investig Drugs* 2:668–673
- Krämer SD, Testa B (2008) The biochemistry of drug metabolism – an introduction part 6. Inter-individual factors affecting drug metabolism. *Chem Biodivers* 5:2465–2578
- Kull B, Arslan G, Nilsson C, Owman C, Lorenzen A, Schwabe U, Fredholm BB (1999) Differences in the order of potency for agonists but not antagonists at human and rat adenosine A<sub>2A</sub> receptors. *Biochem Pharmacol* 57:65–75
- Linden J (1994) Cloned adenosine A<sub>3</sub> receptors: pharmacological properties, species differences and receptor functions. *Trends Pharmacol Sci* 15:298–306
- Linden J, Taylor HE, Robeva AS, Tucker AL, Stehle JH, Rivkees SA, Fink JS, Reppert SM (1993) Molecular cloning and functional expression of a sheep A<sub>3</sub> adenosine receptor with widespread tissue distribution. *Mol Pharmacol* 44:524–532
- Linden J, Thai T, Figler H, Jin X, Robeva AS (1999) Characterization of human A<sub>2B</sub> adenosine receptors: radioligand binding, western blotting, and coupling to G<sub>q</sub> in human embryonic kidney 293 cells and HMC-1 mast cells. *Mol Pharmacol* 56:705–713
- Marcellino D, Carriba P, Filip M, Borgkvist A, Frankowska M, Bellido I, Tanganelli S, Müller CE, Fisone G, Lluis C, Agnati LF, Franco R, Fuxe K (2008) Antagonistic cannabinoid CB<sub>1</sub>/dopamine D<sub>2</sub> receptor interactions in striatal CB<sub>1</sub>/D<sub>2</sub> heteromers. A combined neurochemical and behavioural analysis. *Neuropharmacology* 54:815–823
- Marian T, Boros I, Lengyel Z, Balkay L, Horvath G, Emri M, Sarkadi E, Szentmiklosi AJ, Fekete I, Tron L (1999) Preparation and primary evaluation of [<sup>11</sup>C]CSC as a possible tracer for mapping adenosine A<sub>2A</sub> receptors by PET. *Appl Radiat Isot* 50:887–893

- Martin PL, Wysocki RJ Jr, Barrett RJ, May JM, Linden J (1996) Characterization of 8-(N-methylisopropyl)amino-N<sup>6</sup>-(5'-endohydroxy-endonorbornyl)-9-methyladenine (WRC-0571), a highly potent and selective, non-xanthine antagonist of A<sub>1</sub> adenosine receptors. *J Pharmacol Exp Ther* 276:490–499
- Massip S, Guillon J, Bertarelli D, Bosc JJ, Leger JM, Lacher S, Bontemps C, Dupont T, Müller CE, Jarry C (2006) Synthesis and preliminary evaluation of new 1- and 3-[1-(2-hydroxy-3-phenoxypropyl)]xanthines from 2-amino-2-oxazolines as potential A<sub>1</sub> and A<sub>2A</sub> adenosine receptor antagonists. *Bioorg Med Chem* 14:2697–2719
- Michael S, Warstat C, Michel F, Yan L, Müller CE, Nieber K (2010) Adenosine A<sub>2A</sub> agonist and A<sub>2B</sub> antagonist mediate an inhibition of inflammation-induced contractile disturbance of a rat gastrointestinal preparation. *Purinergic Signal* 6:117–124
- Mishina M, Ishiwata K, Kimura Y, Naganawa M, Oda K, Kobayashi S, Katayama Y, Ishii K (2007) Evaluation of distribution of adenosine A<sub>2A</sub> receptors in normal human brain measured with [<sup>11</sup>C]TMSX PET. *Synapse* 61:778–784
- Moro S, Gao ZG, Jacobson KA, Spalluto G (2006) Progress in the pursuit of therapeutic adenosine receptor antagonists. *Med Res Rev* 26:131–159
- Mott AM, Nunes EJ, Collins LE, Port RG, Sink KS, Hockemeyer J, Müller CE, Salamone JD (2009) The adenosine A<sub>2A</sub> antagonist MSX-3 reverses the effects of the dopamine antagonist haloperidol on effort-related decision making in a T-maze cost/benefit procedure. *Psychopharmacology* 204:103–112
- Müller CE (1994) Formation of oxazolo[3,2-a]purinones from propynyluracils. *J Org Chem* 59:1928–1929
- Müller CE (1997) A<sub>1</sub>-adenosine receptor antagonists. *Expert Opin Ther Patents* 7:419–440
- Müller CE (2000) A<sub>2A</sub> adenosine receptor antagonists - future drugs for Parkinson's disease? *Drugs Future* 25:1043–1052
- Müller CE (2001) A<sub>3</sub> adenosine receptor antagonists. *Mini-Rev Med Chem* 1:417–427
- Müller CE (2003) Medicinal chemistry of adenosine A<sub>3</sub> receptor ligands. *Curr Top Med Chem* 3:445–462
- Müller CE (2009) Prodrug approaches for enhancing the bioavailability of drugs with low solubility. *Chem Biodivers* 6:2071–2083
- Müller CE, Scior T (1993) Adenosine receptors and their modulators. *Pharm Acta Helv* 68:77–111
- Müller CE, Shi D, Manning M Jr, Daly JW (1993) Synthesis of paraxanthine analogs (1,7-disubstituted xanthines) and other xanthines unsubstituted at the 3-position: structure-activity relationships at adenosine receptors. *J Med Chem* 36:3341–3349
- Müller CE, Stein B (1996) Adenosine receptor antagonists: structures and potential therapeutic applications. *Curr Pharm Des* 2:501–530
- Müller CE, Geis U, Hipp J, Schobert U, Frobenius W, Pawlowski M, Suzuki F, Sandoval-Ramirez J (1997a) Synthesis and structure-activity relationships of DMPX (3,7-dimethyl-1-propargyl-xanthine) derivatives, A<sub>2A</sub>-selective adenosine receptor antagonists. *J Med Chem* 40:4396–4405
- Müller CE, Sauer R, Geis U, Frobenius W, Talik P, Pawlowski M (1997b) Aza-analogs of 8-styrylxanthines as A<sub>2A</sub>-adenosine receptor antagonists. *Arch Pharm Pharm Med Chem* 330:181–189
- Müller CE, Schobert U, Hipp J, Geis U, Frobenius W, Pawlowski M (1997c) Configurationally stable analogs of styrylxanthines as A<sub>2A</sub> adenosine receptor antagonist. *Eur J Med Chem* 32:709–719
- Müller CE, Sandoval-Ramirez J, Schobert U, Geis U, Frobenius W, Klotz KN (1998) 8-(Sulfostyryl)xanthines: water-soluble A<sub>2A</sub>-selective adenosine receptor antagonists. *Bioorg Med Chem* 6:707–719
- Müller CE, Maurinsh J, Sauer R (2000) Binding of [<sup>3</sup>H]MSX-2 (3-(3-hydroxypropyl)-7-methyl-8-(*m*-methoxystyryl)-1-propargylxanthine) to rat striatal membranes - a new, selective antagonist radioligand for A<sub>2A</sub> adenosine receptors. *Eur J Pharm Sci* 10:259–265
- Müller CE, Thorand M, Qurishi R, Diekmann M, Jacobson KA, Padgett WL, Daly JW (2002a) Imidazo[2,1-*i*]purin-5-ones and related tricyclic water-soluble purine derivatives: potent A<sub>2A</sub>- and A<sub>3</sub>-adenosine receptor antagonists. *J Med Chem* 45:3440–3450



- Müller CE, Diekmann M, Thorand M, Ozola V (2002b) [3H]8-Ethyl-4-methyl-2-phenyl-(8R)-4,5,7,8-tetrahydro-1H-imidazo[2,1-i]purin-5-one ([3H]PSB-11), a novel high-affinity antagonist radioligand for human A<sub>3</sub> adenosine receptors. *Bioorg Med Chem Lett* 12:501–503
- Müller CE, Ferré S (2007) Blocking striatal adenosine A<sub>2A</sub> receptors: a new strategy for basal ganglia disorders. *Recent Patents CNS Drug Discov* 2:1–21
- Müller CE, Hockemeyer J, Tzvetkov NT, Burbiel JC (2008) Preparation of 8-ethynyl-xanthine derivatives as selective A<sub>2A</sub> receptor antagonists (SANOL Arznei Schwarz GmbH, Germany). *PCT Int Appl; WO 2008077557 A1*
- Nagel J, Schladebach H, Koch M, Schwienbacher I, Müller CE, Hauber W (2003) Effects of an adenosine A<sub>2A</sub> receptor blockade in the nucleus accumbens on locomotion, feeding, and prepulse inhibition in rats. *Synapse* 49:279–286
- Nieto MI, Balo MC, Brea J, Caamano O, Cadavid MI, Fernandez F, Mera XG, Lopez C, Rodriguez-Borges JE (2009) Synthesis of novel 1-alkyl-8-substituted 3-(3-methoxypropyl) xanthines as putative A<sub>2B</sub> receptor antagonists. *Bioorg Med Chem* 17:3426–3432
- Noguchi J, Ishiwata K, Furuat R, J-i S, Kiyosawa M, Ishii S-i, Endo K, Suzuki F, Senda M (1997) Evaluation of carbon-11 labeled KF15372 and its ethyl and methyl derivatives as a potential CNS adenosine A<sub>2</sub> receptor ligand. *Nucl Med Biol* 24:53–59
- Nonaka Y, Shimada J, Nonaka H, Koike N, Aoki N, Kobayashi H, Kase H, Yamaguchi K, Suzuki F (1993) Photoisomerization of a potent and selective adenosine A<sub>2</sub> antagonist, (E)-1,3-dipropyl-8-(3, 4-dimethoxystyryl)-7-methylxanthine. *J Med Chem* 36:3731–3733
- Nonaka H, Ichimura M, Takeda M, Nonaka Y, Shimada J, Suzuki F, Yamaguchi K, Kase H (1994a) KF17837 ((E)-8-(3, 4-dimethoxystyryl)-1,3-dipropyl-7-methylxanthine), a potent and selective adenosine A<sub>2</sub> receptor antagonist. *Eur J Pharmacol* 267:335–341
- Nonaka H, Mori A, Ichimura M, Shindou T, Yanagawa K, Shimada J, Kase H (1994b) Binding of [<sup>3</sup>H]KF17837S, a selective adenosine A<sub>2</sub> receptor antagonist, to rat brain membranes. *Mol Pharmacol* 46:817–822
- Obiefuna PC, Batra VK, Nadeem A, Borron A, Wilson CN, Mustafa SJ (2005) A novel A<sub>1</sub> adenosine receptor antagonist, L-97-1 [3-[2-(4-aminophenyl)-ethyl]-8-benzyl-7-{2-ethyl-(2-hydroxy-ethyl)-amino}-ethyl]-1-propyl-3,7-dihydro-purine-2,6-dione], reduces allergic responses to house dust mite in an allergic rabbit model of asthma. *J Pharmacol Exp Ther* 315:329–336
- Olah ME, Jacobson KA, Stiles GL (1989) Affinity chromatography of the bovine cerebral cortex A<sub>1</sub> adenosine receptor. *FEBS Lett* 257:292–296
- Ozola V, Thorand M, Diekmann M, Qurishi R, Schumacher B, Jacobson KA, Müller CE (2003) 2-Phenylimidazo[2,1-i]purin-5-ones: structure-activity relationships and characterization of potent and selective inverse agonists at human A<sub>3</sub> adenosine receptors. *Bioorg Med Chem* 11:347–356
- Park KS, Hoffmann C, Kim HO, Padgett WL, Daly JW, Brambilla R, Motta C, Abbracchio MP, Jacobson KA (1998) Activation and desensitization of rat A<sub>3</sub>-adenosine receptors by selective adenosine derivatives and xanthine-7-ribosides. *Drug Dev Res* 44:97–105
- Pastorin G, Bolcato C, Cacciari B, Kachler S, Klotz K-N, Montopoli C, Moro S, Spalluto G (2005) Synthesis, biological and modelling studies of 1,3-di-n-propyl-2, 4-dioxo-6-methyl-8-(substituted) 1,2,3,4-tetrahydro[1,2,4]triazolo[3,4-f]purines as adenosine receptor antagonists. *Farmacologia* 60:643–651
- Patel A, Craig RH, Daluge SM, Linden J (1988) 125I-BW-A844U, an antagonist radioligand with high affinity and selectivity for adenosine A<sub>1</sub> receptors, and <sup>125</sup>I-azido-BW-A844U, a photo-affinity label. *Mol Pharmacol* 33:585–591
- Peet NP, Lentz NL, Dudley MW, Ogden AM, McCarty DR, Racke MM (1993) Xanthines with C8 chiral substituents as potent and selective adenosine A<sub>1</sub> antagonists. *J Med Chem* 36:4015–4020
- Petzer JP, Steyn S, Castagnoli KP, Chen JF, Schwarzschild MA, Van der Schyf CJ, Castagnoli N (2003) Inhibition of monoamine oxidase B by selective adenosine A<sub>2A</sub> receptor antagonists. *Bioorg Med Chem* 11:1299–1310

- Petzer JP, Castagnoli N Jr, Schwarzschild MA, Chen J-F, Van der Schyf CJ (2009) Dual-target-directed drugs that block monoamine oxidase B and adenosine A<sub>2A</sub> receptors for Parkinson's disease. *Neurotherapeutics* 6:141–151
- Pfister JR, Belardinelli L, Lee G, Lum RT, Milner P, Stanley WC, Linden J, Baker SP, Schreiner G (1997) Synthesis and biological evaluation of the enantiomers of the potent and selective A<sub>1</sub>-adenosine antagonist 1,3-dipropyl-8-[2-(5,6-epoxynorbornyl)]xanthine. *J Med Chem* 40:1773–1778
- Pretorius J, Malan SF, Castagnoli N Jr, Bergh JJ, Petzer JP (2008) Dual inhibition of monoamine oxidase B and antagonism of the adenosine A<sub>2A</sub> receptor by (E,E)-8-(4-phenylbutadien-1-yl) caffeine analogues. *Bioorg Med Chem* 16:8676–8684
- Priego E-M, von Frijtag Drabbe Kuenzel KJ, IJzerman AP, Camarasa M-J, Pérez-Pérez M-J (2002) Pyrido[2,1-f]purine-2,4-dione derivatives as a novel class of highly potent human A<sub>3</sub> adenosine receptor antagonists. *J Med Chem* 45:3337–3344
- Priego E-M, Pérez-Pérez M-J, von Frijtag Drabbe Kuenzel JK, de Vries H, IJzerman AP, Camarasa M-J, Martín-Santamaría S (2008) Selective human adenosine A<sub>3</sub> antagonists based on pyrido [2,1-f]purine-2,4-diones: novel features of hA<sub>3</sub> antagonist binding. *ChemMedChem* 3:111–119
- Richardson PJ, Kase H, Jenner PG (1997) Adenosine A<sub>2A</sub> receptor antagonists as new agents for the treatment of Parkinson's disease. *Trends Pharmacol Sci* 18:338–344
- Robeva AS, Woodard RL, Jin X, Gao Z, Bhattacharya S, Taylor HE, Rosin DL, Linden J (1996) Molecular characterization of recombinant human adenosine receptors. *Drug Dev Res* 39:243–252
- Saki M, Tsumuki H, Nonaka H, Shimada J, Ichimura M (2002) KF26777 (2-(4-bromophenyl)-7,8-dihydro-4-propyl-1H-imidazo[2,1-i]purin-5(4H)-one dihydrochloride), a new potent and selective adenosine A<sub>3</sub> receptor antagonist. *Eur J Pharmacol* 444:133–141
- Salamone JD, Betz AJ, Ishiwari K, Felsted J, Madson L, Mirante B, Clark K, Font L, Korbey S, Sager TN, Hockemeyer J, Müller CE (2008a) Tremorolytic effects of adenosine A<sub>2A</sub> antagonists: implications for parkinsonism. *Front Biosci* 13:3594–3605
- Salamone JD, Ishiwari K, Betz AJ, Farrar AM, Mingote SM, Font L, Hockemeyer J, Müller CE, Correa M (2008b) Dopamine/adenosine interactions related to locomotion and tremor in animal models: possible relevance to parkinsonism. *Parkinsonism Relat Disord* 14(Suppl 2):S130–134
- Salvatore CA, Jacobson MA, Taylor HE, Linden J, Johnson RG (1993) Molecular cloning and characterization of the human A<sub>3</sub> adenosine receptor. *Proc Natl Acad Sci USA* 90:10365–10369
- Sauer R, Maurinsh J, Reith U, Fülle F, Klotz KN, Müller CE (2000) Water-soluble phosphate prodrugs of 1-propargyl-8-styrylxanthine derivatives, A<sub>2A</sub>-selective adenosine receptor antagonists. *J Med Chem* 43:440–448
- Scammells PJ, Baker SP, Belardinelli L, Olsson RA (1994) Substituted 1,3-dipropylxanthines as irreversible antagonists of A<sub>1</sub> adenosine receptors. *J Med Chem* 37:2704–2712
- Schapira AH, Bezar E, Brotchie J, Calon F, Collingridge GL, Ferger B, Hengerer B, Hirsch E, Jenner P, Le Novere N, Obeso JA, Schwarzschild MA, Spampinato U, Davidai G (2006) Novel pharmacological targets for the treatment of Parkinson's disease. *Nat Rev Drug Discov* 5:845–854
- Schindler CW, Karcz-Kubicha M, Thorndike EB, Müller CE, Tella SR, Goldberg SR, Ferré S (2004) Lack of adenosine A<sub>1</sub> and dopamine D<sub>2</sub> receptor-mediated modulation of the cardiovascular effects of the adenosine A<sub>2A</sub> receptor agonist CGS 21680. *Eur J Pharmacol* 484:269–275
- Schindler CW, Karcz-Kubicha M, Thorndike EB, Müller CE, Tella SR, Ferré S, Goldberg SR (2005) Role of central and peripheral adenosine receptors in the cardiovascular responses to intraperitoneal injections of adenosine A<sub>1</sub> and A<sub>2A</sub> subtype receptor agonists. *Br J Pharmacol* 144:642–650
- Schingnitz G, Küfner-Mühl U, Ensinger H, Lehr E, Kuhn FJ (1991) Selective A<sub>1</sub> antagonists for treatment of cognitive deficits. *Nucleosides Nucleotides* 10:1067–1076
- Schwarzschild MA, Agnati L, Fuxe K, Chen JF, Morelli M (2006) Targeting adenosine A<sub>2A</sub> receptors in Parkinson's disease. *Trends Neurosci* 29:647–654

- Seale TW, Ablu KA, Shamim MT, Carney JM, Daly JW (1988) 3,7-Dimethyl-1-propargylxanthine: a potent and selective in vivo antagonist of adenosine analogs. *Life Sci* 43:1671–1684
- Shamim MT, Ukena D, Padgett WL, Hong O, Daly JW (1988) 8-Aryl and 8-cycloalkyl-1,3-dipropylxanthines: further potent and selective antagonists for A<sub>1</sub>-adenosine receptors. *J Med Chem* 31:613–617
- Shamim MT, Ukena D, Padgett WL, Daly JW (1989) Effects of 8-phenyl and 8-cycloalkyl substituents on the activity of mono-, di-, and trisubstituted alkylxanthines with substitution at the 1-, 3-, and 7-positions. *J Med Chem* 32:1231–1237
- Shimada J, Suzuki F, Nonaka H, Ishii A (1992) 8-Polycycloalkyl-1,3-dipropylxanthines as potent and selective antagonists for A<sub>1</sub>-adenosine receptors. *J Med Chem* 35:924–930
- Shimada J, Koike N, Nonaka H, Shiozaki S, Yanagawa K, Kanda T, Kobayashi H, Ichimura M, Nakamura J, Kase H, Suzuki F (1997) Adenosine A<sub>2A</sub> antagonists with potent anti-cataleptic activity. *Bioorg Med Chem Lett* 7:2349–2352
- Shukla D, Chakraborty S, Singh S, Mishra B (2009) Doxofylline: a promising methylxanthine derivative for the treatment of asthma and chronic obstructive pulmonary disease. *Expert Opin Pharmacother* 10:2343–2356
- Slawsky MT, Givertz MM (2009) Rolofylline: a selective adenosine 1 receptor antagonist for the treatment of heart failure. *Expert Opin Pharmacother* 10:311–322
- Solinas M, Ferré S, Antoniou K, Quarta D, Zustinova Z, Pappas HJ, LA SPN, Wertheim C, Müller CE, Goldberg SR (2005) Involvement of adenosine A<sub>1</sub> receptors in the discriminative-stimulus effects of caffeine in rats. *Psychopharmacology* 179:576–586
- Sorbera LA, Martín L, Castaner J (2000) *Drugs Future* 25:1011–1016
- Soriano A, Ventura R, Molero A, Hoen R, Casadó V, Cortés A, Fanelli F, Albericio F, Lluís C, Franco R, Royo M (2009) Adenosine A<sub>2A</sub> receptor-antagonist/dopamine D<sub>2</sub> receptor agonist bivalent ligands as pharmacological tools to detect A<sub>2A</sub>-D<sub>2</sub> receptor heteromers. *J Med Chem* 52:5590–5602
- Stefanachi A, Brea JM, Cadavid MI, Centeno NB, Esteve C, Loza MI, Martinez A, Nieto R, Ravina E, Sanz F, Segarra V, Sotelo E, Vidal B, Carotti A (2008) 1-, 3- and 8-Substituted 9-deazaxanthines as potent and selective antagonists at the human A<sub>2B</sub> adenosine receptor. *Bioorg Med Chem* 16:2852–2869
- Stefanovich V (1989) The xanthines. *Drug News Perspect* 2:82–88
- Stiles GL, Jacobson KA (1987) A new high affinity, iodinated adenosine receptor antagonist as a radioligand/photoaffinity crosslinking probe. *Mol Pharmacol* 32:184–188
- Stiles GL, Jacobson KA (1988) High affinity acylating antagonists for the A<sub>1</sub> adenosine receptor: identification of binding subunit. *Mol Pharmacol* 34:724–728
- Stone GA, Jarvis MF, Sills M, Weeks B, Snowhill EW, Williams M (1988) Species differences in high affinity adenosine A<sub>2</sub> receptors in striatal membranes from mammalian brain. *Drug Dev Res* 15:31–46
- Strömberg I, Popoli P, Müller CE, Ferré S, Fuxe K (2000) Electrophysiological and behavioural evidence for an antagonistic modulatory role of adenosine A<sub>2A</sub> receptors in dopamine D<sub>2</sub> receptor regulation in the rat dopamine denervated striatum. *Eur J Neurosci* 12:4033–4037
- Suzuki F, Shimada J, Mizumoto H, Karasawa A, Kubo K, Nonaka H, Ishii A, Kawakita T (1992a) Adenosine A<sub>1</sub> antagonists. 2. Structure-activity relationships on diuretic activities and protective effects against acute renal failure. *J Med Chem* 35:3066–3075
- Suzuki F, Shimada J, Nonaka H, Ishii A, Shiozaki S, Ichikawa S, Ono E (1992b) 7, 8-Dihydro-8-ethyl-2-(3-noradamantyl)-4-propyl-1H-imidazo[2,1-*i*]purin-5(4H)-one: a potent and water-soluble adenosine A<sub>1</sub> antagonist. *J Med Chem* 35:3578–3581
- Takahashi RN, Pamplona FA, Prediger RD (2008) Adenosine receptor antagonists for cognitive dysfunction: a review of animal studies. *Front Biosci* 13:2614–2632
- Thorsell A, Johnson J, Heilig M (2007) Effect of the adenosine A<sub>2A</sub> receptor antagonist 3,7-dimethyl-propargylxanthine on anxiety-like and depression-like behavior and alcohol consumption in Wistar rats. *Alcohol Clin Exp Res* 31:1302–1307

- Ukena D, Jacobson KA, Kirk KL, Daly JW (1986a) A [3H]amine congener of 1,3-dipropyl-8-phenylxanthine. A new radioligand for A<sub>2</sub> adenosine receptors of human platelets. *FEBS Lett* 199:269–274
- Ukena D, Jacobson KA, Padgett WL, Ayala C, Shamim MT, Kirk KL, Olsson RA, Daly JW (1986b) Species differences in structure-activity relationships of adenosine agonists and xanthine antagonists at brain A<sub>1</sub> adenosine receptors. *FEBS Lett* 209:122–128
- Ukena D, Daly JW, Kirk KL, Jacobson KA (1986c) Functionalized congeners of 1,3-dipropyl-8-phenylxanthine: potent antagonists for adenosine receptors that modulate membrane adenylate cyclase in pheochromocytoma cells, platelets and fat cells. *Life Sci* 38:797–807
- Ukena D, Schudt C, Sybrecht GW (1993) Adenosine receptor-blocking xanthines as inhibitors of phosphodiesterase isozymes. *Biochem Pharmacol* 45:847–851
- van den Berg D, Zoellner KR, Ogunrombi MO, Malan SF, Terre'Blanche G, Castagnoli N Jr, Bergh JJ, Petzer JP (2007) Inhibition of monoamine oxidase B by selected benzimidazole and caffeine analogues. *Bioorg Med Chem* 15:3692–3702
- van Galen PJM, van Bergen AH, Gallo-Rodriguez C, Melman N, Olah ME, IJzerman AP, Stiles GL, Jacobson KA (1994) A binding site model and structure-activity relationships for the rat A<sub>3</sub> adenosine receptor. *Mol Pharmacol* 45:1101–1111
- van Muijlwijk-Koezen JE, Timmerman H, van der Sluis RP, van de Stolpe AC, Menge WM, Beukers MW, van der Graaf PH, de Groote M, IJzerman AP (2001) Synthesis and use of FSCPX, an irreversible adenosine A<sub>1</sub> antagonist, as a 'receptor knock-down' tool. *Bioorg Med Chem* 11:815–818
- Vlok N, Malan SF, Castagnoli N Jr, Bergh JJ, Petzer JP (2006) Inhibition of monoamine oxidase B by analogues of the adenosine A<sub>2A</sub> receptor antagonist (E)-8-(3-chlorostyryl)caffeine (CSC). *Bioorg Med Chem* 14:3512–2351
- Vollmann K, Qurishi R, Hockemeyer J, Müller CE (2008) Synthesis and properties of a new water-soluble prodrug of the adenosine A<sub>2A</sub> receptor antagonist MSX-2. *Molecules* 13:348–359
- Vu CB (2005) Recent advances in the design and optimization of adenosine A<sub>2A</sub> receptor antagonists. *Curr Opin Drug Discov Dev* 8:458–468
- Vu CB, Kiesman WF, Conlon PR, Lin K-C, Tam M, Petter RC, Smits G, Lutterodt F, Jin X, Chen L (2006) Zhang J (2006) Tricyclic imidazoline derivatives as potent and selective adenosine A<sub>1</sub> receptor antagonists. *J Med Chem* 49:7132–7139
- Weiss HM, Grisshammer R (2002) Purification and characterization of the human adenosine A<sub>2A</sub> receptor functionally expressed in *Escherichia coli*. *Eur J Biochem* 269:82–92
- Weyler S, Fülle F, Diekmann M, Schumacher B, Hinz S, Klotz KN, Müller CE (2006) Improving potency, selectivity, and water-solubility of adenosine A<sub>1</sub> receptor antagonists: xanthines modified at position 3 and related pyrimido[1,2,3-cd]purinediones. *ChemMedChem* 1:891–902
- Worden L, Shahriari M, Farrar A, Sink KS, Hockemeyer J, Müller CE, Salamone JD (2009) The adenosine A<sub>2A</sub> antagonist MSX-3 reverses the effort-related effects of dopamine blockade: differential interaction with D<sub>1</sub> and D<sub>2</sub> family antagonists. *Psychopharmacology* 203:489–499
- Yan L, Müller CE (2004) Preparation, properties, reactions, and adenosine receptor affinities of sulfophenylxanthine nitrophenyl esters: toward the development of sulfonic acid prodrugs with peroral bioavailability. *J Med Chem* 47:1031–1043
- Yan L, Bertarelli CG, Hayallah AM, Meyer H, Klotz KN, Müller CE (2006) A new synthesis of sulfonamides by aminolysis of *p*-nitrophenylsulfonates yielding potent and selective adenosine A<sub>2B</sub> receptor antagonists. *J Med Chem* 49:4384–4391
- Yu L, Shen HY, Coelho JE, Araujo IM, Huang QY, Day YJ, Rebola N, Canas PM, Rapp EK, Ferrara J, Taylor D, Müller CE, Linden J, Cunha RA, Chen JF (2008) Adenosine A<sub>2A</sub> receptor antagonists exert motor and neuroprotective effects by distinct cellular mechanisms. *Ann Neurol* 63:338–346
- Yuzlenko O, Kiec-Kononowicz K (2006) Potent adenosine A<sub>1</sub> and A<sub>2A</sub> receptors antagonists: recent developments. *Curr Med Chem* 13:3609–3625
- Zablocki J, Kalla R, Perry T, Palle V, Varkhedkar V, Xiao D, Piscopio A, Maa T, Gimbel A, Hao J, Chu N, Leung K, Zeng D (2005) The discovery of a selective, high affinity A<sub>2B</sub> adenosine receptor antagonist for the potential treatment of asthma. *Bioorg Med Chem* 15:609–612

# Theobromine and the Pharmacology of Cocoa

Hendrik Jan Smit

## Contents

1	Background .....	202
2	Theobromine .....	203
2.1	Characteristics .....	203
2.2	Natural Occurrence .....	203
2.3	Synthesis, Catabolism and Pharmacokinetics .....	203
2.4	Mechanism of Action .....	206
2.5	Effects in Animals .....	207
2.6	Effects in Man .....	212
2.7	Therapeutic Applications .....	218
3	Caffeine .....	218
4	Biogenic Amines .....	219
4.1	Phenylethylamine .....	220
4.2	Tyramine .....	221
4.3	Serotonin and Tryptophan .....	221
5	Anandamide .....	222
6	Salsolinol and Tetrahydro- $\beta$ -carbolines .....	223
7	Magnesium .....	224
8	Conclusions and Considerations .....	224
	References .....	226

**Abstract** The effects of theobromine in man are underresearched, possibly owing to the assumption that it is behaviourally inert. Toxicology research in animals may appear to provide alarming results, but these cannot be extrapolated to humans for a number of reasons. Domestic animals and animals used for racing competitions need to be guarded from chocolate and cocoa-containing foods, including foods

---

H.J. Smit

Functional Food Centre, Oxford Brookes University, Headington Campus, Gypsy Lane, Oxford OX3 0BP, UK

e-mail: [hsmit@brookes.ac.uk](mailto:hsmit@brookes.ac.uk)

containing cocoa husks. Research ought to include caffeine as a comparative agent, and underlying mechanisms need to be further explored. Of all constituents proposed to play a role in our liking for chocolate, caffeine is the most convincing, though a role for theobromine cannot be ruled out. Most other substances are unlikely to exude a psychopharmacological effect owing to extremely low concentrations or the inability to reach the blood–brain barrier, whilst chocolate craving and addiction need to be explained by means of a culturally determined ambivalence towards chocolate.

**Keywords** Chocolate · Cocoa · Comparative · Craving · Liking · Myths · Pharmacology · Psychology · Theobromine · Toxicology

## 1 Background

Chocolate is an excellent example of a dichotomous food commodity. The current scientific and popular media focus on health issues has produced two conflicting health labels for chocolate – antioxidant benefits versus increased risk of weight gain. This is a change from the 1990s, when the focus was on a search for psychoactive constituents of chocolate that would explain not only its appeal, but also its craving-inducing, even its alleged addictive qualities. First phenylethylamine (PEA; Hamilton 1992) and later anandamide (Tytgat et al. 2000) were at the centre of this debate (see Sects. 4.1, 5, respectively).

The only pharmacologically active substance that has generally been ignored in this respect is theobromine, at least in part because of an early and persistent notion that it does not stimulate the central nervous system (CNS) (e.g. “does not show any central activity worth mentioning” – Czok 1974; “ineffective by itself” – Sprugel et al. 1977; “virtually inactive” – Rall 1980, p. 593; “behaviourally inactive” – Snyder et al. 1981; “possesses little pharmacological activity and is almost devoid of effects on the CNS and cardiovascular system” – Gates and Miners 1999; “does not affect the nervous system” – Bonvehi and Coll 2000). This may explain why relatively few studies or reviews on the effects of theobromine have been published, especially in comparison with caffeine. However, some recent findings have created a renewed interest in theobromine. Indeed, although at first glance there appear to be very few relevant publications on the effects of theobromine, the reader will notice that a surprisingly large number of studies and other communications surfaced as work on this chapter progressed.

The main aim of this chapter is to assess the role theobromine plays in the pharmacological activity of chocolate – the main supplier of theobromine to the human diet. In addition, other (potentially) pharmacologically active chocolate constituents will be discussed.

## 2 Theobromine

### 2.1 Characteristics

As a purified chemical, theobromine is a white powder, and is mainly produced from cocoa husks as a by-product of chocolate manufacture (The Merck Index 2006), although it can also be synthesised from (3-methyl-)uric acid (The Merck Index 2006; Thorpe 1893, p. 697). It is only very slightly soluble in water (1 g/2,000 ml) and alcohol (1 g/2,220 ml 95%), and only slightly more soluble in boiling water (1 g/150 ml), though it dissolves in dilutions of alkali hydroxides and in mineral acids (The Merck Index 2006; European Pharmacopoeia 2005; IARC 1991).

Theobromine is considered a diuretic, a smooth muscle relaxant, a myocardial stimulant and a vasodilator (Dorland's Illustrated Medical Dictionary 2007). Unlike caffeine, it is a very mild CNS stimulant (Mumford et al. 1994), and it has both antioxidant and pro-oxidant characteristics (Azam et al. 2003).

### 2.2 Natural Occurrence

Of all structurally related purine alkaloids (methylxanthines), theobromine is the predominant member present in chocolate (Apgar and Tarka 1998). Therefore, chocolate and other cocoa products are the main sources of theobromine in our Western diet. However, it can also be found in small quantities in tea (*Camilla sinensis*; Hicks et al. 1996), guarana (*Paullinia cupana*; Weckerle et al. 2003), mate (*Ilex paraguariensis*; Cardozo et al. 2007) and cola nut (Souci et al. 1981; Burdock et al. 2009), whilst its presence in coffee is negligible at a mere 10% of that in tea (see Table 1 for a general overview of theobromine content in foods).

Note that different tea varieties contain different typical levels of methylxanthines (Hicks et al. 1996). A relatively recently discovered tea variety, *Camellia ptilophylla*, is naturally free of caffeine, but contains high levels of theobromine instead – around 15–18 times the level in of green tea (Yang et al. 2007; He et al. 2009), hence its familiar name “cocoa tea”. Likewise, cocoa bean varieties differ in their theobromine content, with Forastero varieties generally containing the highest amounts (Brunetto et al. 2007; Timbie et al. 1978), although some results do not agree with this (Hammerstone et al. 1994). See also Ashihara et al. (2010; Sect. 2.3), and Sect. 2.3.1 below.

### 2.3 Synthesis, Catabolism and Pharmacokinetics

The brief overview below cannot pretend to represent the complexities of this topic, although I have attempted to cover the most relevant and informative aspects. For a more detailed and in-depth approach, please refer to Ashihara et al. (2010) and Arnaud (2010).

**Table 1** Theobromine content of various products. (After Smit and Rogers 2001)

Product	Portion size <sup>a</sup>	Concentration (mg per portion)
Chocolate, dark	50 g	378 (237–519) <sup>b</sup> ; 221 <sup>c</sup>
Chocolate, milk	50 g	95 (65–160) <sup>b</sup> ; 94 <sup>c</sup>
Cocoa powder	10 g	189 (146–266) <sup>b</sup> ; 203 <sup>c</sup> ; 260 <sup>d</sup> 206 (178–240) <sup>e</sup> 263 (219–284) <sup>f</sup>
Tea (regular, bag)	230 ml	3.1 (1.4–4.4) <sup>b, g</sup>
Coffee (filter/percolated)	7.6 g/200 ml	0.3 (0.3–0.3) <sup>b</sup>
Coffee (instant)	1.6 g/200 ml	0.2 (0.1–0.5) <sup>b</sup>
Cola drinks	Can (330 ml)	ND <sup>h</sup>

ND not detected

<sup>a</sup>MAFF (1988)

<sup>b</sup>MAFF (1998); figures recalculated using comments in Annex C of this reference where appropriate

<sup>c</sup>Craig and Nguyen (1984)

<sup>d</sup>Risner (2008)

<sup>e</sup>Bonvehí and Coll (2000)

<sup>f</sup>De Vries et al. (1981)

<sup>g</sup>This is in accord with values of first brew in Hicks et al. (1996).

<sup>h</sup>Dried kola nut contains 0.05–0.10% theobromine (Souci et al. 1981; see also Duke 1992 in Burdock et al. 2009)

### 2.3.1 Theobromine Synthesis and Catabolism in *Theobroma cacao*

In the cocoa plant, theobromine accumulates in young leaves, and the concentrations decline as the leaves mature (Koyama et al. 2003). In the cocoa pod, theobromine is synthesised in both the pericarp (fleshy, outer layer) and the cotyledons (seed embryos) of young cocoa fruits, though during the ripening phase, pericarp theobromine concentrations decline sharply, whilst cotyledon (cocoa bean) theobromine concentrations increase. This suggests that the major site of theobromine synthesis is the cocoa bean itself, whilst not excluding a minor role for theobromine migration between pericarp and cocoa bean (Zheng et al. 2004). Whilst theobromine is synthesised from AMP via xanthosine, it is metabolised by demethylation via xanthine, both in the cocoa bean (Zheng et al. 2004; see Ashihara et al. 2008 for a review) as well as in the cocoa leaf (Koyama et al. 2003).

Methylxanthine (including theobromine) concentrations in the cocoa bean are broadly variety-dependent, although publications do not always agree: Brunetto et al. (2007) found cocoa bean theobromine levels varying between 0.7 and 2%, with the highest levels found in the Forastero varieties, whilst the theobromine-to-caffeine ratios varied between 2 and 12, with Criollo, Trinitario and Forastero varieties shown in order of increasing theobromine-to-caffeine ratio. Likewise, Timbie et al. (1978) found cocoa bean theobromine levels of 1.2–3.9%, with the highest average levels found in Forastero, the lowest level in Criollo (which had the highest caffeine content) and increasing theobromine-to-caffeine ratios from Criollo (1.1) through to Forastero (75.1; recalculated from data provided). Hammerstone et al. (1994), however, provided entirely different figures for the same varieties.



Their highest average theobromine content was found in the Criollo varieties (2.3%), with Trinitario, Criollo and Forastero showing increasing theobromine-to-caffeine ratios. Although the analytical procedures are very similar between the publications, minor variations in these procedures may account for some of the differences found. However, the ripening stage at which fruit is picked (Timbie et al. 1978) and possibly also other factors such as growing conditions in terms of soil quality/composition and weather may all affect the methylxanthine content.

Whilst the cocoa beans are being processed (fermentation, roasting, etc.), the theobromine content changes mainly during the fermentation stage. During this stage methylxanthines migrate from the bean into the shell, causing a decrease in cocoa bean theobromine content of around 25% (Timbie et al 1978). Additionally, it is not unreasonable to assume that the microorganisms involved in the fermentation process could further reduce the theobromine content, as is the case with tea (Wang et al. 2008).

### 2.3.2 Theobromine Uptake, Metabolism and Pharmacokinetics in Man

Following oral administration in man, theobromine absorption from the digestive tract is slow, especially compared with caffeine, with an estimated peak plasma time of 2.5 h (compared with 0.5 h for caffeine) (Mumford et al. 1996). Moreover, theobromine absorption is not complete, at least in some people (less than 90%; Cornish and Christman 1957). Interestingly, the theobromine peak plasma time after chocolate consumption is somewhat faster at 2 h after consumption (Mumford et al. 1996). Although this seems counterintuitive because of plausible increases in the release time from the chocolate food matrix and binding to phenolic compounds (Czok 1974), Mumford et al. (1996) suggested the shorter theobromine peak plasma time following chocolate administration may be caused by stimulating bile production, shown in other studies to improve drug absorption. Note, however, that the same study reported slower caffeine uptake from both chocolate and cola. Despite the explanation provided for the latter (delayed gastric emptying), the plasma concentration curves for both foods are strikingly similar, and suggest a possible sucrose-mediated suppression of the excitatory effects of caffeine (Chauchard et al. 1945). Clearly, more research is needed to uncover the factors relevant to methylxanthine absorption from food.

In humans, methylxanthines are metabolised by demethylation (removal of methyl side groups) by the enzyme cytochrome P450 (CYP). Hence, theobromine (3,7-dimethylxanthine) is broken down to 3-methylxanthine and 7-methylxanthine by CYP. 7-Methylxanthine is then further metabolised into 7-methyluric acid by xanthine oxidase (this is not the case for 3-methylxanthine), whilst metabolism of theobromine into 3,7-dimethyluric acid and 3,7-diaminouracil is less well understood, although this is at least in part CYP-mediated (Gates and Miners 1999).

Note that theobromine does not metabolise into other dimethylxanthines (i.e. theophylline or paraxanthine), nor does it “upgrade” to the trimethylxanthine

caffeine (Mumford et al. 1996), although the latter does happen in young leaves of the *Theobroma cacao* plant (Koyama et al. 2003). However, humans are exposed to theobromine through demethylation of caffeine, in addition to the ingestion of theobromine.

The clearance rate for acutely administered theobromine is around 1.2 ml/min/kg, around half of that of caffeine (Lelo et al. 1986), whereas after 4 days of chronic administration, Miners et al. (1982) found a clearance rate of 0.75 ml/min/kg. Likewise, Drouillard et al. (1978) found acute theobromine clearance rates of 0.94 ml/min/kg (1.47 after a 2-week methylxanthine abstinence), reduced to 0.81 after 5 days of chronic administration (figures calculated from published data), suggesting that the chronic exposure-related reduction in theobromine clearance is reversed after dietary theobromine abstinence (Drouillard et al. 1978). Note that interindividual differences in theobromine clearance rates may be substantial, as is the case for caffeine (Lelo et al. 1986 measured a  $1.2 \pm 0.4$  ml/min/kg theobromine clearance rate; Balogh et al. 1992 measured 79% interindividual variance in caffeine clearance rates). Moreover, tobacco smokers have a substantially increased theobromine clearance compared with non-smokers (Miners et al. 1985).

## 2.4 Mechanism of Action

Although various effects of caffeine have in the past been attributed to the release of intracellular calcium and inhibition of cyclic nucleotide phosphodiesterases, ordinary human consumption of dietary methylxanthines would be insufficient to reach the levels needed for these processes to be activated (Fredholm et al. 1999). The main mechanism of action for methylxanthines has long been established as an inhibition of adenosine receptors (Snyder et al 1981; see Fredholm et al. 1999 for an extensive review). A range of secondary effects of adenosine antagonism may explain the variety of effects of methylxanthines on the human system in more detail. The interaction of adenosine A<sub>2A</sub> receptors with dopamine D<sub>2</sub> receptors (Fredholm et al. 1999) is one such example. Interestingly, theobromine shows a much lower affinity for adenosine receptors than caffeine (Daly et al. 1983; Fredholm and Lindström 1999), which may explain why it is generally regarded as behaviourally inert. However, caffeine and theobromine show differential affinities for different adenosine receptor subtypes. Daly et al. (1983) found that theobromine is 2–3 times less active than caffeine as an adenosine A<sub>1</sub> receptor antagonist, but at least 10 times less active than caffeine as an A<sub>2</sub> receptor antagonist. Fredholm and Lindström (1999) gave similar values, but with a clear difference in caffeine-to-theobromine affinity ratios for striatum compared with cortex A<sub>1</sub> receptor antagonism (theobromine was found to be 4.7 and 11.8 times less active than caffeine, respectively). Nevertheless, the authors suggested caffeine and theobromine are non-selective receptor antagonists.

Interestingly, the much higher presence of theobromine in chocolate compared with that of caffeine (theobromine-to-caffeine ratio average 10; milk chocolate

11.3; dark chocolate 14.0; cocoa powder 9.0; recalculated from Tables 1 and 2 in Smit and Rogers 2001), clearly do not make up for the lower average adenosine receptor affinity of caffeine compared with that of theobromine (again, of around a factor 10 in Fredholm and Lindström 1999, akin to the difference in locomotor stimulation threshold between caffeine and theobromine reported by Snyder et al. 1981). Moreover, because  $A_1$  receptors determine the effects of caffeine on fluid intake (Rieg et al. 2007), whilst the  $A_{2A}$  receptors play a role in the desire for caffeine (El Yacoubi et al. 2005), the differential affinities for different receptor types provide a possible explanation for the observation that caffeine and theobromine exert different effects. Note that additionally, the caffeine dimethylxanthine metabolites paraxanthine and theophylline have adenosine receptor affinities even stronger than caffeine (Daly et al. 1983; Fredholm and Lindström 1999), thereby explaining part of the effects of caffeine, whilst theobromine, also a dimethylxanthine, does not have such metabolites. Moreover, the reduced and delayed uptake of theobromine compared with that of caffeine may further diminish the in vivo effect of theobromine as an adenosine receptor antagonist in terms of its central and peripheral effects.

## 2.5 *Effects in Animals*

The effects of theobromine in animals as reported in the scientific literature can broadly be categorised into three groups: (1) toxicology studies; (2) case studies or reports of theobromine poisoning; (3) pharmacology studies; and (4) behavioural studies. Additionally, concern regarding the use of theobromine as a doping agent in equine and related sports has also penetrated the scientific literature.

Dietary theobromine intake in animals originates from two sources: (1) domestic chocolate and chocolate- or cocoa-containing foods as consumed by humans; (2) animal feed containing cocoa shell. The use of cocoa shell in animal has seen a drastic increase since the discovery that (1) it contains high levels of vitamin D, (2) its addition to the cattle's winter diet raised the vitamin D level to that which it typically is during the summer months, and (3) milk fat content was also raised when using this feed (Knapp and Coward 1934; Kon and Henry 1935; Golding and Burr 1937 in Dowden 1938). It is likely that experience from the use of this feed taught the equine sports that it was beneficial to animal performance, though this is not clear from the literature. Note that McDonald et al. (2002; p. 596) mentioned another feed derived from the cocoa bean, that is "extracted cocoa bean meal", which also contains theobromine and which the authors therefore also did not recommend being fed to racing horses. Moreover, the European Food Safety Authority has mentioned cocoa bean meal, cocoa husk meal, cocoa germs, cocoa bean shells and discarded chocolate confectionery as sources for animal feed in Europe (EFSA 2008).

### 2.5.1 Toxicology Studies

Toxicology studies mainly concern teratology and male reproductive toxicology, presumably following a study by Friedman et al. (1979), which reported testicular atrophy in nearly all rats fed caffeine or theobromine at a dietary concentration of 0.5% for over 14 weeks, although the detrimental effects in the caffeine condition were greater. However, Gans (1984) reported the reverse, that is, testicular atrophy and spermatogenic cell destruction following feeding with theobromine were much greater than they were following feeding with caffeine. Though the latter study used a dietary concentration of 0.8% theobromine with an exposure time of 7 weeks, subsequent studies switched to daily doses of 25–500 mg/kg body weight (Wang et al. 1992), and have shown similar effects for a shorter test duration, even after 2 weeks (Funabashi et al. 2000). Lower toxicity has been shown for cocoa powder containing the same amount of theobromine (Wang and Waller 1994). Additionally, Tarka et al. (1981) showed that when rats were fed chow containing 0.6 and 0.8% theobromine for 7 weeks, testicular weight decreased significantly compared with feeding with 0 and 0.2% theobromine. Moreover, they showed this effect was irreversible as measured during the subsequent 7 weeks. Although the underlying mechanism is unclear, its effects are seen also in terms of degeneration and necrosis in spermatogenic cells (Gans 1982; Wang and Waller 1994). Similarly to theobromine, cocoa powder at 5% of the diet showed testicular atrophy and decreased spermatogenesis (Tarka et al. 1991). The effects of theobromine on the male reproduction system described above have been validated in several other publications (Weinberger et al. 1978; Soffiatti et al. 1989; Tarka et al. 1979). Note that similar atrophy effects have been observed for the thymus gland in rats (Tarka et al. 1979), appearing sooner than testicular damage (Gans 1982), and with theobromine producing higher decreases in thymus weight than caffeine (Gans 1984), though these effects were not found in dogs (Gans et al. 1980). Because this gland “plays an important role in cellular immunity by generating circulating T lymphocytes” (Nishino et al. 2006), the effects of theobromine reported on this gland may suggest an increase in overall immune response suppression.

The toxic effects of theobromine also include growth reduction and weight loss, possibly achieved through loss of appetite and food intake (Tarka et al. 1979).

Theobromine doses as low as 6 mg/day in the diet of mother mice reduces embryo weight and embryo tissue angiogenic activity (i.e. the rate at which new blood vessels are formed in growing tissue), and reduces neonatal relative limb size and spleen weight, suggesting that this is caused by a theobromine-induced reduction in the formation of new blood vessels in embryos (Chorostowska-Wynimko et al. 2004). The same research group showed a similar effect of chocolate (Skopinski et al. 2004) but attributed this to its epigallocatechin content owing to the correlations found between effect size and epigallocatechin content. Although this appears strange as the theobromine concentrations would have produced the same conclusion for theobromine and confirmed the results of their other publication of the same year, yet another study confirmed the link between dietary cocoa flavanol dose and embryonic (and tumour) angiogenesis (Wasiutynski et al. 2005).

Further studies will need to point out differential roles or mechanisms for these effects of theobromine and cocoa polyphenols, respectively, and evidence for similar effects in man ought to be sought. Nevertheless, Tarka et al. (1986a, b) pointed out that at much lower doses (25–200 mg/kg body weight/day), only their highest doses showed teratogenic effects (a delay in osteogenesis in rats; maternal toxicity/mortality, fetal malformations and osteogenic delays in rats), whilst the theobromine intake in these doses in rats and rabbits would be equivalent to an unrealistic human consumption of 7.5–10 lb (3.4–4.5 kg) milk chocolate/day, possibly explaining why no human teratogenic effects of theobromine have been reported. Alternatively, a 5% cocoa powder as used by Tarka et al. (1991) would not be impossible to implement in the human diet, though the effects of this on the male (and the female) reproduction system are unknown.

Interestingly, and in line with the findings reported above, angiogenesis in tumour growth is also inhibited by theobromine, an example of how a toxic effect can have a positive outcome (eBarcz et al. 1998; Gil et al. 1993). This effect is mediated through inhibition of adenosine receptors (Barcz et al. 2000) present in the carcinoma itself (Ryzhov et al. 2008) and their role in carcinoma hypoxia (Ryzhov et al. 2007), which would explain why similar effects are found with caffeine (Merighi et al. 2007). Conversely, theobromine intake has been associated with the prevalence of prostate (Slattery and West 1993) and testicular (Giannandrea 2009) cancer, although these associations were inconsistent over several decades Giannandrea (2009), and have not been tested further. Nevertheless, theobromine can reduce copper, thereby generating oxygen radicals (Shamsi and Hadi 1995 in Schmid et al. 2007). Moreover, because caffeine can impair DNA double strand repair (Sarkaria et al. 1999), it is possible this may also apply to theobromine, lending theobromine, as is the case for caffeine (Azam et al. 2003), both pro- and anticarcinogenic properties. Investigating the effects of cocoa powder, Tarka et al. (1991) found no evidence of a carcinogenic effect. However, the phenolic content of cocoa is likely to counteract any carcinogenic activity of other cocoa constituents (Lee et al. 2006; Jourdain et al. 2006).

Note that the toxic effects of theobromine may depend on other dietary constituents (e.g. protein content) and species-specific tolerance levels. Therefore, the marked differences in theobromine's toxic effects observed between animal species may make extrapolations to the human system very complex (Tarka et al. 1979), if not impossible.

### 2.5.2 Case Studies of Animal Poisoning

Many cases of animal poisoning reportedly result from the consumption of chocolate. Dogs, unlike cats, find chocolate a most palatable food, and are therefore most vulnerable to chocolate poisoning, especially when kept indoors. Strachan and Bennett (1994) reported acute cardiac arrest in a dog on the morning after the consumption of cocoa powder on the evening before, with an estimated theobromine exposure of 80 mg/kg body weight. Stidworthy et al. (1997), however, reported

similar symptoms in two dogs who died within 1 h after an estimated consumption of 20–30 g dark chocolate each (using Table 1 and the reported average animal weight of 24 kg, this equates to an estimated theobromine exposure of 8 mg/kg), whilst two similar animals fed the same appeared unaffected. Interestingly, Gans et al. (1980) showed that acute doses of 200 mg/kg and less were not lethal. Other cases of dog poisoning have been reported following the consumption of garden mulch made of chocolate beans and shells, although these animals were successfully treated and recovered within 5 days (Hovda and Kingston 1994). The symptoms are varied, but include vomiting, restlessness, diarrhoea, haematuria (blood in urine), tachycardia (rapid heart beat) and hyperpnoea (deep breaths due to hypoxia) (Hovda and Kingston 1994), shivering and convulsions (Strachan and Bennett 1994), and panting, restlessness and muscle tremors (Gans et al. 1980).

However, deaths following the consumption of chocolate have also been found in wildlife. Reportedly, parrots (Gartrell and Reid 2007), foxes and badgers (Jansson et al. 2001), and undoubtedly more animal species, have been the victim of the consumption of chocolate left unattended.

Even the consumption of cocoa products as an ingredient in cattle feed or other animal feed (i.e. cocoa meal, cocoa husks or chocolate waste from the food or catering industry) can lead to livestock poisoning, even death (e.g. poultry – Black and Barron 1943; calves – Curtis and Griffiths 1972; ducks – Gunning (1950); fowl, ducks and horses – see Blakemore and Shearer 1943 for a review of several early cases).

The toxicity of chocolate to animals has inspired research into coyote pest control in the USA, resulting in an optimal mortality caffeine-to-theobromine ratio of 1:5 (Johnston 2005), not dissimilar to that of chocolate and other cocoa products, reconfirming the danger of this food in domestic animals. Note, however, that the latter publication reiterated the importance of the *combination* of caffeine and theobromine in the effects found, suggesting a focus on theobromine alone as the active toxicant it is not justified when toxic effects or death are caused by the consumption of chocolate.

### 2.5.3 Equine Sports and Theobromine Doping

In equine sports, caffeine and theobromine are considered doping agents owing to their stimulant effects. Hence, horse urine should not contain any caffeine (exposure detection level set at 0.1 µg/ml), whilst theobromine levels should not exceed 2 µg/ml (IFHA 2007). Although this appears to be a fairly generous level for a doping substance, this can be easily exceeded by feeding a horse 20 chocolate-coated peanuts per day (equivalent to 1.5 such peanuts per day for a human being on a weight basis), and could therefore be interpreted as extremely conservative (Budhraj et al. 2007). Logically, the use of by-products from the cocoa industry in horse feed also increases urine theobromine levels (Haywood et al. 1990), again increasing the risk of doping detection. Upon theobromine exposure, Delbecke and Debackere (1991) recommend a 2-day washout period to ensure urinary

theobromine levels are below the legal threshold, although for other methylxanthine-containing foods, e.g., guarana, this may be insufficient: Salvadori et al. (1994) identified theobromine in horse urine up to 318 h (13 days) after guarana administration. Moreover, like many other drugs, toxins and trace elements and/or their metabolites, theobromine can also be detected in equine hair as a means for assessing drug history (Dunnett and Lees 2003). Whilst methylxanthine doping is also an issue in greyhound racing (Wells et al. 1988; Loeffler et al. 2000) it would be interesting to see if the relevant sports organisations will follow the example of the World Anti-Doping Agency of moving caffeine from the “Prohibited List” to the “Monitoring Program” for detecting patterns of misuse rather than imposing a ban. The reasons for this change include (1) the presence of a great interperson variability in caffeine metabolism, (2) the notion that above the traditionally used 12 µg/ml threshold level, caffeine has a detrimental effect on performance, but also (3) that lowering the detection threshold increases the risk of being penalised for consuming caffeine through everyday food and drink (WADA 2008). It is likely that some, if not all, of these arguments are applicable to dogs and horses, where chocolate treats and potential contamination of feed with cacao may impose more problems than the benefits for both racing organisations and competitors.

Much like horse racing in Western countries, camel racing is as important a sport in, for example, the United Arab Emirates, where methylxanthines are assessed in camel urine using a zero-tolerance approach in doping control (Wasfi et al. 2000).

#### **2.5.4 Pharmacology Studies**

Unlike toxicology studies, only a few studies have investigated theobromine metabolism in animals, one of which recorded this in detail in rats (Bonati et al. 1984), and did not find a clear difference between acute and chronic administration on the pharmacokinetics, though the absorption rates declined with increased theobromine doses. Shively and Tarka (1983) found that theobromine metabolism was slower in rats than in humans, whilst in rats it was not affected by pregnancy status. Moreover, in a study comparing five mammalian species (rats, mice, hamsters, rabbits and dogs), Miller et al. (1984) found that theobromine was most extensively metabolised in male mice and rabbits, and that theobromine metabolism shows only quantitative differences between species and sexes.

#### **2.5.5 Behavioural Studies**

Kuribara and Tadokoro (1992) reported that the mean 3-h post-treatment ambulatory activity in mice was increased after oral doses of both 10 mg/kg theobromine and 1 g/kg cocoa powder, whilst response rates were increased in the shuttle avoidance task at 3 mg/kg theobromine. However, the performance in the avoidance tasks was disrupted at 100 mg/kg theobromine or higher (Kuribara and



Tadokoro 1992). Similar results were reported by the same group in a different paper (Kuribara et al. 1992), where only the 10 mg/kg theobromine dose increased the avoidance rate in mice, and where at the 1,000 mg/kg dose, half of the mice died within a few hours. Because the measurements were taken over a 3-h period, this may explain why Snyder et al. (1981) found no effect on locomotor activity in mice at 5–100  $\mu\text{mol/kg}$  (1–18 mg/kg) during their 1-h post-treatment observation. Heim et al. (2009), however, found no effects of 30 mg/kg theobromine or 200 mg/kg cocoa tea (*Camellia pitilophylla*; see Sect. 2.2) on ambulatory behaviour in mice during a 2h post-treatment observation period. Instead, they reported that only in combination with caffeine (as chemicals or as green tea) was a synergistic effect found compared with caffeine alone. Although the caffeine dose (10 mg/kg) at which the synergistic effects with theobromine were shown would have been unusually high in humans and surely not relevant to chocolate consumption, this study provides important evidence for furthering our understanding of the behavioural effects of the methylxanthines in chocolate. Conversely, Heim et al. (1971) and Sprugel et al. (1977) found that locomotive activity, oxygen consumption and brain cyclic GMP and cyclic AMP levels in white mice were affected by caffeine, but that this effect was prevented by theobromine, whilst theobromine itself did not affect these measures (Heim et al. 1971; Sprugel et al. 1977). Only 2–3 h after treatment did effects of theobromine alone occur (Heim et al. 1971). Moreover, after caffeine versus saline discriminative stimulus training in male Sprague-Dawley rats, several methylxanthines, but not theobromine, generalised to the caffeine cue at most doses tested (10–75 mg/kg for theobromine; Carney et al. 1985). These findings suggest that, at least in mice, the theobromine concentrations in chocolate may have a behavioural consequence, that this consequence is of an interactive nature with other methylxanthines, and that behavioural effects of theobromine may be delayed compared with those of caffeine.

Only a few other animal species have been the subject of investigations regarding behavioural effects of theobromine. After previously having identified some purines and other potentially behaviourally active substances from hornet queens, Ishay and Paniry (1979) investigated the effects of the main methylxanthines on hornet behaviour. They found that unlike the effects of purine and hypoxanthine, the effects of caffeine, theobromine and theophylline included nervousness, shaky movement and unsteady gait, reduced physical contact and positive geotropism, with no marked differences between the methylxanthines.

## 2.6 *Effects in Man*

Although theobromine is the most predominant methylxanthine present in chocolate, research into the effects of theobromine in man is relatively scarce compared with that into the effects of caffeine, and compared with research in animals. This section aims to present the research on theobromine in man to date.



### 2.6.1 Psychopharmacological Effects

Several inappropriately substantiated popular claims about the psychopharmacological activity of chocolate constituents (e.g. PEA, see later) resulted in the investigation of the ecological potential of a range of such substances (Smit and Rogers 2001). It was concluded that caffeine and theobromine were the only likely substances to play a role in the psychopharmacological activity of chocolate. This idea was confirmed when the same authors (Smit et al. 2004) showed that the combination of caffeine (19 mg) and theobromine (250 mg) contained in a 2-oz bar (approximately 50 g) of dark chocolate has significant effects on energetic arousal, reaction time and information processing. Subsequent work reported that the same combination of methylxanthines increased the liking for the flavour of a 'novel' drink when combined with the (encapsulated) active substances compared with an encapsulated 'placebo' (Smit and Blackburn 2005). These results show a role for chocolate methylxanthines in our liking for chocolate. Additionally, they provide a very clear explanation for why we prefer milk chocolate over white chocolate, and why dark chocolate is an easily acquired taste. However, a study comparing the individual effects of caffeine and theobromine with the effect of their combination (as used in Smit et al. 2004) using identical, ecologically valid amounts has not been performed to date. Such a study would clarify whether the effects found are either solely or partly attributable to caffeine, and whether caffeine and theobromine provide an additive or synergistic effect.

Only a very few early publications have reported individual and combined effects of caffeine and theobromine. Dorfman and Jarvik (1970) gave volunteers 300 mg caffeine and/or 300 mg theobromine before the volunteers retired for the evening. Those in the caffeine and caffeine + theobromine condition showed a longer sleep latency and lower sleep quality than those in the theobromine condition. Additional data confirmed that sleep latency increases were related to caffeine dose and not to theobromine. Finally, they did not find any interactive effects of the two methylxanthines.

In a study of a more exploratory nature, Mumford et al. (1994) provided some valuable insights into the comparative effects of caffeine and theobromine on mood and cognition by investigating their subjective effects. Despite the small sample size ( $N = 7$ ), and the use of relatively high doses of methylxanthines [the doses used were the lowest discriminable caffeine dose in the least sensitive volunteer (178 mg) and the highest tolerated dose of theobromine by the most sensitive volunteers (560 mg)], this study presented some very interesting and important findings. First of all, it shows how theobromine possesses caffeine-like qualities by means of the subjective effect descriptions of the most theobromine sensitive participant: "Energy", "Motivation to work", "Alert", "Sleepy" (decreased), whilst these effects were emphasised by an additional effect on the measure "Magnitude of drug effect". Second, the discrimination threshold phase of the study showed a wide range of reliable discrimination thresholds amongst the volunteers, although this was not further investigated. This study only provided limited information with

regard to the role of the individual methylxanthines in the psychopharmacological effects of chocolate, although clearly a role for the effects of theobromine cannot be ruled out, and may depend on the individual's sensitivity to these effects.

Further evidence for caffeine-like effects of theobromine, albeit anecdotal, was provided by Ott (1985; pp. 79–80), who replaced his dietary caffeine intake with a daily dose of 600 mg theobromine (200 mg in the morning, afternoon and evening) for 7 days. Upon acute theobromine deprivation, the author described how he “developed a tension headache, muscle tension in his shoulders and neck, and became extremely lethargic” within 16 h. These symptoms were reversed within 60 min of the consumption of another 200-mg dose of theobromine, suggesting that the symptoms were that of theobromine withdrawal. Because this one-man experiment was not performed according to double-blind conventions, Ott advocated that the scientific community carry out a proper study looking into these effects.

In summary, theobromine produces only very minor subjective effects compared with caffeine. In sensitive individuals these effects may be more marked, but can also be detrimental in the form of headaches (Mumford et al. 1994), as can caffeine. However, anecdotal evidence suggests that theobromine behaves like caffeine by means of its capability of producing withdrawal and providing subsequent withdrawal-reversal effects. Unfortunately, no data on the effects of theobromine on mood and cognition in humans other than those presented above have been reported, confirming that this area is seriously underresearched. Although the psychopharmacological effects of theobromine may be smaller than those of caffeine, they have been reported. Taking into account habitual caffeine and theobromine intake, and discriminable and/or tolerable doses, these may help to provide a more sensitive method for uncovering clearer effects of theobromine on mood and mental performance.

## 2.6.2 Physiological Effects

### Cardiovascular

Theobromine is generally regarded both as a bronchodilator and as a vasodilator (Reynolds 1993) and may therefore have an effect on the heart. Indeed, Czok (1974) claimed theobromine provides an effect of medium strength on the heart in general, an effect less strong than the related theophylline, but stronger than caffeine. However, no more precise explanation than that was provided, nor were any citations listed. Effects of theobromine on the heart were confirmed by anecdotal evidence reported in Ott (1985, p. 82), where the author described experiencing cardiac-stimulating effects of an oral dose of 200 mg theobromine within 15 min of administration. Interestingly, theobromine has also been prescribed for relief from pain caused by angina pectoris in some patients, presumably by means of its vasodilating effects (Dock 1926). Although Baron et al. (1999) did not find any cardiac or haemodynamic effects of theobromine, it is possible that the cocoa polyphenols in their chocolate may have obscured any theobromine-related effects.

Note that other studies have also investigated haemodynamic effects of chocolate, but attributed these effects to cocoa polyphenols (Taubert et al. 2003; Grassi et al. 2005) whilst not taking into account the potentially confounding effects of theobromine, although Kelly (2005) argued for this to be addressed.

Geraets et al. (2006) found strong inhibitory effects of theobromine on the activity of the nuclear enzyme poly(ADP-ribose) polymerase-1 (PARP-1), which is implied in acute and chronic inflammatory diseases such as stroke, ischaemia–perfusion and diabetes, and implied in chronic obstructive pulmonary disease. For this reason, they emphasised that methylxanthines (including theobromine) with higher PARP-1 inhibition rates are potentially helpful dietary agents in the treatment of vascular dysfunction and inflammation.

On balance, very few studies have been published investigating cardiovascular effects of theobromine, though some limited evidence suggests that theobromine exerts cardiovascular effects by means of vasodilation and cardiac stimulation. Because the effects of caffeine on cardiovascular functions are expressed through noradrenalin release from sympathetic nerves acting on  $\alpha_2$ -adrenergic receptors, with a possible, but much less important role for adenosine ( $A_1$ ) receptor antagonism (Fredholm et al. 1999), similar effects of theobromine can be expected, although possibly of lower magnitude. Finally, further research is needed to validate the hypothesis that theobromine can be used for the prevention and treatment of vascular dysfunction and inflammation.

## Respiratory

Theobromine improves bronchodilation in asthma patients (Simons et al. 1985), although this effect is stronger with theophylline and caffeine (Becker et al. 1984). However, note that the order of bronchodilation efficacy for these three methylxanthines is different in Apgar and Tarka 1999, who listed theobromine as stronger than caffeine for this effect. Presumably owing to its superior diffusion in bronchial tissue (van Zyl et al. 2008), theophylline (1,3-dimethylxanthine) is still used as a medication for asthma patients but can have serious side effects (Barnes and Pauwels 1994; El-Bitar and Boustany 2009), whilst caffeine and theobromine are not in use as such. Nevertheless, caffeine does improve lung function (Bara and Barley 2001), as is also supported by epidemiological evidence (Pagano et al. 1988), suggesting that asthma and bronchitis patients may be self-dosing on caffeine to relieve symptoms, even if this is subliminally achieved by means of positive reinforcement. Because theobromine *also* improves lung function (10 mg/kg; Simons et al. 1985), there may be a similar role for the consumption of chocolate in the relief of asthmatic symptoms. Indeed, I have anecdotal evidence (from a personal acquaintance whose partner is suffering from asthma) of a clear association between periods of heightened asthmatic symptoms and a marked increase in consumption of both chocolate and cola drinks. Interestingly, it was not the patient herself, but was her partner who became aware of this association. Note that whilst the tobacco industry claims to add cocoa powder to cigarettes as a

flavouring agent, it may also conveniently serve to enhance the uptake of nicotine (and thereby increase the addictive property of tobacco) through the bronchodilating properties of theobromine (Rambali et al. 2002), as well as possibly suppressing smoke-induced cough reflex (see below).

A complementary beneficial effect of theobromine on the airways relates to theobromine's more recently identified cough reflex suppressant ("antitussive") properties through suppression of vagus nerve activity. This effect was shown in response to both inhalation of a citric acid aerosol in guinea pigs and to inhalation of a capsaicin aerosol in humans. Interestingly, although there was no clear difference between theobromine and codeine in suppressing citric acid induced cough in guinea pigs, the suppressant effects of theobromine on cough induced by a capsaicin aerosol in human volunteers were greater than those of codeine. Moreover, and unlike codeine, theobromine was free from side effects (Usmani et al. 2005), an important notion in the context of a strong need for antitussives without side-effects (Chung and Chang 2002). This could lend theobromine a direct medical application in the reduction of cough, as cough is a common symptom in cancer (Walsh et al. 2000), and usually responds well to one or more medications (Table 1 in Estfan and Walsh 2008). This application could be extended to chocolate, where a corresponding portion of dark chocolate could reduce the cost of conventional medicine where proven effective. Hence, dark chocolate is currently being investigated as an alternative to medicine for its potential to reduce cough in cancer patients for whom cough is a troubling symptom (Halfdanarson and Jatoi 2007).

Though seemingly related, not in the least by the effect theobromine has on either, the regulation of smooth muscle relaxation and that of cough suppression are different. Smooth muscle relaxation is regulated by  $\beta_2$ -adrenergic receptors (whereby  $\beta_2$ -adrenergic receptor agonists acutely improve bronchodilation, although chronic exposure can have detrimental effects on the control of asthma; Lipworth and Williamson 2009), and by adenosine receptors (Brown et al. 2008). Although methylxanthines have bronchodilating effects and act as adenosine receptor antagonists, this may not be the main mode of action for theobromine as a bronchodilator or smooth muscle relaxant. Indeed, Lunell et al. (1983) reported strong bronchodilator effects of a xanthine derivative without any CNS effects. However, both  $A_1$  and  $A_{2B}$  receptors have been implied in the pathogenesis of asthma, and although roles for  $A_{2A}$  and  $A_3$  receptors are likely, they are still unclear (Brown et al. 2008).

The cough reflex is triggered by three different kinds of sensory nerve receptors in the respiratory tract, whose signals are relayed via the vagus nerve and the brainstem to the "cough centre" or "central cough generator", where the physical cough response is coordinated (Chung and Pavord 2008). Although current antitussives are mainly opiates and opiate derivatives acting on the central cough pathway, their side effects call for the development of other substances that achieve the same goal through different mechanisms. Therefore, new and proposed antitussives acting centrally may target sigma or GABA receptors, or act through other mechanisms yet to be identified (Reynolds et al. 2004). Alternatively, they may act peripherally by directly targeting neuronal pathways, for example, ion channels, nerve fibres and relevant receptor sites (Chung and Chang 2002; Reynolds et al. 2004). Moreover,

guinea pig sensory nerve activity and human sensory nerve activity in the airways are inhibited by activating cannabinoid CB<sub>2</sub> receptors (Patel et al. 2003; Belvisi et al. 2008). Although Usmani et al. (2005) showed that theobromine also inhibits guinea pig vagus nerve activity, its modus operandi has not been established. Nevertheless, the authors suggested that theobromine is likely to exert its effect through suppression of phosphodiesterase activity and by inhibiting bronchoconstricting adenosine A<sub>1</sub> receptors, though alternative modes of action (e.g. activation of Ca<sup>2+</sup>-activated K<sup>+</sup> channels) cannot be ruled out (Usmani et al. 2005).

Concluding, both the antitussive and the bronchodilating effects of theobromine are at least in part related to the adenosine receptor antagonistic properties of theobromine as part of the methylxanthine family, and it could be that different effects are expressed through different adenosine receptor subtypes and through other receptors, such as β<sub>2</sub>-adrenergic receptors. The exact pathways for the bronchodilating and antitussive effects of theobromine are unclear, and whilst other pathways may be involved, further investigation is clearly needed to clarify this topic.

## Renal

Because adenosine plays an important role in regulating blood flow, it also plays an important role in renal haemodynamics, affecting renal blood flow and glomerular filtration rates. The renal vascular system, however, unlike the main vascular system, is regulated by adenosine A<sub>1</sub> receptors in addition to A<sub>2</sub> receptors (see Hansen and Schnermann 2003; Vallon et al. 2006 for reviews of the role of adenosine in the kidney). The finding that A<sub>1</sub> receptors also determine the effects of caffeine on fluid intake (Rieg et al. 2007) may be related to this. Because theobromine has a lower overall adenosine receptor affinity than caffeine and theophylline, though all three methylxanthines are non-selective adenosine antagonists (Fredholm and Lindström 1999), a small, but possibly insignificant, diuretic effect of theobromine would be predicted not to be functionally different from the other methylxanthines. Indeed, despite a previous and unjustified claim that theobromine has a stronger effect on the kidney than caffeine (Czok 1974), Dorfman and Jarvik (1970) and Massey and Whiting (1993) reported that unlike caffeine, oral administration of 300 mg theobromine did not increase urinary calcium or sodium excretion, although Dorfman and Jarvik (1970) found no change in the overnight urine volume following oral administration of 300 mg caffeine or 300 mg theobromine compared with 'no drug'.

## Dental

The consumption of chocolate, as a sugar-containing confectionery, is inevitably associated with dental caries (i.e. chocolate is seen as a cariogenic food). However, both theobromine (added to the diet in hamsters – Strålfors 1967; applied to human teeth in vitro – Sadeghpour 2007), and cocoa (reviewed in Naylor 1984) reportedly

inhibit dental caries. Sadeghpour (2007) found that regular exposure to theobromine increased the enamel surface microhardness compared with sodium fluoride, and helped surface recrystallisation. Kashket et al. (1985) found that defatted cocoa inhibits dental plaque formation, as did cocoa extracts (Srikanth et al. 2008) and cocoa polyphenol extracts (Percival et al. 2006). Whilst the preparations used in the work reported in the latter publication may or may not have been free of methylxanthines, the authors did not make any reference to this, and also other publications have reported effects of polyphenol-containing drinks on plaque formation without referring to its methylxanthine content (Hannig et al. 2009). Whilst there may also be a role for caffeine in combating dental caries (Strålfors 1967), it is likely that methylxanthines and polyphenols may have an effect on dental caries by means of separate mechanisms, suggesting that a combined application may be more beneficial, although more research is necessary to confirm this. The strong inhibition of the metabolic activity of anaerobic bacteria by fluoride in wastewater treatment (Ochoa-Herrera et al. 2009) may well prove to be another decisive factor for the promotion of theobromine- and polyphenol-containing toothpaste in the near future.

## 2.7 Therapeutic Applications

Theobromine is currently not in use as a medicinal drug. However, *Stedman's Medical Dictionary* (Stedman's Medical Dictionary 1976) describes theobromine as “used as a diuretic, myocardial stimulant, dilator of coronary arteries, and smooth muscle relaxant” and according to Landau (1986), theobromine was used to treat arteriosclerosis and some peripheral vascular diseases, whilst Reynolds (1993) added angina pectoris and hypertension to this list. Rall (1980), however, mentions that it has almost disappeared from the medical scene owing to its low effectiveness in its pharmacological actions compared with caffeine and theophylline, and whilst Tarka (1982) wrote that there was no therapeutic use for theobromine, Simons et al. (1985) mentioned its use in antiasthma medication.

Recent research, however, has identified theobromine as a PARP-1 inhibiting (Geraets et al. 2006), dental enamel strengthening (Sadeghpour 2007) and antitussive (Usmani et al. 2005) agent (see earlier), suggesting there is still a future for theobromine as a medicine, preventative or curative.

## 3 Caffeine

Unlike theobromine, the effects of caffeine have been extensively investigated. Absorption of caffeine is rapid and complete following oral administration, though in the presence of sugar, absorption is slower but still complete (Yesair et al. 1984), and the maximum blood plasma concentrations (peak plasma time) are reached

within 1 h (James 1991). Indeed, after oral administration of 72 mg caffeine, Mumford et al. (1994) found an onset of subjective effects at 21 min (10–45 min) followed by a caffeine peak plasma time at 30 min after treatment. By means of its adenosine receptor antagonistic properties, caffeine stimulates the CNS and increases blood pressure, respiration, lipolysis, renin and catecholamine release, urine output, and intestinal peristalsis (Landau 1986; James 1991).

Consumption of excessive amounts (more than 1 g/day or more than ten cups of strong coffee per day) can result in tachycardia, dyspepsia (disturbed digestion, decreased appetite, oppressive feeling in the stomach and unpleasant taste), irritability and insomnia, also referred to collectively as “caffeinism” (Landau 1986). Other publications have described symptoms following the intake of high doses of caffeine as “signs and symptoms indistinguishable from those of anxiety neurosis”, and nervousness, irritability, tremulousness, occasional muscle twitching, insomnia and sensory disturbances (Tarka 1982) and “a variety of unpleasant subjective states including anxiety, dysphoria and depression” (Mumford and Holtzman 1991).

As a psychostimulant, caffeine increases feelings of energy (more alert, less tired, etc.) and improves other aspects of mood, and enhances psychomotor and cognitive performance when taken in amounts consumed in coffee and tea (Rogers and Dernoncourt 1998; also reviewed in James 1991). Because caffeine reverses overnight caffeine-withdrawal symptoms, which include headache and lethargy (reviewed in Smit and Rogers 2007), it is a powerful (“negative”) reinforcer in learned behaviour as indicated, for example, by its ability to increase flavour preference (Rogers et al. 1995; Yeomans et al. 1998). It is this ability which is thought to lie at the heart of the fact that coffee and tea are the world’s most popular and widely consumed drinks despite their innate bitterness. Because doses as low as 12.5 mg caffeine have shown behavioural effects (Smit and Rogers 2000), and because such amounts are present in easily consumable portions of chocolate (despite their much higher presence in tea and coffee; see Smit and Rogers 2001), one can only assume that caffeine in chocolate has pharmacological activity, and that caffeine reinforcement could contribute to our liking for chocolate.

## 4 Biogenic Amines

Cocoa and cocoa products contain biogenic amines (e.g. PEA, tyramine, tryptamine and serotonin) and their precursors (phenylalanine, tyrosine and tryptophan) in fairly high concentrations, which increase during fermentation of the cocoa beans, and decrease during roasting and alkalisation (Ziegler et al. 1992). In general, these concentrations are irrelevant in healthy people, since biogenic amines are metabolised by the monoamine oxidase (MAO) enzymes in the mucosa of the small intestine, and in the liver and kidneys (Askar and Morad 1980). Because of the endogenous abundance of MAO enzymes, “even the intraduodenal injection of amines in the absence of enzyme inhibition would be unlikely to lead to their absorption and appearance in systemic blood unless the amount was



sufficiently large to swamp the deaminating mechanisms” (Marley and Blackwell 1970). The effects of biogenic amines are therefore only expressed in people with an MAO deficiency, as has been suggested for migraine sufferers (Marley and Blackwell 1970), and in patients receiving medication containing MAO inhibitors (Askar and Morad 1980). These effects, however, can include headaches, increased blood pressure and even a life-threatening “amino shock” (Askar and Morad 1980). Realistically, these adverse effects would presumably lead to the avoidance of chocolate rather than provide an explanation for cravings for chocolate, yet their endogenous biological function may have provided an alleged basis for any wrongfully presumed positive effects. The biogenic amines considered in the following sections have been discussed in the scientific and popular media in this respect.

### **4.1 Phenylethylamine**

2-Phenylethylamine, or  $\beta$ -phenylethylamine (PEA), is the basic molecule or structure for all compounds that make up the PEA family. This includes the stimulant and hallucinogenic substances amphetamine and mescaline, and the endogenous neurotransmitters dopamine, adrenalin and noradrenalin (Passmore and Robson 1970). Although it has been assumed that chocolate contains large amounts of PEA (e.g. 6 mg/100 g according to the British Food Manufacturing Industries Research Association, cited in Sandler et al. 1974), more recent works suggest much smaller amounts (Koehler and Eitenmiller 1978; Ingles et al. 1985; Hurst and Toomey 1981, with a maximum observed concentration of 0.66 mg/100 g for one particular (milk) chocolate sample – Hurst and Toomey 1981).

Endogenously, PEA occurs in minute quantities (single nanograms per gram of nervous tissue) in the mammalian brain, where it is synthesised by decarboxylation of phenylalanine, almost certainly in dopaminergic neurones. It appears to coexist in the brain with dopamine, and is proposed to be a modulator of catecholamine neurotransmission, though it is rapidly metabolised by MAO type B (Paterson et al. 1990).

Although low levels of endogenous PEA have been linked to depression and high levels have been linked with mania, the evidence for this is mixed and inconclusive (Davis and Boulton 1994). Even so, Liebowitz and Klein (1979) identified an affective disorder involving atypical depression and attention-seeking behaviour (“hysteroid dysphoria”) and linked this to an abnormal regulation of PEA. Whilst the authors did not refer to any published evidence, they claimed that “depressed, hysteroid dysphorics often binge on chocolate, which is loaded with phenylethylamine”, and that the production of PEA is “stimulated by positive life events”. Moreover, PEA has been linked to the euphoric feelings that are part of courtship and sexual activity, mainly on the basis of animal experiments where PEA was injected into the brain (Kohl and Francoeur 1995, after Liebowitz 1983; see also Crenshaw 1996). This, in combination with the notion that PEA is the basic structure of all amphetamines, has led the popular media to link PEA with romance, love and sex, branding PEA a “love drug”, making chocolate a “sex substitute”.



Obviously, oral consumption and cerebral injection are entirely different modes of administration, and the idea that people eat chocolate to feel “sexier” or more “sensual” because eating chocolate raises endogenous PEA is simply a myth. However, overlooking this distinction may have been used as a convenient tool for the popular media to promote chocolate as a “sex substitute”, a message further reinforced when a calculation error resulted in suspiciously high PEA concentrations in chocolate (Hamilton 1992; a value of 660 mg/100 g chocolate miscalculated by a factor of 1,000 from either Table 9 in Hurst and Toomey 1981 or from Table 3 in Hurst et al. 1982, same data). Note that PEA is still freely used to commercially promote the sales of PEA as a nutraceutical, e.g. <http://www.americannutrition.com/store/Nootropics.html>, accessed 6 August 2009.

On the basis of the evidence available, it is very doubtful that oral intake of PEA causes any beneficial psychopharmacological effects. Indeed, when assessing the effects of a large variety of synthesised amphetamines, administered (usually orally) in various doses, Shulgin and Shulgin (1991) were surprised to find that only PEA did not induce any subjective effects, either orally (200–1,600 mg) or intravenously (25–50 mg). Clearly, PEA needs side groups to function as an active amphetamine, and these findings further substantiate the “PEA myth” of chocolate.

## 4.2 *Tyramine*

Tyramine is present in a variety of foods, but its levels in chocolate are relatively low and are akin to those of PEA (Koehler and Eitenmiller 1978; Ingles et al. 1985; Hurst and Toomey 1981). Like PEA, tyramine has also been implicated in migraine attacks, and in the “cheese reaction” (*tyros* is Greek for “cheese”; Passmore and Robson 1970): prescribed in the late 1950s and the 1960s for depression and hypertension, MOA inhibitors made patients sensitive to the toxic effects of tyramine, found in some cheeses in relatively high amounts – up to 62.5 mg/100 g was measured by Ingles et al. (1985) and ten Brink et al. (1990). Symptoms of the “cheese reaction” included hypertensive crisis and severe headache, sometimes even leading to intracranial bleeding or cardiac failure (Joosten 1988). However, there appears to be no published evidence suggesting any beneficial effects of tyramine on mood and behaviour.

## 4.3 *Serotonin and Tryptophan*

As a neurotransmitter in the CNS and the peripheral nervous system, serotonin (5-hydroxytryptophan) plays an important role in the regulation of mood and behaviour (Young 1993). Although it has been identified in a range of foods, bananas, pineapples and chocolate contain somewhat higher than average concentrations (2.5, 4.2 and 2.7 mg/100 g – averages calculated from Smith 1981; Marley

and Blackwell 1970; Hurst and Toomey 1981), although the highest concentrations of serotonin have been found in walnuts (55 mg/100g; Smith 1981). Note that as for all biogenic amines, also serotonin is metabolised rapidly after oral intake, and consumption of foods containing serotonin will not directly affect brain levels of serotonin. This fits with the observation that cravings for walnuts are not common, certainly when compared with the prevalence of cravings for chocolate.

As a *precursor* of serotonin, the amino acid tryptophan is *not* prone to deamination. However, large pharmacological doses of tryptophan (much larger than our normal dietary intake of 1–1.5 g/day) can be an effective antidepressant (Young et al. 1986), which is consistent with the idea that a deficit in serotonergic activity is important in the vulnerability to depression (Maes and Meltzer 1995). Likewise, tryptophan has shown improvements in depressive symptoms in seasonal affective disorder (McGrath et al. 1990) and premenstrual syndrome (Steinberg et al. 1986), and people prone to depression show deteriorated mood following the administration of tryptophan-depleted mixtures of amino acids (Young et al. 1986; Benkelfat et al. 1994). Although these studies suggest a clear role for the serotonergic system in the cause of depression, altered brain levels of tryptophan and therefore serotonin are not expected to occur when tryptophan is consumed through the regular diet owing to competition for uptake into the brain with other large neutral amino acids (Young 1993; Rogers 1995). It is therefore extremely unlikely that any mood changes that may arise from the consumption of chocolate are caused by its tryptophan content.

## 5 Anandamide

Anandamide (arachidonyl ethanolamide), an endogenous ligand for the cannabinoid receptor that binds competitively to brain cannabinoid receptors, has been identified in minute concentrations (0.05 µg/g) in chocolate, where this compound is contained in the cocoa solids, as its presence was not confirmed in white chocolate (di Tomaso et al. 1996). Unsubstantiated, the authors suggest that anandamides present in food might “heighten sensitivity and produce euphoria” and in doing so, intensify the orosensory effects of chocolate. However, the bioavailability of anandamide is no more than 5% (Di Marzo et al. 1998). Note also that  $\Delta^9$ -tetrahydrocannabinol, the main psychoactive compound in cannabis, showed a noticeable “high” in human volunteers at doses as low as 18.77 µg/kg body weight (equivalent to 1.3 mg for a 70-kg person) (Perez-Reyes et al. 1973). It then follows that, even if one were to make the generous assumption that anandamide is as bioavailable, stable and potent (magnitude of drug effect) as  $\Delta^9$ -tetrahydrocannabinol, a blood plasma concentration of 18.77 µg/kg body weight can only be achieved by consuming 25 kg chocolate in a single sitting – a most uncomfortable, if not impossible task with potentially lethal consequences. This therefore also contradicts the suggestion of di Tomaso et al. that their findings “point to an unexpected link between non-drug craving and the endogenous cannabinoid system”. The fact that a cannabis user tried to convince the court of having consumed “a massive

amount of chocolate” in defence against the accusation of using and supplying cannabis (this involved a positive routine urine test; Tytgat et al. 2000) only confirms how the discovery of di Tomaso et al. resulted in yet another myth about our liking and cravings for chocolate.

## 6 Salsolinol and Tetrahydro- $\beta$ -carbolines

Salsolinol (SAL) and tetrahydro- $\beta$ -carbolines (THBCs) are neuroactive alkaloids generated endogenously following the consumption of alcohol through a reaction between the primary alcohol metabolite acetaldehyde and dopamine to create SAL,<sup>1</sup> or between acetaldehyde and indoleamines (e.g. serotonin, tryptamine, tryptophan) to create THBCs (Quertemont et al. 2005). Both SAL (Haber et al. 2002) and THBCs (Myers 1989) have been implied as an important factor in alcoholism, and investigated as such (Quertemont et al. 2005).

Additionally, SAL and THBCs have been identified in chocolate: SAL has been found in milk and dark chocolate and cocoa at 5, 20 and 25 mg/kg respectively; Melzig et al. 2000), whilst THBCs were identified in comparable though slightly lower amounts (1.4, 5.5 and 3.3 mg/kg respectively; Herraiz et al. 1993). In part driven by their implication in alcoholism, SAL (Melzig et al. 2000) and THBCs (Herraiz 2000) have independently been named as potentially involved in cravings for chocolate.

Again, also here, a role for SAL and THBCs in the cause of chocolate cravings would require that their consumption results in raised blood plasma levels of these compounds. Unfortunately, the literature is not clear whether this occurs or not. Even if they could be freely absorbed, THBCs are also mild MAO inhibitors (see Sect. 4 for MAO inhibition), potentially amplifying the effects of biogenic amines in chocolate and thereby contributing to migraines following the consumption of THBCs (Baker et al. 1987 in Herraiz 2000; Herraiz and Chaparro 2006) or SAL (Heikkilla et al. 1971 in Melzig et al. 2000) in chocolate. Furthermore, although SAL reportedly shows positive effects on heart rate and muscle contractions (Sokolova et al. 1990 and Chavez-Lara et al. 1989, respectively, in Melzig et al. 2000), again the route of administration is not clear. Finally, a particular THBC has been found in the tubers of the South American maca plant (*Lepidium meyenii* Walp.). Whilst this plant has been ascribed various therapeutic benefits, and is used by athletes as an alternative to anabolic steroids (Brack Egg 1999 in Piacente et al. 2002), it is perfectly possible that these effects are related to one or more of the other maca constituents, especially glucosinolates.

On the basis of the evidence available, it is unlikely that chocolate cravings can be induced by SAL or THBCs. Like PEA, tyramine and anandamide, SAL and

---

<sup>1</sup>SAL is the most widely researched example of a tetrahydroisoquinoline; tetrahydroisoquinolines are formed from acetaldehyde and catecholamines (Quertemont et al. 2005)

THBCs may have to be added to the ever-growing list of myths surrounding this topic as there is no direct evidence indicating biological activity through oral intake of either substance.

## 7 Magnesium

Chocolate has been mentioned as a relevant source of dietary magnesium (Gibson 1990). Indeed, according to some publications, it has one of the highest magnesium levels of all foods listed (Seelig 1989; Rozin et al. 1991). Moreover, magnesium therapy has been claimed to reduce premenstrual tension (Abraham 1980) and to reduce chocolate cravings in women on hormone replacements (Roach 1989; A. Weil, personal communication). Although this appears to indicate an explanation for “why women crave for chocolate at that particular time of the month”, the following findings need to be taken into consideration before any such claims can be made.

First of all, dark chocolate contains 90–100 mg magnesium/100 g, whilst milk chocolate magnesium levels are slightly lower, at 43–50 mg magnesium/100 g (FSA 2002; Souci et al. 1986). Although chocolate may have the potential to contribute to the dietary intake of magnesium, and to even counteract magnesium deficiency, other foods contain similar or even much larger amounts of magnesium, for example Brazil nuts (410 mg/100 g), roasted and salted cashew nuts (250 mg/100 g), peanuts (210 mg/kg) and All-Bran cereal (240 mg/100 g) (FSA 2002). Interestingly, cravings for these foods do not appear to be very common in sufferers from premenstrual tension, as is confirmed by the observation that chocolate is the main target in female food cravings (Hill and Heaton-Brown 1994; Rodin et al. 1991), followed by ice-cream (Rodin et al. 1991). Indeed, “the food cravings reported. . . were hunger-reducing, mood-improving experiences, directed at wanting to consume highly pleasant tasting food” (Hill and Heaton-Brown 1994).

Therefore, despite speculated associations between changes in mood, food preferences and the menstrual cycle (Bancroft et al. 1988; Wurtman and Wurtman 1989), there is no reliable evidence to suggest that magnesium-deficient people show an increased craving or liking for chocolate.

## 8 Conclusions and Considerations

Most of the pharmacologically active substances present in chocolate that have been highlighted by both scientists and the popular media do not exude an effect in man owing to extremely low concentrations, the inability to cross or even reach the blood–brain barrier, or other inabilities that may at times have been conveniently ignored in order to justify a message that appeals to the general public. Of all constituents proposed to play a role in why we like chocolate over and above its

innate appeal as a sweet and creamy tasting food (Drewnowski and Greenwood 1983), caffeine appears to provide the clearest evidence, based on effects found with ecologically relevant doses (Smit and Rogers 2001). Although theobromine is a promising “candidate”, synergistic or detrimental interactions as found in animal research (Heim et al. 1971; He et al. 2009) cannot be ruled out and need to be investigated with doses relevant to generally consumed amounts, as well as the possibility that some people are much more sensitive to the effects of theobromine than others (Mumford et al. 1994; Ott 1985). Research ought to include caffeine as a comparative agent, and underlying mechanisms need to be further explored, especially in the case of theobromine as an antitussive (Usmani et al. 2005) or as a dental enamel protective (Sadeghpour 2007) agent. In animals, theobromine is much better researched, despite this being fairly limited to toxicology studies. Additionally, a range of case studies of animal poisoning point out the dangers of chocolate and other cocoa-containing products to a wide range of animal species. Finally, doping control and therefore potential disqualifications are an issue in animal racing sports. However, the effects found in animals, whether they be of a toxicological, behavioural or other nature, cannot necessarily be translated to the human system (Tarka et al. 1979; Miller et al. 1984).

Clearly, the focus of interest regarding the effects of theobromine has changed over time, and new developments are promising an interesting future for a much underresearched substance. Hence, theobromine is in need of further investigation, and the following points need to be addressed:

1. A theobromine-to-caffeine affinity ratio for adenosine receptors of possibly 1:10 is not compensated for the estimated 10:1 theobromine-to-caffeine prevalence ratio in chocolate in terms of its mood, mental performance, or subjective effects.
2. It appears some individuals may be sensitive to the subjective effects of theobromine, though this needs validating, and a theoretical basis for this needs to be established.
3. Although some interactive effects of caffeine and theobromine have been observed, and although the effects of low doses of caffeine and of relevant caffeine–theobromine combinations on mood and performance have been found, effects of the individual components in relation to their combination have not yet been reported.
4. Although antitussive and enamel-strengthening effects of theobromine may have been found, similar effects of cocoa or chocolate ingestion, or comparative effects with caffeine and maybe other methylxanthines need to be investigated, in part to gain better insight into the possible mechanisms involved.

Note that although the methylxanthines in chocolate appear to represent its pharmacological activity (Smit et al. 2004), the list of minor chocolate constituents presented here is not exhaustive, nor does it address potential interactive effects between compounds that are not explained by their individual effects (Perez-Reyes et al. 1973). Moreover, although pharmacological activity *can* play a role in the liking for a food (see Smit and Blackburn 2005 for the combination of caffeine and theobromine as an example of their role in the liking for chocolate), this does not

translate into *cravings* for such a food. Indeed, Michener and Rozin (1994) showed that only the sensory experience of a food, and not the pharmacologically active constituents, could fulfil such cravings. Note also that cravings for chocolate are usually directed at milk chocolate, containing lower quantities of the active constituents, and that people rarely describe strong urges for the consumption of coffee and tea, even when caffeine intake is reduced because of changes in daily routine (Rogers and Dernoncourt 1998). Finally, unlike chocolate, caffeine intake is rarely resisted (“dietary restraint”). Taking these general observations and experimental findings into account, we must seek the most plausible explanation for the existence of cravings for chocolate and even chocolate “addiction” in a culturally determined ambivalence towards chocolate (Cartwright and Stritzke 2008; Smit and Rogers 2001; Rogers and Smit 2000), and not in a role for any chocolate constituents.

Summarising, despite the assumption of being a behaviourally inert substance, theobromine has shown a range of interesting effects, both in man and in other animal species. Novel findings may have caused a renewed interest in this caffeine-related compound, and much is yet to be clarified. Also, with regard to my personal interest in the psychopharmacological effects of chocolate, I can only conclude that the last word on theobromine has not yet been heard.

## References

- Abraham GE (1980) Premenstrual tension. *Curr Probl Obstet Gynecol* 3:1–39
- Apgar JL, Tarka SM Jr (1998) Methylxanthine composition and consumption patterns of cocoa and chocolate products. In: Spiller GA (ed) *Caffeine*, 1st edn. CRC, Boca Raton
- Apgar JL, Tarka SM Jr (1999) Methylxanthines. In: Knight I (ed) *Chocolate & cocoa: health and nutrition*, 1st edn. Blackwell Science, Oxford
- Arnaud M (2010) Pharmacokinetics and metabolism of natural methylxanthines in animal and man. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Ashihara H, Sano H, Crozier A (2008) Caffeine and related purine alkaloids: biosynthesis, catabolism, function and genetic engineering. *Phytochemistry* 69:841–856
- Ashihara H, Kato M, Crozier A (2010) Distribution, biosynthesis and catabolism of methylxanthines in plants. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Askar A, Morad MM (1980) Lebensmittelvergiftung. I. Toxine in Natürlichen Lebensmitteln. *Alimenta* 19:59–66
- Azam S, Hadi N, Khan NU et al (2003) Antioxidant and prooxidant properties of caffeine, theobromine and xanthine. *Med Sci Monit* 9:BR325–BR330
- Balogh A, Harder S, Vollandt R et al (1992) Intra-individual variability of caffeine elimination. *Int J Clin Pharmacol* 30:383–387
- Bancroft J, Cook A, Williamson L (1988) Food craving, mood and the menstrual cycle. *Psychol Med* 18:855–860
- Bara A, Barley E (2001) Caffeine for asthma. *Cochrane Database Syst Rev* (4) Art No CD001112. doi:001110.001002/14651858
- Barcz E, Sommer E, Janik P et al (2000) Adenosine receptor antagonism causes inhibition of angiogenic activity of human ovarian cancer cells. *Oncol Rep* 7:1285–1291
- Barcz E, Sommer E, Sokolnicka I et al (1998) The influence of theobromine on angiogenic activity and proangiogenic cytokines production of human ovarian cancer cells. *Oncol Rep* 5:517–520

- Barnes PJ, Pauwels RA (1994) Theophylline in the management of asthma: time for reappraisal? *Eur Respir J* 7:579–591
- Baron AM, Donnerstein RL, Samson RA et al (1999) Hemodynamic and electrophysiologic effects of acute chocolate ingestion in young adults. *Am J Cardiol* 84:370–373
- Becker AB, Simons KJ, Gillespie CA et al (1984) The bronchodilator effects and pharmacokinetics of caffeine in asthma. *N Engl J Med* 310:743–746
- Belvisi MG, Patel HJ, Freund-Michel V et al (2008) Inhibitory activity of the novel CB2 receptor agonist, GW833972A, on guinea-pig and human sensory nerve function in the airways. *Br J Pharmacol* 155:547–557
- Benkelfat C, Ellenbogen MA, Dean P et al (1994) Mood-lowering effect of tryptophan depletion. *Arch Gen Psychiatry* 51:687–697
- Black DJG, Barron NS (1943) Observations on the feeding of a cacao waste product to poultry. *Vet Rec* 55:166–167
- Blakemore F, Shearer GD (1943) The poisoning of livestock by cacao products. *Vet Rec* 55:165
- Bonati M, Latini R, Sadurska B et al (1984) Kinetics and metabolism of theobromine in male rats. *Toxicol* 30:327–341
- Bonvehi JS, Coll FV (2000) Evaluation of purine alkaloids and diketopiperazines contents in processed cocoa powder. *Eur Food Res Technol* 210:189–195
- Brown RA, Spina D, Page CP (2008) Adenosine receptors and asthma. *Br J Pharmacol* 153: S446–S456
- Brunetto MdR, Gutiérrez L, Delgado Y et al (2007) Determination of theobromine, theophylline and caffeine in cocoa samples by a high-performance liquid chromatographic method with on-line sample cleanup in a switching-column system. *Food Chem* 100:459–467
- Budhraj A, Camargo FC, Hughes C et al (2007) Caffeine and theobromine identifications in post-race urines: threshold levels and regulatory significance of such identifications. *AAEP Proc* 53:87–94
- Burdock GA, Carabin IG, Crincoli CM (2009) Safety assessment of kola nut extract as a food ingredient. *Food Chem Toxicol* 47:1725–1732
- Cardozo EL Jr, Cardozo-Filho L, Filho OF et al (2007) Selective liquid CO<sub>2</sub> extraction of purine alkaloids in different *Ilex paraguariensis* progenies grown under environmental influences. *J Agric Food Chem* 22:6835–6841
- Carney JM, Holloway FA, Modrow HE (1985) Discriminative stimulus properties of methylxanthines and their metabolites in rats. *Life Sci* 36:913–920
- Cartwright F, Stritzke WG (2008) A multidimensional ambivalence model of chocolate craving: construct validity and associations with chocolate consumption and disordered eating. *Eat Behav* 9:1–12
- Chauchard P, Mazoué H, Lecoq R (1945) Inhibition par les sucres de l'effet excitant qu'exercent les bases puriques sur le système nerveux. *Soc Biol Seance* 12–13
- Chorostowska-Wynimko J, Skopinska-Rózewska E, Sommer E et al (2004) Multiple effects of theobromine on fetus development and postnatal status of the immune system. *Int J Tissue React* 26:53–60
- Chung KF, Chang AB (2002) Therapy for cough: active agents. *Pulm Pharmacol Ther* 15:335–338
- Chung KF, Pavord ID (2008) Prevalence, pathogenesis, and causes of chronic cough. *Lancet* 371:1364–1374
- Cornish HH, Christman AA (1957) A study of the metabolism of theobromine, theophylline, and caffeine in man. *J Biol Chem* 228:315–323
- Craig WJ, Nguyen TT (1984) Caffeine and theobromine levels in cocoa and carob products. *J Food Sci* 49:302–305
- Crenshaw TL (1996) *Why we love and lust: how our sex hormones influence our relationships*. HarperCollins, London
- Curtis PE, Griffiths JE (1972) Suspected chocolate poisoning of calves. *Vet Rec* 90:313–314
- Czok G (1974) Zur Frage der biologischen Wirksamkeit von Methylxanthinen in Kakaoprodukten. *Z Ernahrungswiss* 13:165–171

- Daly JW, Butts-Lamb P, Padgett W (1983) Subclasses of adenosine receptors in the central nervous system: interaction with caffeine and related methylxanthines. *Cell Mol Neurobiol* 3:69–80
- Davis BA, Boulton AA (1994) The trace amines and their acidic metabolites in depression - an overview. *Prog Neuropsychopharmacol Biol Psychiatry* 18:17–45
- de Vries JW, Johnson KD, Heroff JC (1981) HPLC determination of caffeine and theobromine content of various natural and red dutched cocoas. *J Food Sci* 46:1968–1969
- Delbeke FT, Debackere M (1991) Urinary excretion of theobromine in horses given contaminated pelleted food. *Vet Res Commun* 15:107–116
- Di Marzo V, Sepe N, De Petrocellis L et al (1998) Trick or treat from food endocannabinoids? *Nature* 396:636
- di Tomaso E, Beltramo M, Piomelli D (1996) Brain cannabinoids in chocolate. *Nature* 382:677–678
- Dock W (1926) The use of theobromine for pain of arteriosclerotic origin. *Cal West Med* 25:636–638
- Dorfman LJ, Jarvik ME (1970) Comparative stimulant and diuretic actions of caffeine and theobromine in man. *Clin Pharmacol Ther* 11:869–872
- Dorland's Illustrated Medical Dictionary (2007) Saunders Elsevier, Philadelphia
- Dowden HC (1938) Note on the quantity of theobromine in the milk of cows fed on a diet including this alkaloid. *Biochem J* 32:71–73
- Drouillard DD, Vesell ES, Dvorchik BH (1978) Studies on theobromine disposition in normal subjects. *Clin Pharmacol Ther* 23:296–302
- Dunnett M, Lees P (2003) Trace elements, toxin and drug elimination in hair with particular reference to the horse. *Res Vet Sci* 75:89–101
- El Yacoubi M, Ledent C, Parmentier M et al (2005) Reduced appetite for caffeine in adenosine A<sub>2A</sub> receptor knockout mice. *Eur J Pharmacol* 519:290–291
- El-Bitar MK, Boustany RM (2009) Common causes of uncommon seizures. *Pediatr Neurol* 41:83–87
- Estfan B, Walsh D (2008) The cough from hell: diazepam for intractable cough in a patient with renal cell carcinoma. *J Pain Symptom Manage* 36:553–558
- European Food Safety Authority (2008) Theobromine as undesirable substances in animal feed. *EFSA J* 725:1–66
- European Pharmacopoeia 5.0 (2005) Monographs: 0298 Theobromine:2554
- Food Standards Agency (2002) McCance and Widdowson's the composition of foods. Sixth summary edition. Royal Society of Chemistry, Cambridge
- Fredholm BB, Bättig K, Holmén J et al (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fredholm BB, Lindström K (1999) Autoradiographic comparison of the potency of several structurally unrelated adenosine receptor antagonists at adenosine A<sub>1</sub> and A<sub>2A</sub> receptors. *Eur J Pharmacol* 380:197–202
- Friedman L, Weinberger MA, Farber TM et al (1979) Testicular atrophy and impaired spermatogenesis in rats fed high levels of the methylxanthines caffeine, theobromine, or theophylline. *J Environ Path Toxicol* 2:287–706
- Funabashi H, Fujioka M, Kohchi M et al (2000) Collaborative work to evaluate toxicity on male reproductive organs by repeated dose studies in rats 22). Effects of 2- and 4-week administration of theobromine on the testis. *J Toxicol Sci* 25:211–221
- Gans JH (1982) Dietary influences on theobromine-induced toxicity in rats. *Toxicol Appl Pharmacol* 63:312–320
- Gans JH (1984) Comparative toxicities of dietary caffeine and theobromine in the rat. *Food Chem Toxicol* 22:365–369
- Gans JH, Korson R, Cater MR et al (1980) Effects of short-term and long-term theobromine administration to male dogs. *Toxicol Appl Pharmacol* 53:481–496
- Gartrell BD, Reid C (2007) Death by chocolate: a fatal problem for an inquisitive wild parrot. *N Z Vet J* 55:149–151



- Gates S, Miners JO (1999) Cytochrome P450 isoform selectivity in human hepatic theobromine metabolism. *Br J Clin Pharmacol* 47:299–305
- Geraets L, Moonen HJ, Wouters EF et al (2006) Caffeine metabolites are inhibitors of the nuclear enzyme poly(ADP-ribose)polymerase-1 at physiological concentrations. *Biochem Pharmacol* 72:902–910
- Giannandrea F (2009) Correlation analysis of cocoa consumption data with worldwide incidence rates of testicular cancer and hypospadias. *Int J Environ Res Public Health* 6:568–578
- Gibson RS (1990) Assessment of calcium, phosphorus, and magnesium status. Principles of nutritional assessment. Oxford University Press, Oxford
- Gil M, Skopinska-Różewska E, Radomska D et al (1993) Effect of purinergic receptor antagonists suramin and theobromine on tumor-induced angiogenesis in balb/c mice. *Folia Biol (Praha)* 39:63–68
- Grassi D, Lippi C, Necozione S et al (2005) Short-term administration of dark chocolate is followed by a significant increase in insulin sensitivity and a decrease in blood pressure in healthy persons. *Am J Clin Nutr* 81:611–614
- Gunning OV (1950) Theobromine poisoning in ducks due to the feeding of cacao waste products. *Br Vet J* 106:31–32
- Haber H, Jahn H, Ehrenreich H et al (2002) Assay of salsolinol in peripheral blood mononuclear cells of alcoholics and healthy subjects by gas chromatography-mass spectrometry. *Addict Biol* 7:403–407
- Halfdanarson TR, Jatoti A (2007) Chocolate as a cough suppressant: rationale and justification for an upcoming clinical trial. *Support Cancer Ther* 4:119–122
- Hamilton S (1992) Why the lady loves C<sub>6</sub>H<sub>5</sub>(CH<sub>2</sub>)<sub>2</sub>NH<sub>2</sub>. *New Sci* 132:26–28
- Hammerstone JF Jr, Romanczyk LJ Jr, Aitken WM (1994) Purine alkaloid distribution within *Herrania* and *Theobroma*. *Phytochem* 35:1237–1240
- Hannig C, Sorg J, Spitzmüller B et al (2009) Polyphenolic beverages reduce initial bacterial adherence to enamel in situ. *J Dent* 37:560–566
- Hansen PB, Schnermann J (2003) Vasoconstrictor and vasodilator effects of adenosine in the kidney. *Am J Physiol Renal Physiol* 285:F590–F599
- Haywood PE, Teale P, Moss MS (1990) The excretion of theobromine in thoroughbred racehorses after feeding compounded cubes containing cocoa husk establishment of a threshold value in horse urine. *Equine Vet J* 22:244–246
- He R, Xie G, Yao X-S et al (2009) Effect of cocoa tea (*Camellia pitlophylla*) co-administrated with green tea on ambulatory behaviors. *Biosci Biotechnol Biochem* 73:957–960
- Heim F, Hach B, Mitznegg P et al (1971) Coffein-antagonistische Wirkungen des Theobromins und coffeinartige Eigenschaften von Theobromin-Metaboliten. *Arzneimittelforschung (Drug Res)* 21:1039–1043
- Herraiz T (2000) Tetrahydro-β-carbolines, potential neuroactive alkaloids, in chocolate and cocoa. *J Agric Food Chem* 48:4900–4904
- Herraiz T, Chaparro C (2006) Human monoamine oxidase enzyme inhibition by coffee and h-carbolines norharman and harman isolated from coffee. *Life Sci* 78:795–802
- Herraiz T, Huang Z, Ough CS (1993) 1,2,3,4-Tetrahydro-β-carboline-3-carboxylic acid and 1-methyl-1,2,3,4-tetrahydro-β-carboline-3-carboxylic acid in wines. *J Agric Food Chem* 41:455–459
- Hicks MB, Hsieh Y-HP, Bell LN (1996) Tea preparation and its influence on methylxanthine concentration. *Food Res Int* 29:325–330
- Hill AJ, Heaton-Brown L (1994) The experience of food craving; a prospective investigation in healthy women. *J Psychosom Res* 38:801–814
- Hovda LR, Kingston RL (1994) Cocoa bean mulch poisoning in dogs. *Vet Hum Toxicol* 35:357
- Hurst WJ, Martin RA, Zoumas BL et al (1982) Biogenic amines in chocolate - a review. *Nutr Rep Int* 26:1081–1086
- Hurst WJ, Toomey PB (1981) High-performance liquid chromatographic determination of four biogenic amines in chocolate. *Analyst* 106:394–402

- IARC (1991) Theobromine. IARC Monogr Eval Carcinog Risks Hum 51:421–441
- Ingles DL, Back JF, Gallimore D et al (1985) Estimation of biogenic amines in foods. *J Sci Food Agric* 36:402–406
- International Federation of Horseracing Authorities (2007) International agreement on breeding, racing and wagering. Article 6. [http://www.horseracingintfed.com/resources/2007\\_choose\\_eng.pdf](http://www.horseracingintfed.com/resources/2007_choose_eng.pdf). Accessed 12 Jun 2009
- Ishay JS, Paniry VA (1979) Effects of caffeine and various xanthines on hornets and bees. *Psychopharmacology* 65:299–309
- James JE (1991) Caffeine and health. Academic, London
- Jansson DS, Galgan V, Schubert B et al (2001) Theobromine intoxication in a red fox and a European badger. *J Wildl Dis* 37:362–365
- Johnston JJ (2005) Evaluation of cocoa- and coffee-derived methylxanthines as toxicants for the control of pest coyotes. *J Agric Food Chem* 53:4069–4075
- Joosten HMLJ (1988) The biogenic amine contents of Dutch cheese and their toxicological significance. *Neth Milk Dairy J* 42:25–42
- Jourdain C, Tenca G, Deguercy A et al (2006) In-vitro effects of polyphenols from cocoa and beta-sitosterol on the growth of human prostate cancer and normal cells. *Eur J Cancer Prev* 15:353–361
- Kashket S, Paolino VJ, Lewis DA et al (1985) In-vitro inhibition of glucosyltransferase from the dental plaque bacterium *Streptococcus mutans* by common beverages and food extracts. *Arch Oral Biol* 30:821–826
- Kelly CJ (2005) Effects of theobromine should be considered in future studies. *Am J Clin Nutr* 82:486–487
- Koehler PE, Eitenmiller RR (1978) High pressure liquid chromatographic analysis of tyramine, phenylethylamine and tryptamine in sausage, cheese and chocolate. *J Food Sci* 43:1245–1247
- Kohl JV, Francoeur RT (1995) The scent of Eros. Continuum, New York
- Koyama Y, Tomoda Y, Kato M et al (2003) Metabolism of purine bases, nucleosides and alkaloids in theobromine-forming *Theobroma cacao* leaves. *Plant Physiol Biochem* 41:977–984
- Kuribara H, Asahi T, Tadokoro S (1992) Behavioral evaluation of psycho-pharmacological and psychotoxic actions of methylxanthines by ambulatory activity and discrete avoidance in mice. *J Toxicol Sci* 17:81–90
- Kuribara H, Tadokoro S (1992) Behavioral effects of cocoa and its main active compound theobromine: evaluation by ambulatory activity and discrete avoidance in mice. *Jpn J Alcohol Drug Depend* 27:168–179
- Landau SI (ed) (1986) International dictionary of medicine and biology. Wiley, New York
- Lee KW, Kundu JK, Kim SO et al (2006) Cocoa polyphenols inhibit phorbol ester-induced superoxide anion formation in cultured HL-60 cells and expression of cyclooxygenase-2 and activation of NF- $\kappa$ B and MAPKs in mouse skin in vivo. *J Nutr* 136:1150–1155
- Lelo A, Birkett DJ, Robson RA et al (1986) Comparative pharmacokinetics of caffeine and its primary demethylated metabolites paraxanthine, theobromine and theophylline in man. *Br J Clin Pharmacol* 22:177–182
- Liebowitz MR (1983) The chemistry of love. Little Brown, Boston
- Liebowitz MR, Klein DF (1979) Hysteroid dysphoria. *Psychiatr Clin North Am* 2:555–575
- Lipworth BJ, Williamson PA (2009) Beta blockers for asthma: a double-edged sword. *Lancet* 373:104–105
- Loeffler B, Kluge K, Ungemach FR et al (2000) [Concentrations of caffeine, theophylline and theobromine in plasma and urine of dogs after application of coffee, tea and chocolate and its relevance to doping]. *Tierärztl Prax (K)* 28:79–85
- Lunell E, Svedmyr N, Andersson KE et al (1983) A novel bronchodilator xanthine apparently without adenosine receptor antagonism and tremorogenic effect. *Eur J Respir Dis* 64:333–339
- Maes M, Meltzer HY (1995) The serotonin hypothesis of major depression. In: Bloom FE, Kupfer DJ (eds) *Psychopharmacology: the fourth generation of progress*. Raven, New York
- MAFF UK (1988) Food portion sizes. HMSO, London

- MAFF UK (1998) Survey of caffeine and other methylxanthines in energy drinks and other caffeine-containing products (updated). Food surveillance information sheet 144
- Marley E, Blackwell B (1970) Interactions of monoamine oxidase inhibitors, amines, and food-stuffs. *Adv Pharmacol Chemother* 8:185–239
- Massey LK, Whiting SJ (1993) Caffeine, urinary calcium, calcium metabolism and bone. *J Nutr* 123:1611–1614
- McDonald P, Edwards RA, Greenhalgh JFD et al (2002) Animal nutrition. Prentice Hall, Harlow
- McGrath RE, Buckwald B, Resnick EV (1990) The effect of L-tryptophan on seasonal affective disorder. *J Clin Psychiatry* 51:162–163
- Melzig MF, Putscher I, Henklein P et al (2000) In vitro pharmacological activity of the tetrahydroisoquinoline salsolinol present in products from *Theobroma cacao* L. like cocoa and chocolate. *J Ethnopharmacol* 73:153–159
- Merighi S, Benini A, Mirandola P et al (2007) Caffeine inhibits adenosine-induced accumulation of hypoxia-inducible factor-1 $\alpha$ , vascular endothelial growth factor, and interleukin-8 expression in hypoxic human colon cancer cells. *Mol Pharmacol* 72:395–406
- Michener W, Rozin P (1994) Pharmacological versus sensory factors in the satiation of chocolate craving. *Physiol Behav* 56:419–422
- Miller GE, Radulovic LL, Dewit RH et al (1984) Comparative theobromine metabolism in five mammalian species. *Drug Metab Dispos* 12:154–160
- Miners JO, Attwood J, Birkett DJ (1982) Theobromine metabolism in man. *Drug Metab Dispos* 10:672–675
- Miners JO, Attwood J, Wing LM et al (1985) Influence of cimetidine, sulfapyrazone, and cigarette smoking on theobromine metabolism in man. *Drug Metab Dispos* 13:598–601
- Mumford GK, Benowitz NL, Evans SM et al (1996) Absorption rate of methylxanthines following capsules, cola and chocolate. *Eur J Clin Pharmacol* 51:319–325
- Mumford GK, Evans SM, Kaminski BJ et al (1994) Discriminative stimulus and subjective effects of theobromine and caffeine in humans. *Psychopharmacology* 115:1–8
- Mumford GK, Holtzman G (1991) Qualitative differences in the discriminative stimulus effects of low and high doses of caffeine in the rat. *J Pharmacol Exp Ther* 258:857–865
- Myers RD (1989) Isoquinolines, beta-carbolines and alcohol drinking: involvement of opioid and dopaminergic mechanisms. *Experientia* 45:436–443
- Naylor MN (1984) Nutrition and dental decay. *Proc Nutr Soc* 43:257–263
- Nishino M, Ashiku SK, Kocher ON et al (2006) The thymus: a comprehensive review. *Radiographics* 26:335–348
- Ochoa-Herrera V, Banihani Q, León G et al (2009) Toxicity of fluoride to microorganisms in biological wastewater treatment systems. *Water Res* 43:3177–3186
- Ott J (1985) The cacahuatl eater. Natural Products, Vashon
- Pagano R, Negri E, Decarli A et al (1988) Coffee drinking and prevalence of bronchial asthma. *Chest* 94:386–389
- Passmore R, Robson JS (1970) A companion to medical studies in three volumes. Pharmacology, microbiology, general pathology and related subjects, vol 2. Blackwell, Oxford
- Patel HJ, Birrell MA, Crispino N et al (2003) Inhibition of guinea-pig and human sensory nerve activity and the cough reflex in guinea-pigs by cannabinoid (CB2) receptor activation. *Br J Pharmacol* 140:261–268
- Paterson IA, Juorio AV, Boulton AA (1990) 2-Phenylethylamine: a modulator of catecholamine transmission in the mammalian central nervous system? *J Neurochem* 55:1827–1837
- Percival RS, Devine DA, Duggal MS et al (2006) The effect of cocoa polyphenols on the growth, metabolism and biofilm formation by *Streptococcus mutans* and *Streptococcus sanguinis*. *Eur J Oral Sci* 114:343–348
- Perez-Reyes M, Timmons MC, Davis KH et al (1973) A comparison of the pharmacological activity in man of intravenously administered  $\Delta^9$ -tetrahydrocannabinol, cannabinol, and cannabidiol. *Experientia* 29:1368–1369
- Piacente S, Carbone V, Plaza A et al (2002) Investigation of the tuber constituents of maca (*Lepidium meyenii* Walp.). *J Agric Food Chem* 50:5621–5625

- Quertemont E, Tambour S, Tirelli E (2005) The role of acetaldehyde in the neurobehavioral effects of ethanol: a comprehensive review of animal studies. *Prog Neurobiol* 75:247–274
- Rall TW (1980) Central nervous system stimulants [continued]: the xanthines. In: Goodman Gilman A, Goodman LS, Gilman A (eds) *The pharmacological basis of therapeutics*, 6th edn. Macmillan, New York
- Rambali B, Van Anel I, Schenk E et al (2002) The contribution of cocoa additive to cigarette smoking addiction. RIVM, Bilthoven
- Reynolds JEFE (1993) *Martindale: the extra pharmacopoeia*. The Pharmaceutical Press, London
- Reynolds SM, Mackenzi AJ, Spina D et al (2004) The pharmacology of cough. *Trends Pharmacol Sci* 25:569–576
- Rieg T, Schnermann J, Vallon V (2007) Adenosine A1 receptors determine effects of caffeine on total fluid intake but not caffeine appetite. *Eur J Pharmacol* 555:174–177
- Risner CH (2008) Simultaneous determination of theobromine, (+)-catechin, caffeine, and (–)-epicatechin in standard reference material baking chocolate 2384, cocoa, cocoa beans, and cocoa butter. *J Chromatogr Sci* 46:892–899
- Roach M (1989) More reasons to love chocolate. *New Woman Febr*:135–136
- Rodin J, Mancuso J, Granger J et al (1991) Food cravings in relation to body mass index, restraint and estradiol levels: a repeated measures study in healthy women. *Appetite* 17:177–185
- Rogers PJ (1995) Food, mood and appetite. *Nutr Res Rev* 8:243–269
- Rogers PJ, Derroncourt C (1998) Regular caffeine consumption: a balance of adverse and beneficial effects for mood and psychomotor performance. *Pharmacol Biochem Behav* 59:1039–1045
- Rogers PJ, Richardson NJ, Elliman NA (1995) Overnight caffeine abstinence and negative reinforcement of preference for caffeine-containing drinks. *Psychopharmacology* 120:457–462
- Rogers PJ, Smit HJ (2000) Food craving and food “addiction”: A critical review of the evidence from a biopsychosocial perspective. *Pharmacol Biochem Behav* 66:3–14
- Rozin P, Levine E, Stoess C (1991) Chocolate craving and liking. *Appetite* 17:199–212
- Ryzhov S, McCaleb JL, Goldstein AE et al (2007) Role of adenosine receptors in the regulation of angiogenic factors and neovascularization in hypoxia. *J Pharmacol Exp Ther* 320:565–572
- Ryzhov S, Novitskiy SV, Zaynagetdinov R et al (2008) Host A(2B) adenosine receptors promote carcinoma growth. *Neoplasia* 10:987–995
- Sadeghpour A (2007) A neural network analysis of theobromine vs. fluoride on the enamel surface of human teeth: an experimental case study with strong implications for the production of a new line of revolutionary and natural non-fluoride based dentifrices. Tulane University, New Orleans
- Salvadori MC, Rieser EM, Ribeiro Neto LM et al (1994) Determination of xanthines by high-performance liquid chromatography and thin-layer chromatography in horse urine after ingestion of guaraná powder. *Analyst* 119:2701–2703
- Sandler M, Youdim MBH, Hanington E (1974) A phenylethylamine oxidising defect in migraine. *Nature* 250:335–337
- Sarkaria JN, Busby EC, Tibbetts RS et al (1999) Inhibition of ATM and ATR kinase activities by the radiosensitizing agent, caffeine. *Cancer Res* 59:4375–4382
- Schmid TE, Eskenazi B, Baumgartner A et al (2007) The effects of male age on sperm DNA damage in healthy non-smokers. *Hum Reprod* 22:180–187
- Seelig M (1989) Cardiovascular consequences of magnesium deficiency and loss: pathogenesis, prevalence and manifestations - magnesium and chloride loss in refractory potassium repletion. *Am J Cardiol* 63:4G–21G
- Shively CA, Tarka SM Jr (1983) Theobromine metabolism and pharmacokinetics in pregnant and nonpregnant Sprague-Dawley rats. *Toxicol Appl Pharmacol* 67:376–382
- Shulgin A, Shulgin A (1991) *PIHKAL: a chemical love story*. Transform, Berkeley
- Simons FER, Becker AB, Simons KJ et al (1985) The bronchodilator effect and pharmacokinetics of theobromine in young patients with asthma. *J Allergy Clin Immunol* 76:703–707
- Skopinski P, Skopinska-Rózewska E, Kaminski A et al (2004) Chocolate feeding of pregnant mice resulted in epigallocatechin-related embryonic angiogenesis suppression and bone mineralization disorder. *Pol J Vet Sci* 7:131–133

- Slattery ML, West DW (1993) Smoking, alcohol, coffee, tea, caffeine, and theobromine: risk of prostate cancer in Utah (United States). *Cancer Causes Control* 4:559–563
- Smit HJ, Blackburn RJ (2005) Reinforcing effects of caffeine and theobromine as found in chocolate. *Psychopharmacology* 181:101–106
- Smit HJ, Gaffan EA, Rogers PJ (2004) Methylxanthines are the psycho-pharmacologically active constituents of chocolate. *Psychopharmacology* 176:412–419
- Smit HJ, Rogers PJ (2000) Effects of low doses of caffeine on cognitive performance, mood and thirst in low and higher caffeine consumers. *Psychopharmacology* 152:167–173
- Smit HJ, Rogers PJ (2001) Potentially psychoactive constituents of cocoa-containing products. In: Hetherington MM (ed) *Food cravings and addiction*. Leatherhead Food RA Publishing, Leatherhead
- Smit HJ, Rogers PJ (2007) Effects of caffeine on mood. In: Smith BD, Gupta U, Gupta BS (eds) *Caffeine and activation theory*. CRC, Boca Raton
- Smith TA (1981) Amines in food. *Food Chem* 6:169–200
- Snyder SH, Katims JJ, Annau Z et al (1981) Adenosine receptors and behavioral actions of methylxanthines. *Proc Natl Acad Sci USA* 78:3260–3264
- Soffietti MG, Nebbia C, Valenza F et al (1989) Toxic effects of theobromine on mature and immature male rabbits. *J Comp Pathol* 100:47–58
- Souci SW, Fachmann W, Kraut H (1981) *Food composition and nutrition tables 1981/1982*. Wissenschaftliche Verlagsgesellschaft, Stuttgart
- Souci SW, Fachmann W, Kraut H (1986) *Food composition and nutrition tables 1986/1987*. Wissenschaftliche Verlagsgesellschaft, Stuttgart
- Sprugel W, Mitznegg P, Heim F (1977) The influence of caffeine and theobromine on locomotive activity and the brain cGMP/cAMP ratio in white mice. *Biochem Pharmacol* 26:1723–1724
- Srikanth RK, Shashikiran ND, Subba Reddy VV (2008) Chocolate mouth rinse: effect on plaque accumulation and mutans streptococci counts when used by children. *J Indian Soc Pedod Prev Dent* 26:67–70
- Stedman's Medical Dictionary (1976). The Williams & Wilkins Company, Baltimore
- Steinberg S, Annable L, Young SN et al (1986) Tryptophan in the treatment of late luteal phase dysphoric disorder: a pilot study. *J Psychiatry Neurosci* 19:114–119
- Stidworthy MF, Bleakley JS, Cheeseman MT et al (1997) Chocolate poisoning in dogs. *Vet Rec* 141:28
- Strachan ER, Bennett A (1994) Theobromine poisoning in dogs. *Vet Rec* 134:284
- Strålfors A (1967) Effect on hamster caries by purine derivatives vanillin and some tannin-containing materials. *Arch Oral Biol* 12:321–332
- Tarka SM Jr (1982) The toxicology of cocoa and methylxanthines: a review of the literature. *CRC Crit Rev Toxicol* 9:275–312
- Tarka SM Jr, Applebaum RS, Borzelleca JF (1986a) Evaluation of the perinatal, postnatal and teratogenic effects of cocoa powder and theobromine in Sprague-Dawley/CD rats. *Food Chem Toxicol* 24:375–382
- Tarka SM Jr, Applebaum RS, Borzelleca JF (1986b) Evaluation of the teratogenic potential of cocoa powder and theobromine in New Zealand white rabbits. *Food Chem Toxicol* 24:363–374
- Tarka SM Jr, Morrissey RB, Apgar JL et al (1991) Chronic toxicity/carcinogenicity studies of cocoa powder in rats. *Food Chem Toxicol* 29:7–19
- Tarka SM Jr, Zoumas BL, Gans JH (1979) Short-term effects of graded levels of theobromine in laboratory rodents. *Toxicol Appl Pharmacol* 49:127–149
- Tarka SM Jr, Zoumas BL, Gans JH (1981) Effects of continuous administration of dietary theobromine on rat testicular weight and morphology. *Toxicol Appl Pharmacol* 58:76–82
- Taubert D, Berkels R, Roesen R et al (2003) Chocolate and blood pressure in elderly individuals with isolated systolic hypertension. *JAMA* 290:1029–1030
- ten Brink B, Damink C, Joosten JMLJ et al (1990) Occurrence and formation of biologically active amines in foods. *Int J Food Microbiol* 11:73–84
- The Merck Index (2006). Merck & Co Inc., Whitehouse Station, NJ

- Thorpe TE (1893) On the rise and development of synthetical chemistry. *Fortn Rev* 53:691–701
- Timbie DJ, Sechrist L, Keeney PG (1978) Application of high-pressure liquid chromatography to the study of variables affecting theobromine and caffeine concentrations in cocoa beans. *J Food Sci* 43(560–562):565
- Tytgat J, Mv B, Daenens P (2000) Cannabinoid mimics in chocolate utilized as an argument in court. *Int J Legal Med* 113:137–139
- Usmani OS, Belvisi MG, Patel HJ et al (2005) Theobromine inhibits sensory nerve activation and cough. *FASEB J* 19:231–233
- Vallon V, Mühlbauer B, Osswald H (2006) Adenosine and kidney function. *Physiol Rev* 86: 901–940
- van Zyl JM, Derendinger B, Seifart HI et al (2008) Comparative diffusion of drugs through bronchial tissue. *Int J Pharm* 357:32–36
- Walsh D, Donnelly S, Rybicki L (2000) The symptoms of advanced cancer: relationship to age, gender, and performance status in 1,000 patients. *Support Care Cancer* 8:175–179
- Wang X, Wan X, Hu S et al (2008) Study on the increase mechanism of the caffeine content during the fermentation of tea with microorganisms. *Food Chem* 107:1086–1091
- Wang Y, Waller DP (1994) Theobromine toxicity on Sertoli cells and comparison with cocoa extract in male rats. *Toxicol Lett* 70:155–164
- Wang Y, Waller DP, Hikim AP et al (1992) Reproductive toxicity of theobromine and cocoa extract in male rats. *Reprod Toxicol* 6:347–353
- Wasfi IA, Boni NS, Elghazali M et al (2000) The pharmacokinetics, metabolism and urinary detection time of caffeine in camels. *Res Vet Sci* 69:69–74
- Wasiutynski A, Siwicki AK, Balan BJ et al (2005) Inhibitory effect of cocoa catechins on embryonic and tumor angiogenesis in mice. *Pol J Environ Stud* 14:800–805
- Weckerle CS, Stutz MA, Baumann TW (2003) Purine alkaloids in *Paullinia*. *Phytochemistry* 64:735–742
- Weinberger MA, Friedman L, Farber TM et al (1978) Testicular atrophy and impaired spermatogenesis in rats fed high levels of the methylxanthines caffeine, theobromine, or theophylline. *J Environ Path Toxicol* 1:669–688
- Wells DJ, Hanks BM, Yarbrough CS et al (1988) Determination of methylxanthine stimulants in urine of racing greyhounds by high-performance liquid chromatography. Resolution of a contested drug administration case. *J Anal Toxicol* 12:30–32
- World Anti-Doping Agency (2008) Q&A: 2009 prohibited list. [http://www.fibt.com/fileadmin/Medical/2009-2010/QA\\_List\\_OR.pdf](http://www.fibt.com/fileadmin/Medical/2009-2010/QA_List_OR.pdf). Accessed 8 Apr 2010
- Wurtman RJ, Wurtman JJ (1989) Carbohydrates and depression. *Sci Am* 260:68–75
- Yang XR, Ye CX, Xu JK et al (2007) Simultaneous analysis of purine alkaloids and catechins in *Camellia sinensis*, *Camellia ptilophylla* and *Camellia assamica* var. *kucha* by HPLC. *Food Chem* 100:1132–1136
- Yeomans MR, Spetch H, Rogers PJ (1998) Conditioned flavor preference negatively reinforced by caffeine in human volunteers. *Psychopharmacology* 137:401–409
- Yesair DW, Branfman AR, Callahan MM (1984) Human disposition and some biochemical aspects of methylxanthines. In: Spiller GA (ed) *The methylxanthine beverages and foods. Chemistry, consumption and health effects*. Liss, New York
- Young SN (1993) The use of diet and dietary components in the study of factors controlling affect in humans: a review. *J Psychiatry Neurosci* 18:235–244
- Young SN, Pihl RO, Ervin FR (1986) The effect of altered tryptophan levels on mood and behavior in normal human males. *Clin Neuropharmacol* 9:516–518
- Zheng X-Q, Koyama Y, Nagai C et al (2004) Biosynthesis, accumulation and degradation of theobromine in developing *Theobroma cacao* fruits. *J Plant Physiol* 161:363–369
- Ziegleder G, Stojacic E, Stumpf B (1992) Vorkommen von beta-Phenylethylamin und seinen Derivaten in Kakao und Kakaoerzeugnissen. *Z Lebensm Unters Forsch* 195:235–238

# Propentofylline: Glial Modulation, Neuroprotection, and Alleviation of Chronic Pain

Sarah Sweitzer and Joyce De Leo

## Contents

1	Introduction .....	236
1.1	PPF in Neuroprotection .....	237
1.2	PPF as a Glial Modulator in Ischemia .....	237
1.3	PPF as an Antiallodynic Agent .....	239
2	Attenuation of Morphine Tolerance and Hyperalgesia in Acute and Chronic Pain .....	243
3	PPF increases GLT-1 Expression <i>In Vitro</i> and <i>In Vivo</i> : Novel Mechanisms of Action ...	244
4	Previous Clinical Data Summary .....	245
5	Conclusions .....	246
	References .....	248

**Abstract** Propentofylline is a unique methylxanthine with clear cyclic AMP, phosphodiesterase, and adenosine actions, including enhanced synaptic adenosine signaling. Both *in vitro* and *in vivo* studies have demonstrated profound neuroprotective, antiproliferative, and anti-inflammatory effects of propentofylline. Propentofylline has shown efficacy in preclinical models of stroke, opioid tolerance, and acute and chronic pain. Clinically, propentofylline has shown efficacy in degenerative and vascular dementia, and as a potential adjuvant treatment for schizophrenia and multiple sclerosis. Possible mechanisms of action include a direct glial modulation to decrease a reactive phenotype, decrease glial production and release of damaging proinflammatory factors, and enhancement of

---

This chapter is dedicated to Peter Schubert for introducing propentofylline to J. De Leo and for 23 years of scientific and personal mentorship and friendship.

S. Sweitzer

Department of Pharmacology, Physiology and Neuroscience, University of South Carolina, USC School of Medicine, Columbia, SC 29208, USA

J. De Leo (✉)

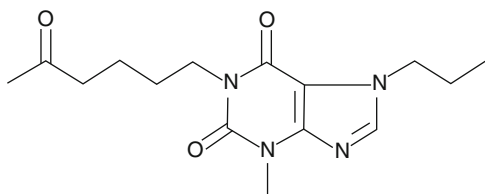
Department of Pharmacology and Toxicology, Dartmouth Medical School, Hanover, NH 03755, USA  
e-mail: joyce.a.deleo@dartmouth.edu

astrocyte-mediated glutamate clearance. This chapter reviews the literature that supports a myriad of protective actions of this small molecule and implicates propentofylline as a potential therapeutic for the treatment of chronic pain. From these studies, we propose a CNS multipartite synaptic action of propentofylline that includes modulation of pre- and postsynaptic neurons, astrocytes, and microglia in the treatment of chronic pain syndromes, including, but not limited to, neuropathic pain.

**Keywords** Astrocytes · Cytokines · Glia · Microglia · Neuropathic pain · Opioids · Tolerance

## 1 Introduction

Propentofylline (PPF) is an atypical synthetic methylxanthine [1-(5'-oxohexyl)-3-methyl-7-propylxanthine]. In this chapter, we will review the 30-year history of this compound from small-molecule development to clinical trials, describing its actions both in animals and in patients. PPF is closely related to pentoxifylline, which has been used for decades for intermittent claudication. PPF, previously known as HWA 285 (see Fig. 1), has a profile distinct from the profiles of other typical methylxanthines. Although it inhibits cyclic AMP (cAMP) and cyclic GMP phosphodiesterases, it is also a weak antagonist of the adenosine A1 receptor and blocks adenosine transport. However, the elucidation of its cellular and molecular mechanisms of action is still under way. A recent PubMed search that spanned 1981–2009 resulted in 215 publications that demonstrate PPF's diversity of actions, including, but are not limited to, vasodilatory, anti-inflammatory, antiproliferative, neuroprotective, platelet aggregation inhibitor, glial modulator, and glutamate inhibitor. These diverse actions translate into a plethora of potential clinical uses, including for ischemia, dementia, spinal cord injury, multiple sclerosis, transplantation, and chronic pain. We will take the reader on a journey highlighting the history of PPF focusing on its neuroprotection and inhibition of pain states in the quest for both a mechanism of action and a clinical translation of PPF for the treatment of chronic pain.



**Fig. 1** Chemical structure of propentofylline (PPF)



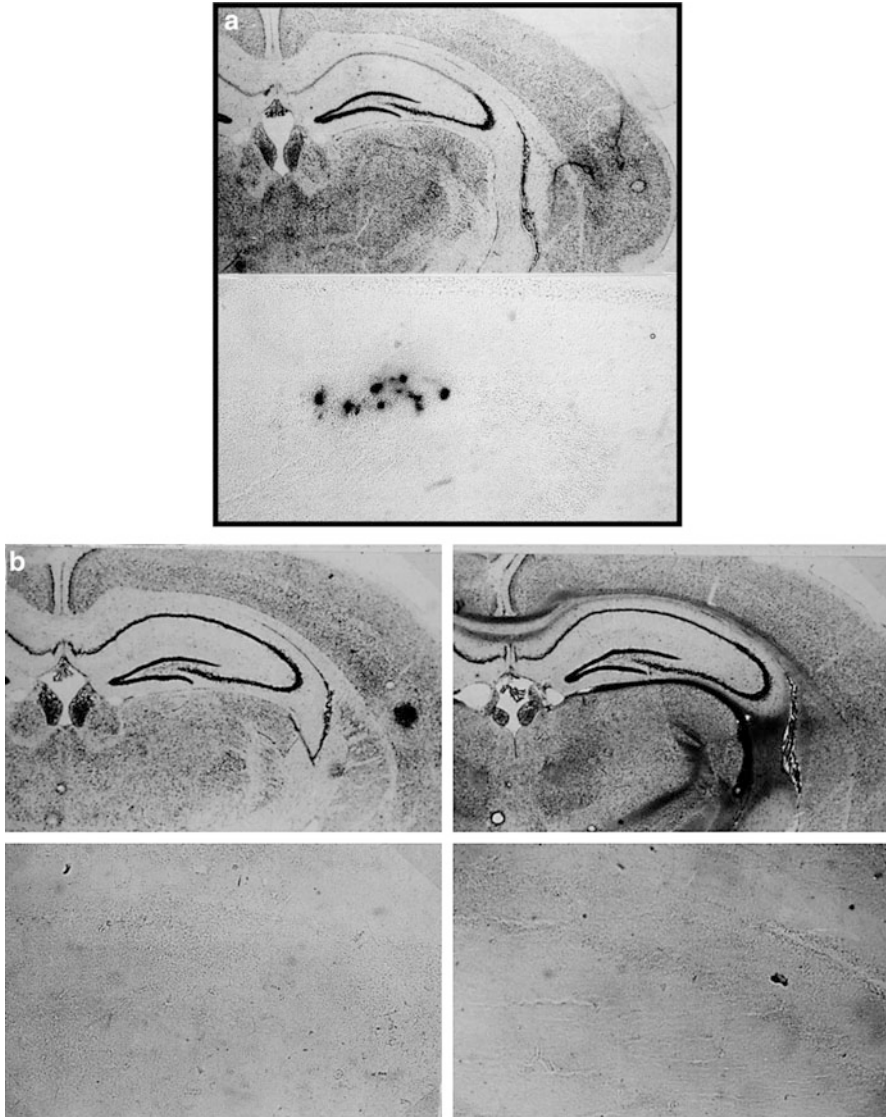
## ***1.1 PPF in Neuroprotection***

PPF is a potent inhibitor of cAMP and phosphodiesterases (Nagata et al. 1985). In the mid-1980s, PPF was found to increase cerebral blood flow without a concomitant increase in cerebral glucose utilization (Grome and Stefanovich 1985). Distinct from many classic methylxanthines, PPF also enhances the actions of adenosine partly as an inhibitor of adenosine uptake (Fredholm and Lindstrom 1986). It was postulated that PPF may reduce neuronal activity and increase the vasodilatory effects of adenosine on blood vessels. Thus, it was questioned whether such a drug would have clinical utility (Fredholm and Lindstrom 1986).

In 1987, we demonstrated a neuronal protective action of PPF in a gerbil model of cerebral ischemia (DeLeo et al. 1987). In addition to a decrease of Nissl staining in the CA1 region of the hippocampus, systemic PPF also inhibited the marked glial fibrillary acidic protein (GFAP) astrocytic response and selective calcium accumulation in the transition zone between the hippocampal CA1 and CA3 areas (see Fig. 2). The most remarkable finding of that study was that treatment with PPF exerted a protective action against postischemic damage. In an effort to investigate mechanisms of neuronal protection, the efficacy of PPF was compared with that of pentobarbital. In contrast to PPF, pentobarbital afforded no protection when administered 1 h after bilateral carotid occlusion, suggesting divergent drug mechanisms of PPF and pentobarbital (DeLeo et al. 1988b). Electrophysiological experiments demonstrated no evidence of a direct depressant action of PPF on neuronal firing (P. Schubert, unpublished data). In a later study, we determined that PPF protects hippocampal neurons against ischemic damage in the presence of the adenosine antagonist theophylline, supporting a non-adenosine-mediated action (DeLeo et al. 1988a). Interestingly, the mechanism of action of this neuroprotection has not been clearly delineated and only recently has a novel mechanism been uncovered (see Sect. 2.1).

## ***1.2 PPF as a Glial Modulator in Ischemia***

Of importance to future research directions utilizing PPF is the novel finding in 1987 that PPF decreased GFAP immunoreactivity in the CA1 region of the hippocampus following transient cerebral ischemia (Fig. 3). This was the first indication of a direct glial modulating action which would not be fully realized until almost a decade later. It was proposed by Schubert and others that pathological microglial activation contributes to progressive neuronal damage in neurodegenerative diseases by the release of potentially toxic agents and by triggering reactive astrocytic changes (Schubert et al. 1997). With use of primary neonatal cultured microglia, it was demonstrated that PPF enhanced cAMP-dependent intracellular signaling (Si et al. 1998). PPF dose-dependently inhibited lipopolysaccharide-induced release of both tumor necrosis factor  $\alpha$  (TNF $\alpha$ ) and interleukin (IL)-1 $\beta$

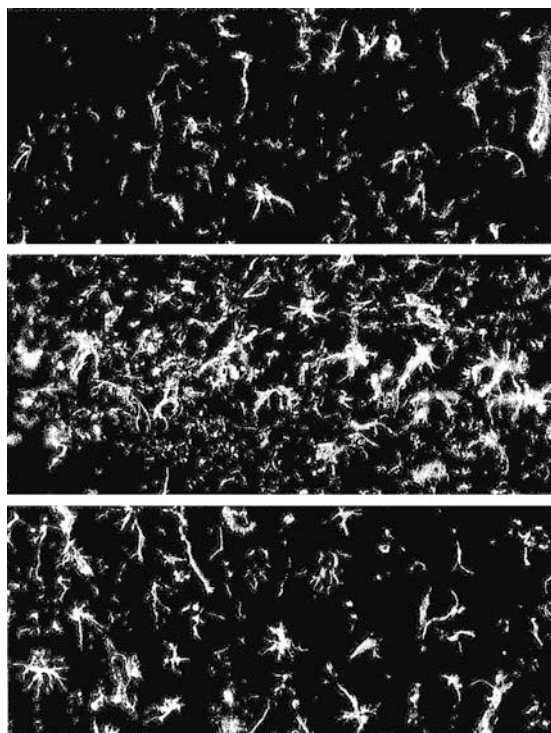


**Fig. 2** Two days following 5-min bilateral carotid occlusion. **a** Selective decrease of Nissl staining in the CA1 region of the hippocampus and calcium accumulation in the CA1/CA2 transition zone. **b** Effect of PPF treatment ( $10 \text{ mg kg}^{-1}$ , intraperitoneal) 15 min prior to cerebral ischemia on Nissl staining and calcium accumulation. *Left*: 5-min period of bilateral carotid occlusion. *Right*: 12-min period of bilateral carotid occlusion

(Si et al. 1998). Microglial proliferation was also dose-dependently inhibited by PPF, demonstrating a direct action of PPF on microglial function (Si et al. 1996).

These results suggest that PPF, probably via increases in cAMP intracellular signaling, alters the profile of newly adopted glial/immune properties in a way that

**Fig. 3** Glial fibrillary acidic protein (GFAP) immunoreactivity in the CA1 region of the hippocampus. *Top*: control animal. *Middle*: 2 days following 5 min of cerebral ischemia. *Bottom*: 2 days following 5 min of cerebral ischemia after treatment with PPF (10 mg kg<sup>-1</sup>, intraperitoneal)



inhibits potentially neurotoxic functions while maintaining beneficial functions. This differential regulation of microglial activation may explain the neuroprotective mechanism exerted by PPF. Schubert further posited that glial functional changes are controlled by an altered balance of the second messengers Ca<sup>2+</sup> and cAMP which can be inhibited by the endogenous cell modulator adenosine via enhanced cAMP-dependent signaling (Schubert et al. 1998; Schubert and Rudolphi, 1998). The homeostatic adenosine effects on glial cells by PPF may prove to be a mediator of neuroprotection. Furthermore, the glial modulatory effect of PPF was observed in the spinal cord following cerebral ischemia, demonstrating PPF's effects at a site distant from the original site of injury, and in a site long considered to be the “gate” for pain transmission from the periphery to the CNS (Wu et al. 1999). These concepts laid the groundwork for investigations into the role of glial modulators, namely, PPF, in reducing CNS sensitization, the electrophysiological correlate to chronic pain via a direct glial–neuronal interaction.

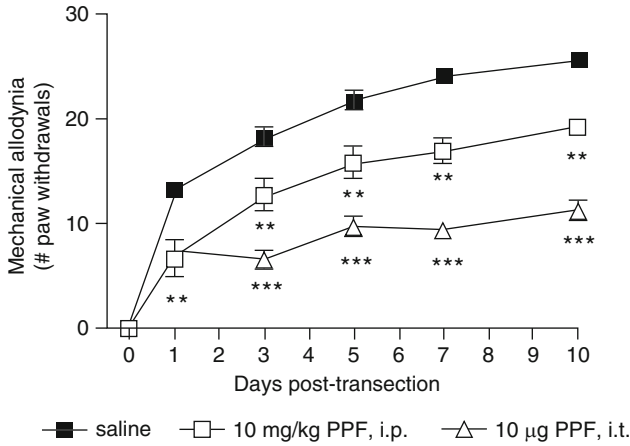
### 1.3 PPF as an Antiallodynic Agent

Pain is defined as “an unpleasant sensory and emotional experience associated with actual or potential tissue damage, or described in terms of such damage”

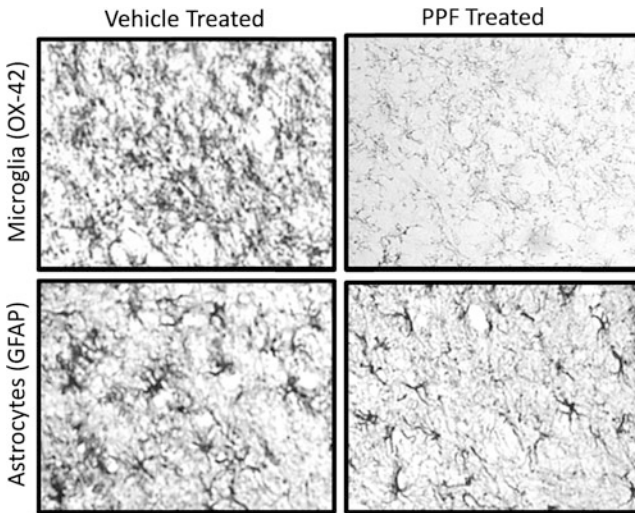
(International Association for the Study of Pain 1986). Acute pain serves a very important physiological function to protect the body from tissue damage and to prevent further tissue damage following injury. In contrast, pain of greater than 3 months in duration is termed “chronic pain.” Chronic pain can develop in the absence of identifiable injury or as a result of infection (herpes zoster, HIV), physical injury (disc herniation, spinal cord injury, surgical incision, complex regional pain syndrome), chemical injury (alcohol, chemotherapy), autoimmune disease (multiple sclerosis), inflammatory disease (rheumatoid arthritis, osteoarthritis), metabolic disorders (diabetes), and cancer. One common form of chronic pain is neuropathic pain that arises from an injury or dysfunction in the peripheral nervous system or the CNS. Neuropathic pain has a prevalence of 3–8% in the US population and is characterized by paresthesia (abnormal skin sensations such as burning and shooting pain), dysesthesia (uncomfortable sensation from light touch), allodynia (pain in response to a normally nonpainful stimuli), and hyperalgesia (increased sensitivity to painful stimuli). Neuropathic pain is often refractory to current drug therapies. Over the last decade evidence has grown that reactive microglia and astrocytes modulate neuronal responsiveness to painful sensory stimuli. Seminal studies using PPF in preclinical models of neuropathic pain have not only highlighted the importance of reactive microglia and astrocytes in the transition from acute to chronic pain, but also the potential clinical utility for PPF in the treatment of chronic neuropathic pain.

Early studies suggesting the involvement of reactive glia in pain and nociception used the glial metabolic inhibitor fluorocitrate to attenuate acute formalin-induced nociception (Watkins et al. 1997) and zymosan-induced inflammatory hyperalgesia (Meller et al. 1994) or the NMDA antagonist MK-801 to attenuate chronic sciatic nerve constriction induced astrocyte activation (Garrison et al. 1991, 1994). A subsequent study demonstrated that the degree of glial changes in the dorsal horn of the spinal cord (a site where primary nociceptive afferents synapse on ascending spinothalamic tract neurons) correlated with the duration and severity of mechanical allodynia. This study showed mild and transient allodynia and glial reactivity following an acute noxious stimulus, moderate allodynia and glial reactivity following an intermediate duration inflammatory insult, and robust and long-lasting allodynia and glial reactivity following a spinal nerve transection (Sweitzer et al. 1999).

With the growing evidence for phenotypic changes in glial protein expression in the dorsal horn of the spinal cord in a number of peripheral nerve injury models of neuropathic pain (Colburn et al. 1997, 1999; Hashizume et al. 2000), we undertook a study of PPF in a preclinical model of chronic neuropathic pain (Sweitzer et al. 2001b). Daily systemic or intrathecal administration of PPF initiated prior to nerve injury prevented the development of mechanical allodynia (Fig. 4). Impressively, daily administration of PPF produced an approximately 50% reduction in astrocytic and microglial reactive changes (Fig. 5). Similarly, daily systemic administration of PPF reduced astrocytic and microglial reactive changes and attenuated mechanical allodynia in a vincristine model of chemotherapy-induced neuropathic pain (Sweitzer et al. 2006). In the L5 spinal nerve transection model,



**Fig. 4** PPF attenuates mechanical allodynia in a preclinical model of neuropathic pain. Daily intraperitoneal or intrathecal administration of PPF beginning on the day of nerve transection reduced mechanical allodynia over the 10 days of the study. \*\* $p < 0.01$ , \*\*\* $p < 0.005$  compared with saline-vehicle-treated controls



**Fig. 5** Daily PPF treatment attenuated microglial (OX-42) and astrocytic (GFAP) reactivity in the dorsal horn of the spinal cord on day 10 after nerve transection as compared with saline vehicle controls

systemic administration of PPF starting on day 4 after injury prevented the further development of allodynia (Sweitzer et al. 2001a). In a crossover study, discontinuation of PPF at day 7 after injury resulted in the rapid return of allodynia, while initiation of therapy on day 7 produced only a small drop in established allodynia (Sweitzer et al. 2001a).

In a follow-up study, minocycline, a microglial specific inhibitor in *in vitro* studies, attenuated the development of mechanical allodynia and thermal hyperalgesia, but was unable to attenuate existing allodynia and hyperalgesia. While both pre- and posttreatment with minocycline reduced microglial reactivity, only pretreatment with minocycline reduced astrocytic reactivity (Raghavendra et al. 2003a). This suggests that preventative treatment with PPF (Sweitzer et al. 2001b) was much more effective by inhibiting both microglial and astrocytic changes that may promote a proinflammatory phenotype. A subsequent study showed a second phase of microglial involvement in neuropathic pain-associated behaviors that could be attenuated with daily systemic treatment with PPF for 14 days beginning 2 weeks after nerve transection (Tawfik et al. 2007). These studies highlight the temporal importance of astrocytes and microglia in chronic neuropathic pain. The question of how glial inhibition attenuates behavioral hypersensitivity remained unknown.

A subsequent study with daily intrathecal administration of PPF confirmed a decrease in the levels of CD11b, a marker of microglial reactivity, and GFAP, a marker of astrocytic reactivity at both the messenger RNA (mRNA) level and the protein level (Raghavendra et al. 2003b). As mentioned above, PPF had been shown to reduce the levels of proinflammatory cytokines in microglial cultures (Si et al. 1996), and thus cytokine expression was examined in the spinal cord of neuropathic animals treated with PPF. PPF decreased mRNA and protein expression for the proinflammatory cytokines TNF $\alpha$ , IL-1 $\beta$ , and IL-6 in the spinal cord. It had previously been shown that inhibition of these three proinflammatory cytokines attenuated allodynia (Arruda et al. 2000; Sweitzer et al. 2001a). Furthermore, PPF pretreatment was shown to attenuate IL-1 $\beta$ -induced C-fiber mediated windup in dorsal horn neurons, an electrophysiological measure of CNS sensitization (Arriagada et al. 2007). Together these findings suggest that PPF inhibits glial reactivity, decreases synthesis and release of proinflammatory cytokines, and prevents cytokine-induced sensitization of central nociceptive neurons and attenuation of neuropathic pain-associated behaviors.

More recent studies support this mechanism involving a multipartite synapse involving microglia, astrocytes, presynaptic neurons, and postsynaptic neurons. Acute bolus administration of PPF has been shown to reverse thermal and mechanical hyperalgesia and decreases phosphorylation of p38 and p42/44 mitogen-activated protein kinases in the spinal cord in the chronic constriction model of neuropathic pain (Garry et al. 2005). Similarly, direct spinal application of PPF attenuates neuronal hyperexcitability and pain that originates below the level of the spinal cord lesion (Gwak and Hulsebosch 2009). At presynaptic neuron terminals, PPF may prevent the downregulation of glutamic acid decarboxylase 65, which is the rate-limiting enzyme in the production of the inhibitory neurotransmitter GABA, thus increasing endogenous inhibitory tone (Gwak et al. 2008).

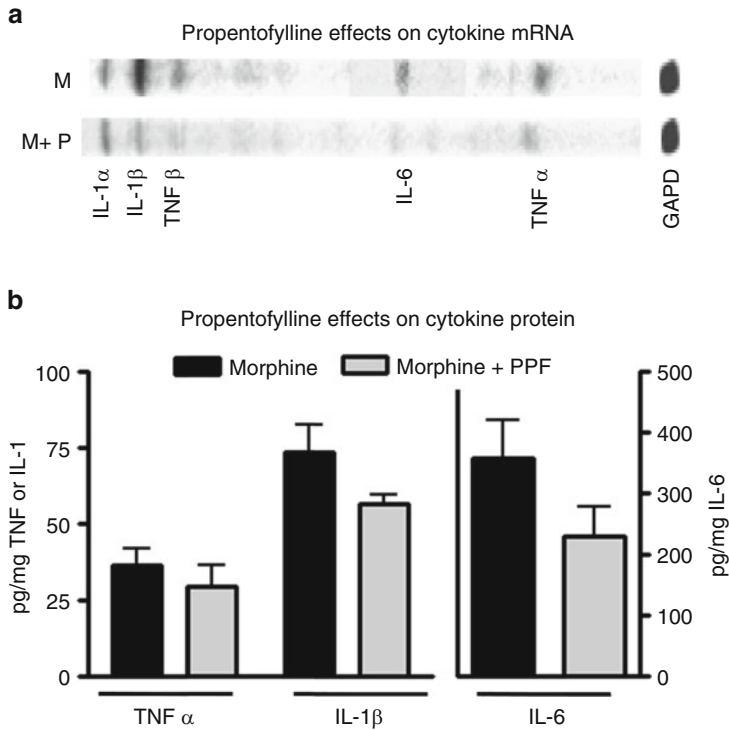
While the focus of the majority of pain research using PPF has examined activity at the level of the spinal cord, there is some evidence for PPF activity



outside the spinal cord. A potential local antinociceptive action of PPF on phase II formalin-induced nociception was reported following PPF administration in the hind paw either before or after formalin injury (Dorazil-Dudzic et al. 2004). A supraspinal site of action of PPF is suggested with the report of a prolonged antihyperalgesic/allodynic effect of a single administration of PPF in the cingulate cortex 24 h prior to nerve injury (Kuzumaki et al. 2007). It remains to be seen whether post-treatment is similarly effective in this paradigm. Follow-up studies are eagerly awaited to identify the mechanism(s) of action by which PPF produces antinociception in neuropathic pain following systemic, local, spinal, and brain-site administration. Within the spinal cord, attenuation of central sensitization by decreasing proinflammatory cytokine production/release and a reactive glial phenotype, and increasing endogenous inhibitory tone are potential mechanisms for PPF efficacy in neuropathic pain. However, there may be additional mechanisms involved within the peripheral skin microenvironment and brain.

## **2 Attenuation of Morphine Tolerance and Hyperalgesia in Acute and Chronic Pain**

The previous section detailed the mounting evidence for an antinociceptive action of PPF in acute and chronic pain. There is also growing evidence that PPF may have a second, equally important beneficial effect in both acute and chronic pain treatment by preventing opioid tolerance. Owing to robust efficacy in a variety of acute and chronic severe pains, opioids have been the mainstay for the treatment of pain. Unfortunately, the use of opioids in the treatment of chronic pain remains limited by the need for high doses, which are associated with undesirable side effects, and the development of tolerance, which necessitates dose escalation and further adverse side effects. Daily intrathecal administration of PPF prevents morphine tolerance in a preclinical model of neuropathic pain (Raghavendra et al. 2003b) or in normal animals (Raghavendra et al. 2004). Similarly, intrathecal coadministration of PPF is able to prevent stereo-selective cross-tolerance between dextromorphine and levomorphine (Wu et al. 2005) and morphine tolerance induced enhancement of  $\delta$ -opioid receptor analgesia (Holdridge et al. 2007). Systemic PPF administration has been shown to delay the onset of morphine (Narita et al. 2006; Shumilla et al., 2005) or entorphine (Narita et al. 2006) tolerance. Similar to preclinical models of neuropathic pain, PPF attenuated spinal GFAP and CD11b expression (Holdridge et al. 2007; Raghavendra et al. 2004) and proinflammatory cytokine expression in animals repeatedly exposed to morphine (Fig. 6). These findings provide further evidence that neuropathic pain and opioid tolerance share similar mechanisms, including the importance of glia and the potential for PPF to be both antinociceptive and opioid-sparing in the treatment of neuropathic pain.



**Fig. 6** Daily intrathecal PPF administration decreased morphine-induced upregulation of proinflammatory cytokines in the dorsal horn of the spinal cord. (A) With use of RNase protection assay, administration of morphine with PPF (M+P) decreased messenger RNA levels for interleukin (IL)-1 $\beta$ , tumor necrosis factor  $\alpha$  (TNF $\alpha$ ), and IL-6 as compared with morphine alone (M). (B) Administration of morphine with PPF decreased protein expression of IL-1 $\beta$ , TNF $\alpha$ , and IL-6 as compared with morphine alone (M)

### 3 PPF increases GLT-1 Expression *In Vitro* and *In Vivo*: Novel Mechanisms of Action

In a series of more recent *in vitro* and *in vivo* studies, we sought to further elucidate PPF's mechanism of action to attenuate nerve-injury-induced behavioral hypersensitivity. One of the key unifying neurotransmitters in the pathogenesis of neurodegeneration and neuronal sensitization is glutamate. PPF has previously been shown to reduce glutamate release in gerbil hippocampus following transient forebrain ischemia (Miyashita et al. 1992). In relation to chronic pain, peripheral nerve injury results in a decrease in glutamate uptake (Binns et al. 2005). Mature, differentiated astrocytes are known to express the GLT-1 (EAAT2) transporter, which is responsible for over 90% of synaptic glutamate clearance (Tanaka et al. 1997). Reactive astrocytes display decreased levels of glutamate transporters such as GLT-1 and as a result, synaptic glutamate clearance is impaired. In addition, these reactive astrocytes



become immunocompetent and release algogenic mediators which can sensitize neurons in the spinal cord. It was previously demonstrated that the level of GLT-1 is reduced in the dorsal horn of the spinal cord in rodent models of neuropathic pain (Cata et al. 2006; Sung et al. 2003; Weng et al. 2005). We evaluated the effect of PPF on cultured cortical astrocytes. Primary astrocyte cultures, which represent an activated phenotype with a polygonal morphology and low GLT-1 expression, were treated for 3 or 7 days with 10, 100, or 1,000  $\mu\text{M}$  PPF or db-cAMP, a known inducer of GLT-1 expression. PPF dose-dependently induced astrocytes to display a mature phenotype, with elongated processes and a stellate shape. In addition, PPF dose-dependently increased GLT-1 immunoreactivity. Real-time reverse transcription PCR and western blot analysis clearly demonstrated that PPF caused a potent dose-dependent induction of GLT-1 mRNA and protein in astrocytes. Importantly, the observed increase in the level of glutamate transporters was found to have a functional effect. PPF significantly enhanced glutamate uptake in astrocytes at both 100 and 1,000  $\mu\text{M}$  concentrations, which was sensitive to dihydrokainate inhibition, suggesting a robust GLT-1-mediated effect. In addition, PPF decreased both MCP-1 (CCL2) and MIP-2 (CXCL2) release from astrocytes, while db-cAMP significantly enhanced this chemokine expression. These findings suggest that PPF is capable of differentiating astrocytes to a homeostatic, mature phenotype, competent for glutamate clearance and distinct from that induced by db-cAMP.

In a parallel series of experiments, we determined whether PPF-induced glial modulation alters the levels of spinal glutamate transporters, GLT-1 and GLAST *in vivo*, which may contribute to reduced behavioral hypersensitivity after nerve injury. Rats received PPF (10  $\mu\text{g}$ ) or saline via lumbar puncture starting 1 h prior to L5 spinal nerve transection and then daily for 4 or 12 days. In addition to a marked suppression of mechanical allodynia, PPF increased mRNA levels for GLT-1, but not GLAST after injury. In support of previous data, western blot analysis demonstrated a decrease in the level of GLT-1 at day 12 after nerve injury, which was reversed with PPF treatment. In order to specifically examine the expression of spinal glutamate transporters, a novel line of double transgenic eGFP-GLT-1/DsRed-GLAST reporter mice were used (Tawfik et al. 2008). PPF restored transporter levels on the injured side as evidenced by an equal number of GLT-1 and GLAST puncta in both dorsal horns. As demonstrated in previous studies, PPF induced a concomitant reversal of L5 spinal nerve transection-induced expression of GFAP, a marker of astrocytic activation. The ability of PPF to alter the levels of glial glutamate transporters highlights the importance of controlling aberrant glial activation in neuropathic pain and suggests one possible mechanism for the antiallodynic action of this drug.

## 4 Previous Clinical Data Summary

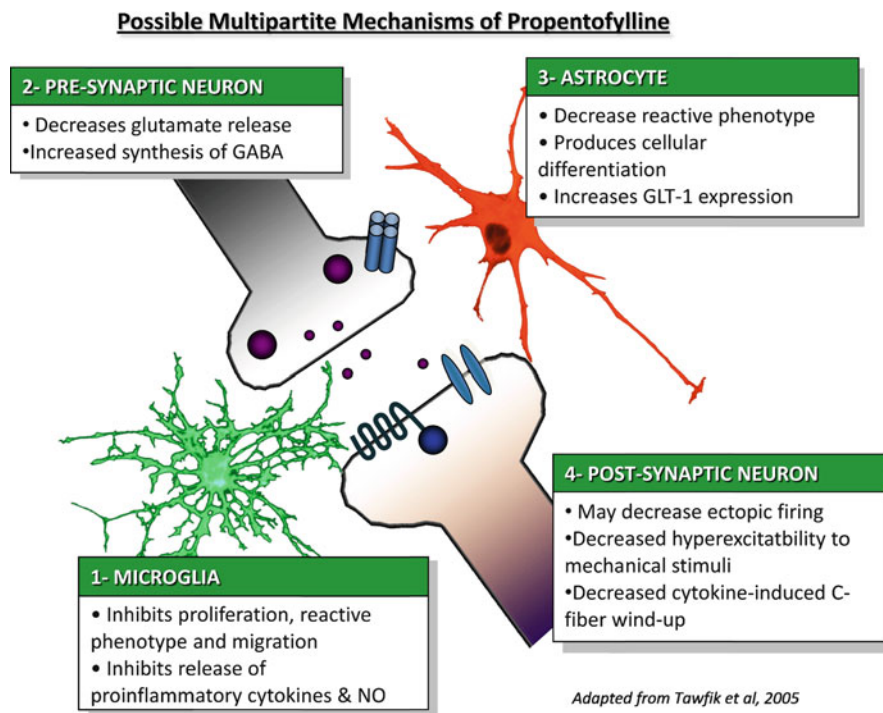
During the late 1990s, there were extensive clinical trials testing PPF in patients with mild to moderate Alzheimer's disease and vascular dementia. The phase II and phase III placebo-controlled trials were conducted by Aventis and Hoechst

Marion Roussel Pharmaceuticals in Europe, Canada, Japan, and the USA. PPF was also marketed in Japan from 1988 to 1998 for the symptoms of cerebrovascular disease. Therefore, PPF has been administered to thousands of patients over more than a decade. The efficacy in the Alzheimer's disease and vascular dementia trials varied among studies, with decreased efficacy attributed to drug–food interactions resulting in poor PPF absorption and, thus, decreased therapeutic plasma levels. It is difficult to determine the exact cause of reduced efficacy retrospectively given the long period since these trials. In addition, the primary end points used to assess efficacy were very different from the parameters that would be used today to assess treatment in Alzheimer's disease and vascular dementia. Even considering these caveats, the data demonstrate modest but positive effects on cognition with PPF compared with placebos and effects equivalent to those of other drugs clinically available for Alzheimer's disease. Considering the cost of these large trials, it is understandable but unfortunate that further testing has not been performed. Importantly, the overall profile of the safety data in thousands of elderly patients with underlying health problems supports future clinical trials.

## 5 Conclusions

This chapter highlighted key experimental findings demonstrating diverse actions and mechanisms of the atypical methylxanthine, PPF, in a variety of preclinical and clinical situations. Our focus was on its actions to reduce behavioral hypersensitivity in acute and chronic pain animal models and to attenuate opioid tolerance. These preclinical data from both our laboratory as well as many other investigators have culminated in an ongoing multicenter, multicountry (USA, Germany, Russia, and India) clinical trial for the treatment of postherpetic neuralgia. The importance of this trial is underscored by the first attempt to utilize a CNS glial modulator for the treatment of a chronic pain syndrome. As shown in Fig. 7, we describe a multipartite synapse where PPF may have distinct actions at each component of the synapse. There are extensive data demonstrating PPF's direct actions on microglia, including the inhibition of proliferation, migration, and a reactive phenotype, including suppression of proinflammatory factors. In addition, we and others have shown that PPF also decreases the levels of astrocytic cytosolic markers of reactivity, produces astrocyte differentiation, and increases GLT-1 expression after injury. Direct effects on neurons have also been described herein, which may lead to an inhibitory action partly due to enhancing GABAergic transmission. Further studies are needed to determine the specific effects of PPF on glial–neuronal signaling that would result in decreased CNS sensitization.

As mentioned previously, the potent neuroprotective effects we observed in a gerbil model of global ischemia were not reversed by an adenosine antagonist. In addition, PPF's actions in producing neuroprotection, glial modulation, and antiallodynia are significantly greater than the actions reported with other



**Fig. 7** Conceptual diagram of proposed PPF mechanisms of action on neurons, astrocytes, and microglia

methylxanthines or phosphodiesterase inhibitors. Similarly, the astrocytic GLT-1 enhancement we demonstrated with PPF was not seen with the typical methylxanthine, caffeine (Tawfik et al. 2006). Therefore, it seems unlikely we can attribute the mechanisms of action of PPF to a direct potentiation of adenosine or via phosphodiesterase inhibition. PPF does appear to have a pharmacodynamic profile distinct from that of other methylxanthines in many aspects, which may afford exciting drug development opportunities. As a potent glial modulator, cytokine release and functional migration assays in glial cultures are used as screening tools to enhance potency and improve pharmacokinetics of future compounds.

In conclusion, this small molecule has withstood the test of time by demonstrating both efficacy and safety in a plethora of preclinical and clinical studies. Fortuitously, PPF has more recently stimulated the fields of neuroimmunology and glial biology. As a powerful tool, it has elucidated novel molecular and cellular mechanisms of injury-induced CNS glial responses. These studies may one day lead to the utilization of glial modulators as novel pharmacological agents for the treatment of many neurodegenerative and chronic pain disorders.

## References

- Arriagada O, Constandil L, Hernandez A, Barra R, Soto-Moyano R, Laurido C (2007) Effects of interleukin-1beta on spinal cord nociceptive transmission in intact and propentofylline-treated rats. *Int J Neurosci* 117(5):617–625
- Arruda JL, Sweitzer S, Rutkowski MD, DeLeo JA (2000) Intrathecal anti-IL-6 antibody and IgG attenuates peripheral nerve injury-induced mechanical allodynia in the rat: possible immune modulation in neuropathic pain. *Brain Res* 879(1–2):216–225
- Binns BC, Huang Y, Goettl VM, Hackshaw KV, Stephens RL Jr (2005) Glutamate uptake is attenuated in spinal deep dorsal and ventral horn in the rat spinal nerve ligation model. *Brain Res* 1041(1):38–47
- Cata JP, Weng HR, Chen JH, Dougherty PM (2006) Altered discharges of spinal wide dynamic range neurons and down-regulation of glutamate transporter expression in rats with paclitaxel-induced hyperalgesia. *Neuroscience* 138(1):329–338
- Colburn RW, DeLeo JA, Rickman AJ, Yeager MP, Kwon P, Hickey WF (1997) Dissociation of microglial activation and neuropathic pain behaviors following peripheral nerve injury in the rat. *J Neuroimmunol* 79(2):163–175
- Colburn RW, Rickman AJ, DeLeo JA (1999) The effect of site and type of nerve injury on spinal glial activation and neuropathic pain behavior. *Exp Neurol* 157(2):289–304
- DeLeo J, Toth L, Schubert P, Rudolph K, Kreutzberg GW (1987) Ischemia-induced neuronal cell death, calcium accumulation, and glial response in the hippocampus of the Mongolian gerbil and protection by propentofylline (HWA 285). *J Cereb Blood Flow Metab* 7(6):745–751
- DeLeo J, Schubert P, Kreutzberg GW (1988a) Propentofylline (HWA 285) protects hippocampal neurons of Mongolian gerbils against ischemic damage in the presence of an adenosine antagonist. *Neurosci Lett* 84(3):307–311
- DeLeo J, Schubert P, Kreutzberg GW (1988b) Protection against ischemic brain damage using propentofylline in gerbils. *Stroke* 19(12):1535–1539
- Dorazil-Dudzik M, Mika J, Schafer MK, Li Y, Obara I, Wordliczek J, Przewlocka B (2004) The effects of local pentoxifylline and propentofylline treatment on formalin-induced pain and tumor necrosis factor-alpha messenger RNA levels in the inflamed tissue of the rat paw. *Anesth Analg* 98(6):1566–1573
- Fredholm BB, Lindstrom K (1986) The xanthine derivative 1-(5'-oxohexyl)-3-methyl-7-propyl xanthine (HWA 285) enhances the actions of adenosine. *Acta Pharmacol Toxicol (Copenh)* 58(3):187–192
- Garrison CJ, Dougherty PM, Kajander KC, Carlton SM (1991) Staining of glial fibrillary acidic protein (GFAP) in lumbar spinal cord increases following a sciatic nerve constriction injury. *Brain Res* 565(1):1–7
- Garrison CJ, Dougherty PM, Carlton SM (1994) GFAP expression in lumbar spinal cord of naive and neuropathic rats treated with MK-801. *Exp Neurol* 129(2):237–243
- Garry EM, Delaney A, Blackburn-Munro G, Dickinson T, Moss A, Nakalembe I, Robertson DC, Rosie R, Robberecht P, Mitchell R, Fleetwood-Walker SM (2005) Activation of p38 and p42/44 MAP kinase in neuropathic pain: involvement of VPAC2 and NK2 receptors and mediation by spinal glia. *Mol Cell Neurosci* 30(4):523–537
- Grome J, Stefanovich V (1985) Differential effects of xanthine derivatives on local cerebral blood flow and glucose utilization in the conscious rat. In: Stefanovich V, Rudolph R, Schubert P (eds) Adenosine: receptors and modulation of cell function. IRL, Oxford, pp 453–460
- Gwak YS, Hulsebosch CE (2009) Remote astrocytic and microglial activation modulates neuronal hyperexcitability and below-level neuropathic pain after spinal injury in rat. *Neuroscience* 161(3):895–903
- Gwak YS, Crown ED, Unabia GC, Hulsebosch CE (2008) Propentofylline attenuates allodynia, glial activation and modulates GABAergic tone after spinal cord injury in the rat. *Pain* 138(2):410–422

- Hashizume H, DeLeo JA, Colburn RW, Weinstein JN (2000) Spinal glial activation and cytokine expression after lumbar root injury in the rat. *Spine* 25(10):1206–1217
- Holdridge SV, Armstrong SA, Taylor AM, Cahill CM (2007) Behavioural and morphological evidence for the involvement of glial cell activation in delta opioid receptor function: implications for the development of opioid tolerance. *Mol Pain* 3:7
- Kuzumaki N, Narita M, Narita M, Hareyama N, Niikura K, Nagumo Y, Nozaki H, Amano T, Suzuki T (2007) Chronic pain-induced astrocyte activation in the cingulate cortex with no change in neural or glial differentiation from neural stem cells in mice. *Neurosci Lett* 415(1):22–27
- Meller ST, Dykstra C, Grzybycki D, Murphy S, Gebhart GF (1994) The possible role of glia in nociceptive processing and hyperalgesia in the spinal cord of the rat. *Neuropharmacology* 33(11):1471–1478
- Miyashita K, Nakajima T, Ishikawa A, Miyatake T (1992) An adenosine uptake blocker, propentofylline, reduces glutamate release in gerbil hippocampus following transient forebrain ischemia. *Neurochem Res* 17(2):147–150
- Nagata K, Ogawa T, Omosu M, Fujimoto K, Hayashi S (1985) *In vitro* and *in vivo* inhibitory effects of propentofylline on cyclic AMP phosphodiesterase activity. *Arzneimittelforschung* 35(7):1034–1036
- Narita M, Suzuki M, Narita M, Niikura K, Nakamura A, Miyatake M, Yajima Y, Suzuki T (2006) mu-Opioid receptor internalization-dependent and -independent mechanisms of the development of tolerance to mu-opioid receptor agonists: comparison between etorphine and morphine. *Neuroscience* 138(2):609–619
- Raghavendra V, Tanga F, DeLeo JA (2003a) Inhibition of microglial activation attenuates the development but not existing hypersensitivity in a rat model of neuropathy. *J Pharmacol Exp Ther* 306(2):624–630
- Raghavendra V, Tanga F, Rutkowski MD, DeLeo JA (2003b) Anti-hyperalgesic and morphine-sparing actions of propentofylline following peripheral nerve injury in rats: mechanistic implications of spinal glia and proinflammatory cytokines. *Pain* 104(3):655–664
- Raghavendra V, Tanga FY, DeLeo JA (2004) Attenuation of morphine tolerance, withdrawal-induced hyperalgesia, and associated spinal inflammatory immune responses by propentofylline in rats. *Neuropsychopharmacology* 29(2):327–334
- Schubert P, Rudolphi K (1998) Interfering with the pathologic activation of microglial cells and astrocytes in dementia. *Alzheimer Dis Assoc Disord* 12(Suppl 2):S21–S28
- Schubert P, Ogata T, Rudolphi K, Marchini C, McRae A, Ferroni S (1997) Support of homeostatic glial cell signaling: a novel therapeutic approach by propentofylline. *Ann N Y Acad Sci* 826:337–347
- Schubert P, Ogata T, Miyazaki H, Marchini C, Ferroni S, Rudolphi K (1998) Pathological immuno-reactions of glial cells in Alzheimer's disease and possible sites of interference. *J Neural Transm Suppl* 54:167–174
- Shumilla JA, Samuels I, Johnson KW, Forsayeth JR (2005) Systemic administration of propentofylline does not attenuate morphine tolerance in non-injured rodents. *Neurosci Lett* 384(3):344–348
- Si QS, Nakamura Y, Schubert P, Rudolphi K, Kataoka K (1996) Adenosine and propentofylline inhibit the proliferation of cultured microglial cells. *Exp Neurol* 137(2):345–349
- Si Q, Nakamura Y, Ogata T, Kataoka K, Schubert P (1998) Differential regulation of microglial activation by propentofylline via cAMP signaling. *Brain Res* 812(1–2):97–104
- Sung B, Lim G, Mao J (2003) Altered expression and uptake activity of spinal glutamate transporters after nerve injury contribute to the pathogenesis of neuropathic pain in rats. *J Neurosci* 23(7):2899–2910
- Sweitzer SM, Colburn RW, Rutkowski M, DeLeo JA (1999) Acute peripheral inflammation induces moderate glial activation and spinal IL-1beta expression that correlates with pain behavior in the rat. *Brain Res* 829(1–2):209–221
- Sweitzer S, Martin D, DeLeo JA (2001a) Intrathecal interleukin-1 receptor antagonist in combination with soluble tumor necrosis factor receptor exhibits an anti-allodynic action in a rat model of neuropathic pain. *Neuroscience* 103(2):529–539

- Sweitzer SM, Schubert P, DeLeo JA (2001b) Propentofylline, a glial modulating agent, exhibits antiallodynic properties in a rat model of neuropathic pain. *J Pharmacol Exp Ther* 297(3):1210–1217
- Sweitzer SM, Pahl JL, DeLeo JA (2006) Propentofylline attenuates vincristine-induced peripheral neuropathy in the rat. *Neurosci Lett* 400(3):258–261
- Tanaka K, Watase K, Manabe T, Yamada K, Watanabe M, Takahashi K, Iwama H, Nishikawa T, Ichihara N, Kikuchi T, Okuyama S, Kawashima N, Hori S, Takimoto M, Wada K (1997) Epilepsy and exacerbation of brain injury in mice lacking the glutamate transporter GLT-1. *Science* 276(5319):1699–1702
- Tawfik VL, Nutile-McMenemy N, Lacroix-Fralish ML, Deleo JA (2007) Efficacy of propentofylline, a glial modulating agent, on existing mechanical allodynia following peripheral nerve injury. *Brain Behav Immun* 21(2):238–246
- Tawfik VL, Regan MR, Haenggeli C, Lacroix-Fralish ML, Nutile-McMenemy N, Perez N, Rothstein JD, DeLeo JA (2008) Propentofylline-induced astrocyte modulation leads to alterations in glial glutamate promoter activation following spinal nerve transection. *Neuroscience* 152(4):1086–1092
- Tawfik VL, LaCroix-Fralish ML, Bercery KK, Nutile-McMenemy N, Harris BT, DeLeo JA (2006) Induction of astrocyte differentiation by propentofylline increases glutamate transporter expression in vitro: Heterogeneity of the quiescent phenotype. *Glia* 54(3):193–203
- Watkins LR, Martin D, Ulrich P, Tracey KJ, Maier SF (1997) Evidence for the involvement of spinal cord glia in subcutaneous formalin induced hyperalgesia in the rat. *Pain* 71(3):225–235
- Weng HR, Aravindan N, Cata JP, Chen JH, Shaw AD, Dougherty PM (2005) Spinal glial glutamate transporters downregulate in rats with taxol-induced hyperalgesia. *Neurosci Lett* 386(1):18–22
- Wu YP, McRae A, Rudolphi K, Ling EA (1999) Propentofylline attenuates microglial reaction in the rat spinal cord induced by middle cerebral artery occlusion. *Neurosci Lett* 260(1):17–20
- Wu HE, Thompson J, Sun HS, Terashvili M, Tseng LF (2005) Antianalgesia: stereoselective action of dextro-morphine over levo-morphine on glia in the mouse spinal cord. *J Pharmacol Exp Ther* 314(3):1101–1108

# Methylxanthines, Seizures, and Excitotoxicity

Detlev Boison

## Contents

1	Introduction .....	252
2	Clinical Findings .....	252
3	Experimental Findings .....	254
4	Adenosine, Seizures, and Excitotoxicity .....	255
4.1	Adenosine Deficiency and Seizure Generation .....	255
4.2	Adenosine Deficiency and Excitotoxicity .....	256
4.3	Adenosine-Based Therapeutic Approaches .....	257
5	Methylxanthines, Seizures, and Excitotoxicity .....	257
5.1	Acute Versus Chronic Caffeine .....	258
5.2	Caffeine: A <sub>1</sub> R- and A <sub>2A</sub> R-Mediated Actions .....	259
5.3	GABA <sub>A</sub> Receptor and Phosphodiesterase Inhibition .....	259
5.4	Ryanodine-Receptor-Activated Calcium-Induced Calcium Release .....	260
5.5	Free Radicals in Theophylline-Induced Seizures .....	260
5.6	Inhibition of TREK-1 Channels by Methylxanthines .....	260
6	Conclusions and Outlook .....	261
	References .....	261

**Abstract** Clinical evidence, in particular the wide use of theophylline as a bronchodilator, suggests that methylxanthines can cause seizures in patients without known underlying epilepsy. Theophylline is also known to be an added risk factor for seizure exacerbation in patients with epilepsy. The proconvulsant activity of methylxanthines can best be explained by their antagonizing the brain's own anticonvulsant adenosine. Recent evidence suggests that adenosine dysfunction is a pathological hallmark of epilepsy contributing to seizure generation and seizure spread. Conversely, adenosine augmentation therapies are effective in seizure suppression and prevention, whereas adenosine receptor antagonists such as

---

D. Boison

R.S. Dow Neurobiology Laboratories, Legacy Research, Portland, OR 97232, USA

methylxanthines generally exacerbate seizures. The impact of the methylxanthines caffeine and theophylline on seizures and excitotoxicity depends on timing, dose, and acute versus chronic use. New findings suggest a role of free radicals in theophylline-induced seizures, and adenosine-independent mechanisms for seizure generation have been proposed.

**Keywords** Adenosine · Adenosine kinase · Caffeine · Epilepsy · Toxicity

## 1 Introduction

Seizures, ranging from altered states of consciousness to clonic and/or tonic convulsions, are commonly encountered in patients who do not have epilepsy (Delanty et al. 1998). Among other potential triggers, such nonepileptic seizures can be provoked by medication or medication withdrawal. Within this context, seizures are potentially severe or fatal complications of theophylline therapy. Theophylline can trigger seizures in patients without known underlying epilepsy and is an added risk factor for seizure exacerbation in patients with epilepsy. Most of these seizures result from toxic theophylline serum concentrations and are difficult to control. Nevertheless, clinical diagnosis and management of theophylline-induced seizures are underappreciated compared with clinical diagnosis and management of other drug-induced seizures. Despite a long clinical history of theophylline-induced seizures, relatively little is known about the underlying molecular mechanisms that contribute to methylxanthine-induced seizure generation. Knowledge gained from patient data, but most notably from animal or *in vitro* studies aimed at elucidating the role of endogenous adenosine in seizure control, contributes to our current understanding of how methylxanthines influence the excitability of the brain.

## 2 Clinical Findings

Anecdotally, caffeinated beverages are “known” to lower seizure thresholds in patients with epilepsy and the avoidance of excessive caffeine has been recommended in patients with epilepsy (Kaufman and Sachdeo 2003). However, owing to the lack of well-designed, randomized, and placebo-controlled clinical trials, this concept has been challenged (Asadi-Pooya et al. 2008). Clinical findings in support of a proconvulsant role of methylxanthines are largely based on theophylline (or aminophylline, a mixture of theophylline with ethylenediamine that is 20 times more soluble than theophylline alone), which, clinically, is widely used to manage bronchospasms in reversible airway obstruction associated with stable asthma and chronic bronchitis (Barnes 2005; Van Dellen 1979). In addition, aminophylline



is indicated in asystolic cardiac arrest and periarrest bradycardia refractory to atropine, whereas caffeine is used to treat diabetic cardiac autonomic neuropathy (Duby et al. 2004). Theophylline has a narrow therapeutic window, with an optimal plasma concentration of 10–20 mg l<sup>-1</sup> (55–110 mmol l<sup>-1</sup>). Above this concentration, side effects such as arrhythmias and convulsions may occur, especially when theophylline is given rapidly by intravenous injection (Nolan et al. 2005). Theophylline-associated seizures (TAS) are considered a neurological emergency with potentially fatal outcome (Nakada et al. 1983). These seizures – largely focal onset generalized motor seizures – tend to be the only sign of theophylline toxicity, and can occur in neurologically intact patients (Aminoff and Simon 1980; Nakada et al. 1983). Remarkably, anticonvulsant therapy is ineffective in controlling these seizures, which often progress to status epilepticus and become intractable (Nakada et al. 1983; Yoshikawa 2007). In a recent clinical study the usual first-line treatment of diazepam was found to be more likely to be ineffective in TAS cases compared with non-TAS cases (Yoshikawa 2007); the failure of diazepam to stop those seizures might be based on interactions of theophylline with benzodiazepines (see later) (Yoshikawa 2007).

Interestingly, TAS is most common in pediatric patients under 5 years of age (Korematsu et al. 2008; Yoshikawa 2007), who can be considered to be naïve to theophylline or caffeine. In a recent study of eight pediatric TAS cases without underlying epilepsy, all the patients had fever at the onset of TAS (more than 38°C), and six of them had a family history of febrile seizures and/or idiopathic epilepsy (Korematsu et al. 2008). The authors of this study concluded that in infants with an idiopathic reduced seizure threshold and fever, theophylline administration might possibly be sufficient to trigger a seizure. Apart from TAS discussed here, methylxanthine-induced seizures have also been described after the consumption of caffeinated energy drinks (Iyadurai and Chung 2007), and theophylline, caffeine, and aminophylline are used clinically to prolong seizure durations in electroconvulsive therapy for major depression (Stern et al. 1999). The potential risks associated with theophylline therapy are now well recognized. Owing to concerns of CNS stimulant effects, theophylline use in patients with insomnia was included in the 2002 Criteria for Potentially Inappropriate Medication Use in Older Adults (Fick et al. 2003).

Pharmacokinetic drug interactions of methylxanthines also need to be considered. Theophylline is largely metabolized by the hepatic enzyme CYP1A2, which is induced not only by a variety of antibiotics (Gillum et al. 1993) but also by the commonly used enzyme-inducing antiepileptic drugs phenobarbital, phenytoin, carbamazepine, and primidone, and might require an increase in the therapeutic dose of theophylline (Patsalos et al. 2002; Spina et al. 1996). In view of the potential seizure-inducing effects of theophylline, the use of theophylline in patients with epilepsy is now limited despite the fact that second-generation antiepileptic drugs do not interfere with the pharmacokinetics of theophylline (Patsalos et al. 2002). Of note, caffeine comedication in combination with phenobarbital during the first trimester of pregnancy leads to a significant increase in the number of congenital malformations in offspring (Samren et al. 1999).

### 3 Experimental Findings

The proconvulsant potential of methylxanthines has been corroborated in countless animal studies that go back more than 35 years (Roussinov et al. 1974). Early studies suggested slight differences in the convulsant role of methylxanthines: Intraperitoneal administration of caffeine produced immediate excitation and seizures followed by an encephalopathy, whereas progression from encephalopathy to seizures was observed following aminophylline administration (Chu 1981). The proconvulsant and convulsant effects of methylxanthines generally depend on the dose and mode of application: Aminophylline at 100 mg kg<sup>-1</sup> is known to increase the susceptibility of rats to pilocarpine- or pentylenetetrazole-induced seizures (Chakrabarti et al. 1997; Turski et al. 1989), whereas higher doses of aminophylline (250 mg kg<sup>-1</sup>) lead to seizures and death in rats (Chakrabarti et al. 1997). These detrimental effects of high doses of aminophylline could be avoided by using equivalent doses of theophylline in preparations of acepifylline (theophylline ethanoate of piperazine) (Chakrabarti et al. 1997). Aminophylline-induced seizures directly depended on cerebrospinal fluid concentrations of theophylline and were not influenced by metabolites of theophylline (Ramzan and Levy 1986). In several experimental combinations it was shown that methylxanthines reduce or abolish the anticonvulsant activity of several antiepileptic drugs (Kulkarni et al. 1991). In contrast, the anticonvulsant effectiveness of felbamate was only affected at higher doses of aminophylline and caffeine (Gasior et al. 1998), and aminophylline did not alter the ability of gabapentin to protect mice against seizures induced by electroconvulsive shock (Luszczki et al. 2007). The concept that methylxanthines can exacerbate seizures in epilepsy has recently been challenged by Loscher (2009), arguing that CNS stimulants exert (pro)convulsant activity only at supratherapeutic doses.

Whereas methylxanthine-induced seizures are refractory to diazepam in patients, it is important to point out that levetiracetam and several other anti-epileptic drugs that do not act via activation of GABA<sub>A</sub> receptors are highly effective in suppressing caffeine-induced seizures in mice (Klitgaard et al. 1998). Astemizole, a novel histamine H<sub>1</sub> receptor antagonist, at a dose of 2 mg kg<sup>-1</sup> increased the threshold for aminophylline-induced seizures (Swiader et al. 2005), an interesting observation since these drugs are usually combined during the treatment of asthma.

Pharmacokinetic and pharmacodynamic drug interactions have also been studied in animal models. Of note are interactions of the fluoroquinolone class of antibacterials with theophylline. In one study, chronic pretreatment of rats with the fluoroquinolone pefloxacin was shown to exacerbate aminophylline-induced seizures without altering brain concentrations of theophylline (Imperatore et al. 1997). Likewise, certain environmental toxins, such as toluene, were shown to reduce the thresholds for methylxanthine-induced seizures (Chan and Chen 2003).

## 4 Adenosine, Seizures, and Excitotoxicity

Several potential mechanisms have been discussed that could explain the proconvulsant role of acute theophylline (Yoshikawa 2007): (1) general decrease of seizure thresholds; (2) inhibition of adenosine A<sub>1</sub> receptors (A<sub>1</sub>Rs) that normally suppress seizures by blocking the release of excitatory amino acids; (3) inhibition of cerebral blood flow via adenosine antagonism (Puiroud et al. 1988); (4) inhibition of 5'-nucleotidase and decrease in endogenous adenosine production; (5) inhibition of pyridoxal kinase, an enzyme needed for the synthesis of GABA; (6) increase in cyclic GMP that is involved in maintaining the epileptic discharge; and (7) a presumed direct inhibition of the GABA<sub>A</sub> receptor (Sugimoto et al. 2001), although interactions between GABA<sub>A</sub> receptors and the adenosine system might also be involved (Bonfiglio and Dasta 1991; Phillis 1979). Overall, it appears that theophylline does not trigger seizures as such, but rather potentiates preexisting brain hyperexcitability, a mechanism consistent with the role of A<sub>1</sub>Rs in preventing seizure spread and in mediating seizure arrest (Fedele et al. 2006; Lado and Moshe 2008; Young and Dragunow 1994). Given the dominant role of the adenosine system in seizure control within the context of theophylline toxicity, the following sections focus on the role of adenosine in epilepsy.

### 4.1 Adenosine Deficiency and Seizure Generation

The role of adenosine as an endogenous regulator of hippocampal excitability was first recognized by Dunwiddie (1980) almost 30 years ago. In a subsequent study it was shown that theophylline and other alkylxanthines antagonized electrophysiological responses to adenosine and adenosine-stimulated cyclic AMP formation, indicating that alkylxanthines increase hippocampal excitability by antagonizing the actions of adenosine (Dunwiddie et al. 1981). Several adenosine receptor agonists that activate the A<sub>1</sub>R were shown to suppress seizures in a variety of models, albeit accompanied by sedative and hypothermic side effects (Dunwiddie and Worth 1982). Endogenous adenosine is a potent regulator of hippocampal activity and was recently shown to control hippocampal sharp waves in CA3 via activation of A<sub>1</sub>Rs (Wu et al. 2009). It is now well recognized that adenosine is an endogenous anticonvulsant and regulator of brain activity (Boison 2005; Dunwiddie and Masino 2001; Fredholm et al. 2005a, b; Ribeiro et al. 2002). The anticonvulsant activity of adenosine is largely mediated by activation of A<sub>1</sub>Rs, since A<sub>1</sub>R knockout mice experience spontaneous seizures (Li et al. 2007a) and are highly susceptible to seizure spread (Fedele et al. 2006). Conversely, A<sub>1</sub>R agonists are highly effective in the suppression of seizures (Benarroch 2008; Fredholm 2003; Jacobson and Gao 2006), and have been demonstrated to suppress seizures that are resistant to conventional antiepileptic drugs (Gouder et al. 2003). Decreased extracellular adenosine levels and reduced

A<sub>1</sub>R activation as a consequence of kindling or caused by hypercapnia in a hippocampal slice preparation provide a plausible mechanisms for seizure generation (Dulla et al. 2005; Rebola et al. 2003).

In adult brain, synaptic levels of adenosine are largely regulated by an astrocyte-based adenosine cycle (Boison 2008). Under physiological conditions, synaptic adenosine is largely derived from vesicular release of ATP from astrocytes followed by extracellular cleavage into adenosine (Pascual et al. 2005), although astrocytic release of ATP via hemichannels has been demonstrated (Iglesias et al. 2009; Kang et al. 2008). In adult brain, adenosine is rapidly phosphorylated into AMP by the astrocyte-based enzyme adenosine kinase (ADK; EC 2.7.1.20) (Boison 2006, 2008). In contrast to conventional neurotransmitters, such as glutamate and glycine, there is no transporter-based regulatory mechanism to terminate the synaptic activity of adenosine. Owing to the presence of two types of equilibrative nucleoside transporters in the astrocyte membrane (Baldwin et al. 2004), intracellular ADK is able to fulfill the role of a metabolic reuptake system for adenosine (Boison 2008). On the basis of its low  $K_M$  for adenosine, ADK is the key regulator for ambient concentrations of adenosine (Boison 2006; Etherington et al. 2009; Lloyd and Fredholm 1995).

ADK has recently been identified as a molecular link between astrogliosis and neuronal hyperexcitability in epilepsy (Li et al. 2008). Astrogliosis – a pathological hallmark of the epileptic brain – is associated with upregulation of the adenosine-removing enzyme ADK (Gouder et al. 2004; Li et al. 2008). Remarkably, the development of spontaneous electrographic seizures coincides both spatially (Li et al. 2008) as well as temporally (Li et al. 2007a) with astrogliosis and upregulated ADK. Uncoupling of astrogliosis from epileptogenesis in ADK-transgenic mice (Adk-tg) (Li et al. 2009) has demonstrated that overexpression of ADK, rather than astrogliosis per se, can be the cause for seizures. In line with these findings, Adk-tg mice express spontaneous recurrent electrographic seizures (Li et al. 2007a). Conversely, therapeutic augmentation of the adenosine system is very effective in suppressing seizures (Boison 2009). Together, these findings demonstrate that adenosine deficiency and therefore deficient activation of A<sub>1</sub>Rs can be a direct cause for seizures. This conclusion supports the notion that methylxanthines have proconvulsant activity owing to antagonizing the function of the endogenous anticonvulsant adenosine.

## 4.2 Adenosine Deficiency and Excitotoxicity

Adenosine, acting via A<sub>1</sub>Rs, is not only an endogenous anticonvulsant of the brain, but also a powerful neuroprotectant (Cunha 2005; Fredholm 1997). Thus, in addition to a proconvulsant role of A<sub>1</sub>R deficiency or increased adenosine clearance (overexpression of ADK), these conditions lead to increased vulnerability to excitotoxic injury. Consequently, A<sub>1</sub>R knockout mice are highly susceptible to seizure-induced (Fedele et al. 2006) or traumatic (Kochanek et al. 2006) brain injury

and they experience highly aggravated neuronal cell loss after status epilepticus (Li et al. 2007a).

Pharmacological studies in a model of oxygen glucose deprivation suggest that whereas  $A_1$ Rs desensitize after prolonged agonist exposure, adenosine  $A_{2A}$  receptor ( $A_{2A}R$ )-mediated facilitation of glutamate release by endogenous adenosine remains fully operational under long-term oxygen glucose deprivation (Sperlagh et al. 2007). Thus, the inhibition of  $A_{2A}Rs$  might be a more effective approach to attenuate glutamatergic excitotoxicity than the stimulation of  $A_1Rs$  (Cunha 2005). Consequently,  $A_{2A}R$  antagonists are actively investigated clinically for their neuroprotective potential (Chase et al. 2003; Hauser et al. 2003).

### 4.3 Adenosine-Based Therapeutic Approaches

Given the prominent role of adenosine as an endogenous anticonvulsant and neuroprotectant, adenosine augmentation therapies are highly effective in preventing seizures (Boison 2009). Pharmacologically, seizures can be suppressed by  $A_1R$  agonists (Benarroch 2008) or by ADK inhibitors (McGaraughty et al. 2005); however, systemic augmentation of the adenosine system is associated with significant side effects, including the suppression of cardiac function and depression of blood pressure, and is therefore not a therapeutic option (Dunwiddie and Masino 2001). Alternatives are focal adenosine augmentation therapies to avoid systemic side effects and to restore adenosinergic signaling within a localized area of adenosine dysfunction, which can be equated with an epileptogenic focus (Li et al. 2008). Strategies that have been explored include the implantation of adenosine-releasing silk-based polymers into the infrahippocampal fissure in kindled rats. Rats treated with these polymers were protected both from established seizures as well as from developing epilepsy (Szybala et al. 2009). Likewise, rats with focal implants of adenosine-releasing encapsulated fibroblasts or ADK-deficient stem cells were protected from kindled seizures or kindling development, respectively (Huber et al. 2001; Li et al. 2007b). Stem-cell-derived adenosine-releasing implants that were placed into the infrahippocampal fissure in mice were shown to suppress acute chemoconvulsant-induced seizures with associated injury (Ren et al. 2007), and to suppress epilepsy development and spontaneous seizure expression in a model of CA3-selective focal epileptogenesis (Li et al. 2008). Together, these data demonstrate that focal reconstitution of adenosine signaling within an area of acquired adenosine dysfunction (i.e., within an epileptogenic focus) constitutes a powerful approach to suppress seizures.

## 5 Methylxanthines, Seizures, and Excitotoxicity

The previous sections suggest that methylxanthines – via antagonizing adenosine's anticonvulsant and neuroprotective actions (Fredholm et al. 1999; Nehlig et al. 1992) – are proconvulsants that aggravate excitotoxicity. There are, however,

additional interactions that need to be considered: the influence of methylxanthines on seizures and excitotoxicity is context- and receptor-dependent, and appears to be influenced by pathways not related to adenosine.

### 5.1 *Acute Versus Chronic Caffeine*

Whereas the proconvulsant activity of acute methylxanthines has long been recognized (see earlier), the chronic dosing of caffeine has different effects. Caffeine administered at a dose of 60–70 mg kg<sup>-1</sup> per day in mice over a period of 2 weeks (resulting in plasma levels of caffeine in the range 6–14 μM, corresponding to chronic caffeine use in humans) reduced *N*-methyl-D-aspartate-, bicuculline-, and pentylenetetrazol-induced seizures in mice in the absence of changes in A<sub>1</sub>R, A<sub>2A</sub>Rs, or GABA<sub>A</sub> receptors (Georgiev et al. 1993; Johansson et al. 1996). The effect was due to the combined effects of theophylline, to which caffeine is metabolized in the brain, and caffeine itself, but could not be ascribed to changes in A<sub>1</sub>Rs, A<sub>2A</sub>Rs, or GABA<sub>A</sub> receptors (Johansson et al. 1996). In contrast, higher plasma concentrations of caffeine (100 μM) after chronic dosage for 12 days resulted in increased A<sub>1</sub>R densities, whereas messenger RNA levels or A<sub>2A</sub>Rs were not affected (Johansson et al. 1993). Remarkably, chronic caffeine administration in rats (40 mg kg<sup>-1</sup>, twice daily for 7 days) increased the thresholds for subsequent theophylline-induced seizures (Zhi and Levy 1990). This phenomenon of effect inversion might be an explanation why children (who are considered to be caffeine-naïve) appear to be more sensitive to TAS (see earlier). Effect inversion of chronic adenosine receptor antagonists has also been described within the context of ischemic excitotoxicity (de Mendonca et al. 2000). Whereas acute methylxanthines generally aggravate ischemic injury, the chronic use of caffeine or of the A<sub>1</sub>R-selective antagonist DPCPX protects the brain from ischemic injury (de Mendonca et al. 2000). The phenomenon of effect inversion of acute versus chronic caffeine has been studied intensively and has been explained by antagonism of an endogenous agonist that downregulates A<sub>1</sub>Rs without affecting gene transcription (Jacobson et al. 1996). Evidence for effect inversion by caffeine or adenosine receptor ligands has been obtained through changes in physiological outcome parameters such as susceptibility to seizures or to seizure- and ischemia-induced neuronal cell death (Jacobson et al. 1996). Despite these clear physiological changes, the molecular mechanisms behind this phenomenon appear to be more complex since upregulation of A<sub>1</sub>Rs as a consequence of chronic caffeine was not always observed (Georgiev et al. 1993; Johansson et al. 1996). Later studies have ruled out upregulation of A<sub>1</sub>Rs as a consequence of the long-term use of caffeine or theophylline in reasonably normal doses, indicating that upregulation of A<sub>1</sub>Rs is triggered only by excessively high or toxic doses of methylxanthines (Svenningsson et al. 1999). Thus, selected doses and durations of exposure and withdrawal as well as A<sub>2A</sub>R-mediated effects (see later) might play an important role.

In a recent study a single dose of acute caffeine (40 mg kg<sup>-1</sup> intraperitoneally) given *after* the onset of seizures in a new mouse model of sudden unexplained death

in epilepsy (SUDEP) significantly increased the survival time from 24 to 55 min (Shen et al. 2009). This protective effect of acute caffeine can best be explained by caffeine antagonizing a seizure-induced surge of adenosine, which had experimentally been exacerbated by pharmacological disruption of adenosine clearance. In this model of SUDEP, excessive seizure-induced concentrations of adenosine are thought to induce cardiac and respiratory failure by overstimulation of brainstem adenosine receptors, an effect that can be ameliorated by caffeine-induced blockade of these receptors (Shen et al. 2009).

## 5.2 Caffeine: $A_1R$ - and $A_{2A}R$ -Mediated Actions

Whereas the anticonvulsant role of  $A_1R$ s is well established, newer findings suggest that  $A_{2A}R$ s play an important role in modulating the susceptibility to seizures. Thus,  $A_{2A}R$  knockout mice are partially resistant to limbic seizures induced by chomoconvulsants or to seizures induced by ethanol withdrawal (El Yacoubi et al. 2001, 2009). Interestingly, the attenuation of clonic pentylenetetrazole-induced seizures in  $A_{2A}R$  knockout mice could be mimicked in wild-type mice exposed to chronic caffeine (0.3 g l<sup>-1</sup> caffeine in drinking water) during a period of 14 days prior to the seizure tests (El Yacoubi et al. 2008). However,  $A_{2A}R$  knockout mice under chronic caffeine were less protected from clonic seizures than water-treated  $A_{2A}R$  knockout mice, a conflicting result that was not further addressed (El Yacoubi et al. 2008). Together, these findings indicate that the protective effects of chronic caffeine might best be explained by antagonizing the  $A_{2A}R$  and thus causing a state of decreased neuronal excitability; however, these studies also indicate a proconvulsant role of chronic caffeine under conditions during which  $A_{2A}R$ -dependent signaling is abolished.

## 5.3 $GABA_A$ Receptor and Phosphodiesterase Inhibition

In contrast to adenosine receptors, which are affected by caffeine plasma concentrations attainable by normal human caffeine consumption, 10–100 times higher concentrations are needed to inhibit  $GABA_A$  receptors or phosphodiesterase (Fredholm et al. 1999). Therefore, a direct proconvulsant role of “physiological” doses of methylxanthines via  $GABA_A$  receptors or phosphodiesterase appears to be unlikely. However, caffeine can inhibit the binding of benzodiazepines to the  $GABA_A$  receptor (Marangos et al. 1979), which might contribute to a convulsant role of high or toxic doses of methylxanthines. Inhibition of benzodiazepine binding to  $GABA_A$  receptors might be an explanation for the clinical findings that TAS are usually refractory to treatment with diazepam or other drugs that act via the  $GABA_A$  receptor (Yoshikawa 2007).



#### **5.4 *Ryanodine-Receptor-Activated Calcium-Induced Calcium Release***

Changes in  $\text{Ca}^{2+}$  homeostasis and persistent increases in the intracellular  $\text{Ca}^{2+}$  level contribute to the initiation and maintenance of acquired epilepsy (DeLorenzo et al. 2005). Ryanodine-receptor-mediated calcium-induced calcium release plays a key role in regulating intracellular calcium concentrations in epileptic conditions (Pal et al. 2001). The brain ryanodine receptor is a caffeine-sensitive calcium-release channel and mediates the caffeine-induced mobilization of  $\text{Ca}^{2+}$  from internal stores (McPherson et al. 1991; Usachev et al. 1993). The caffeine-induced release of  $\text{Ca}^{2+}$  from ryanodine-sensitive calcium stores in the neuronal endoplasmic reticulum and pathological mechanisms that potentiate this response may render neurons more vulnerable to excitotoxicity and to the expression of seizures (Chan et al. 2000; Verkhratsky 2005). Interestingly, in cultured hippocampal neurons, the newer antiepileptic drug levetiracetam led to a 61% decrease in caffeine-induced peak height of the intracellular  $\text{Ca}^{2+}$  level (Nagarkatti et al. 2008), indicating that levetiracetam might interact with adenosine-related signaling.

#### **5.5 *Free Radicals in Theophylline-Induced Seizures***

A possible role of free radicals in theophylline-induced seizures was recently suggested (Gulati et al. 2005, 2007; Ray et al. 2005). In the underlying studies, aminophylline ( $50\text{--}250\text{ mg kg}^{-1}$ ) dose-dependently induced convulsions and mortality in rats. Seizures and mortality were attenuated by antioxidants (melatonin, *N*-acetylcysteine) and by nitric oxide (NO) synthase inhibitors [*N*<sup>o</sup>-nitro-*L*-arginine methyl ester (L-NAME), 7-nitroindazole]. Combination of antioxidant and NO-reducing treatments augmented the anticonvulsant effects of single treatments. Further, the authors found increased concentrations of malondialdehyde and NO metabolites in brain homogenates of mice with aminophylline-induced seizures; accumulation of these metabolites could be attenuated by melatonin or L-NAME pretreatment. These studies suggest the contribution of free radicals in the mechanism of theophylline-induced ictogenesis.

#### **5.6 *Inhibition of TREK-1 Channels by Methylxanthines***

TREK-1, a member of the two-pore-domain  $\text{K}^+$  channel superfamily, plays a major role in regulating the resting membrane potential of neurons, and thus contributes to controlling neuronal excitability (Honore 2007). Using whole-cell patch-clamp recordings on human TREK-1 channel expressing Chinese hamster ovary cells, Harinath and Sikdar (2005) demonstrated reversible inhibition of the channels, and



depolarization of the membrane potential, by caffeine and theophylline in a concentration-dependent manner. Inhibition by caffeine and theophylline was attenuated in channels with a mutation of a protein kinase A (PKA) consensus sequence, indicating involvement of the cyclic AMP/PKA pathway. Thus, inhibition of TREK-1-dependent membrane depolarization may contribute to seizure generation by toxic doses of caffeine or theophylline.

## 6 Conclusions and Outlook

Although adenosine-independent mechanisms have been proposed, the majority of evidence indicates that the proconvulsant roles of methylxanthines are based on antagonism of the brain's endogenous adenosine-based seizure control system. Whereas inhibition of A<sub>1</sub>Rs by methylxanthines can directly contribute to ictogenesis and seizure spread, under certain conditions methylxanthines can also contribute to seizure suppression. First, this can be the case after chronic drug exposure leading to inversion and alterations in gene expression (Svenningsson et al. 1999). Second, antagonism of A<sub>2A</sub>Rs by methylxanthines may have direct anticonvulsant and neuroprotective consequences.

A detailed understanding of the convulsant role of methylxanthines is of importance since many new drugs that act on adenosine receptors are in clinical trials. For example, in recent clinical trials conducted with the A<sub>1</sub>R antagonist rolofylline, which facilitates diuresis and preserves renal function in patients with acute heart failure with renal impairment, the occurrence of seizures was described in some patients who were treated with higher doses of the drug (Cotter et al. 2008). This example demonstrates that caution is needed when evaluating the clinical use of new adenosine-related therapeutic agents; however, understanding the mechanisms involved in the adenosine-related control of seizure mechanisms will allow the safe use of novel drugs that act on new therapeutic principles. New approaches using gene-array-based strategies might unravel novel pathways and interactions that might help explain the complex role of methylxanthines in determining the brain's susceptibility to seizures and excitotoxicity (Yu et al. 2009).

**Acknowledgements** The work of the author is supported by grants R01NS058780, R01NS061844, R01MH083973, R21NS057475-01, and R21NS057538-01 from the National Institutes of Health (NIH), from Citizens United in Research against Epilepsy (CURE) in collaboration with the Department of Defense (DoD), and from the Legacy Hospital Foundation.

## References

- Aminoff MJ, Simon RP (1980) Status epilepticus. Causes, clinical features and consequences in 98 patients. *Am J Med* 69:657–666
- Asadi-Pooya AA, Mintzer S, Sperling MR (2008) Nutritional supplements, foods, and epilepsy: is there a relationship? *Epilepsia* 49:1819–1827

- Baldwin SA, Beal PR, Yao SY, King AE, Cass CE, Young JD (2004) The equilibrative nucleoside transporter family, SLC29. *Pflugers Arch* 447:735–743
- Barnes PJ (2005) Targeting histone deacetylase 2 in chronic obstructive pulmonary disease treatment. *Expert Opin Ther Targets* 9:1111–1121
- Benarroch EE (2008) Adenosine and its receptors: multiple modulatory functions and potential therapeutic targets for neurologic disease. *Neurology* 70:231–236
- Boison D (2005) Adenosine and epilepsy: from therapeutic rationale to new therapeutic strategies. *Neuroscientist* 11:25–36
- Boison D (2006) Adenosine kinase, epilepsy and stroke: mechanisms and therapies. *Trends Pharmacol Sci* 27:652–658
- Boison D (2008) The adenosine kinase hypothesis of epileptogenesis. *Prog Neurobiol* 84:249–262
- Boison D (2009) Adenosine augmentation therapies (AATs) for epilepsy: prospect of cell and gene therapies. *Epilepsy Res* 85:131–141
- Bonfiglio MF, Dasta JF (1991) Clinical significance of the benzodiazepine-theophylline interaction. *Pharmacotherapy* 11:85–87
- Chakrabarti A, Saini HK, Garg SK (1997) A comparative study of aminophylline- and acepifylline-induced seizures and death in the chemoconvulsion model in rats. *J Pharm Pharmacol* 49:812–815
- Chan MH, Chen HH (2003) Toluene exposure increases aminophylline-induced seizure susceptibility in mice. *Toxicol Appl Pharmacol* 193:303–308
- Chan SL, Mayne M, Holden CP, Geiger JD, Mattson MP (2000) Presenilin-1 mutations increase levels of ryanodine receptors and calcium release in PC12 cells and cortical neurons. *J Biol Chem* 275:18195–18200
- Chase TN, Bibbiani F, Bara-Jimenez W, Dimitrova T, Oh-Lee JD (2003) Translating A2A antagonist KW6002 from animal models to parkinsonian patients. *Neurology* 61:S107–S111
- Chu NS (1981) Caffeine- and aminophylline-induced seizures. *Epilepsia* 22:85–94
- Cotter G, Dittrich HC, Weatherley BD, Bloomfield DM, O'Connor CM, Metra M, Massie BM (2008) The PROTECT pilot study: a randomized, placebo-controlled, dose-finding study of the adenosine A(1) receptor antagonist rolofylline in patients with acute heart failure and renal impairment. *J Card Fail* 14:631–640
- Cunha RA (2005) Neuroprotection by adenosine in the brain: from A1 receptor activation to A2A receptor blockade. *Purinergic Signal* 1:111–134
- de Mendonca A, Sebastiao AM, Ribeiro JA (2000) Adenosine: does it have a neuroprotective role after all? *Brain Res Brain Res Rev* 33:258–274
- Delanty N, Vaughan CJ, French JA (1998) Medical causes of seizures. *Lancet* 352:383–390
- DeLorenzo RJ, Sun DA, Deshpande LS (2005) Cellular mechanisms underlying acquired epilepsy: the calcium hypothesis of the induction and maintenance of epilepsy. *Pharmacol Ther* 105:229–266
- Duby JJ, Campbell RK, Setter SM, White JR, Rasmussen KA (2004) Diabetic neuropathy: an intensive review. *Am J Health Syst Pharm* 61:160–73
- Dulla CG, Dobelis P, Pearson T, Frenguelli BG, Staley KJ, Masino SA (2005) Adenosine and ATP link P-CO<sub>2</sub> to cortical excitability via pH. *Neuron* 48:1011–1023
- Dunwiddie TV (1980) Endogenously released adenosine regulates excitability in the in vitro hippocampus. *Epilepsia* 21:541–548
- Dunwiddie TV, Masino SA (2001) The role and regulation of adenosine in the central nervous system. *Annu Rev Neurosci* 24:31–55
- Dunwiddie TV, Worth T (1982) Sedative and anticonvulsant effects of adenosine analogs in mouse and rat. *J Pharmacol Exp Ther* 220:70–76
- Dunwiddie TV, Hoffer BJ, Fredholm BB (1981) Alkylxanthines elevate hippocampal excitability. Evidence for a role of endogenous adenosine. *Naunyn Schmiedebergs Arch Pharmacol* 316:326–330
- El Yacoubi M, Ledent C, Parmentier M, Daoust M, Costentin J, Vaugeois J (2001) Absence of the adenosine A(2A) receptor or its chronic blockade decrease ethanol withdrawal-induced seizures in mice. *Neuropharmacology* 40:424–432

- El Yacoubi M, Ledent C, Parmentier M, Costentin J, Vaugeois JM (2008) Evidence for the involvement of the adenosine A<sub>2A</sub> receptor in the lowered susceptibility to pentylenetetrazol-induced seizures produced in mice by long-term treatment with caffeine. *Neuropharmacology* 55:35–40
- El Yacoubi M, Ledent C, Parmentier M, Costentin J, Vaugeois JM (2009) Adenosine A<sub>2A</sub> receptor deficient mice are partially resistant to limbic seizures. *Naunyn-Schmiedeberg Arch Pharmacol* 380:223–232
- Etherington LA, Patterson GE, Meechan L, Boison D, Irving AJ, Dale N, Frenguelli B (2009) Astrocytic adenosine kinase regulates basal synaptic adenosine levels and seizure activity but not activity-dependent adenosine release in the hippocampus. *Neuropharmacology* 56:429–437
- Fedele DE, Li T, Lan JQ, Fredholm BB, Boison D (2006) Adenosine A<sub>1</sub> receptors are crucial in keeping an epileptogenic focus localized. *Exp Neurol* 200:184–190
- Fick DM, Cooper JW, Wade WE, Waller JL, Maclean JR, Beers MH (2003) Updating the Beers criteria for potentially inappropriate medication use in older adults: results of a US consensus panel of experts. *Arch Intern Med* 163:2716–2724
- Fredholm BB (1997) Adenosine and neuroprotection. *Int Rev Neurobiol* 40:259–280
- Fredholm BB (2003) Adenosine receptors as targets for drug development. *Drug News Perspect* 16:283–289
- Fredholm BB, Battig K, Holmen J, Nehlig A, Zvartau EE (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fredholm BB, Chen JF, Cunha RA, Svenningsson P, Vaugeois JM (2005a) Adenosine and brain function. *Int Rev Neurobiol* 63:191–270
- Fredholm BB, Chen JF, Masino SA, Vaugeois JM (2005b) Actions of adenosine at its receptors in the CNS: insights from knockouts and drugs. *Annu Rev Pharmacol Toxicol* 45:385–412
- Gasior M, Swiader M, Przybylko M, Borowicz K, Turski WA, Kleinrok Z, Czuczwar SJ (1998) Felbamate demonstrates low propensity for interaction with methylxanthines and Ca<sup>2+</sup> channel modulators against experimental seizures in mice. *Eur J Pharmacol* 352:207–214
- Georgiev V, Johansson B, Fredholm BB (1993) Long-term caffeine treatment leads to a decreased susceptibility to NMDA-induced clonic seizures in mice without changes in adenosine A<sub>1</sub> receptor number. *Brain Res* 612:271–277
- Gillum JG, Israel DS, Polk RE (1993) Pharmacokinetic drug interactions with antimicrobial agents. *Clin Pharmacokinet* 25:450–482
- Gouder N, Fritschy JM, Boison D (2003) Seizure suppression by adenosine A<sub>1</sub> receptor activation in a mouse model of pharmacoresistant epilepsy. *Epilepsia* 44:877–885
- Gouder N, Scheurer L, Fritschy J-M, Boison D (2004) Overexpression of adenosine kinase in epileptic hippocampus contributes to epileptogenesis. *J Neurosci* 24:692–701
- Gulati K, Ray A, Pal G, Vijayan VK (2005) Possible role of free radicals in theophylline-induced seizures in mice. *Pharmacol Biochem Behav* 82:241–245
- Gulati K, Ray A, Vijayan VK (2007) Free radicals and theophylline neurotoxicity: an experimental study. *Cell Mol Biol (Noisy-le-grand)* 53:42–52
- Harinath S, Sikdar SK (2005) Inhibition of human TREK-1 channels by caffeine and theophylline. *Epilepsy Res* 64:127–135
- Hauser RA, Hubble JP, Truong DD (2003) Randomized trial of the adenosine A<sub>2A</sub> receptor antagonist istradefylline in advanced PD. *Neurology* 61:297–303
- Honore E (2007) The neuronal background K<sub>2P</sub> channels: focus on TREK1. *Nat Rev Neurosci* 8:251–261
- Huber A, Padrun V, Deglon N, Aebischer P, Mohler H, Boison D (2001) Grafts of adenosine-releasing cells suppress seizures in kindling epilepsy. *Proc Natl Acad Sci USA* 98:7611–7616
- Iglesias R, Dahl G, Qiu F, Spray DC, Scemes E (2009) Pannexin 1: the molecular substrate of astrocyte “hemichannels”. *J Neurosci* 29:7092–7097
- Imperatore C, Trimarchi GR, De Sarro A (1997) Interaction between pefloxacin and aminophylline in genetically epilepsy-prone rats. *J Pharm Pharmacol* 49:1025–1029

- Iyadurai SJ, Chung SS (2007) New-onset seizures in adults: possible association with consumption of popular energy drinks. *Epilepsy Behav* 10:504–508
- Jacobson KA, Gao ZG (2006) Adenosine receptors as therapeutic targets. *Nat Rev Drug Discov* 5:247–264
- Jacobson KA, von Lubitz DKJE, Daly JW, Fredholm BB (1996) Adenosine receptor ligands: differences with acute versus chronic treatment. *Trends Pharmacol Sci* 17:108–113
- Johansson B, Ahlberg S, van der Ploeg I, Brené S, Lindefors N, Persson H, Fredholm BB (1993) Effect of long term caffeine treatment on A1 and A2 adenosine receptor binding and on mRNA levels in rat brain. *Naunyn Schmiedeberg's Arch Pharmacol* 347:407–414
- Johansson B, Georgiev V, Kuosmanen T, Fredholm BB (1996) Long-term treatment with some methylxanthines decreases the susceptibility to bicuculline- and pentylenetetrazol-induced seizures in mice. Relationship to c-fos expression and receptor binding. *Eur J Neurosci* 8:2447–2458
- Kang J, Kang N, Lovatt D, Torres A, Zhao Z, Lin J, Nedergaard M (2008) Connexin 43 hemichannels are permeable to ATP. *J Neurosci* 28:4702–4711
- Kaufman KR, Sachdeo RC (2003) Caffeinated beverages and decreased seizure control. *Seizure* 12: 519–521
- Klitgaard H, Matagne A, Gobert J, Wulfert E (1998) Evidence for a unique profile of levetiracetam in rodent models of seizures and epilepsy. *Eur J Pharmacol* 353:191–206
- Kochanek PM, Vagni VA, Janesko KL, Washington CB, Crumrine PK, Garman RH, Jenkins LW, Clark RS, Homanics GE, Dixon CE, Schnermann J, Jackson EK (2006) Adenosine A1 receptor knockout mice develop lethal status epilepticus after experimental traumatic brain injury. *J Cereb Blood Flow Metab* 26:565–575
- Korematsu S, Miyahara H, Nagakura T, Suenobu S, Izumi T (2008) Theophylline-associated seizures and their clinical characterizations. *Pediatr Int* 50:95–98
- Kulkarni C, Joseph T, David J (1991) Influence of adenosine receptor antagonists, aminophylline and caffeine, on seizure protective ability of antiepileptic drugs in rats. *Indian J Exp Biol* 29: 751–754
- Lado FA, Moshe SL (2008) How do seizures stop? *Epilepsia* 49:1651–1664
- Li T, Lan JQ, Fredholm BB, Simon RP, Boison D (2007a) Adenosine dysfunction in astrogliosis: cause for seizure generation? *Neuron Glia Biol* 3:353–366
- Li T, Steinbeck JA, Lusardi T, Koch P, Lan JQ, Wilz A, Segsneider M, Simon RP, Brustle O, Boison D (2007b) Suppression of kindling epileptogenesis by adenosine releasing stem cell-derived brain implants. *Brain* 130:1276–1288
- Li T, Ren G, Lusardi T, Wilz A, Lan JQ, Iwasato T, Itohara S, Simon RP, Boison D (2008) Adenosine kinase is a target for the prediction and prevention of epileptogenesis in mice. *J Clin Inv* 118:571–582
- Li T, Lan JQ, Boison D (2009) Uncoupling of astrogliosis from epileptogenesis in adenosine kinase (ADK) transgenic mice. *Neuron Glia Biol* 4:91–99
- Lloyd HG, Fredholm BB (1995) Involvement of adenosine deaminase and adenosine kinase in regulating extracellular adenosine concentration in rat hippocampal slices. *Neurochem Int* 26:387–395
- Loscher W (2009) Preclinical assessment of proconvulsant drug activity and its relevance for predicting adverse events in humans. *Eur J Pharmacol* 610:1–11
- Luszczki JJ, Jankiewicz K, Jankiewicz M, Czuczwar SJ (2007) Influence of aminophylline on the anticonvulsive action of gabapentin in the mouse maximal electroshock seizure threshold model. *J Neural Transm* 114:1539–1545
- Marangos PJ, Paul SM, Parma AM, Goodwin FK, Syapin P, Skolnick P (1979) Purinergic inhibition of diazepam binding to rat brain (in vitro). *Life Sci* 24:851–857
- McGaraughey S, Cowart M, Jarvis MF, Berman RF (2005) Anticonvulsant and antinociceptive actions of novel adenosine kinase inhibitors. *Curr Top Med Chem* 5:43–58
- McPherson PS, Kim YK, Valdivia H, Knudson CM, Takekura H, Franzini-Armstrong C, Coronado R, Campbell KP (1991) The brain ryanodine receptor: a caffeine-sensitive calcium release channel. *Neuron* 7:17–25

- Nagarkatti N, Deshpande LS, DeLorenzo RJ (2008) Levetiracetam inhibits both ryanodine and IP<sub>3</sub> receptor activated calcium induced calcium release in hippocampal neurons in culture. *Neurosci Lett* 436:289–293
- Nakada T, Kwee IL, Lerner AM, Remler MP (1983) Theophylline-induced seizures: clinical and pathophysiological aspects. *West J Med* 138:371–374
- Nehlig A, Daval JL, Debry G (1992) Caffeine and the central nervous system: mechanisms of action, biochemical, metabolic and psychostimulant effects. *Brain Res Brain Res Rev* 17:139–170
- Nolan JP, Deakin CD, Soar J, Bottiger BW, Smith G (2005) European Resuscitation Council guidelines for resuscitation 2005. Section 4. Adult advanced life support. *Resuscitation* 67(Suppl 1):S39–S86
- Pal S, Sun D, Limbrick D, Rafiq A, DeLorenzo RJ (2001) Epileptogenesis induces long-term alterations in intracellular calcium release and sequestration mechanisms in the hippocampal neuronal culture model of epilepsy. *Cell Calcium* 30:285–296
- Pascual O, Casper KB, Kubera C, Zhang J, Revilla-Sanchez R, Sul JY, Takano H, Moss SJ, McCarthy K, Haydon PG (2005) Astrocytic purinergic signaling coordinates synaptic networks. *Science* 310:113–116
- Patsalos PN, Froscher W, Pisani F, van Rijn CM (2002) The importance of drug interactions in epilepsy therapy. *Epilepsia* 43:365–385
- Phillis JW (1979) Diazepam potentiation of purinergic depression of central neurons. *Can J Physiol Pharmacol* 57:432–435
- Puiroud S, Pinard E, Seylaz J (1988) Dynamic cerebral and systemic circulatory effects of adenosine, theophylline and dipyridamole. *Brain Res* 453:287–298
- Ramzan IM, Levy G (1986) Kinetics of drug action in disease states. XVI. Pharmacodynamics of theophylline-induced seizures in rats. *J Pharmacol Exp Ther* 236:708–713
- Ray A, Gulati K, Anand S, Vijayan VK (2005) Pharmacological studies on mechanisms of aminophylline-induced seizures in rats. *Indian J Exp Biol* 43:849–853
- Rebola N, Coelho JE, Costenla AR, Lopes LV, Parada A, Oliveira CR, Soares-da-Silva P, de Mendonca A, Cunha RA (2003) Decrease of adenosine A<sub>1</sub> receptor density and of adenosine neuromodulation in the hippocampus of kindled rats. *Eur J Neurosci* 18:820–828
- Ren G, Li T, Lan JQ, Wilz A, Simon RP, Boison D (2007) Lentiviral RNAi-induced down-regulation of adenosine kinase in human mesenchymal stem cell grafts: a novel perspective for seizure control. *Exp Neurol* 208:26–37
- Ribeiro JA, Sebastiao AM, de Mendonca A (2002) Adenosine receptors in the nervous system: pathophysiological implications. *Prog Neurobiol* 68:377–392
- Roussinov KS, Lazarova MB, Atanassova-Shopova S (1974) Experimental study of the effect of lithium, haloperidol, caffeine and theophylline on convulsive seizure reactions. *Acta Physiol Pharmacol Bulg* 2:67–75
- Samren EB, van Duijn CM, Christiaens GC, Hofman A, Lindhout D (1999) Antiepileptic drug regimens and major congenital abnormalities in the offspring. *Ann Neurol* 46:739–746
- Shen H-Y, Li T, Boison D (2009) A novel mouse model for sudden unexpected death in epilepsy (SUDEP): role of impaired adenosine clearance. *Epilepsia* 51:485–488, doi:10.1111/j.1528-1167.2009.02248.x
- Sperlagh B, Zsilla G, Baranyi M, Illes P, Vizi ES (2007) Purinergic modulation of glutamate release under ischemic-like conditions in the hippocampus. *Neuroscience* 149:99–111
- Spina E, Pisani F, Perucca E (1996) Clinically significant pharmacokinetic drug interactions with carbamazepine. An update. *Clin Pharmacokinet* 31:198–214
- Stern L, Dannon PN, Hirschmann S, Schriber S, Amytal D, Dolberg OT, Grunhaus L (1999) Aminophylline increases seizure length during electroconvulsive therapy. *J ECT* 15:252–257
- Sugimoto T, Sugimoto M, Uchida I, Mashimo T, Okada S (2001) Inhibitory effect of theophylline on recombinant GABA(A) receptor. *Neuroreport* 12:489–493
- Svenningsson P, Nomikos GG, Fredholm BB (1999) The stimulatory action and the development of tolerance to caffeine is associated with alterations in gene expression in specific brain regions. *J Neurosci* 19:4011–4022

- Swiader MJ, Luszczyk JJ, Wielosz M, Czuczwar SJ (2005) Effect of histamine receptor antagonists on aminophylline-induced seizures and lethality in mice. *Pharmacol Rep* 57:531–535
- Szybala C, Pritchard EM, Wilz A, Kaplan DL, Boison D (2009) Antiepileptic effects of silk-polymer based adenosine release in kindled rats. *Exp Neurol* 219:126–135
- Turski L, Ikonomidou C, Turski WA, Bortolotto ZA, Cavalheiro EA (1989) Review: cholinergic mechanisms and epileptogenesis. The seizures induced by pilocarpine: a novel experimental model of intractable epilepsy. *Synapse* 3:154–171
- Usachev Y, Shmigol A, Pronchuk N, Kostyuk P, Verkhratsky A (1993) Caffeine-induced calcium release from internal stores in cultured rat sensory neurons. *Neuroscience* 57:845–859
- Van Dellen RG (1979) Clinical pharmacology. Series on pharmacology in practice. 4. Theophylline. Practical application of new knowledge. *Mayo Clin Proc* 54:733–745
- Verkhratsky A (2005) Physiology and pathophysiology of the calcium store in the endoplasmic reticulum of neurons. *Physiol Rev* 85:201–279
- Wu C, Wong T, Wu X, Sheppy E, Zhang L (2009) Adenosine as an endogenous regulating factor of hippocampal sharp waves. *Hippocampus* 19:205–220
- Yoshikawa H (2007) First-line therapy for theophylline-associated seizures. *Acta Neurol Scand* 115:57–61
- Young D, Dracunow M (1994) Status epilepticus may be caused by loss of adenosine anticonvulsant mechanisms. *Neuroscience* 58:245–261
- Yu L, Coelho JE, Zhang X, Fu Y, Tillman A, Karaoz U, Fredholm BB, Weng Z, Chen JF (2009) Uncovering multiple molecular targets for caffeine using a drug target validation strategy combining A<sub>2A</sub> receptor knockout mice with microarray profiling. *Physiol Genomics* 37:199–210
- Zhi JG, Levy G (1990) Effect of chronic caffeine administration on theophylline concentrations required to produce seizures in rats. *Proc Soc Exp Biol Med* 193:210–213

# Impacts of Methylxanthines and Adenosine Receptors on Neurodegeneration: Human and Experimental Studies

Jiang-Fan Chen and Yijuang Chern

## Contents

1	Introduction .....	268
2	Molecular Targets of Methylxanthines, Including Caffeine .....	270
2.1	Non-adenosine Receptors .....	270
2.2	Adenosine Receptors .....	271
3	Molecular and Cellular Mechanisms of Neuroprotection by Caffeine and Adenosine Receptors .....	273
3.1	Control of Glutamate Release at Presynaptic Sites .....	273
3.2	Modulation of Cellular Survival Signals at Postsynaptic Sites .....	274
3.3	Control of Neuroinflammation in the CNS .....	275
3.4	Interaction with Neurotrophic Factors .....	276
3.5	Regulation of Blood–Brain Barrier Integrity .....	276
4	Methylxanthine/Caffeine and Neurodegenerative Disorders .....	276
4.1	Stroke .....	276
4.2	Traumatic Brain Injury .....	280
4.3	Alzheimer’s Disease .....	282
4.4	Parkinson’s Disease .....	285
4.5	Huntington’s Disease .....	289
4.6	Multiple Sclerosis .....	291
5	Concluding Remarks .....	293
	References .....	294

**Abstract** Neurodegenerative disorders are some of the most feared illnesses in modern society, with no effective treatments to slow or halt this neurodegeneration. Several decades after the earliest attempt to treat Parkinson’s disease using caffeine, tremendous amounts of information regarding the potential beneficial effect

---

J.-F. Chen (✉)

Department of Neurology, 715 Albany Street, Boston, MA, USA

e-mail: chenjf@bu.edu

Y. Chern

Institute of Biomedical Sciences, Academia Sinica, Taipei 11529, Taiwan

of caffeine as well as adenosine drugs on major neurodegenerative disorders have accumulated. In the first part of this review, we provide general background on the adenosine receptor signaling systems by which caffeine and methylxanthine modulate brain activity and their role in relationship to the development and treatment of neurodegenerative disorders. The demonstration of close interaction between adenosine receptor and other G protein coupled receptors and accessory proteins might offer distinct pharmacological properties from adenosine receptor monomers. This is followed by an outline of the major mechanism underlying neuroprotection against neurodegeneration offered by caffeine and adenosine receptor agents. In the second part, we discuss the current understanding of caffeine/methylxanthine and its major target adenosine receptors in development of individual neurodegenerative disorders, including stroke, traumatic brain injury Alzheimer's disease, Parkinson's disease, Huntington's disease and multiple sclerosis. The exciting findings to date include the specific *in vivo* functions of adenosine receptors revealed by genetic mouse models, the demonstration of a broad spectrum of neuroprotection by *chronic* treatment of caffeine and adenosine receptor ligands in animal models of neurodegenerative disorders, the encouraging development of several A<sub>2A</sub> receptor selective antagonists which are now in advanced clinical phase III trials for Parkinson's disease. Importantly, increasing body of the human and experimental studies reveals encouraging evidence that regular human consumption of caffeine in fact may have several beneficial effects on neurodegenerative disorders, from motor stimulation to cognitive enhancement to potential neuroprotection. Thus, with regard to neurodegenerative disorders, these potential benefits of methylxanthines, caffeine in particular, strongly argue against the common practice by clinicians to discourage regular human consumption of caffeine in aging populations.

**Keywords** A<sub>1</sub> receptor · A<sub>2A</sub> receptor · Alzheimer's disease · Adenosine receptors · Huntington's disease · Caffeine · Cognitive enhancer · Multiple sclerosis · Neuroprotection · Parkinson's disease · Stroke · Traumatic brain injury

## 1 Introduction

Strictly on the basis of the duration of the insults, neurodegeneration may be broadly divided into acute degeneration, such as stroke and traumatic brain injury (TBI), and chronic degeneration, such as Alzheimer's disease (AD), Parkinson's disease (PD), and Huntington's disease (HD) (Mattson 2000). Neurodegenerative disorders are some of the most feared illnesses in modern society. Ischemic stroke is the second most common cause of death in the USA/Europe and the third most common in Japan (Lo et al. 2003). TBI is the leading cause of death in the USA for individuals under 37 years old (Rutland-Brown et al. 2006; Moppett 2007).



Ischemic and traumatic insults produce necrosis and apoptosis as a result of overstimulation of neurons due to excitotoxicity and calcium overload (Lo et al. 2003). Chronic neurodegenerative disorders generally occur in mid to late life with both genetic and environmental etiological factors that result in severe and progressive cognitive and motor deficits. AD and PD are the first and second most common neurodegenerative disorders: AD affects 5% of people over 65, and another 1% of this population has PD (Lang and Lozano 1998b; Olanow 2004; Perrin et al. 2009). The pathological hallmark of chronic neurodegenerative disorders is neuronal cell death within specific regions and cell subpopulations, frequently characterized by prominent proteinaceous inclusions that accumulate in the extracellular milieu or within intercellular compartments of affected neurons (Braak and Braak 1997; Hardy and Gwinn-Hardy 1998; Dauer and Przedborski 2003; Perrin et al. 2009). Unfortunately, there are no successful treatments currently available that can slow or halt this chronic neurodegeneration (Shoulson 1998; Mattson 2004; Olanow 2004; Lopez-Diego and Weiner 2008; Mestre et al. 2009). Meanwhile, the number of people with neurodegenerative disorders is rapidly increasing as average lifespan increases. For example, the number of individuals afflicted by PD is expected to double by 2030 in line with the aging population and increases in life expectancy (Dorsey et al. 2007). Thus, there are critical and immediate medical needs for new pharmacological approaches to neurodegenerative disorders.

In the absence of an effective treatment for neurodegenerative disorders, epidemiological and experimental investigations into potential risk factors (including dietary factors) that may allow individuals to decrease their risk for neurodegenerative disorders become compelling. Caffeine is doubtless the most widely consumed psychoactive substance. It is estimated that more than 50% of the world's adult population consume caffeine on a daily basis (Fredholm et al. 1999). One of the main reasons for such prevalent use is the well-known psychostimulant (cognitive enhancement) effect of caffeine. Caffeine's psychostimulant effect is most obvious at low doses (Nehlig et al. 1992; Daly et al. 1999; Fredholm et al. 1999). At higher doses, the effects of caffeine vary among individuals and may lead to various untoward effects (Fredholm et al. 1999). Human intake of coffee/caffeine appears to decline during aging in Western cultures. This is in part due to undesirable side effects of high doses of caffeine, including increased anxiety, increased blood pressure, headache, and confusion (Nehlig et al. 1992; Daly et al. 1999; Fredholm et al. 1999), and in part due to caffeine-intake restrictions suggested by health care professions. Such restrictions appear unwarranted, based on the comprehensive literature review here indicating that regular human consumption of caffeine does not impose significant adverse effects on the cardiovascular system, bone status, or the incidence of cancer (Fredholm et al. 1999; Winkelmayr et al. 2005; Higdon and Frei 2006; van Dam et al. 2006; Cadden et al. 2007; Daly 2007), and in fact regular caffeine consumption may be associated with reduced risk for some neurodegenerative disorders such as PD and AD during aging (Fredholm et al. 1999; Ross et al. 2000; Daly 2007; Ritchie et al. 2007).

In this review, we will first provide a general background on the signaling systems by which caffeine and methylxanthine modulate brain activity and their role in the relationship to the development and treatment of neurodegenerative disorders. This is followed by an outline of the major mechanism underlying neuroprotection against neurodegeneration offered by caffeine and adenosine receptor agents. In the second part, we discuss the current understanding of caffeine/xanthine and its major target adenosine receptors in the development of individual neurodegenerative disorders, including PD, stroke, AD, HD, TBI, and multiple sclerosis (MS).

## 2 Molecular Targets of Methylxanthenes, Including Caffeine

### 2.1 *Non-adenosine Receptors*

Pharmacologically, caffeine produces complex actions through multiple molecular targets (Nehlig et al. 1992; Daly et al. 1999; Fredholm et al. 1999). Historically, four different molecular responses to caffeine have been proposed to underlie its psychostimulant effects: calcium release, phosphodiesterase (PDE) inhibition, GABA<sub>A</sub> receptor inhibition, and antagonism of adenosine receptors. Initially, calcium release was thought to be the major mechanism for the action of caffeine and other methylxanthine-based compounds, because caffeine stimulates calcium release from intracellular storage at a threshold concentration of 250  $\mu\text{M}$  (with maximal effect at 5–20 mM) (McPherson et al. 1991). Somewhat later it was found that these compounds also inhibit PDE, the enzyme which degrades cyclic AMP (cAMP), and it was proposed that caffeine operates by elevating intracellular cAMP concentrations (Choi et al. 1988). However, both calcium release and PDE inhibition are unlikely to account for the effects of caffeine in humans because PDE inhibition requires caffeine concentrations 10–100 times higher than those achieved by typical dietary intake (Daly et al. 1999), and caffeine is even less potent as a releaser of calcium. In the 1970s, caffeine was also found to inhibit binding of benzodiazepines to sites on GABA<sub>A</sub> receptors, raising a third possibility for the mechanism of caffeine's actions in humans (Marangos et al. 1979). The interaction between caffeine and GABA<sub>A</sub> receptors might explain the anxiogenic and convulsant effects induced by high doses of caffeine. However, the low affinity of caffeine for GABA<sub>A</sub> receptors ( $K_i = 280 \mu\text{M}$ ) suggested that normal human caffeine consumption is unlikely to produce caffeine doses high enough to inhibit GABA<sub>A</sub> receptors. Consistent with multiple molecular targets of caffeine, a novel drug target validation strategy coupled with microarray profiling recently showed that a high dose of caffeine (50 mg  $\text{kg}^{-1}$ ) induced complex expression patterns with *three* distinct sets of striatal genes associated with three distinct molecular targets: adenosine A<sub>2A</sub> receptor (A<sub>2A</sub>R), non-A<sub>2A</sub>R, and a group requiring interaction of the A<sub>2A</sub>R inactivation and non-A<sub>2A</sub>R targets (Yu et al. 2009). Furthermore, the involvement of

PDE and GABA<sub>A</sub> receptor are also indicated as components of the non-A<sub>2A</sub>R targets by the overlapping of the striatal gene sets elicited by the PDE inhibitor rolipram or by the GABA<sub>A</sub> receptor antagonist bicucullin with distinct subsets of striatal genes elicited by caffeine (50 mg kg<sup>-1</sup>) administered to A<sub>2A</sub>R-knockout mice. This supports the findings of earlier work suggesting there are multiple molecular targets for caffeine, including the contribution of the A<sub>2A</sub>R and non-A<sub>2A</sub>R targets such as PDE and GABA<sub>A</sub> receptors. More recently, other novel targets for caffeine in mediating the modulatory effect of caffeine on neuronal survival, including ryanodine receptor channels (Guerreiro et al. 2008) and nuclear enzyme poly(ADP-ribose)polymerase-1 (Geraets et al. 2006; Szabo et al. 2006), are suggested.

## 2.2 Adenosine Receptors

In the late 1970s, it became increasingly obvious that caffeine's actions in the brain are better explained by its antagonism of adenosine receptors (Fredholm 1980). Adenosine receptor blockade takes place at concentrations many times lower than those required for calcium release, PDE inhibition, or inhibition of GABA<sub>A</sub> receptor binding (Fredholm et al. 1999; Daly 2007). The caffeine concentrations required for adenosine receptor blockade can be attained after drinking even a single cup of coffee (containing 40–180 mg of caffeine), which can produce plasma concentrations of 2–10 μM (Fredholm et al. 1999; Daly 2007). Thus, of these possible mechanisms, only adenosine receptor blockade occurs at an affinity (2–50 μM) compatible with the caffeine plasma concentrations attained by normal human caffeine consumption (i.e., 250 mg day<sup>-1</sup>). In addition, caffeine's psychostimulant effects are best correlated with its blockade of brain adenosine receptors (Fredholm et al. 1999; Daly 2007). It is now the accepted view that caffeine, in doses consumed habitually by humans, binds both adenosine A<sub>1</sub> receptors (A<sub>1</sub>Rs) and A<sub>2A</sub>Rs and, by blocking these receptors at pre- and postsynaptic sites, respectively, relieves the largely inhibitory tone of endogenous adenosine and enhances brain dopaminergic activity, resulting in psychostimulant action (Fredholm et al. 1999; Daly 2007). This is supported by pharmacological evidence and by behavioral studies performed in A<sub>2A</sub>R-knockout mice (Nehlig et al. 1992; Daly et al. 1999; Fredholm et al. 1999; El Yacoubi et al. 2000; Chen et al. 2001). The requirement of the A<sub>2A</sub>R for caffeine's action in the brain is validated by the finding that caffeine-induced striatal gene expression is abolished in A<sub>2A</sub>R-knockout mice (Yu et al. 2009). Interestingly, despite caffeine's similar affinities for the A<sub>1</sub>R and the A<sub>2A</sub>R in brain, both pharmacological (Nehlig et al. 1992; Fredholm 1995; Fredholm et al. 1999) and genetic knockout (Fredholm et al. 1999; El Yacoubi et al. 2000; Chen et al. 2001) studies have revealed that caffeine's psychostimulant effect is better correlated with its blockade of brain A<sub>2A</sub>R. It must be emphasized that although this evidence is suggestive, the conclusion is not yet firm and there is evidence that other targets may also play a role, particularly at the higher end of the range of doses taken by humans.

### 2.2.1 Interactions Between Adenosine Receptors and Other G-Protein-Coupled Receptors

Adenosine receptors are capable of forming oligomers among themselves or with other G-protein-coupled receptors (GPCRs). The  $A_1R$  forms an oligomer with the  $D_1$  dopamine receptor which can be regulated by agonist binding (Gines et al. 2000). The  $A_{2A}R$  can form higher-order oligomers with itself (Vidi et al. 2008), and has multiple binding partners, including the  $A_1R$  (Ciruela et al. 2006), the  $D_2$  dopamine receptor ( $D_2R$ ) (Hillion et al. 2002; Kamiya et al. 2003), the  $D_3$  dopamine receptor (Torvinen et al. 2005), the metabotropic glutamate type 5 receptor (mGlu5R) (Diaz-Cabiale et al. 2002; Kachroo et al. 2005), and the cannabinoid  $CB_1$  receptor (Carriba et al. 2007). With use of high-resolution immunoelectron microscopy and immunoprecipitation, higher-order oligomer complexes containing the  $A_{2A}R$ , the  $D_2R$ , the mGlu5R, or  $CB_1$  receptors were found to exist in native tissues (Navarro et al. 2008; Cabello et al. 2009; Ferre et al. 2009). Formation of these homo- or hetero-GPCR oligomers usually affects the pharmacological properties, intracellular signaling, and/or surface expression of both receptors (Vidi et al. 2008; Cabello et al. 2009; Navarro et al. 2009). Years after the initial discovery of these receptor oligomers, evidence began to emerge from different laboratories to support their physiological relevance. For example, with use of competitive peptides in brain slices, the  $A_{2A}R$  was found to suppress  $D_2R$ -mediated regulation of membrane potential transitions and firing patterns in striatal neurons through the formation of  $A_{2A}R$ - $D_2R$  heteromerization (Azdad et al. 2009). Direct protein-protein interactions between the  $A_{2A}R$  and the  $D_2R$  may provide a mechanistic basis for the involvement of the  $D_2R$  in caffeine's action (Zahniser et al. 2000) and the ability of caffeine to affect the action of antipsychotic drugs that target the  $D_2R$ , including haloperidol (Varty et al. 2008; Salamone et al. 2009). It will of great interest to further characterize the pharmacological properties of these adenosine-receptor-containing heteromers toward adenosine drugs and methylxanthine in vivo. Along this line, a few drugs targeted toward receptor dimers or oligomers were reported in the past few years. For example, a dendrimer of the  $A_{2A}R$ -selective agonist CGS21680, which molecular modeling analysis predicts will bind to the homodimer of the  $A_{2A}R$ , was shown to effectively inhibit platelet aggregation (Ivanov and Jacobson 2008; Kim et al. 2008). Heterobivalent reagents composed of an  $A_{2A}R$  antagonist and a  $D_2R$  agonist bound to membranes containing both receptors with higher affinities than monovalent ligands, and were implicated in drug development for PD (Soriano et al. 2009).

### 2.2.2 Interacting Accessory Proteins

Besides binding to GPCRs, the C terminus of the  $A_{2A}R$  is also directly associated with a handful of interacting proteins, including the F-actin-cross-linking protein ( $\alpha$ -actinin), which is important for  $A_{2A}R$  internalization (Burgueno et al. 2003), a deubiquitination enzyme (Usp4) that controls  $A_{2A}R$  expression level in plasma

membranes (Milojevic et al. 2006), a binding protein of Translin (Trax) that is known to transport the brain-derived neurotrophic factor (BDNF) transcript in neuronal processes (Sun et al. 2006; Chiaruttini et al. 2009), a nucleotide exchange factor (ARNO) which mediates sustained mitogen-activated protein kinase (MAPK) signaling (Gsandtner et al. 2005), a calcium-binding protein (calmodulin) critical for the formation of the A<sub>2A</sub>R–D<sub>2</sub>R heterodimer (Navarro et al. 2009), and the fibroblast growth factor (FGF) receptor (Flajolet et al. 2008). These binding proteins have different natures and are involved in various novel functions of the A<sub>2A</sub>R. Further studies are required to demonstrate how these binding proteins function in *in vivo* pathophysiological condition and to determine whether they might be useful targets for therapeutic development. In particular, the finding that coactivation of the A<sub>2A</sub>R and the FGF receptor plays a crucial role in the synaptic plasticity of the corticostriatopallidal pathway may have important clinical implications for major neurodegenerative diseases (Flajolet et al. 2008).

The multiplicity of adenosine receptor subtypes (particularly A<sub>1</sub>Rs and A<sub>2A</sub>Rs) and the complex interactions between adenosine receptors, other GPCRs, and accessory signaling proteins in the brain may partly explain the heterogeneity and complexity of caffeine's action, and may underlie the biphasic motor and cardiovascular responses to increasing doses of caffeine in rodents (Svenningsson et al. 1995, 1999).

### **3 Molecular and Cellular Mechanisms of Neuroprotection by Caffeine and Adenosine Receptors**

#### ***3.1 Control of Glutamate Release at Presynaptic Sites***

Adenosine receptors may influence the outcome of brain injury by modulating glutamate and aspartate release in the brain. A<sub>1</sub>Rs are detected at high density at presynaptic nerve terminals and their activation efficiently inhibits the action of almost all neurotransmitters via G-protein-mediated inhibition of calcium channels in nerve endings (Dunwiddie and Masino 2001). The inhibition appears most prominent in excitatory glutamatergic systems where synaptic neurotransmission is almost completely blocked (Dunwiddie et al. 1981). Thus, by blocking A<sub>1</sub>R-mediated presynaptic inhibition of excitatory neurotransmission, caffeine increases the general excitability of systems in the brain. On the other hand, A<sub>2A</sub>R agonists enhance the release of glutamate under ischemic and nonischemic conditions (O'Regan et al. 1992; Simpson et al. 1992; Sebastiao and Ribeiro 1996; Dunwiddie and Masino 2001). For example, the A<sub>2A</sub>R agonist CGS21680 can enhance glutamate release at concentrations as low as 10<sup>-12</sup> M in synaptosomal and slice preparations (Sebastiao and Ribeiro 1996; Marchi et al. 2002). Of note, the concentration needed to enhance glutamate release (EC<sub>50</sub> = 1 pM) is almost 1,000-fold lower than is required for GABA release (EC<sub>50</sub> = 1 nM) (Cunha and Ribeiro 2000).

In vivo studies have shown that the neuroprotective effect of SCH58261 is observed only at a very low dose ( $0.01 \text{ mg kg}^{-1}$ ), 100–1,000 fold-lower than the dose used to stimulate motor activity ( $1\text{--}10 \text{ mg kg}^{-1}$ ) (Popoli et al. 2002, 2003). The enhancement of glutamate release by the  $A_{2A}R$  may be due to its positive coupling to the cAMP–protein kinase A (PKA) signaling pathway, leading to increased  $\text{Ca}^{2+}$  influx (Gubitz et al. 1996; Dunwiddie and Fredholm 1997). This facilitating effect of  $A_{2A}R$  agonists has been attributed to either an effect on glutamatergic terminals (Nikbakht and Stone 2001; Rosin et al. 2003) or an indirect effect via downregulation of  $A_1R$ -mediated inhibition (Lopes et al. 2002). Recent studies have indicated that glial  $A_{2A}Rs$  may also play an important role in the control of glutamate efflux by regulation of a specific glial glutamate transporter (GLT-1). The  $A_{2A}R$  agonist CGS21680 enhances glutamate efflux on cultured astrocytic glial cells from the cortex or brainstem (Li et al. 2001; Nishizaki et al. 2002). Voltage-clamp recording suggests that this effect occurs without affecting presynaptic glutamate release or postsynaptic glutamatergic conductance (Nishizaki et al. 2002). Thus,  $A_{2A}Rs$  significantly modulate glutamate release by presynaptic and glial mechanisms.

### ***3.2 Modulation of Cellular Survival Signals at Postsynaptic Sites***

Protection by adenosine receptor activity may also result from direct action on receptors at postsynaptic sites on neurons. While adenosine-mediated inhibition of neural activity in the central nervous system (CNS) is believed to be largely a product of  $A_1R$ -mediated presynaptic inhibition, activation of  $A_1R$  also results in G-protein-dependent activation of inwardly rectifying potassium channels at postsynaptic sites, leading to hyperpolarization of the resting membrane potential of postsynaptic neurons (Dunwiddie 1997). This could potentially explain how caffeine, via blockade of  $A_1Rs$ , increases firing of different neurons, including the cholinergic neurons regulating sleep–wakefulness (Rainnie et al. 1994; Oishi et al. 2008). In addition, this effect may be why high doses of caffeine can induce seizures. Such a view is supported by in vitro data demonstrating that caffeine acting at  $A_1Rs$  on glutamatergic neurons produces epileptiform activity in vitro (Dunwiddie 1980; Dunwiddie et al. 1981). On the other hand, activation of  $A_{2A}Rs$  inhibits NMDA-receptor-mediated synaptic currents in rat neostriatal neurons (Norenberg et al. 1997, 1998; Wirkner et al. 2000). This inhibition of NMDA receptor activity is mediated by the PKA pathway (Wirkner et al. 2000). Consequently, blockade of postsynaptic  $A_{2A}Rs$  is likely detrimental to striatal neurons. Consistent with this notion, recent studies have demonstrated that the  $A_{2A}R$  antagonists ZM241385 and SCH58261 potentiate quinolinic acid induced intracellular calcium efflux in striatal neurons (Popoli et al. 2002) and significantly amplify the excitotoxic effect of direct NMDA receptor stimulation, while reducing neurotransmitter release. Similarly, the  $A_{2A}R$  antagonist 8-(3-chlorostyryl)caffeine (CSC) has been found to potentiate NMDA-induced hippocampal toxicity (Robledo et al. 1999).

In addition, stimulation of the postsynaptic  $A_{2A}Rs$  was proposed to activate PKA to promote cell survival signals (Blum et al. 2003a). In PC12 cells, atypical protein

kinase C was found to act downstream of PKA to prevent apoptosis induced by serum withdrawal (Huang et al. 2001). In addition,  $A_{2A}R$  stimulation rescues the blockage of nerve growth factor (NGF)-induced neurite outgrowth by enhancing the phosphorylation of the cAMP-response element-binding (CREB) protein when the NGF-evoked MAPK pathway is damaged (Cheng et al. 2002), or by direct binding to TRAX when the p53-mediated pathway is blocked (Sun et al. 2006). Consistent with a protective role of the  $A_{2A}R$ , an  $A_{2A}R$ -selective agonist (CGS21680) effectively ameliorates several major symptoms of HD in a transgenic mouse model (R6/2) of HD (Chou et al. 2005). At least part of the beneficial effect of CGS21680 in HD mice can be attributed to the PKA-mediated enhancement of ubiquitin proteasome activity (Chiang et al. 2009).

### 3.3 Control of Neuroinflammation in the CNS

Vascular and inflammatory effects, including  $A_{2A}R$ -mediated vasodilation (Winn et al. 1985; Ngai et al. 2001), inhibition of platelet aggregation (Sandoli et al. 1994; Ledent et al. 1997), and suppression of superoxide species generation by neutrophils (Cronstein et al. 1983, 1990) may underlie the protection offered by  $A_{2A}R$  agonists. Particularly, activation of  $A_{2A}Rs$  inhibits the production of proinflammatory cytokines such as tumor necrosis factor  $\alpha$ , interleukin-6, and interleukin-12 (Hasko et al. 2000; Mayne et al. 2001; Ohta and Sitkovsky 2001; Day et al. 2003, 2004; Sitkovsky et al. 2004). This effect could be responsible for the neuroprotection afforded by  $A_{2A}R$  agonists in cerebral hemorrhage (Mayne et al. 2001), spinal cord injuries (Cassada et al. 2002b), and other tissue-damaging insults (Ohta and Sitkovsky 2001; Day et al. 2003). Largely on the basis of  $A_{2A}R$  action in peripheral tissues, extracellular adenosine acting at  $A_{2A}Rs$  has been proposed as an endogenous circuit breaker that inhibits inflammation and limits extensive inflammatory tissue damage (Sitkovsky et al. 2004). On the other hand, in a recent study using chimeric mice, selective inactivation of  $A_{2A}Rs$  in bone marrow cells (generated by transplantation) attenuated ischemia-induced expression of proinflammatory cytokines and reduced ischemic brain injury (Yu et al. 2004). This may reflect differential effects of  $A_{2A}R$  activation on inflammation in the peripheral nervous system versus the CNS. It may also reflect complex (both potentially deleterious and neuroprotective) effects of  $A_{2A}R$  activation in glial cells, including regulation of glutamate efflux (as described above), upregulation of cyclooxygenase 2, modulation of the activities of nitric oxide synthase, production of proinflammatory prostaglandins and cytokines, as well as microglial activation (Fiebich et al. 1996; Brodie et al. 1998). The precise influence of  $A_{2A}R$  activation on brain glial cells remains to be determined. Recently, it was proposed that caffeine metabolites can inhibit the nuclear enzyme poly(ADP-ribose)polymerase-1 at concentrations attainable by normal human caffeine consumption (Geraets et al. 2006), suggesting another possible mechanism for caffeine modulation of the inflammatory response in the brain.



### **3.4 Interaction with Neurotrophic Factors**

The Trk receptors for neurotrophic factors (such as NGF, BDNF, and glial-cell-line-derived neurotrophic factor, GDNF) are an important class of membrane receptors that involve autophosphorylation in tyrosine residues as a result of ligand binding, triggering a signaling cascade associated with regulation of cell death, survival, and differentiation (Hu and Russek 2008). The cross talk between adenosine receptors, particularly the A<sub>2A</sub>R, and the receptor for neurotrophic factors involving tyrosine receptor kinase, has been demonstrated (for a review, see Sebastiao and Ribeiro 2009b). The A<sub>2A</sub>R was also shown to transactivate neurotrophin receptors and enhance their trophic functions (Lee and Chao 2001; Wiese et al. 2007; Sebastiao and Ribeiro 2009a) on BDNF-mediated synaptic transmission (Diogenes et al. 2004; Tebano et al. 2008) and long-term potentiation (Fontinha et al. 2008) and GDNF-mediated action in striatal dopaminergic terminals (Gomes et al. 2006). Indeed, systemic administration of an A<sub>2A</sub>-selective antagonist (SCH58261) decreased the level of BDNF in the brain (Domenici et al. 2007). These findings collectively warrant further evaluation of the complex roles of adenosine drugs and methylxanthine in neurodegenerative diseases in the future.

### **3.5 Regulation of Blood–Brain Barrier Integrity**

The blood–brain barrier (BBB) is a physical, immune, and metabolic barrier to protect the microenvironment of brain from the systemic circulation. BBB breakdown occurs in a variety of neurological disorders, including stroke (Latour et al. 2004), TBI (Hoane et al. 2006), AD (Zipser et al. 2007), PD (Kortekaas et al. 2005; Rite et al. 2007), and MS (Minagar and Alexander 2003), and may contribute to PD pathogenesis (Rite et al. 2007). Thus, improving BBB integrity represents an important mechanism by which neuroprotective agents such as caffeine may exert neuroprotective effects. Indeed, caffeine has been shown to protect against BBB dysfunction induced by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) and high cholesterol in animal models of PD and AD (Chen et al. 2008a, b).

## **4 Methylxanthine/Caffeine and Neurodegenerative Disorders**

### **4.1 Stroke**

During ischemic insults, the level of extracellular adenosine increases rapidly and markedly (Hagberg et al. 1987; Matsumoto et al. 1992; Latini and Pedata 2001). An increased level of extracellular adenosine is generally considered an important neuroprotective mechanism against ischemic injury (Rudolphi et al. 1992; de Mendonça et al. 2000; Latini and Pedata 2001; Boison 2006; Cunha et al. 2006;



Chen et al. 2007). Indeed, reduction of the level of extracellular adenosine by transgenic overexpression of adenosine kinase – the primary adenosine-metabolizing enzyme – rendered the mouse brain more susceptible to ischemic cell death (Pignataro et al. 2007).

#### 4.1.1 Human Studies of Methylxanthine/Caffeine

Given the widespread and extensive use of dietary caffeine, the impacts of coffee consumption on stroke have been extensively investigated in many different settings, with somewhat inconsistent findings. Some of the ambiguity as to whether caffeine is a risk factor for stroke might result from poor estimation of dietary caffeine consumption and/or a lack of information on systemic levels of caffeine and its metabolites in the body (James 2004). The potential contribution of caffeine to stroke is of particular interest because caffeine at dietary doses enhances blood pressure owing to direct vasoconstriction caused by adenosine receptor blockage (Pincomb et al. 1988; James 2004), and therefore caffeine is considered a cardiovascular risk factor (Greenland 1993). Interestingly, an earlier study of 45,589 men showed that total caffeine consumption was not associated with the risk of coronary heart disease or stroke (Grobbee et al. 1990). Consistent with this notion, a recent report of a 24-year follow-up study of 83,076 women also suggested that chronic coffee consumption is not a risk factor for stroke in women. However, a higher dose of coffee intake by hypertensive men (55–68 years old, 499 patients) was associated with a higher risk of thromboembolic stroke, but not of hemorrhagic stroke (Grobbee et al. 1990). In addition, heavy caffeine use (five drinks per day or pharmaceutical products) was found to be linked to intracerebral hemorrhage (18–49 years old, 217 patients; Feldmann et al. 2005) and subarachnoid hemorrhage (18–49 years old, 312 cases; Broderick et al. 2003). Since caffeine at dietary doses suppresses cerebral blood flow, caution may be required for patients recovering from acute ischemia stroke (Ragab et al. 2004). However, these effects of caffeine on cerebral blood flow may not occur after long-term use (Addicott et al. 2009).

Owing to their vascular effects, a combination of caffeine and ethanol (the two most commonly consumed substances by humans, designated caffeineol) was shown to reduce cortical infarct areas in rat ischemia models (Strong et al. 2000; Aronowski et al. 2003). Although the protection by caffeineol in cortical areas is effective, caffeineol exerts no beneficial effects in subcortical regions, where most human strokes occur (Belayev et al. 2004; Hoyte et al. 2004). In a rabbit embolic ischemia model which is more similar to human stroke than the rat model (Lapchak et al. 2002), caffeineol by itself exhibited no neuroprotective function (Lapchak et al. 2004). This discrepancy between the effects of caffeineol in rat and rabbit models might result from the fact that the stroke induced in rabbits was more heterogeneous than that in the rat model. Moreover, when caffeineol was combined with a plasminogen activator (t-PA, an FDA-approved treatment for acute ischemic stroke; Lapchak et al. 2002), the risk of hemorrhage increased, which somewhat dampened the enthusiasm for using caffeineol in stroke patients

(Lapchak et al. 2004). Fortunately, in pilot studies of patients with acute ischemia, caffeinol alone, caffeinol plus t-PA, or caffeine plus tissue-type plasminogen activator plus hypothermia evoked no adverse side effects, and therefore might be safe for patients with acute stroke (Piriyawat et al. 2003; Martin-Schild et al. 2009). Further studies are required to evaluate the efficacy of the above-mentioned combined treatments for acute stroke. Theophylline and its synthetic analog aminophylline, when administered alone to stroke patients, did not produce significant beneficial effects on ischemic stroke (Geismar et al. 1976; Britton et al. 1980).

#### 4.1.2 Experimental Studies on Methylxanthine

A major part of methylxanthine's function can be attributed to adenosine receptors (particularly the  $A_1R$  and  $A_{2A}R$ ) (Fredholm et al. 2001; Yang et al. 2009a). Indeed, mice expressing half the normal levels of the  $A_1R$  and the  $A_{2A}R$  behave very similarly to those exposed to chronic caffeine (Yang et al. 2009b). In a rat cerebral ischemia model created by occlusion of the carotid arteries, theophylline worsened injury in the hippocampus with both normoglycemic and hyperglycemic ischemia (Zhou et al. 1994; Higashi et al. 2002). This experimental paradigm is of great interest because the amount of adenosine released during hyperglycemic ischemia is much lower than that released during normoglycemic ischemia (Hsu et al. 1991). Note that theophylline is a nonselective antagonist of  $A_1R$  and  $A_{2A}R$ . In the same ischemia model, an  $A_1R$ -predominant agonist (cyclohexyladenosine) markedly reduced hippocampal injury under normoglycemic and hyperglycemic ischemia (Zhou et al. 1994), while an  $A_{2A}R$ -selective antagonist (ZM 241385) reduced brain damage and memory function loss in hyperglycemic ischemia (but not in normoglycemia) (Higashi et al. 2002). The authors speculated that the  $A_1R$  is likely to exert a protective function against ischemic damage, and that the beneficial effects of  $A_{2A}R$  blockage during hyperglycemic ischemia might be the result of potentiating the activity of the  $A_1R$  which attenuates glutamate release during ischemia (Heron et al. 1993; Pedata et al. 2001; Higashi et al. 2002). Effective protection against overall ischemia by other  $A_{2A}R$  antagonists (e.g., SCH58261, CSC, and 4-amino-1-phenyl[1,2,4]-triazolo[4,3-*a*] quinoxaline) was reported in rodent ischemia models (Phillis 1995; Monopoli et al. 1998). Conversely, antagonists (e.g., 8-cyclopentyl-1,3- dipropylxanthine, DPCPX) of the  $A_1R$  were found to exacerbate ischemia-evoked damage (Phillis 1995), which suggested a protective role of the  $A_1R$  in ischemia. Nonetheless, although glutamatergic transmission is markedly affected by antagonists of the  $A_1R$ , elimination of the  $A_1R$  did not affect ischemia outcome owing to compensatory effects (Olsson et al. 2004). In addition, glutamate release and neuroprotection by (-)- $N^6$ -(*R*)-(phenylisopropyl) adenosine do not correlate well. Glutamate release, therefore, might not be the sole pathway underlying the neuroprotective effect of the  $A_1R$  (Heron et al. 1994). An intriguing finding is that chronic treatment with  $A_1R$  agonists or antagonists produced effects opposite to those of acute treatment (Von Lubitz et al. 1994). In a gerbil forebrain ischemia model, acute treatment with an  $A_1R$ -selective agonist

(*N*<sup>6</sup>-cyclopentyladenosine, CPA) protected neurons in the hippocampal CA1 region, while chronic treatment with CPA (15 days) shortened lifespan and decreased neuronal survival (Von Lubitz et al. 1994). The molecular mechanisms underlying the distinct effects of acute and chronic treatment with A<sub>1</sub>R agonists are currently unknown. The change in response to chronic treatment might reflect compensatory adaptations. In contrast, the acute and chronic effects of A<sub>2A</sub>R drugs on stroke/ischemia are similar (Von Lubitz et al. 1995), probably owing to the relatively stable expression of the A<sub>2A</sub>R during chronic treatment with agonists or antagonists (Lupica et al. 1991; Abbracchio et al. 1992; Chern et al. 1993; Adami et al. 1995). Collectively, these data indicate that adenosine receptors are major players in modulating ischemic injury in the brain and in mediating the actions of methylxanthines during ischemia.

The A<sub>2A</sub>R is considered critically important in stroke because genetic inactivation of the A<sub>2A</sub>R attenuates cerebral infarction and decreases the severity of neurological deficits caused by focal ischemia (Chen et al. 1999, 2007). Acute administration of two A<sub>2A</sub>R antagonists (CSC and CGS15943) (Gao and Phillis 1994; Von Lubitz et al. 1995) was found to significantly suppress neurological deficits in a global hypoxia model in gerbils. Similarly, another A<sub>2A</sub>R antagonist (SCH58261) attenuated the release of excitatory amino acids, reduced infarct volume, and ameliorated turning behavior (Monopoli et al. 1998; Melani et al. 2003) in a rat focal ischemia model. In addition to inhibiting the release of glutamate, blocking the A<sub>2A</sub>R modulated the activities of other brain cells. For example, inhibition of the A<sub>2A</sub>R in microglia after ischemia blunted activation of MAPK and might reduce the damaging inflammatory response evoked by ischemia (Melani et al. 2006). Moreover, blocking the A<sub>2A</sub>R in oligodendrocytes suppresses the c-Jun N-terminal kinase MAPK pathway, obstructs the activity of glial scar inhibitor molecules, and might consequently prohibit myelin disorganization (Melani et al. 2009). Surprisingly, brain cells are not the only or even the major cell types responsible for the beneficial functions of A<sub>2A</sub>R inhibition during ischemia. Using A<sub>2A</sub>R-knockout mice, Yu et al. (2004) demonstrated that the A<sub>2A</sub>R on bone-marrow-derived cells plays the key role in the detrimental function of the A<sub>2A</sub>R during ischemia, further strengthening the primary role of proinflammatory mediators in stroke.

Because of the importance of both the A<sub>1</sub>R and the A<sub>2A</sub>R in stroke, the expression and function of both adenosine receptors have been actively investigated. In young animals (7 days old) subjected to unilateral ischemia expression of both receptors was markedly reduced in the cortex and hippocampus of the damaged hemisphere (Aden et al. 1994). This would clearly affect the efficacy of adenosine drugs for treating ischemia in young animals. In adult animals, expression of A<sub>1</sub>R during ischemia appears to depend on the experimental paradigm and the extent of ischemic damage. Brief hypoxia evoked internalization and desensitization of the A<sub>1</sub>R in hippocampal slices (Coelho et al. 2006), but there was no significant change in A<sub>1</sub>R function or protein levels in the hippocampus following a four-vessel-occlusion ischemia model (Shen et al. 2002). It should also be pointed out that chronic treatment with caffeine alters the expression profiles of many

important signaling molecules, including the  $A_1R$  and several non-adenosine receptors (Shi et al. 1993; Justinova et al. 2009), which might also contribute to the long-term impact of caffeine consumption.

As suggested in the previous paragraph, the function, regulation, and expression of the  $A_{2A}R$  in young animals might be distinct from what has been observed in adult animals. This is an important issue because cerebral hypoxic ischemia is one of the major brain injuries of newborns. Genetic inactivation of the  $A_{2A}R$  exacerbates the brain damage evoked by cerebral hypoxic ischemia in young mice (7 days old), suggesting a protective role for the  $A_{2A}R$  in young animals (Aden et al. 2003). This finding is consistent with an earlier study showing that caffeine ( $50 \text{ mg kg}^{-1}$  by intraperitoneal injection three times a day) caused apoptosis in multiple brain areas in young rats (7 days old) and in primary cortical cell culture (Kang et al. 2002). Similarly, in contrast to the protective role of the  $A_1R$  in adult animals, the  $A_1R$  in the brain of neonatal mice mediates the hypoxia-induced ventriculomegaly (Turner et al. 2003). Studies on the distribution and signaling pathways of adenosine receptors (particularly the  $A_1R$  and the  $A_{2A}R$ ) in the brain of neonatal animals are required to evaluate whether there is any potential therapeutic use for adenosine drugs and methylxanthines in treating hypoxic–ischemic brain damage in newborns.

## 4.2 Traumatic Brain Injury

TBI remains one of the leading causes of disability and death in modern society (Marshall 2000; Rutland-Brown et al. 2006; Moppett 2007). Despite extensive efforts, no effective neuroprotective therapy is currently available for TBI. The neurological deficits found following TBI result from both primary and secondary (delayed) events (Graham et al. 2000; Leker and Shohami 2002; Moppett 2007). Primary damage to the brain cells and tissues is not reversible, while secondary damage is reversible and potentially preventable. The potential for reversing damage closely correlates with prognosis. Three possible mechanisms have been proposed for secondary brain injury in TBI: glutamate release, cytokine production, and  $\text{Ca}^{2+}$  overloading (Graham et al. 2000; Marshall 2000; Leker and Shohami 2002). Interfering with any of these pathological processes may reduce brain damage.

Adenosine levels are markedly increased both in brain interstitial fluid and in cerebrospinal fluid (CSF) after severe TBI in humans (Clark et al. 1997; Bell et al. 1998; Robertson et al. 2001). Adenosine has complex effects in experimental models of CNS trauma – both neuroprotection and neurotoxicity have been reported (Cassada et al. 2002a; Varma et al. 2002).

### 4.2.1 Human Studies

In clinical studies, Sachse et al. (2008) assessed concentrations of caffeine and its metabolites (theobromine, paraxanthine, and theophylline) in 97 ventricular CSF samples from an established bank of samples from 30 adults with severe TBI. Nine

of the 24 TBI patients with detectable caffeine levels had levels of  $1 \mu\text{M}$  or greater ( $194 \text{ ng mL}^{-1}$ ), the concentration chosen as the clinically significant threshold. The study found that the caffeine levels in CSF correlated with prognosis, with higher caffeine level predicting better clinical outcome.

#### 4.2.2 Experimental Studies

In experimental TBI, caffeine and adenosine receptor agents can be neuroprotective or neurotoxic, depending on the dose, model, and timing. In rat models, acute administration of extremely large doses of caffeine ( $100\text{--}150 \text{ mg kg}^{-1}$ , which equates with a human acutely consuming more than cups of caffeinated beverage) immediately before injury worsened the outcome (Al Moutaery et al. 2003). In contrast, in mice, chronic pretreatment with caffeine ( $5\text{--}50 \text{ mg kg}^{-1}$ ) reduced hippocampal neuronal cell death after experimental TBI (Washington et al. 2005). Similarly, Li et al. (2008) demonstrated that at 24 h after a cortical impact brain injury, neurological deficits, cerebral edema, and inflammatory cell infiltration were all significantly attenuated in mice pretreated chronically (for 3 weeks) with caffeine in their drinking water. By contrast, acute treatment with caffeine (30 min before TBI) either had no effect (Li et al. 2008) or exacerbated TBI-induced brain injury (Al Moutaery et al. 2003). Thus, acute caffeine treatment adversely affects outcome after TBI, while chronic treatment provides a consistent neuroprotective effect. Lastly, the combination of caffeine and ethanol (“caffeinol”) produces a beneficial effect after experimental TBI (Dash et al. 2004), mirroring its effect on stroke (Aronowski et al. 2003; Belayev et al. 2004) and indicating the importance of the interaction between caffeine and ethanol. Thus, chronic caffeine consumption may confer some neuroprotection against TBI and an epidemiological investigation into the possible association between human caffeine intake and TBI outcome is warranted.

It is suggested that the neuroprotective effect of caffeine may be due to long-term upregulation of  $A_1\text{Rs}$  or acute inhibition of  $A_{2A}\text{Rs}$ , along with other potential mechanisms. The attenuation of brain injury by chronic (but not acute) caffeine treatment suggests that enhanced  $A_1\text{R}$ -mediated suppression of excessive glutamate release and production of inflammatory cytokines may be responsible for this effect. In support of this notion, chronic caffeine treatment attenuated glutamate release and the production of inflammatory cytokines and upregulated  $A_1\text{R}$  messenger RNA in the brain (Li et al. 2008). Additional support for the role of the  $A_1\text{R}$  comes from the demonstration that administration of an  $A_1\text{R}$  agonist attenuates the behavioral deficits and contusion volume associated with brain injury, while an  $A_1\text{R}$  antagonist exacerbates them (Varma et al. 2002). Similarly,  $A_1\text{R}$ -knockout mice develop lethal acute status epilepticus after experimental TBI (Kochanek et al. 2006). On the other hand, the  $A_{2A}\text{R}$  activity may also contribute to the protection against TBI. Administration of an  $A_{2A}\text{R}$  agonist conferred beneficial effects on outcome and cerebral blood flow after experimental spinal cord injury or TBI (Cassada et al. 2002a; Reece et al. 2004). However, genetic inactivation of

A<sub>2A</sub>Rs reduced neuronal apoptosis in a moderate cortical impact model (Li et al. 2009) and in a spinal cord compression injury model of mice (Li et al. 2006). Thus, both activation and inactivation of A<sub>2A</sub>Rs have been shown to produce a neuroprotective effect in the early stage of TBI. The mechanism underlying these apparent paradoxical effects of the A<sub>2A</sub>R activity in TBI remains unclear.

### 4.3 Alzheimer's Disease

AD is the most common cause of dementia, afflicting about 5% of the population older than 65 years and 10–20% at age 80 (Braak and Braak 1997; Allouf et al. 1998). While a small fraction of autosomal dominant, early onset forms of AD are caused by genetic mutations of genes encoding the amyloid precursor protein (APP), presenilin 1 (PS-1), and presenilin 2 (PS-2), the cause of the most common form of AD – sporadic, late-onset AD – remains unknown (Selkoe 2004; Bertram and Tanzi 2008). Polymorphisms of other genes, notably apolipoprotein E (APOE ε4), have been associated with the late form of AD (Strittmatter et al. 1993; Bertram and Tanzi 2008). These predisposing gene variations likely interact with environmental factors, including dietary factors, in the pathogenesis of AD. There are now treatments to ameliorate the cognitive symptoms of AD, but no interventions yet exist that slow or reverse the progressive pathogenic changes of AD (Mattson 2004; Selkoe 2004). Recent efforts to identify potential risk factors, as well as factors that may decrease risk or prolong autonomy, have provided epidemiological evidence that caffeine, which is known to have positive effects on vigilance, attention, mood, and arousal, may also be neuroprotective in AD.

#### 4.3.1 Human Studies

Early studies of brain tissue from patients who died with a confirmed diagnosis of AD revealed a loss of A<sub>1</sub>R in the hippocampus, a brain region critically involved in learning and memory (Jansen et al. 1990; Jaarsma et al. 1991; Ulas et al. 1993), suggesting possible involvement of the adenosine receptor in the development of AD. This reduction of A<sub>1</sub>R in the hippocampus was recently confirmed by a PET study of AD patients using the A<sub>1</sub>R antagonist <sup>11</sup>C-8-dicyclopropylmethyl-3-propylxanthine as a ligand (Fukumitsu et al. 2008). In contrast, the levels of A<sub>1</sub>R and A<sub>2A</sub>R appear to be increased in the frontal cortex in AD, in parallel with increased functional activity of these receptors (Albasanz et al. 2008). Finally, a strong negative correlation between increased plasma levels of homocysteine and reduced levels of adenosine has been reported in AD (Selley 2004). This is due at least in part to formation of *S*-adenosylhomocysteine, suggesting the possibility that a deficiency of adenosine may contribute to neurological manifestation of increased homocysteine levels.

Several longitudinal studies have reported recently that a daily caffeine intake equivalent to three or more cups of coffee reduces cognitive decline in “non-demented” elderly men and women. For example, a significant association between regular coffee intake and improvement of cognitive performance in older subjects

(55+) and in women was found in two large cross-sectional population studies (Jarvis 1993; Johnson-Kozlow et al. 2002). This relationship is further substantiated by the prospective population study, Maastricht Aging Study (MAAS Study) in southern Netherlands (van Boxtel et al. 2003). Despite inadequate adjustment for other potential causes of cognitive change, the study found a cross-sectional association between caffeine consumption and improvements in psychomotor speed long term (Hameleers et al. 2000; van Boxtel et al. 2003) and verbal memory performance (van Boxtel et al. 2003), supporting the enhancement of cognitive function by caffeine in the “nondemented” elderly population.

Maia and de Mendonca (2002) first reported in a small case-control study that 78 AD patients had consumed markedly less caffeine during the 20 years preceding AD diagnosis than age-matched individuals without AD. Four recent prospective studies of large cohorts also support the inverse relationship between caffeine consumption and reduced risk of developing AD. The Canadian Study of Health and Aging (CSHA) is a large, nationwide, multicenter, longitudinal study of dementia in elderly people focusing on risk factors for AD. After surveying 4,615 subjects at 5-year follow-up, including 194 with AD and 3,894 cognitively normal controls, this population-based, prospective study found that regular consumption of caffeine is associated with a reduced risk of developing AD (Lindsay et al. 2002). In addition, the 10-year follow-up of the FINE study, involving 667 healthy men born in Finland, Italy, and the Netherlands between 1900 and 1920, suggests that consuming coffee reduces cognitive decline in elderly men, with the least cognitive decline for men consuming three cups of coffee per day. This inverse relationship has now been confirmed by yet another large population-based prospective study, the French Three Cities study involving 4,197 women and 2,820 men (Ritchie et al. 2007). This large prospective study examined the impact of caffeine use on cognitive functioning over time, taking into account multiple possible codeterminants of cognitive decline and thus clarifying previous small cross-sectional control studies. The study found that women (but not men) drinking three cups or more of coffee exhibited less decline in verbal retrieval and visuospatial memory over 4 years, even adjusting for other multiple factors contributing to cognitive decline. Lastly, the Cardiovascular Risk Factors, Aging, and Dementia (CAIDE) study addresses the possible relationship between coffee/tea drinking at midlife and development of dementia later in life (Eskelinen et al. 2009). This study involved 1,409 individuals aged 65–79 after an average 21-year follow-up (total of 61 cases identified as demented, 48 with AD) and found that individuals identified as coffee drinkers at midlife were 65% less likely to develop dementia and AD later in life compared with those drinking little or no coffee. This association is observed after adjusting for demographic, lifestyle, and vascular factors, as well as APOE  $\epsilon$ 4 allele and depressive symptoms. Thus, caffeine consumption may represent a potential prevention strategy for dementia/AD.

Interestingly, the xanthine derivative propentofylline has been specifically developed for and tested in clinical trials of AD. Propentofylline, a weak antagonist at adenosine receptors and an effective inhibitor of adenosine reuptake and of PDE, has been shown to enhance cognition in patients with vascular dementias (Mielke et al. 1996a, b). The initial study suggested that propentofylline might attenuate



progression of AD (Kittner et al. 1997; Marcusson et al. 1997), but a subsequent large clinical trial has not confirmed these encouraging findings (Propentylfilline Plus Study). The lack of efficacy of this drug may be due to its relatively short-term use in this study. In AD, neurodegeneration may evolve for many years before the emergence of clinical symptoms and short-term treatment with neuroprotective agents is unlikely to markedly modify the progression of neurodegeneration.

### 4.3.2 Experimental Studies

Despite the considerable strength of the correlation in these large, longitudinal studies, epidemiological investigations cannot definitively isolate caffeine intake from other lifestyle choices that potentially affect cognition. Studies in cellular and animal models of AD have now provided evidence to address the causal relationship between caffeine's protective effect and reduced cognitive decline in humans. The first evidence of neuroprotection by caffeine in AD models was in cultured cerebellum granular cells where caffeine and the  $A_{2A}R$  antagonist SCH58261 reduced amyloid  $\beta$  ( $A\beta$ ) peptide induced aggregation, a key event associated with AD pathogenesis (Dall'Igna et al. 2003). This protective effect was substantiated in an animal study in which caffeine and  $A_{2A}R$  antagonists protected against loss of learning and memory induced by intracerebroventricular infusion of  $A\beta$  peptide (Dall'Igna et al. 2003; Cunha et al. 2008). These findings are also in agreement with pharmacology studies showing that caffeine and  $A_{2A}R$  antagonists reverse memory loss induced by aging (Prediger et al. 2005) and by spontaneous hypertension (Prediger and Takahashi 2005) and also attenuate neurochemical modifications in the hippocampus of streptozotocin-induced diabetic rats (Duarte et al. 2009). Studies of aged AD transgenic (APPsw, Swedish mutation) mice with high levels of brain  $A\beta$  and widespread cognitive impairment provide evidence for the therapeutic benefit of caffeine. The study found that long-term (between 4 and 9 months) administration of a 1.5 mg daily dose of caffeine (equivalent to 500 mg in human) to APPsw mice can reduce brain  $A\beta$  levels and protect against certain cognitive impairments (Arendash et al. 2006). This protective effect of caffeine is associated with reduced expression of both PS-1 and  $\beta$ -secretase (BACE1), thereby suppressing production of  $A\beta$  in the brain (Arendash et al. 2006). Furthermore, in aged (18–19-month-old) APPsw mice, which already exhibit decreased cognitive function, caffeine treatment enhanced working memory compared with nontreated APPsw mice (Arendash et al. 2009). Moreover, acute caffeine treatment rapidly reduces production of  $A\beta$  in both brain interstitial fluid and plasma without affecting  $A\beta$  elimination (Cao et al. 2009). Long-term oral caffeine treatment not only sustainably reduces the plasma levels of  $A\beta$ , but also decreases the levels of both soluble and deposited  $A\beta$  in hippocampus and cortex in aged AD mice. Intriguingly, caffeine's ability to improve cognitive performance in individual aged AD mice did not correlate with reduced plasma  $A\beta$  levels, but was closely associated with reduced inflammatory cytokine levels in hippocampus (Arendash et al. 2009; Cao et al. 2009). Lastly, caffeine was recently shown to reduce disruption



of the BBB induced by a high-cholesterol diet (Chen et al. 2008b), suggesting additional mechanisms for the therapeutic effect of caffeine in AD and other neurodegenerative disorders.

#### 4.4 Parkinson's Disease

PD is the second most common neurodegenerative disorder, affecting 1% of the world population aged 65 and older (Lang and Lozano 1998a, b). Dopamine replacement, such as L-dopa, remains the mainstay in treatment to control motor symptoms even 40 years after its discovery (Rascol et al. 2002; Lang and Obeso 2004; Goetz et al. 2005). As a consequence, the clinical problems associated with treatment, particularly with long-term management of PD, remain largely unchanged. After 5–10 years of L-dopa treatment, loss of L-dopa/dopamine agonist efficacy and the onset of debilitating motor complications (dyskinesia, wearing off, and on–off) are almost inevitable (Ahlskog and Muentner 2001). Other side effects of the dopamine therapy, including psychosis and dopamine dysregulation syndromes (e.g., compulsive gambling, hypersexuality), are difficult to manage (Stamey and Jankovic 2008; Yamamoto and Schapira 2008). Furthermore, non-motor symptoms of PD (e.g., cognitive dysfunction, fatigue, balance impairment, sleep disturbance, autonomic dysfunction) are recognized as a key component of the illness and present another unmet therapeutic need (Chaudhuri et al. 2006). There is no effective therapy to slow or halt progression of PD.

The A<sub>2A</sub>R recently emerged as a leading nondopaminergic therapeutic target in PD. This is primarily the result of three lines of experimental and clinical investigation. First, the unique colocalization of A<sub>2A</sub>R–D<sub>2</sub>R in striatopallidal neurons and the antagonistic interaction between A<sub>2A</sub>R and D<sub>2</sub>R, which may occur through A<sub>2A</sub>R–D<sub>2</sub>R heterodimer formation, provide a strong anatomical and molecular basis for the motor benefit of A<sub>2A</sub>R antagonists in PD (Ferre et al. 1991; Richardson et al. 1999; Schwarzschild et al. 2006). Second, on the basis of decade-long preclinical studies, A<sub>2A</sub>R antagonists such as KW-6002 (istradefylline) and SCH420814 have now completed clinical phase II–III trials, successfully confirming a motor benefit in advanced PD patients and leading to FDA filing of KW-6002 for PD patients (April, 2007, see <http://www.kyowa-kpi.com>). Third, the recent convergence of epidemiology and animal studies strongly suggests that A<sub>2A</sub>R antagonists also confer a neuroprotective effect in PD.

##### 4.4.1 Human Studies

###### Relief of Motor Symptoms

The initial results showed that the A<sub>2A</sub>R antagonist KW-6002 (istradefylline, 20–80 mg day<sup>-1</sup>) enhanced motor activity and potentiated the motor stimulant effect elicited by low doses of L-dopa, as evidenced by reduced “off” time and

increased “on” time in advanced PD patients (Bara-Jimenez et al. 2003; Hauser et al. 2003). To confirm these initial findings, several double-blind, placebo-controlled, clinical phase IIB and III trials of KW-6002 in advanced PD patients were conducted. Four clinical IIB and III trials (Stacy et al. 2008; Jenner et al. 2009), with a total of about 1,500 advanced PD patients, have been reported so far, with a reduction in the average “off” time of about 1.7 h, compared with the “optimal” L-dopa dose regime (Stacy et al. 2008; Jenner et al. 2009). Thus, these results demonstrate the motor benefits of KW-6002 in advanced PD patients. As a result of these clinical findings, KW-6002 has been filed for FDA (USA) approval for the treatment of advanced PD. Similarly, a preliminary report by the Schering-Plough Research Institute at the conference “Targeting the  $A_{2A}R$  to PD and Other CNS Disorders” (Boston, May, 2006) showed that the  $A_{2A}R$  antagonist SCH420814 also produced motor benefits in advanced PD patients in a clinical phase IIA trial (see <http://www.a2apd.org> for the abstract). Despite some limitations of these clinical trials, such as a relatively large drop-off rate, and some unrealistic clinical trial settings, and the admittedly modest effects (Jankovic 2008), these studies support the concept that selective  $A_{2A}R$  antagonists can stimulate motor activity by potentiating the L-dopa effect in advanced PD patients. Recent clinical trials indicate that KW-6002 administered with the existing L-dopa treatment appears to increase the incidence of dyskinesia in advanced PD patients; however, this increased dyskinesia was reported as benign by most patients (Stacy et al. 2008). Thus, it is not clear whether  $A_{2A}R$  antagonists can modify L-dopa-induced dyskinesia (LID).

### Potential Disease Modifying Effect

The most exciting prospective role for  $A_{2A}R$  antagonists as a novel therapy for PD is their potential to attenuate dopaminergic neurodegeneration, as suggested by convergent epidemiological and experimental evidence. Ross et al. (2000) reported an inverse relationship between consumption of the nonselective adenosine antagonist caffeine and the risk of developing PD in a 30-year follow-up study in a large prospective study of 8,004 Japanese-American men in the Honolulu Heart Program. The age- and smoking-adjusted risk of PD was 5 times higher among men who reported no coffee consumption compared with men who reported a daily consumption of 28 oz or more of coffee. This finding was substantiated by a similar inverse relationship between the consumption of caffeinated (but not decaffeinated) coffee and the risk of developing PD in two larger, more ethnically diverse cohorts – the Health Professionals’ Follow-Up Study and the Nurses’ Health Study – involving 47,351 men and 88,565 women (Ascherio et al. 2001) and also more recently in the Finnish Mobile Clinic Health Examination Survey involving 19,518 men and women (Saaksjarvi et al. 2007). These studies firmly established a relationship between increased caffeine consumption and decreased risk of developing PD in men.

Despite the strong epidemiological evidence for a neuroprotective effect of caffeine, two recent pilot studies did not detect a positive correlation between caffeine consumption and PD progression in 1-year clinical trials involving a total of 413 early PD subjects (Simon et al. 2008). However, these studies are limited by short duration (1 year) of the follow-up and relatively small numbers of PD cases. In another study, no significant association was noted in a small case-control study of PD (94 cases) and  $A_{2A}R$  gene polymorphism ( $A_{2A}R$  1976T>C) (Hong et al. 2005). Additional clinical studies are clearly warranted to assess this critical issue.

#### 4.4.2 Experimental Studies

Over the last decade, several specific  $A_{2A}R$  antagonists (such as KW-6002 and SCH58261, ST1535) have been developed and shown to enhance motor activity in animal models of PD (Richardson et al. 1997; Schwarzschild et al. 2002; Chen 2003; Tronci et al. 2007; Varty et al. 2008; Trevitt et al. 2009). In rodents depleted of dopamine by MPTP or reserpine treatment or rendered cataleptic by haloperidol, the administration of selective  $A_{2A}R$  antagonists increases motor activity. Animals treated with  $A_{2A}R$  antagonists following unilateral lesion with 6-hydroxydopamine (6-OHDA) (hemiparkinsonian) demonstrated an increase in contralateral rotation (Ferre et al. 1992, 1997; Shiozaki et al. 1999; Tronci et al. 2007). Similarly,  $A_{2A}R$  antagonists stimulate motor activity in MPTP-treated nonhuman primates (Kanda et al. 1998a; Grondin et al. 1999; Varty et al. 2008). Thus,  $A_{2A}R$  antagonists can stimulate motor activity in dopamine-depleted animals either alone or in synergy with L-dopa and other dopaminergic agonists. These preclinical studies set the stage for clinical trials to evaluate the ability of  $A_{2A}R$  antagonists to relieve motor symptoms in PD patients.

To establish the causal relationship between caffeine consumption and neuroprotection against dopaminergic neurodegeneration, studies with animal models of PD provide a compelling clue about the potentially protective effects of caffeine by demonstrating that pharmacological blockade (by caffeine or selective  $A_{2A}R$  antagonists) or genetic depletion of the  $A_{2A}R$  attenuates dopaminergic neurotoxicity and neurodegeneration (Chen et al. 2001; Ikeda et al. 2002; Xu et al. 2002). Administration of caffeine following MPTP treatment attenuates the MPTP-induced reduction in dopamine content and loss of dopaminergic terminals in the striatum (Chen et al. 2001; Xu et al. 2002) as well as the loss of dopaminergic neurons in the substantia nigra (Ikeda et al. 2002; Oztas et al. 2002). This neuroprotection was seen after acute coadministration and after repeated injection of caffeine (Xu et al. 2002). Various  $A_{2A}R$  antagonists (including SCH58261, KW-6002, 3,7-dimethyl-1-propargylxanthine, and CSC), attenuate MPTP-induced dopaminergic neurotoxicity, suggesting that the protective effects of caffeine are due to its action at the  $A_{2A}R$  (Chen et al. 2001; Alfinito et al. 2003; Pierrri et al. 2005). In contrast, there is no protective effect of the  $A_1R$  antagonist DPCPX (Chen et al. 2001), despite an early report that adenosine and an  $A_1R$  agonist protect

against methamphetamine-induced neurotoxicity (Delle Donne and Sonsalla 1994). Finally, genetic inactivation of  $A_{2A}$ Rs also reduces MPTP-induced dopaminergic neurotoxicity (Chen et al. 2001). These studies provide a neurobiological basis for the inverse relationship between increased caffeine consumption and reduced risk of developing PD. The convergence of epidemiological evidence and findings from animal studies also raises the exciting possibility that  $A_{2A}$ R antagonists, including caffeine, may slow or halt dopaminergic neuronal degeneration.

Despite the consistent demonstration that  $A_{2A}$ R antagonists afford neuroprotection against dopaminergic neurotoxicity, the mechanism by which  $A_{2A}$ R inactivation protects against the loss of dopaminergic neurons remains unknown. The particular challenge lies in explaining the apparent dichotomy between restricted expression of the  $A_{2A}$ R in striatopallidal neurons and neuroprotection against degeneration of dopaminergic neurons in the substantia nigra, where only scattered expression of  $A_{2A}$ Rs is detected. A partial answer to this mechanistic question comes from a study by Chen's group, who employed a forebrain-neuron-specific  $A_{2A}$ R-knockout model to demonstrate the distinct cellular mechanisms underlying motor stimulant and neuroprotective effects by  $A_{2A}$ R antagonists. While forebrain neuronal  $A_{2A}$ Rs are responsible for the motor effect,  $A_{2A}$ Rs in other cellular elements, such as microglial cells, may be associated with neuroprotection (Yu et al. 2008). An additional challenge is to identify the cellular mechanism which allows  $A_{2A}$ R inactivation to provide neuroprotection against a broad spectrum of brain insults, from ischemia to excitotoxicity to mitochondrial toxicity (for a recent review, see Chen et al. 2007). However, it should be emphasized that targets other than adenosine receptors should also be considered. For example, it was recently proposed that caffeine-mediated neuroprotection may be associated with expression of cytochrome P450 (Singh et al. 2009) or cytochrome oxidase (Jones et al. 2008) and with stimulation of ryanodine receptor channels (Guerreiro et al. 2008). Lastly, a recent study also suggested that caffeine may confer neuroprotection by increasing BBB integrity since MPTP-induced leakage in Evans blue dye and TITC-albumin in the striatum was attenuated by caffeine treatment (Chen et al. 2008a). This effect is further associated with increased expression of tight junction proteins (Chen et al. 2008a).

L-dopa-induced motor complications, dyskinesia in particular, are a major limiting factor in the management of advanced PD and are intrinsically linked with the chronic stimulation of dopamine receptors. Thus, nondopaminergic agents, such as  $A_{2A}$ R antagonists, may exert similar beneficial motor effects but have a low propensity to induce dyskinesia. Indeed, in L-dopa-sensitized, dopamine-depleted nonhuman primates, KW-6002 treatment reverses motor deficits but does not induce a dyskinesia score (Kanda et al. 1998b, 2000; Grondin et al. 1999). Genetic and pharmacology studies further suggest that the  $A_{2A}$ R antagonists may also modify development of LID. Genetic inactivation of the  $A_{2A}$ R attenuated the sensitization of rotational behavior induced by repeated L-dopa treatment, indicating that activation of  $A_{2A}$ Rs is required for the *development* of behavioral sensitization by chronic treatment with L-dopa (Fredduzzi et al. 2002) and with amphetamine (Xiao et al. 2006). Similarly, L-dopa-induced abnormal involuntary

movements (AIMs) are attenuated by selective inactivation of  $A_{2A}$ Rs in forebrain neurons (Xiao et al. 2006) or by a *low* dose of KW-6002 (Bastia et al. 2005). Additionally, coadministration of KW-6002 and apomorphine ( $1 \text{ mg kg}^{-1}$ ) to MPTP-treated monkeys completely prevented the development of dyskinesia, while L-dopa alone produced typical dyskinesia (Bibbiani et al. 2002). The role of the  $A_{2A}$ R in the development of LID is supported by the increased level of  $A_{2A}$ Rs in striatum of 6-OHDA-lesioned rats, dyskinesic nonhuman primates, and dyskinesic PD patients after chronic L-dopa treatment (Morelli et al. 2007). However, a recent study measuring AIMs, showed that coadministration of KW-6002 at regular motor stimulant doses with L-dopa did not modify the AIM behavioral score in rats (Lundblad et al. 2003). Thus, it remains to be determined whether  $A_{2A}$ R antagonists can modify LID in PD models. Intriguingly, the  $A_{2A}$ R– $D_2$ R model predicts that  $A_{2A}$ R antagonists should enhance rather than attenuate LID. However, a recent study has demonstrated the critical role of the  $A_{2A}$ R in forebrain outside the striatum in modulation of dopamine-mediated behavior (Shen et al. 2008). It is proposed that additional mechanisms (such as adenosine–glutamate interaction) in the cerebral cortex may be responsible for the possible antidyskinesic effects of  $A_{2A}$ R antagonists (Shen et al. 2008).

## 4.5 *Huntington's Disease*

HD (affecting one in 10,000 individuals) is an autosomal dominant disease characterized by chorea, dementia, psychiatric symptoms, and eventual death (Martin and Gusella 1986). The causative mutation is a CAG trinucleotide expansion in exon 1 of the huntingtin (Htt) gene (Group 1993). When the number of CAG repeats exceeds 36, Htt forms aggregates in the nuclei and cytoplasm of neurons, glia, and several different types of peripheral cells (e.g., liver, muscles, and adipocytes), hijacks a wide variety of proteins, and causes neuronal degeneration and metabolic dysfunction (Group 1993; Sugars and Rubinsztein 2003; Li and Li 2004; Chiang et al. 2005, 2009; Chou et al. 2005; Ryu et al. 2005; Chiang et al. 2007b; Wang et al. 2008b). Because the major clinical presentations predominantly appear in the CNS, HD was initially considered a neuronal degenerative disorder. Nonetheless, accumulating evidence suggests that defects in peripheral tissues also significantly contribute to HD pathogenesis (Sathasivam et al. 1999; Djousse et al. 2002; Ribchester et al. 2004; Chiang et al. 2007a; Mihm et al. 2007; Valenza et al. 2007; Bjorkqvist et al. 2008; Phan et al. 2009). To date, there is no effective treatment to prevent the progression of this dreadful disease (Mestre et al. 2009).

### 4.5.1 Human Studies

Given the concentrated expression of  $A_{2A}$ Rs in striatopallidal neurons, it is not surprising that a characteristic loss of  $A_{2A}$ R binding is detected in the very early

stage of HD (grade 0) (Glass et al. 2000). Interestingly,  $A_{2A}R$  expression and function in peripheral blood cells in HD was found to increase in 48 heterozygous and three homozygous patients compared with 58 healthy subjects (Varani et al. 2003). Furthermore,  $A_{2A}R$  binding density in blood platelets of HD patients apparently correlates with age at onset anticipation (Maglione et al. 2006). It is suggested that these changes in  $A_{2A}R$  binding in peripheral blood platelets likely reflect the status of inflammation and oxidative events associated with HD pathogenesis. Thus, if these findings can be confirmed, the  $A_{2A}R$  in peripheral platelets could be a useful biomarker for HD.

A recent genetic association study of 791 unrelated HD patients found that the single nuclear polymorphism in the  $A_{2A}R$  gene (1976C/T, rs5751876) is associated with the residual age at onset of the disease of 3.8 years (Dhaenens et al. 2009). This finding needs to be confirmed by a follow-up study with large cohorts.

#### 4.5.2 Experimental Studies

The two primary targets of caffeine, the  $A_{2A}R$  and the  $A_1R$ , have been investigated as targets for drug development for treatment of HD. The  $A_1R$  is of interest because of its well-established protective role in ischemia and epileptic conditions as described already herein. Consistent with the hypothesis that the  $A_1R$  might confer a protective role in HD, blockage of the  $A_1R$  using DPCPX exacerbated the damage to GABAergic neurons caused by a mitochondrial toxin (malonate; Alfinito et al. 2003) in a model of HD. In addition, with use of a 3-nitropropionic acid (3NP)-induced rat model of HD (Blum et al. 2001), an  $A_1R$ -selective agonist (adenosine amine congener, ADAC) devoid of cardiovascular side effects was shown to protect against striatal lesions and motor impairment caused by 3NP (Blum et al. 2002). Similar to what was observed in ischemia/stroke models, chronic use of ADAC in the test protocol did not have any beneficial effects, owing to desensitization of the  $A_1R$  (Abbracchio et al. 1992).

The  $A_{2A}R$  has attracted much attention as a potential drug target for HD because of its expression in enkephalin-containing striatal neurons as well as at glutamatergic terminals in the corticostriatal pathway of the brain. In particular, stimulation of the presynaptic  $A_{2A}R$  triggers glutamate release, and therefore is generally believed to be detrimental to neuronal survival. In contrast, the  $A_{2A}R$  located on postsynaptic GABAergic terminals is considered protective (Corsi et al. 2000; Blum et al. 2003b; Fink et al. 2004). Mutant Htt suppresses CREB binding to its core promoter; thus, expression of the  $A_{2A}R$  in the striatum is markedly decreased during HD progression (Ferre et al. 1993; Glass et al. 2000; Wyttenbach et al. 2001; Chiang et al. 2005). Nonetheless, the ability of the striatal  $A_{2A}R$  in HD mice to evoke cAMP signaling is similar to that of wild-type mice (Chou et al. 2005), indicating aberrantly amplified signaling from the  $A_{2A}R$  as was observed in peripheral blood cells of HD patients (Varani et al. 2001, 2003; Maglione et al. 2006). The  $A_{2A}R$  is therefore a potential therapeutic target for HD despite its reduced expression.

In genetic mouse models of HD, both agonists and antagonists of the  $A_{2A}R$  were tested for beneficial effects. Systemic delivery of an  $A_{2A}R$  antagonist (SCH58261) for 1 week reduced NMDA-induced toxicity in a transgenic mouse model of HD (R6/2), but worsened motor coordination (Domenici et al. 2007). Intriguingly, in the same mouse model, chronic treatment (5 weeks) with an  $A_{2A}R$  agonist (CGS21680) ameliorated several major symptoms [e.g., brain atrophy, striatal aggregates, deteriorated motor coordination, urea cycle deficiency, and poor ubiquitin proteasome system (UPS) activity] (Chou et al. 2005; Chiang et al. 2009). It appears that the disease stage, the drug administration protocol, and the clinical manifestations might play critical roles in evaluating the future therapeutic potential of  $A_{2A}R$  drugs (Popoli et al. 2008). The ability of CGS21680 to enhance the UPS activity via a cAMP–PKA-dependent pathway is of particular interest (Chiang et al. 2009), because aggregate formation is a major hallmark of HD, and a dysregulated UPS is closely associated with the formation of Htt aggregates and HD pathogenesis (Zhou et al. 2003; Seo et al. 2004; Bennett et al. 2007; Hunter et al. 2007; Wang et al. 2008a). The proteasome-activating (or proteasome-modulating) capacity of  $A_{2A}R$  agonists provides a new means of treating HD, and merits further evaluation. Because the  $A_{2A}R$  is expressed in multiple tissues in which mHtt is present and forms aggregates, systematic administration of  $A_{2A}R$  agonists is expected to boost the inferior UPS activities in both the CNS and peripheral tissues of HD patients, and might be more effective than interventions which only treat the CNS. Unfortunately, certain adverse effects of  $A_{2A}R$  drugs on the cardiovascular system were reported (Gordi et al. 2006; Mingote et al. 2008), inevitably dampening the enthusiasm for their potential clinical application. Partial  $A_{2A}R$  agonists with good BBB penetration might reduce or eliminate unfavorable side effects as has been recommended for other adenosine drugs (Gao and Jacobson 2004). Moreover, epidemiological investigation into the long-term effects of chronic caffeine consumption on the progress of HD patients is warranted.

## 4.6 Multiple Sclerosis

Multiple sclerosis (MS) is a common autoimmune disorder of the CNS with pathological characteristics that include lymphocyte and macrophage infiltration, CNS demyelination, and axonal damage, resulting in recurrent impairment of brain and spinal cord function (Prineas and Wright 1978; Hafler 2004). Its cause remains unknown, but MS is thought to be a prototypic autoimmune disease mediated by T lymphocytes with the participation of B lymphocytes targeted against myelin protein (Noseworthy et al. 2000; Keegan and Noseworthy 2002; Meinl et al. 2006). Current immunosuppression therapies such as glucocorticosteroids have significant side effects and their effectiveness is limited (a 30–60% reduction in the frequency of relapse). Thus, there is an ongoing search for a more effective treatment with fewer side effects.



### 4.6.1 Human Studies

Recently, several lines of clinical and experimental evidence have suggested that adenosine receptors may modulate neuroinflammation in MS. Adenosine levels in blood plasma decrease greatly in MS, accompanied by an increase in the level of tumor necrosis factor (Mayne et al. 1999). Furthermore, downregulation of A<sub>1</sub>Rs has been reported in mononuclear cells in the peripheral blood and CD45-positive glial cells in the brain of MS patients (Johnston et al. 2001). These studies suggest that dysfunction of A<sub>1</sub>Rs may contribute to the pathogenesis in MS patients.

### 4.6.2 Animal Studies

Myelin oligodendroglia glycoprotein (MOG) induces a combined autoimmune pathogenic T-cell and B-cell response (Adelmann et al. 1995), leading to demyelination in the brain, a hallmark of human MS lesions (Adelmann et al. 1995; Berger et al. 1997). Experimental autoimmune encephalomyelitis (EAE) induced by immunization with MOG is a widely used animal model of MS (Steinman and Zamvil 2006). A recent study showed that mice with a genetic deficiency in extracellular nucleotidase CD73, a molecule critical for generating extracellular adenosine in various cells, including T cells, are largely resistant to MOG-induced brain and spinal cord injury (Mills et al. 2008). Furthermore, genetic deficiency in A<sub>1</sub>R exacerbated spinal cord injury with extensive inflammation and demyelination (Tsutsui et al. 2004). Conversely, treatment with the A<sub>1</sub>R agonist ADAC reduces spinal cord inflammation and demyelination in EAE mice (Tsutsui et al. 2004). These results suggest that adenosine acting at the A<sub>1</sub>R suppresses the inflammatory response, contributing to the pathogenesis of MS. The critical role of the A<sub>1</sub>R in development and treatment of MS is further supported by the recent finding that suppression of EAE-induced neuroinflammation and neurobehavioral deficits by glucocorticoid treatment is accompanied by a concurrent increase in A<sub>1</sub>R expression. This modulation in A<sub>1</sub>R expression by glucocorticoids in the EAE model may be related to the reciprocal interactions between the A<sub>1</sub>R and  $\beta$ -arrestin-1 reported in monocytoid cells (Tsutsui et al. 2008). In addition to these effects of the A<sub>1</sub>R in experimental models of MS, pharmacological blockade of the A<sub>2A</sub>R has been shown to attenuate EAE (Mills et al. 2008). Thus, adenosine acting at the A<sub>2A</sub>R may facilitate inflammation and EAE. While the precise roles of adenosine and adenosine receptors in the development of EAE remain to be clarified, these findings clearly highlight the critical involvement of adenosine and adenosine receptors in modulation of EAE.

Two recent reports showed that caffeine can attenuate MOG-induced EAE in mice. Mice chronically treated with caffeine displayed significantly fewer clinical symptoms of motor impairment and reduced microglial activation in cerebral cortex after MOG immunization (Tsutsui et al. 2004; Mills et al. 2008). This finding has recently been extended to an EAE model induced by guinea pig spinal cord



homogenates (GPSCH) with a more extensive inflammatory response, less extensive demyelination, but a more chronic disease course than MOG-induced EAE (Raine 1984; Smith et al. 2005). In the GPSCH model of EAE, chronic caffeine imparts neuroprotection against EAE in rats with both decreased incidence of EAE and attenuated EAE (Chen et al. 2010). Interestingly, chronic treatment with caffeine apparently exerts a neuroprotective effect against EAE through an A<sub>1</sub>R-mediated shift from Th1 to Th2 cell function (Chen et al. 2010). These animal studies provide a neurobiological basis for epidemiological investigation into the possible relationship between caffeine consumption and development of MS in humans.

## 5 Concluding Remarks

Several decades after the earliest attempt to treat PD using caffeine (Shoulson and Chase 1975), tremendous amounts of information regarding the potential beneficial effect of caffeine as well as adenosine drugs on major neurodegenerative disorders have accumulated. In addition to the encouraging development of several A<sub>2A</sub>R selective antagonists which are now in advanced clinical phase III trials, detailed characterization of the primary targets of caffeine (A<sub>1</sub>R and A<sub>2A</sub>R) provides sound mechanistic bases for the action of caffeine. The exciting findings to date include the specific *in vivo* functions of adenosine receptors revealed by genetic mouse models, and the awareness of profound interaction between adenosine receptors and other GPCR and accessory proteins which might exhibit pharmacological properties distinct from those of adenosine receptor monomers. An increasing body of human and experimental studies provided encouraging evidence that regular human consumption of caffeine may, in fact, have several beneficial effects on neurodegenerative disorders, from motor stimulation to cognitive enhancement to potential neuroprotection. Importantly, neuroprotection by *chronic* treatment with caffeine can be demonstrated in animal models of PD, AD, TBI, stroke, and MS, highlighting a broad spectrum of neuroprotection with possibly common mechanisms of caffeine. Thus, with regard to neurodegeneration, these potential benefits of methylxanthines, caffeine in particular, strongly argue against the common practice by clinicians to discourage regular human consumption of caffeine. Additional studies are warranted to confirm neuroprotective and cognitive enhancement effects of caffeine in large, longitudinal clinical trials for these neurodegenerative disorders. This encouraging development invites further investigation into the action of caffeine and adenosine drugs to define the molecular basis by which methylxanthines and adenosine exert their neuroprotective or cognitive effects. This knowledge not only provides a neurobiological basis for guidelines for healthy usage of caffeine as a stimulant to improve human performance, but also opens up a real and novel possibility to develop methylxanthine-based treatment for these neurodegenerative diseases.

## References

- Abbracchio MP, Fogliatto G, Paoletti AM, Rovati GE, Cattabeni F (1992) Prolonged in vitro exposure of rat brain slices to adenosine analogues: selective desensitization of adenosine A1 but not A2 receptors. *Eur J Pharmacol* 227:317–324
- Adami M, Bertorelli R, Ferri N, Foddi MC, Ongini E (1995) Effects of repeated administration of selective adenosine A1 and A2A receptor agonists on pentylenetetrazole-induced convulsions in the rat. *Eur J Pharmacol* 294:383–389
- Addicott MA, Yang LL, Peiffer AM, Burnett LR, Burdette JH, Chen MY, Hayasaka S, Kraft RA, Maldjian JA, Laurienti PJ (2009) The effect of daily caffeine use on cerebral blood flow: How much caffeine can we tolerate? *Hum Brain Mapp* 30:3102–3114
- Adelmann M, Wood J, Benzl I, Fiori P, Lassmann H, Matthieu JM, Gardinier MV, Dormmair K, Linington C (1995) The N-terminal domain of the myelin oligodendrocyte glycoprotein (MOG) induces acute demyelinating experimental autoimmune encephalomyelitis in the Lewis rat. *J Neuroimmunol* 63:17–27
- Aden U, Lindstrom K, Bona E, Hagberg H, Fredholm BB (1994) Changes in adenosine receptors in the neonatal rat brain following hypoxic ischemia. *Brain Res Mol Brain Res* 23:354–358
- Aden U, Halldner L, Lagercrantz H, Dalmau I, Ledent C, Fredholm BB (2003) Aggravated brain damage after hypoxic ischemia in immature adenosine A2A knockout mice. *Stroke* 34:739–744
- Ahlskog JE, Muentner MD (2001) Frequency of levodopa-related dyskinesias and motor fluctuations as estimated from the cumulative literature. *Mov Disord* 16:448–458
- Al Moutaery K, Al Deeb S, Ahmad Khan H, Tariq M (2003) Caffeine impairs short-term neurological outcome after concussive head injury in rats. *Neurosurgery* 53:704–711, discussion 711–712
- Albasanz JL, Perez S, Barrachina M, Ferrer I, Martin M (2008) Up-regulation of adenosine receptors in the frontal cortex in Alzheimer's disease. *Brain Pathol* 18:211–219
- Alfinito PD, Wang SP, Manzano L, Rijhsinghani S, Zeevalk GD, Sonsalla PK (2003) Adenosinergic protection of dopaminergic and GABAergic neurons against mitochondrial inhibition through receptors located in the substantia nigra and striatum, respectively. *J Neurosci* 23:10982–10987
- Allouf K, Sauriol L, Kennedy W, Laurier C, Tessier G, Novosel S, Contandriopoulos A (1998) Alzheimer's disease: a review of the disease, its epidemiology and economic impact. *Arch Gerontol Geriatr* 27:189–221
- Arendash GW, Schleif W, Rezai-Zadeh K, Jackson EK, Zacharia LC, Cracchiolo JR, Shippy D, Tan J (2006) Caffeine protects Alzheimer's mice against cognitive impairment and reduces brain beta-amyloid production. *Neuroscience* 142:941–952
- Arendash GW, Mori T, Cao C, Mamcarz M, Runfeldt M, Dickson A, Rezai-Zadeh K, Tane J, Citron BA, Lin X, Echeverria V, Potter H (2009) Caffeine reverses cognitive impairment and decreases brain amyloid-beta levels in aged Alzheimer's disease mice. *J Alzheimers Dis* 17:661–680
- Aronowski J, Strong R, Shirzadi A, Grotta JC (2003) Ethanol plus caffeine (caffeinol) for treatment of ischemic stroke: preclinical experience. *Stroke* 34:1246–1251
- Ascherio A, Zhang SM, Hernan MA, Kawachi I, Colditz GA, Speizer FE, Willett WC (2001) Prospective study of caffeine consumption and risk of Parkinson's disease in men and women. *Ann Neurol* 50:56–63
- Azad K, Gall D, Woods AS, Ledent C, Ferre S, Schiffmann SN (2009) Dopamine D2 and adenosine A2A receptors regulate NMDA-mediated excitation in accumbens neurons through A2A-D2 receptor heteromerization. *Neuropsychopharmacology* 34:972–986
- Bara-Jimenez W, Sherzai A, Dimitrova T, Favit A, Bibbiani F, Gillespie M, Morris MJ, Mouradian MM, Chase TN (2003) Adenosine A(2A) receptor antagonist treatment of Parkinson's disease. *Neurology* 61:293–296
- Bastia E, Xu YH, Scibelli AC, Day YJ, Linden J, Chen JF, Schwarzschild MA (2005) A crucial role for forebrain adenosine A(2A) receptors in amphetamine sensitization. *Neuropsychopharmacology* 30:891–900

- Belayev L, Khoutorova L, Zhang Y, Belayev A, Zhao W, Busto R, Ginsberg MD (2004) Caffeinol confers cortical but not subcortical neuroprotection after transient focal cerebral ischemia in rats. *Brain Res* 1008:278–283
- Bell MJ, Kochanek PM, Carcillo JA, Mi Z, Schiding JK, Wisniewski SR, Clark RS, Dixon CE, Marion DW, Jackson E (1998) Interstitial adenosine, inosine, and hypoxanthine are increased after experimental traumatic brain injury in the rat. *J Neurotrauma* 15:163–170
- Bennett EJ, Shaler TA, Woodman B, Ryu K-Y, Zaitseva TS, Becker CH, Bates GP, Schulman H, Kopito RR (2007) Global changes to the ubiquitin system in Huntington's disease. *Nature* 448:704–708
- Berger T, Weerth S, Kojima K, Lington C, Wekerle H, Lassmann H (1997) Experimental autoimmune encephalomyelitis: the antigen specificity of T lymphocytes determines the topography of lesions in the central and peripheral nervous system. *Lab Invest* 76:355–364
- Bertram L, Tanzi RE (2008) Thirty years of Alzheimer's disease genetics: the implications of systematic meta-analyses. *Nat Rev Neurosci* 9:768–778
- Bibbiani F, Oh JD, Petzer JP, Castagnoli N Jr, Chen J-F, Schwarzschild MA, Chase T (2002) A2A receptor antagonist prevents the development of dopamine agonist-induced motor complications in primate and rodent models of Parkinson's disease. In: Annual meeting of Society for Neuroscience. Orlando, FL
- Bjorkqvist M et al (2008) A novel pathogenic pathway of immune activation detectable before clinical onset in Huntington's disease. *J Exp Med* 205:1869–1877
- Blum D, Gall D, Cuvelier L, Schiffmann SN (2001) Topological analysis of striatal lesions induced by 3-nitropropionic acid in the Lewis rat. *Neuroreport* 12:1769–1772
- Blum D, Hourez R, Galas MC, Popoli P, Schiffmann SN (2003a) Adenosine receptors and Huntington's disease: implications for pathogenesis and therapeutics. *Lancet Neurol* 2:366–374
- Blum D, Gall D, Galas MC, d'Alcantara P, Bantubungi K, Schiffmann SN (2002) The adenosine A1 receptor agonist adenosine amine congener exerts a neuroprotective effect against the development of striatal lesions and motor impairments in the 3-nitropropionic acid model of neurotoxicity. *J Neurosci* 22:9122–9133
- Blum D, Galas MC, Pintor A, Brouillet E, Ledent C, Muller CE, Bantubungi K, Galluzzo M, Gall D, Cuvelier L, Rolland AS, Popoli P, Schiffmann SN (2003b) A dual role of adenosine A2A receptors in 3-nitropropionic acid-induced striatal lesions: implications for the neuroprotective potential of A2A antagonists. *J Neurosci* 23:5361–5369
- Boison D (2006) Adenosine kinase, epilepsy and stroke: mechanisms and therapies. *Trends Pharmacol Sci* 27:652–658
- Braak H, Braak E (1997) Frequency of stages of Alzheimer-related lesions in different age categories. *Neurobiol Aging* 18:351–357
- Britton M, de Faire U, Helmers C, Miah K, Rane A (1980) Lack of effect of theophylline on the outcome of acute cerebral infarction. *Acta Neurol Scand* 62:116–123
- Broderick JP, Viscoli CM, Brott T, Kernan WN, Brass LM, Feldmann E, Morgenstern LB, Wilterdink JL, Horwitz RI (2003) Major risk factors for aneurysmal subarachnoid hemorrhage in the young are modifiable. *Stroke* 34:1375–1381
- Brodie C, Blumberg PM, Jacobson KA (1998) Activation of the A2A adenosine receptor inhibits nitric oxide production in glial cells. *FEBS Lett* 429:139–142
- Burgueno J, Blake DJ, Benson MA, Tinsley CL, Esapa CT, Canela EI, Penela P, Mallol J, Mayor F Jr, Lluís C, Franco R, Ciruela F (2003) The adenosine A2A receptor interacts with the actin-binding protein alpha-actinin. *J Biol Chem* 278:37545–37552
- Cabello N, Gandia J, Bertarelli DC, Watanabe M, Lluís C, Franco R, Ferre S, Lujan R, Ciruela F (2009) Metabotropic glutamate type 5, dopamine D2 and adenosine A2a receptors form higher-order oligomers in living cells. *J Neurochem* 109:1497–1507
- Cadden IS, Partovi N, Yoshida EM (2007) Review article: possible beneficial effects of coffee on liver disease and function. *Aliment Pharmacol Ther* 26:1–8
- Cao C, Cirrito JR, Lin X, Wang L, Verges DK, Dickson A, Mamcarz M, Zhang C, Mori T, Arendash GW, Holtzman DM, Potter H (2009) Caffeine suppresses amyloid-beta levels in plasma and brain of Alzheimer's disease transgenic mice. *J Alzheimers Dis* 17:681–697

- Carriba P, Ortiz O, Patkar K, Justinova Z, Stroik J, Themann A, Muller C, Woods AS, Hope BT, Ciruela F, Casado V, Canela EI, Lluís C, Goldberg SR, Moratalla R, Franco R, Ferré S (2007) Striatal adenosine A2A and cannabinoid CB1 receptors form functional heteromeric complexes that mediate the motor effects of cannabinoids. *Neuropsychopharmacology* 32: 2249–2259
- Cassada DC, Tribble CG, Young JS, Gangemi JJ, Gohari AR, Butler PD, Rieger JM, Kron IL, Linden J, Kern JA (2002a) Adenosine A2A analogue improves neurologic outcome after spinal cord trauma in the rabbit. *J Trauma* 53:225–229, discussion 229–231
- Cassada DC, Tribble CG, Long SM, Laubach VE, Kaza AK, Linden J, Nguyen BN, Rieger JM, Fiser SM, Kron IL, Kern JA (2002b) Adenosine A2A analogue ATL-146e reduces systemic tumor necrosis factor- $\alpha$  and spinal cord capillary platelet-endothelial cell adhesion molecule-1 expression after spinal cord ischemia. *J Vasc Surg* 35:994–998
- Chaudhuri KR, Healy DG, Schapira AH (2006) Non-motor symptoms of Parkinson's disease: diagnosis and management. *Lancet Neurol* 5:235–245
- Chen GQ, Chen YY, Wang XS, Wu SZ, Yang HM, Xu HQ, He JC, Wang XT, Chen JF, Zheng RY (2010) Chronic caffeine treatment attenuates experimental autoimmune encephalomyelitis induced by guinea pig spinal cord homogenates in Wistar rats. *Brain Res* 1309:116–125
- Chen JF (2003) The adenosine A(2A) receptor as an attractive target for Parkinson's disease treatment. *Drug News Perspect* 16:597–604
- Chen JF, Huang Z, Ma J, Zhu J, Moratalla R, Standaert D, Moskowitz MA, Fink JS, Schwarzschild MA (1999) A(2A) adenosine receptor deficiency attenuates brain injury induced by transient focal ischemia in mice. *J Neurosci* 19:9192–9200
- Chen JF, Sonsalla PK, Pedata F, Melani A, Domenici MR, Popoli P, Geiger J, Lopes LV, de Mendonca A (2007) Adenosine A2A receptors and brain injury: broad spectrum of neuroprotection, multifaceted actions and "fine tuning" modulation. *Prog Neurobiol* 83:310–331
- Chen JF, Xu K, Petzer JP, Staal R, Xu YH, Beilstein M, Sonsalla PK, Castagnoli K, Castagnoli N Jr, Schwarzschild MA (2001) Neuroprotection by caffeine and A(2A) adenosine receptor inactivation in a model of Parkinson's disease. *J Neurosci* 21:RC143
- Chen X, Lan X, Roche I, Liu R, Geiger JD (2008a) Caffeine protects against MPTP-induced blood-brain barrier dysfunction in mouse striatum. *J Neurochem* 107:1147–1157
- Chen X, Gawryluk JW, Wagener JF, Ghribi O, Geiger JD (2008b) Caffeine blocks disruption of blood brain barrier in a rabbit model of Alzheimer's disease. *J Neuroinflammation* 5:12
- Cheng HC, Shih HM, Chern Y (2002) Essential role of cAMP-response element-binding protein activation by A2A adenosine receptors in rescuing the nerve growth factor-induced neurite outgrowth impaired by blockage of the MAPK cascade. *J Biol Chem* 277:33930–33942
- Chern Y, Lai HL, Fong JC, Liang Y (1993) Multiple mechanisms for desensitization of A2A adenosine receptor-mediated cAMP elevation in rat pheochromocytoma PC12 cells. *Mol Pharmacol* 44:950–958
- Chiang M-C, Chen H-M, Lee Y-H, Chang H-H, Wu Y-C, Soong B-W, Chen C-M, Wu Y-R, Liu C-S, Niu D-M, Wu J-Y, Chen Y-T, Chern Y (2007) Dysregulation of C/EBP $\alpha$  by mutant Huntingtin causes the urea cycle deficiency in Huntington's disease. *Hum Mol Genet* 16:483–498
- Chiang MC, Lee YC, Huang CL, Chern Y (2005) cAMP-response element-binding protein contributes to suppression of the A2A adenosine receptor promoter by mutant Huntingtin with expanded polyglutamine residues. *J Biol Chem* 280:14331–14340
- Chiang MC, Chen HM, Lai HL, Chen HW, Chou SY, Chen CM, Tsai FJ, Chern Y (2009) The A2A adenosine receptor rescues the urea cycle deficiency of Huntington's disease by enhancing the activity of the ubiquitin-proteasome system. *Hum Mol Genet* 18:2929–2942
- Chiaruttini C, Vicario A, Li Z, Baj G, Braiuca P, Wu Y, Lee FS, Gardossi L, Baraban JM, Tongiorgi E (2009) Dendritic trafficking of BDNF mRNA is mediated by translin and blocked by the G196A (Val66Met) mutation. *Proc Natl Acad Sci USA* 106:16481–16486
- Choi OH, Shamim MT, Padgett WL, Daly JW (1988) Caffeine and theophylline analogues: correlation of behavioral effects with activity as adenosine receptor antagonists and as phosphodiesterase inhibitors. *Life Sci* 43:387–398

- Chou SY, Lee YC, Chen HM, Chiang MC, Lai HL, Chang HH, Wu YC, Sun CN, Chien CL, Lin YS, Wang SC, Tung YY, Chang C, Chern Y (2005) CGS21680 attenuates symptoms of Huntington's disease in a transgenic mouse model. *J Neurochem* 93:310–320
- Ciruela F, Casado V, Rodrigues RJ, Lujan R, Burgueno J, Canals M, Borycz J, Rebola N, Goldberg SR, Mallol J, Cortes A, Canela EI, Lopez-Gimenez JF, Milligan G, Lluís C, Cunha RA, Ferre S, Franco R (2006) Presynaptic control of striatal glutamatergic neurotransmission by adenosine A1-A2A receptor heteromers. *J Neurosci* 26:2080–2087
- Clark RS, Carcillo JA, Kochanek PM, Obrist WD, Jackson EK, Mi Z, Wisniewski SR, Bell MJ, Marion DW (1997) Cerebrospinal fluid adenosine concentration and uncoupling of cerebral blood flow and oxidative metabolism after severe head injury in humans. *Neurosurgery* 41:1284–1292, discussion 1292–1283
- Coelho JE, Rebola N, Fragata I, Ribeiro JA, de Mendonça A, Cunha RA (2006) Hypoxia-induced desensitization and internalization of adenosine A1 receptors in the rat hippocampus. *Neuroscience* 138:1195–1203
- Corsi C, Melani A, Bianchi L, Pedata F (2000) Striatal A2A adenosine receptor antagonism differentially modifies striatal glutamate outflow in vivo in young and aged rats. *Neuroreport* 11:2591–2595
- Cronstein BN, Kramer SB, Weissmann G, Hirschhorn R (1983) Adenosine: a physiological modulator of superoxide anion generation by human neutrophils. *J Exp Med* 158:1160–1177
- Cronstein BN, Daguma L, Nichols D, Hutchison AJ, Williams M (1990) The adenosine/neutrophil paradox resolved: human neutrophils possess both A1 and A2 receptors that promote chemotaxis and inhibit O2 generation, respectively. *J Clin Invest* 85:1150–1157
- Cunha GM, Canas PM, Melo CS, Hockemeyer J, Muller CE, Oliveira CR, Cunha RA (2008) Adenosine A2A receptor blockade prevents memory dysfunction caused by beta-amyloid peptides but not by scopolamine or MK-801. *Exp Neurol* 210:776–781
- Cunha GMA, Canas PM, Chen JF, Oliveira CR, Cunha RA (2006) Blockade of adenosine A2A receptors prevents  $\beta$ -amyloid ( $A\beta_{1-42}$ )-induced synaptotoxicity and memory impairment in rodents. *Purinergic Signal* 2:135
- Cunha RA, Ribeiro JA (2000) Adenosine A2A receptor facilitation of synaptic transmission in the CA1 area of the rat hippocampus requires protein kinase C but not protein kinase A activation. *Neurosci Lett* 289:127–130
- Dall'igna OP, Porciuncula LO, Souza DO, Cunha RA, Lara DR (2003) Neuroprotection by caffeine and adenosine A(2A) receptor blockade of beta-amyloid neurotoxicity. *Br J Pharmacol* 138:1207–1209
- Daly JW (2007) Caffeine analogs: biomedical impact. *Cell Mol Life Sci* 64:2153–2169
- Daly JW, Shi D, Nikodijevic O, Jacobson KA (1999) The role of adenosine receptors in the central action of caffeine. In: Gupta BS, Gupta U (eds) *Caffeine and behavior-current views and research trends*. CRC, Boca Raton, pp 1–16
- Dash PK, Moore AN, Moody MR, Treadwell R, Felix JL, Clifton GL (2004) Post-trauma administration of caffeine plus ethanol reduces contusion volume and improves working memory in rats. *J Neurotrauma* 21:1573–1583
- Dauer W, Przedborski S (2003) Parkinson's disease: mechanisms and models. *Neuron* 39:889–909
- Day YJ, Marshall MA, Huang L, McDuffie MJ, Okusa MD, Linden J (2004) Protection from ischemic liver injury by activation of A2A adenosine receptors during reperfusion: inhibition of chemokine induction. *Am J Physiol Gastrointest Liver Physiol* 286:G285–G293
- Day YJ, Huang L, McDuffie MJ, Rosin DL, Ye H, Chen JF, Schwarzschild MA, Fink JS, Linden J, Okusa MD (2003) Renal protection from ischemia mediated by A2A adenosine receptors on bone marrow-derived cells. *J Clin Invest* 112:883–891
- de Mendonça A, Sebastião AM, Ribeiro JA (2000) Adenosine: does it have a neuroprotective role after all? *Brain Res Brain Res Rev* 33:258–274
- Delle Donne KT, Sonsalla PK (1994) Protection against methamphetamine-induced neurotoxicity to neostriatal dopaminergic neurons by adenosine receptor activation. *J Pharmacol Exp Ther* 271:1320–1326

- Dhaenens CM et al (2009) A genetic variation in the ADORA2A gene modifies age at onset in Huntington's disease. *Neurobiol Dis* 35:474–476
- Diaz-Cabiale Z, Vivo M, Del Arco A, O'Connor WT, Harte MK, Muller CE, Martinez E, Popoli P, Fuxe K, Ferre S (2002) Metabotropic glutamate mGlu5 receptor-mediated modulation of the ventral striopallidal GABA pathway in rats. Interactions with adenosine A(2A) and dopamine D(2) receptors. *Neurosci Lett* 324:154–158
- Diogenes MJ, Fernandes CC, Sebastiao AM, Ribeiro JA (2004) Activation of adenosine A2A receptor facilitates brain-derived neurotrophic factor modulation of synaptic transmission in hippocampal slices. *J Neurosci* 24:2905–2913
- Djousse L, Knowlton B, Cupples LA, Marder K, Shoulson I, Myers RH (2002) Weight loss in early stage of Huntington's disease. *Neurology* 59:1325–1330
- Domenici MR, Scattoni ML, Martire A, Lastoria G, Potenza RL, Borioni A, Venerosi A, Calamandrei G, Popoli P (2007) Behavioral and electrophysiological effects of the adenosine A2A receptor antagonist SCH 58261 in R6/2 Huntington's disease mice. *Neurobiol Dis* 28:197–205
- Dorsey ER, Constantinescu R, Thompson JP, Biglan KM, Holloway RG, Kieburtz K, Marshall FJ, Ravina BM, Schifitto G, Siderowf A, Tanner CM (2007) Projected number of people with Parkinson disease in the most populous nations, 2005 through 2030. *Neurology* 68:384–386
- Duarte JM, Carvalho RA, Cunha RA, Gruetter R (2009) Caffeine consumption attenuates neurochemical modifications in the hippocampus of streptozotocin-induced diabetic rats. *J Neurochem* 111:368–379
- Dunwiddie TV (1980) Endogenously released adenosine regulates excitability in the in vitro hippocampus. *Epilepsia* 21:541–548
- Dunwiddie TV, Fredholm BB (1997) Adenosine neuromodulation. In: Jacobson KA, Jarvis MF (eds) *Purinergic approaches in experimental therapeutics*. Wiley-Liss, New York, pp 359–382
- Dunwiddie TV, Masino SA (2001) The role and regulation of adenosine in the central nervous system. *Annu Rev Neurosci* 24:31–55
- Dunwiddie TV, Hoffer BJ, Fredholm BB (1981) Alkylxanthines elevate hippocampal excitability. Evidence for a role of endogenous adenosine. *Naunyn Schmiedebergs Arch Pharmacol* 316:326–330
- El Yacoubi M, Ledent C, Menard JF, Parmentier M, Costentin J, Vaugeois JM (2000) The stimulant effects of caffeine on locomotor behaviour in mice are mediated through its blockade of adenosine A(2A) receptors. *Br J Pharmacol* 129:1465–1473
- Eskelinen MH, Ngandu T, Tuomilehto J, Soininen H, Kivipelto M (2009) Midlife coffee and tea drinking and the risk of late-life dementia: a population-based CAIDE study. *J Alzheimers Dis* 16:85–91
- Feldmann E, Broderick JP, Kernan WN, Viscoli CM, Brass LM, Brott T, Morgenstern LB, Wilterdink JL, Horwitz RI (2005) Major risk factors for intracerebral hemorrhage in the young are modifiable. *Stroke* 36:1881–1885
- Ferre S, O'Connor WT, Fuxe K, Ungerstedt U (1993) The striopallidal neuron: a main locus for adenosine-dopamine interactions in the brain. *J Neurosci* 13:5402–5406
- Ferre S, Goldberg SR, Lluis C, Franco R (2009) Looking for the role of cannabinoid receptor heteromers in striatal function. *Neuropharmacology* 56(Suppl 1):226–234
- Ferre S, von Euler G, Johansson B, Fredholm BB, Fuxe K (1991) Stimulation of high-affinity adenosine A2 receptors decreases the affinity of dopamine D2 receptors in rat striatal membranes. *Proc Natl Acad Sci USA* 88:7238–7241
- Ferre S, Fuxe K, von Euler G, Johansson B, Fredholm BB (1992) Adenosine-dopamine interactions in the brain. *Neuroscience* 51:501–512
- Ferre S, Fredholm BB, Morelli M, Popoli P, Fuxe K (1997) Adenosine-dopamine receptor-receptor interactions as an integrative mechanism in the basal ganglia. *Trends Neurosci* 20:482–487
- Fiebich BL, Biber K, Lieb K, van Calker D, Berger M, Bauer J, Gebicke-Haerter PJ (1996) Cyclooxygenase-2 expression in rat microglia is induced by adenosine A2a-receptors. *Glia* 18:152–160

- Fink JS, Kalda A, Ryu H, Stack EC, Schwarzschild MA, Chen JF, Ferrante RJ (2004) Genetic and pharmacological inactivation of the adenosine A2A receptor attenuates 3-nitropropionic acid-induced striatal damage. *J Neurochem* 88:538–544
- Flajolet M, Wang Z, Futter M, Shen W, Nuangchamngong N, Bendor J, Wallach I, Nairn AC, Surmeier DJ, Greengard P (2008) FGF acts as a co-transmitter through adenosine A(2A) receptor to regulate synaptic plasticity. *Nat Neurosci* 11:1402–1409
- Fontinha BM, Diogenes MJ, Ribeiro JA, Sebastiao AM (2008) Enhancement of long-term potentiation by brain-derived neurotrophic factor requires adenosine A2A receptor activation by endogenous adenosine. *Neuropharmacology* 54:924–933
- Fredduzzi S, Moratalla R, Monopoli A, Cuellar B, Xu K, Ongini E, Impagnatiello F, Schwarzschild MA, Chen JF (2002) Persistent behavioral sensitization to chronic L-DOPA requires A2A adenosine receptors. *J Neurosci* 22:1054–1062
- Fredholm BB (1980) Are methylxanthine effects due to antagonism of endogenous adenosine? *Trends Pharmacol Sci* 1:129–132
- Fredholm BB (1995) Astra Award Lecture. Adenosine, adenosine receptors and the actions of caffeine. *Pharmacol Toxicol* 76:93–101
- Fredholm BB, Irenius E, Kull B, Schulte G (2001) Comparison of the potency of adenosine as an agonist at human adenosine receptors expressed in Chinese hamster ovary cells. *Biochem Pharmacol* 61:443–448
- Fredholm BB, Battig K, Holmen J, Nehlig A, Zvartau EE (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fukumitsu N, Ishii K, Kimura Y, Oda K, Hashimoto M, Suzuki M, Ishiwata K (2008) Adenosine A (1) receptors using 8-dicyclopropylmethyl-1-[(11)C]methyl-3-propylxanthine PET in Alzheimer's disease. *Ann Nucl Med* 22:841–847
- Gao Y, Phillis JW (1994) CGS 15943, an adenosine A2 receptor antagonist, reduces cerebral ischemic injury in the Mongolian gerbil. *Life Sci* 55:PL61–PL65
- Gao ZG, Jacobson KA (2004) Partial agonists for A(3) adenosine receptors. *Curr Top Med Chem* 4:855–862
- Geismar P, Marquardsen J, Sylvest J (1976) Controlled trial of intravenous aminophylline in acute cerebral infarction. *Acta Neurol Scand* 54:173–180
- Geraets L, Moonen HJ, Wouters EF, Bast A, Hageman GJ (2006) Caffeine metabolites are inhibitors of the nuclear enzyme poly(ADP-ribose)polymerase-1 at physiological concentrations. *Biochem Pharmacol* 72:902–910
- Gines S, Hillion J, Torvinen M, Le Crom S, Casado V, Canela EI, Rondin S, Lew JY, Watson S, Zoli M, Agnati LF, Verniera P, Lluís C, Ferré S, Fuxe K, Franco R (2000) Dopamine D1 and adenosine A1 receptors form functionally interacting heteromeric complexes. *Proc Natl Acad Sci USA* 97:8606–8611
- Glass M, Dragunow M, Faull RL (2000) The pattern of neurodegeneration in Huntington's disease: a comparative study of cannabinoid, dopamine, adenosine and GABA(A) receptor alterations in the human basal ganglia in Huntington's disease. *Neuroscience* 97:505–519
- Goetz CG, Poewe W, Rascol O, Sampaio C (2005) Evidence-based medical review update: pharmacological and surgical treatments of Parkinson's disease: 2001 to 2004. *Mov Disord* 20:523–539
- Gomes CA, Vaz SH, Ribeiro JA, Sebastiao AM (2006) Glial cell line-derived neurotrophic factor (GDNF) enhances dopamine release from striatal nerve endings in an adenosine A2A receptor-dependent manner. *Brain Res* 1113:129–136
- Gordi T, Frohna P, Sun HL, Wolff A, Belardinelli L, Lieu H (2006) A population pharmacokinetic/pharmacodynamic analysis of regadenoson, an adenosine A2A-receptor agonist, in healthy male volunteers. *Clin Pharmacokinet* 45:1201–1212
- Graham DI, McIntosh TK, Maxwell WL, Nicoll JA (2000) Recent advances in neurotrauma. *J Neuropathol Exp Neurol* 59:641–651
- Greenland S (1993) A meta-analysis of coffee, myocardial infarction, and coronary death. *Epidemiology* 4:366–374

- Grobbee DE, Rimm EB, Giovannucci E, Colditz G, Stampfer M, Willett W (1990) Coffee, caffeine, and cardiovascular disease in men. *N Engl J Med* 323:1026–1032
- Grondin R, Bedard PJ, Hadj Tahar A, Gregoire L, Mori A, Kase H (1999) Antiparkinsonian effect of a new selective adenosine A2A receptor antagonist in MPTP-treated monkeys. *Neurology* 52:1673–1677
- Gsandtner I, Charalambous C, Stefan E, Ogris E, Freissmuth M, Zezula J (2005) Heterotrimeric G protein-independent signaling of a G protein-coupled receptor. Direct binding of ARNO/cytohesin-2 to the carboxyl terminus of the A2A adenosine receptor is necessary for sustained activation of the ERK/MAP kinase pathway. *J Biol Chem* 280:31898–31905
- Gubitz AK, Widdowson L, Kurokawa M, Kirkpatrick KA, Richardson PJ (1996) Dual signalling by the adenosine A2a receptor involves activation of both N- and P-type calcium channels by different G proteins and protein kinases in the same striatal nerve terminals. *J Neurochem* 67:374–381
- Guerreiro S, Toulorge D, Hirsch E, Marien M, Sokoloff P, Michel PP (2008) Paraxanthine, the primary metabolite of caffeine, provides protection against dopaminergic cell death via stimulation of ryanodine receptor channels. *Mol Pharmacol* 74:980–989
- Hafler DA (2004) Multiple sclerosis. *J Clin Invest* 113:788–794
- Hagberg H, Andersson P, Kjellmer I, Thiringer K, Thordstein M (1987) Extracellular overflow of glutamate, aspartate, GABA and taurine in the cortex and basal ganglia of fetal lambs during hypoxia-ischemia. *Neurosci Lett* 78:311–317
- Hameleers PA, Van Boxtel MP, Hogervorst E, Riedel WJ, Houx PJ, Buntinx F, Jolles J (2000) Habitual caffeine consumption and its relation to memory, attention, planning capacity and psychomotor performance across multiple age groups. *Hum Psychopharmacol* 15:573–581
- Hardy J, Gwinn-Hardy K (1998) Genetic classification of primary neurodegenerative disease. *Science* 282:1075–1079
- Hasko G, Kuhel DG, Chen JF, Schwarzschild MA, Deitch EA, Mabley JG, Marton A, Szabo C (2000) Adenosine inhibits IL-12 and TNF- $\alpha$  production via adenosine A2a receptor-dependent and independent mechanisms. *FASEB J* 14:2065–2074
- Hauser RA, Hubble JP, Truong DD (2003) Randomized trial of the adenosine A(2A) receptor antagonist istradefylline in advanced PD. *Neurology* 61:297–303
- Heron A, Lasbennes F, Seylaz J (1993) Adenosine modulation of amino acid release in rat hippocampus during ischemia and veratridine depolarization. *Brain Res* 608:27–32
- Heron A, Lekieffre D, Le Peillet E, Lasbennes F, Seylaz J, Plotkine M, Boulu RG (1994) Effects of an A1 adenosine receptor agonist on the neurochemical, behavioral and histological consequences of ischemia. *Brain Res* 641:217–224
- Higashi H, Meno JR, Marwaha AS, Winn HR (2002) Hippocampal injury and neurobehavioral deficits following hyperglycemic cerebral ischemia: effect of theophylline and ZM 241385. *J Neurosurg* 96:117–126
- Higdon JV, Frei B (2006) Coffee and health: a review of recent human research. *Crit Rev Food Sci Nutr* 46:101–123
- Hillion J, Canals M, Torvinen M, Casado V, Scott R, Terasmaa A, Hansson A, Watson S, Olah ME, Mallol J, Canela EI, Zoli M, Agnati LF, Ibanez CF, Lluis C, Franco R, Ferre S, Fuxe K (2002) Coaggregation, cointernalization, and codesensitization of adenosine A2A receptors and dopamine D2 receptors. *J Biol Chem* 277:18091–18097
- Hoane MR, Kaplan SA, Ellis AL (2006) The effects of nicotinamide on apoptosis and blood-brain barrier breakdown following traumatic brain injury. *Brain Res* 1125:185–193
- Hong CJ, Liu HC, Liu TY, Liao DL, Tsai SJ (2005) Association studies of the adenosine A2a receptor (1976T > C) genetic polymorphism in Parkinson's disease and schizophrenia. *J Neural Transm* 112:1503–1510
- Hoyte L, Kaur J, Buchan AM (2004) Lost in translation: taking neuroprotection from animal models to clinical trials. *Exp Neurol* 188:200–204
- Hsu SS, Meno JR, Zhou JG, Gordon EL, Winn HR (1991) Influence of hyperglycemia on cerebral adenosine production during ischemia and reperfusion. *Am J Physiol* 261:H398–H403



- Hu Y, Russek SJ (2008) BDNF and the diseased nervous system: a delicate balance between adaptive and pathological processes of gene regulation. *J Neurochem* 105:1–17
- Huang NK, Lin YW, Huang CL, Messing RO, Chern Y (2001) Activation of protein kinase A and atypical protein kinase C by A(2A) adenosine receptors antagonizes apoptosis due to serum deprivation in PC12 cells. *J Biol Chem* 276:13838–13846
- Hunter JM, Lesort M, Johnson GV (2007) Ubiquitin-proteasome system alterations in a striatal cell model of Huntington's disease. *J Neurosci Res* 85:1774–1788
- Ikeda K, Kurokawa M, Aoyama S, Kuwana Y (2002) Neuroprotection by adenosine A2A receptor blockade in experimental models of Parkinson's disease. *J Neurochem* 80:262–270
- Ivanov AA, Jacobson KA (2008) Molecular modeling of a PAMAM-CGS21680 dendrimer bound to an A2A adenosine receptor homodimer. *Bioorg Med Chem Lett* 18:4312–4315
- Jaarsma D, Sebens JB, Korf J (1991) Reduction of adenosine A1-receptors in the perforant pathway terminal zone in Alzheimer hippocampus. *Neurosci Lett* 121:111–114
- James JE (2004) Critical review of dietary caffeine and blood pressure: a relationship that should be taken more seriously. *Psychosom Med* 66:63–71
- Jankovic J (2008) Are adenosine antagonists, such as istradefylline, caffeine, and chocolate, useful in the treatment of Parkinson's disease? *Ann Neurol* 63:267–269
- Jansen KL, Faull RL, Dragunow M, Synek BL (1990) Alzheimer's disease: changes in hippocampal N-methyl-D-aspartate, quisqualate, neurotensin, adenosine, benzodiazepine, serotonin and opioid receptors—an autoradiographic study. *Neuroscience* 39:613–627
- Jarvis MJ (1993) Does caffeine intake enhance absolute levels of cognitive performance? *Psychopharmacology (Berl)* 110:45–52
- Jenner P, Mori A, Hauser R, Morelli M, Fredholm BB, Chen JF (2009) Adenosine, adenosine A2A antagonists, and Parkinson's disease. *Parkinsonism Relat Disord* 15:406–413
- Johnson-Kozlow M, Kritz-Silverstein D, Barrett-Connor E, Morton D (2002) Coffee consumption and cognitive function among older adults. *Am J Epidemiol* 156:842–850
- Johnston JB, Silva C, Gonzalez G, Holden J, Warren KG, Metz LM, Power C (2001) Diminished adenosine A1 receptor expression on macrophages in brain and blood of patients with multiple sclerosis. *Ann Neurol* 49:650–658
- Jones FS, Jing J, Stonehouse AH, Stevens A, Edelman GM (2008) Caffeine stimulates cytochrome oxidase expression and activity in the striatum in a sexually dimorphic manner. *Mol Pharmacol* 74:673–684
- Justinova Z, Ferre S, Barnes C, Wertheim CE, Pappas LA, Goldberg SR, Le Foll B (2009) Effects of chronic caffeine exposure on adenosinergic modulation of the discriminative-stimulus effects of nicotine, methamphetamine, and cocaine in rats. *Psychopharmacology* 203:355–367
- Kachroo A, Orlando LR, Grandy DK, Chen JF, Young AB, Schwarzschild MA (2005) Interactions between metabotropic glutamate 5 and adenosine A2A receptors in normal and parkinsonian mice. *J Neurosci* 25:10414–10419
- Kamiya T, Saitoh O, Yoshioka K, Nakata H (2003) Oligomerization of adenosine A2A and dopamine D2 receptors in living cells. *Biochem Biophys Res Commun* 306:544–549
- Kanda T, Tashiro T, Kuwana Y, Jenner P (1998a) Adenosine A2A receptors modify motor function in MPTP-treated common marmosets. *Neuroreport* 9:2857–2860
- Kanda T, Jackson MJ, Smith LA, Pearce RK, Nakamura J, Kase H, Kuwana Y, Jenner P (1998b) Adenosine A2A antagonist: a novel antiparkinsonian agent that does not provoke dyskinesia in parkinsonian monkeys. *Ann Neurol* 43:507–513
- Kanda T, Jackson MJ, Smith LA, Pearce RK, Nakamura J, Kase H, Kuwana Y, Jenner P (2000) Combined use of the adenosine A(2A) antagonist KW-6002 with L-DOPA or with selective D1 or D2 dopamine agonists increases antiparkinsonian activity but not dyskinesia in MPTP-treated monkeys. *Exp Neurol* 162:321–327
- Kang SH, Lee YA, Won SJ, Rhee KH, Gwag BJ (2002) Caffeine-induced neuronal death in neonatal rat brain and cortical cell cultures. *Neuroreport* 13:1945–1950
- Keegan BM, Noseworthy JH (2002) Multiple sclerosis. *Annu Rev Med* 53:285–302

- Kim Y, Hechler B, Klutz AM, Gachet C, Jacobson KA (2008) Toward multivalent signaling across G protein-coupled receptors from poly(amidoamine) dendrimers. *Bioconjug Chem* 19:406–411
- Kittner B, Rossner M, Rother M (1997) Clinical trials in dementia with propentofylline. *Ann N Y Acad Sci* 826:307–316
- Kochanek PM, Vagni VA, Janesko KL, Washington CB, Crumrine PK, Garman RH, Jenkins LW, Clark RS, Homanics GE, Dixon CE, Schnermann J, Jackson EK (2006) Adenosine A1 receptor knockout mice develop lethal status epilepticus after experimental traumatic brain injury. *J Cereb Blood Flow Metab* 26:565–575
- Kortekaas R, Leenders KL, van Oostrom JC, Vaalburg W, Bart J, Willemsen AT, Hendrikse NH (2005) Blood-brain barrier dysfunction in parkinsonian midbrain in vivo. *Ann Neurol* 57:176–179
- Lang AE, Lozano AM (1998a) Parkinson's disease. Second of two parts. *N Engl J Med* 339:1130–1143
- Lang AE, Lozano AM (1998b) Parkinson's disease. First of two parts. *N Engl J Med* 339:1044–1053
- Lang AE, Obeso JA (2004) Challenges in Parkinson's disease: restoration of the nigrostriatal dopamine system is not enough. *Lancet Neurol* 3:309–316
- Lapchak PA, Song D, Wei J, Zivin JA (2004) Pharmacology of caffeine in embolized rabbits: clinical rating scores and intracerebral hemorrhage incidence. *Exp Neurol* 188:286–291
- Lapchak PA, Araujo DM, Pakola S, Song D, Wei J, Zivin JA (2002) Microplasmin: a novel thrombolytic that improves behavioral outcome after embolic strokes in rabbits. *Stroke* 33:2279–2284
- Latini S, Pedata F (2001) Adenosine in the central nervous system: release mechanisms and extracellular concentrations. *J Neurochem* 79:463–484
- Latour LL, Kang DW, Ezzeddine MA, Chalela JA, Warach S (2004) Early blood-brain barrier disruption in human focal brain ischemia. *Ann Neurol* 56:468–477
- Ledent C, Vaugeois JM, Schiffmann SN, Pedrazzini T, El Yacoubi M, Vanderhaeghen JJ, Costentin J, Heath JK, Vassart G, Parmentier M (1997) Aggressiveness, hypoalgesia and high blood pressure in mice lacking the adenosine A2a receptor. *Nature* 388:674–678
- Lee FS, Chao MV (2001) Activation of Trk neurotrophin receptors in the absence of neurotrophins. *Proc Natl Acad Sci USA* 98:3555–3560
- Leker RR, Shohami E (2002) Cerebral ischemia and trauma-different etiologies yet similar mechanisms: neuroprotective opportunities. *Brain Res Brain Res Rev* 39:55–73
- Li SH, Li XJ (2004) Huntingtin-protein interactions and the pathogenesis of Huntington's disease. *Trends Genet* 20:146–154
- Li W, Dai S, An J, Xiong R, Li P, Chen X, Zhao Y, Liu P, Wang H, Zhu P, Chen J, Zhou Y (2009) Genetic inactivation of adenosine A2A receptors attenuates acute traumatic brain injury in the mouse cortical impact model. *Exp Neurol* 215:69–76
- Li W, Dai S, An J, Li P, Chen X, Xiong R, Liu P, Wang H, Zhao Y, Zhu M, Liu X, Zhu P, Chen JF, Zhou Y (2008) Chronic but not acute treatment with caffeine attenuates traumatic brain injury in the mouse cortical impact model. *Neuroscience* 151:1198–1207
- Li XX, Nomura T, Aihara H, Nishizaki T (2001) Adenosine enhances glial glutamate efflux via A2a adenosine receptors. *Life Sci* 68:1343–1350
- Li Y, Oskouian RJ, Day YJ, Rieger JM, Liu L, Kern JA, Linden J (2006) Mouse spinal cord compression injury is reduced by either activation of the adenosine A2A receptor on bone marrow-derived cells or deletion of the A2A receptor on non-bone marrow-derived cells. *Neuroscience* 141:2029–2039
- Lindsay J, Laurin D, Verreault R, Hebert R, Helliwell B, Hill GB, McDowell I (2002) Risk factors for Alzheimer's disease: a prospective analysis from the Canadian Study of Health and Aging. *Am J Epidemiol* 156:445–453
- Lo EH, Dalkara T, Moskowitz MA (2003) Mechanisms, challenges and opportunities in stroke. *Nat Rev Neurosci* 4:399–415

- Lopes LV, Cunha RA, Kull B, Fredholm BB, Ribeiro JA (2002) Adenosine A(2A) receptor facilitation of hippocampal synaptic transmission is dependent on tonic A(1) receptor inhibition. *Neuroscience* 112:319–329
- Lopez-Diego RS, Weiner HL (2008) Novel therapeutic strategies for multiple sclerosis—a multifaceted adversary. *Nat Rev Drug Discov* 7:909–925
- Lundblad M, Vaudano E, Cenci MA (2003) Cellular and behavioural effects of the adenosine A2a receptor antagonist KW-6002 in a rat model of L-DOPA-induced dyskinesia. *J Neurochem* 84:1398–1410
- Lupica CR, Berman RF, Jarvis MF (1991) Chronic theophylline treatment increases adenosine A1, but not A2, receptor binding in the rat brain: an autoradiographic study. *Synapse* 9:95–102
- Maglione V, Cannella M, Martino T, De Blasi A, Frati L, Squitieri F (2006) The platelet maximum number of A2A-receptor binding sites (Bmax) linearly correlates with age at onset and CAG repeat expansion in Huntington's disease patients with predominant chorea. *Neurosci Lett* 393:27–30
- Maia L, de Mendonca A (2002) Does caffeine intake protect from Alzheimer's disease? *Eur J Neurol* 9:377–382
- Marangos PJ, Paul SM, Parma AM, Goodwin FK, Syapin P, Skolnick P (1979) Purinergic inhibition of diazepam binding to rat brain (in vitro). *Life Sci* 24:851–857
- Marchi M, Raiteri L, Risso F, Vallarino A, Bonfanti A, Monopoli A, Ongini E, Raiteri M (2002) Effects of adenosine A1 and A2A receptor activation on the evoked release of glutamate from rat cerebrocortical synaptosomes. *Br J Pharmacol* 136:434–440
- Marcusson J, Rother M, Kittner B, Rossner M, Smith RJ, Babic T, Folnegovic-Smalc V, Moller HJ, Labs KH (1997) A 12-month, randomized, placebo-controlled trial of propentofylline (HWA 285) in patients with dementia according to DSM III-R. The European Propentofylline Study Group. *Dement Geriatr Cogn Disord* 8:320–328
- Marshall LF (2000) Head injury: recent past, present, and future. *Neurosurgery* 47:546–561
- Martin-Schild S, Hallevi H, Shaltoni H, Barreto AD, Gonzales NR, Aronowski J, Savitz SI, Grotta JC (2009) Combined neuroprotective modalities coupled with thrombolysis in acute ischemic stroke: a pilot study of caffeine and mild hypothermia. *J Stroke Cerebrovasc Dis* 18:86–96
- Martin JB, Gusella JF (1986) Huntington's disease. Pathogenesis and management. *N Engl J Med* 315:1267–1276
- Matsumoto K, Graf R, Rosner G, Shimada N, Heiss WD (1992) Flow thresholds for extracellular purine catabolite elevation in cat focal ischemia. *Brain Res* 579:309–314
- Mattson MP (2000) Apoptosis in neurodegenerative disorders. *Nat Rev Mol Cell Biol* 1:120–129
- Mattson MP (2004) Pathways towards and away from Alzheimer's disease. *Nature* 430:631–639
- Mayne M, Shepel PN, Jiang Y, Geiger JD, Power C (1999) Dysregulation of adenosine A1 receptor-mediated cytokine expression in peripheral blood mononuclear cells from multiple sclerosis patients. *Ann Neurol* 45:633–639
- Mayne M, Fotheringham J, Yan HJ, Power C, Del Bigio MR, Peeling J, Geiger JD (2001) Adenosine A2A receptor activation reduces proinflammatory events and decreases cell death following intracerebral hemorrhage. *Ann Neurol* 49:727–735
- McPherson PS, Kim YK, Valdivia H, Knudson CM, Takekura H, Franzini-Armstrong C, Coronado R, Campbell KP (1991) The brain ryanodine receptor: a caffeine-sensitive calcium release channel. *Neuron* 7:17–25
- Meinl E, Krumbholz M, Hohlfeld R (2006) B lineage cells in the inflammatory central nervous system environment: migration, maintenance, local antibody production, and therapeutic modulation. *Ann Neurol* 59:880–892
- Melani A, Gianfriddo M, Vannucchi MG, Cipriani S, Baraldi PG, Giovannini MG, Pedata F (2006) The selective A2A receptor antagonist SCH 58261 protects from neurological deficit, brain damage and activation of p38 MAPK in rat focal cerebral ischemia. *Brain Res* 1073–1074:470–480
- Melani A, Cipriani S, Vannucchi MG, Nosi D, Donati C, Bruni P, Giovannini MG, Pedata F (2009) Selective adenosine A2a receptor antagonism reduces JNK activation in oligodendrocytes after cerebral ischaemia. *Brain* 132:1480–1495

- Melani A, Pantoni L, Bordoni F, Gianfriddo M, Bianchi L, Vannucchi MG, Bertorelli R, Monopoli A, Pedata F (2003) The selective A2A receptor antagonist SCH 58261 reduces striatal transmitter outflow, turning behavior and ischemic brain damage induced by permanent focal ischemia in the rat. *Brain Res* 959:243–250
- Mestre T, Ferreira J, Coelho MM, Rosa M, Sampaio C (2009) Therapeutic interventions for disease progression in Huntington's disease. *Cochrane Database Syst Rev* CD006455
- Mielke R, Kessler J, Szelies B, Herholz K, Wienhard K, Heiss WD (1996a) Vascular dementia: perfusional and metabolic disturbances and effects of therapy. *J Neural Transm Suppl* 47:183–191
- Mielke R, Kittner B, Ghaemi M, Kessler J, Szelies B, Herholz K, Heiss WD (1996b) Propentofylline improves regional cerebral glucose metabolism and neuropsychologic performance in vascular dementia. *J Neurol Sci* 141:59–64
- Mihm MJ, Amann DM, Schanbacher BL, Altschuld RA, Bauer JA, Hoyt KR (2007) Cardiac dysfunction in the R6/2 mouse model of Huntington's disease. *Neurobiol Dis* 25:297–308
- Mills JH, Thompson LF, Mueller C, Waickman AT, Jalkanen S, Niemela J, Airas L, Bynoe MS (2008) CD73 is required for efficient entry of lymphocytes into the central nervous system during experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci USA* 105:9325–9330
- Milojevic T, Reiterer V, Stefan E, Korkhov VM, Dorostkar MM, Ducza E, Ogris E, Boehm S, Freissmuth M, Nanoff C (2006) The ubiquitin-specific protease Usp4 regulates the cell surface level of the A2A receptor. *Mol Pharmacol* 69:1083–1094
- Minagar A, Alexander JS (2003) Blood-brain barrier disruption in multiple sclerosis. *Mult Scler* 9:540–549
- Mingote S, Pereira M, Farrar AM, McLaughlin PJ, Salamone JD (2008) Systemic administration of the adenosine A(2A) agonist CGS 21680 induces sedation at doses that suppress lever pressing and food intake. *Pharmacol Biochem Behav* 89:345–351
- Monopoli A, Lozza G, Forlani A, Mattavelli A, Ongini E (1998) Blockade of adenosine A2A receptors by SCH 58261 results in neuroprotective effects in cerebral ischaemia in rats. *Neuroreport* 9:3955–3959
- Moppett IK (2007) Traumatic brain injury: assessment, resuscitation and early management. *Br J Anaesth* 99:18–31
- Morelli M, Di Paolo T, Wardas J, Calon F, Xiao D, Schwarzschild MA (2007) Role of adenosine A2A receptors in parkinsonian motor impairment and l-DOPA-induced motor complications. *Prog Neurobiol* 83:293–309
- Navarro G, Carriba P, Gandia J, Ciruela F, Casado V, Cortes A, Mallol J, Canela EI, Lluís C, Franco R (2008) Detection of heteromers formed by cannabinoid CB1, dopamine D2, and adenosine A2A G-protein-coupled receptors by combining bimolecular fluorescence complementation and bioluminescence energy transfer. *ScientificWorldJournal* 8:1088–1097
- Navarro G, Aymerich MS, Marcellino D, Cortes A, Casado V, Mallol J, Canela EI, Agnati L, Woods AS, Fuxe K, Lluís C, Lanciego JL, Ferre S, Franco R (2009) Interactions between calmodulin, adenosine A2A, and dopamine D2 receptors. *J Biol Chem* 284:28058–28068
- Nehlig A, Daval JL, Debry G (1992) Caffeine and the central nervous system: mechanisms of action, biochemical, metabolic and psychostimulant effects. *Brain Res Brain Res Rev* 17:139–170
- Ngai AC, Coyne EF, Meno JR, West GA, Winn HR (2001) Receptor subtypes mediating adenosine-induced dilation of cerebral arterioles. *Am J Physiol Heart Circ Physiol* 280: H2329–H2335
- Nikbakht MR, Stone TW (2001) Suppression of presynaptic responses to adenosine by activation of NMDA receptors. *Eur J Pharmacol* 427:13–25
- Nishizaki T, Nagai K, Nomura T, Tada H, Kanno T, Tozaki H, Li XX, Kondoh T, Kodama N, Takahashi E, Sakai N, Tanaka K, Saito N (2002) A new neuromodulatory pathway with a glial contribution mediated via A(2a) adenosine receptors. *Glia* 39:133–147
- Norenberg W, Wirkner K, Illes P (1997) Effect of adenosine and some of its structural analogues on the conductance of NMDA receptor channels in a subset of rat neostriatal neurones. *Br J Pharmacol* 122:71–80

- Norenberg W, Wirkner K, Assmann H, Richter M, Illes P (1998) Adenosine A2A receptors inhibit the conductance of NMDA receptor channels in rat neostriatal neurons. *Amino Acids* 14:33–39
- Noseworthy JH, Lucchinetti C, Rodriguez M, Weinschenker BG (2000) Multiple sclerosis. *N Engl J Med* 343:938–952
- O'Regan MH, Simpson RE, Perkins LM, Phillis JW (1992) The selective A2 adenosine receptor agonist CGS 21680 enhances excitatory transmitter amino acid release from the ischemic rat cerebral cortex. *Neurosci Lett* 138:169–172
- Ohta A, Sitkovsky M (2001) Role of G-protein-coupled adenosine receptors in downregulation of inflammation and protection from tissue damage. *Nature* 414:916–920
- Oishi Y, Huang ZL, Fredholm BB, Urade Y, Hayaishi O (2008) Adenosine in the tuberomammillary nucleus inhibits the histaminergic system via A1 receptors and promotes non-rapid eye movement sleep. *Proc Natl Acad Sci USA* 105:19992–19997
- Olanow CW (2004) The scientific basis for the current treatment of Parkinson's disease. *Annu Rev Med* 55:41–60
- Olsson T, Cronberg T, Rytter A, Asztely F, Fredholm BB, Smith ML, Wieloch T (2004) Deletion of the adenosine A1 receptor gene does not alter neuronal damage following ischaemia in vivo or in vitro. *Eur J Neurosci* 20:1197–1204
- Oztas E, Xu K, Kalda A, Irrizary M, Schwarzschild MA, Chen J-F (2002) Caffeine attenuates MPTP-induced loss of dopaminergic neurons in substantial nigra in mice. In: Annual meeting of Society for Neuroscience. Orlando, FL
- Pedata F, Corsi C, Melani A, Bordoni F, Latini S (2001) Adenosine extracellular brain concentrations and role of A2A receptors in ischemia. *Ann N Y Acad Sci* 939:74–84
- Perrin RJ, Fagan AM, Holtzman DM (2009) Multimodal techniques for diagnosis and prognosis of Alzheimer's disease. *Nature* 461:916–922
- Phan J, Hickey MA, Zhang P, Chesselet MF, Reue K (2009) Adipose tissue dysfunction tracks disease progression in two Huntington's disease mouse models. *Hum Mol Genet* 18:1006–1016
- Phillis JW (1995) The effects of selective A1 and A2a adenosine receptor antagonists on cerebral ischemic injury in the gerbil. *Brain Res* 705:79–84
- Pierri M, Vaudano E, Sager T, Englund U (2005) KW-6002 protects from MPTP induced dopaminergic toxicity in the mouse. *Neuropharmacology* 48:517–524
- Pignataro G, Simon RP, Boison D (2007) Transgenic overexpression of adenosine kinase aggravates cell death in ischemia. *J Cereb Blood Flow Metab* 27:1–5
- Pincomb GA, Lovallo WR, Passey RB, Wilson MF (1988) Effect of behavior state on caffeine's ability to alter blood pressure. *Am J Cardiol* 61:798–802
- Piriyawat P, Labiche LA, Burgin WS, Aronowski JA, Grotta JC (2003) Pilot dose-escalation study of caffeine plus ethanol (caffeinol) in acute ischemic stroke. *Stroke* 34:1242–1245
- Popoli P, Blum D, Domenici MR, Burnouf S, Chern Y (2008) A critical evaluation of adenosine A2A receptors as potentially "druggable" targets in Huntington's disease. *Curr Pharm Des* 14:1500–1511
- Popoli P, Frank C, Tebano MT, Potenza RL, Pintor A, Domenici MR, Nazzicone V, Pezzola A, Reggio R (2003) Modulation of glutamate release and excitotoxicity by adenosine A2A receptors. *Neurology* 61:S69–S71
- Popoli P, Pintor A, Domenici MR, Frank C, Tebano MT, Pezzola A, Scarchilli L, Quarta D, Reggio R, Malchiodi-Albedi F, Falchi M, Massotti M (2002) Blockade of striatal adenosine A2A receptor reduces, through a presynaptic mechanism, quinolinic acid-induced excitotoxicity: possible relevance to neuroprotective interventions in neurodegenerative diseases of the striatum. *J Neurosci* 22:1967–1975
- Prediger RD, Takahashi RN (2005) Modulation of short-term social memory in rats by adenosine A1 and A(2A) receptors. *Neurosci Lett* 376:160–165
- Prediger RD, Batista LC, Takahashi RN (2005) Caffeine reverses age-related deficits in olfactory discrimination and social recognition memory in rats. Involvement of adenosine A1 and A2A receptors. *Neurobiol Aging* 26:957–964

- Prineas JW, Wright RG (1978) Macrophages, lymphocytes, and plasma cells in the perivascular compartment in chronic multiple sclerosis. *Lab Invest* 38:409–421
- Ragab S, Lunt M, Birch A, Thomas P, Jenkinson DF (2004) Caffeine reduces cerebral blood flow in patients recovering from an ischaemic stroke. *Age Ageing* 33:299–303
- Raine CS (1984) Biology of disease. Analysis of autoimmune demyelination: its impact upon multiple sclerosis. *Lab Invest* 50:608–635
- Rainnie DG, Grunze HC, McCarley RW, Greene RW (1994) Adenosine inhibition of mesopontine cholinergic neurons: implications for EEG arousal. *Science* 263:689–692
- Rascol O, Goetz C, Koller W, Poewe W, Sampaio C (2002) Treatment interventions for Parkinson's disease: an evidence based assessment. *Lancet* 359:1589–1598
- Reece TB, Davis JD, Okonkwo DO, Maxey TS, Ellman PI, Li X, Linden J, Tribble CG, Kron IL, Kern JA (2004) Adenosine A2A analogue reduces long-term neurologic injury after blunt spinal trauma. *J Surg Res* 121:130–134
- Ribchester RR, Thomson D, Wood NI, Hinks T, Gillingwater TH, Wishart TM, Court FA, Morton AJ (2004) Progressive abnormalities in skeletal muscle and neuromuscular junctions of transgenic mice expressing the Huntington's disease mutation. *Eur J Neurosci* 20:3092–3114
- Richardson PJ, Kase H, Jenner PG (1997) Adenosine A2A receptor antagonists as new agents for the treatment of Parkinson's disease. *Trends Pharmacol Sci* 18:338–344
- Richardson PJ, Gubitz AK, Freeman TC, Dixon AK (1999) Adenosine receptor antagonists and Parkinson's disease: actions of the A2A receptor in the striatum. *Adv Neurol* 80:111–119
- Ritchie K, Carrière I, Portet F, de Mendonca A, Dartigues JF, Rouaud O, Barberger-Gateau P, Ancelin ML (2007) The neuro-protective effects of caffeine: a prospective population study (the Three City Study). *Neurology* 69:536–545
- Rite I, Machado A, Cano J, Venero JL (2007) Blood-brain barrier disruption induces in vivo degeneration of nigral dopaminergic neurons. *J Neurochem* 101:1567–1582
- Robertson CL, Bell MJ, Kochanek PM, Adelson PD, Ruppel RA, Carcillo JA, Wisniewski SR, Mi Z, Janesko KL, Clark RS, Marion DW, Graham SH, Jackson EK (2001) Increased adenosine in cerebrospinal fluid after severe traumatic brain injury in infants and children: association with severity of injury and excitotoxicity. *Crit Care Med* 29:2287–2293
- Robledo P, Ursu G, Mahy N (1999) Effects of adenosine and gamma-aminobutyric acid A receptor antagonists on N-methyl-D-aspartate induced neurotoxicity in the rat hippocampus. *Hippocampus* 9:527–533
- Rosin DL, Hettinger BD, Lee A, Linden J (2003) Anatomy of adenosine A2A receptors in brain: morphological substrates for integration of striatal function. *Neurology* 61:S12–S18
- Ross GW, Abbott RD, Petrovitch H, Morens DM, Grandinetti A, Tung KH, Tanner CM, Masaki KH, Blanchette PL, Curb JD, Popper JS, White LR (2000) Association of coffee and caffeine intake with the risk of Parkinson disease. *JAMA* 283:2674–2679
- Rudolphi KA, Schubert P, Parkinson FE, Fredholm BB (1992) Neuroprotective role of adenosine in cerebral ischaemia. *Trends Pharmacol Sci* 13:439–445
- Rutland-Brown W, Langlois JA, Thomas KE, Xi YL (2006) Incidence of traumatic brain injury in the United States, 2003. *J Head Trauma Rehabil* 21:544–548
- Ryu H, Rosas HD, Hersch SM, Ferrante RJ (2005) The therapeutic role of creatine in Huntington's disease. *Pharmacol Ther* 108:193–207
- Saaksjarvi K, Knekt P, Rissanen H, Laaksonen MA, Reunanen A, Mannisto S (2007) Prospective study of coffee consumption and risk of Parkinson's disease. *Eur J Clin Nutr* 62:908–915
- Sachse KT, Jackson EK, Wisniewski SR, Gillespie DG, Puccio AM, Clark RS, Dixon CE, Kochanek PM (2008) Increases in cerebrospinal fluid caffeine concentration are associated with favorable outcome after severe traumatic brain injury in humans. *J Cereb Blood Flow Metab* 28:395–401
- Salamone JD, Farrar AM, Font L, Patel V, Schlar DE, Nunes EJ, Collins LE, Sager TN (2009) Differential actions of adenosine A1 and A2A antagonists on the effort-related effects of dopamine D2 antagonism. *Behav Brain Res* 201:216–222

- Sandoli D, Chiu PJ, Chintala M, Dionisotti S, Ongini E (1994) In vivo and ex vivo effects of adenosine A1 and A2 receptor agonists on platelet aggregation in the rabbit. *Eur J Pharmacol* 259:43–49
- Sathasivam K, Hobbs C, Turmaine M, Mangiarini L, Mahal A, Bertaux F, Wanker EE, Doherty P, Davies SW, Bates GP (1999) Formation of polyglutamine inclusions in non-CNS tissue. *Hum Mol Genet* 8:813–822
- Schwarzschild MA, Chen JF, Ascherio A (2002) Caffeinated clues and the promise of adenosine A (2A) antagonists in PD. *Neurology* 58:1154–1160
- Schwarzschild MA, Agnati L, Fuxe K, Chen JF, Morelli M (2006) Targeting adenosine A2A receptors in Parkinson's disease. *Trends Neurosci* 29:647–654
- Sebastiao AM, Ribeiro JA (1996) Adenosine A2 receptor-mediated excitatory actions on the nervous system. *Prog Neurobiol* 48:167–189
- Sebastiao AM, Ribeiro JA (2009a) Triggering neurotrophic factor actions through adenosine A2A receptor activation: implications for neuroprotection. *Br J Pharmacol* 158:15–22
- Sebastiao AM, Ribeiro JA (2009b) Adenosine receptors and the central nervous system. *Handb Exp Pharmacol* 193:471–534
- Selkoe DJ (2004) Alzheimer disease: mechanistic understanding predicts novel therapies. *Ann Intern Med* 140:627–638
- Selley ML (2004) Increased homocysteine and decreased adenosine formation in Alzheimer's disease. *Neurol Res* 26:554–557
- Seo H, Sonntag KC, Isacson O (2004) Generalized brain and skin proteasome inhibition in Huntington's disease. *Ann Neurol* 56:319–328
- Shen H, Zhang L, Yuen D, Logan R, Jung BP, Zhang G, Eubanks JH (2002) Expression and function of A1 adenosine receptors in the rat hippocampus following transient forebrain ischemia. *Neuroscience* 114:547–556
- Shen HY, Coelho JE, Ohtsuka N, Canas PM, Day YJ, Huang QY, Rebola N, Yu L, Boison D, Cunha RA, Linden J, Tsien JZ, Chen JF (2008) A critical role of the adenosine A2A receptor in extrastriatal neurons in modulating psychomotor activity as revealed by opposite phenotypes of striatum and forebrain A2A receptor knock-outs. *J Neurosci* 28:2970–2975
- Shi D, Nikodijevic O, Jacobson KA, Daly JW (1993) Chronic caffeine alters the density of adenosine, adrenergic, cholinergic, GABA, and serotonin receptors and calcium channels in mouse brain. *Cell Mol Neurobiol* 13:247–261
- Shiozaki S, Ichikawa S, Nakamura J, Kitamura S, Yamada K, Kuwana Y (1999) Actions of adenosine A2A receptor antagonist KW-6002 on drug-induced catalepsy and hypokinesia caused by reserpine or MPTP. *Psychopharmacology (Berl)* 147:90–95
- Shoulson I (1998) Experimental therapeutics of neurodegenerative disorders: unmet needs. *Science* 282:1072–1074
- Shoulson I, Chase T (1975) Caffeine and the antiparkinsonian response to levodopa or piribedil. *Neurology* 25:722–724
- Simon DK, Swearingen CJ, Hauser RA, Trugman JM, Aminoff MJ, Singer C, Truong D, Tilley BC (2008) Caffeine and progression of Parkinson disease. *Clin Neuropharmacol* 31:189–196
- Simpson RE, O'Regan MH, Perkins LM, Phillis JW (1992) Excitatory transmitter amino acid release from the ischemic rat cerebral cortex: effects of adenosine receptor agonists and antagonists. *J Neurochem* 58:1683–1690
- Singh S, Singh K, Gupta SP, Patel DK, Singh VK, Singh RK, Singh MP (2009) Effect of caffeine on the expression of cytochrome P450 1A2, adenosine A2A receptor and dopamine transporter in control and 1-methyl 4-phenyl 1,2,3,6-tetrahydropyridine treated mouse striatum. *Brain Res* 1283:115–126
- Sitkovsky MV, Lukashev D, Apasov S, Kojima H, Koshiba M, Caldwell C, Ohta A, Thiel M (2004) Physiological control of immune response and inflammatory tissue damage by hypoxia-inducible factors and adenosine A2A receptors. *Annu Rev Immunol* 22:657–682
- Smith PA, Heijmans N, Ouwerling B, Breij EC, Evans N, van Noort JM, Plomp AC, Delarasse C, 't Hart B, Pham-Dinh D, Amor S (2005) Native myelin oligodendrocyte glycoprotein promotes

- severe chronic neurological disease and demyelination in Biozzi ABH mice. *Eur J Immunol* 35:1311–1319
- Soriano A, Ventura R, Molero A, Hoen R, Casado V, Cortes A, Fanelli F, Albericio F, Lluís C, Franco R, Royo M (2009) Adenosine A2A receptor-antagonist/dopamine D2 receptor-agonist bivalent ligands as pharmacological tools to detect A2A-D2 receptor heteromers. *J Med Chem* 52:5590–5602
- Stacy M, Silver D, Mendis T, Sutton J, Mori A, Chaikin P, Sussman NM (2008) A 12-week, placebo-controlled study (6002-US-006) of istradefylline in Parkinson disease. *Neurology* 70:2233–2240
- Stamey W, Jankovic J (2008) Impulse control disorders and pathological gambling in patients with Parkinson disease. *Neurologist* 14:89–99
- Steinman L, Zamvil SS (2006) How to successfully apply animal studies in experimental allergic encephalomyelitis to research on multiple sclerosis. *Ann Neurol* 60:12–21
- Strittmatter WJ, Saunders AM, Schmechel D, Pericak-Vance M, Enghild J, Salvesen GS, Roses AD (1993) Apolipoprotein E: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc Natl Acad Sci USA* 90:1977–1981
- Strong R, Grotta JC, Aronowski J (2000) Combination of low dose ethanol and caffeine protects brain from damage produced by focal ischemia in rats. *Neuropharmacology* 39:515–522
- Sugas KL, Rubinsztein DC (2003) Transcriptional abnormalities in Huntington disease. *Trends Genet* 19:233–238
- Sun CN, Cheng HC, Chou JL, Lee SY, Lin YW, Lai HL, Chen HM, Chern Y (2006) Rescue of p53 blockage by the A2A adenosine receptor via a novel interacting protein, Translin-associated protein X. *Mol Pharmacol* 70:454–466
- Svenningsson P, Nomikos GG, Fredholm BB (1995) Biphasic changes in locomotor behavior and in expression of mRNA for NGFI-A and NGFI-B in rat striatum following acute caffeine administration. *J Neurosci* 15:7612–7624
- Svenningsson P, Le Moine C, Fisone G, Fredholm BB (1999) Distribution, biochemistry and function of striatal adenosine A2A receptors. *Prog Neurobiol* 59:355–396
- Szabo C, Pacher P, Swanson RA (2006) Novel modulators of poly(ADP-ribose) polymerase. *Trends Pharmacol Sci* 27:626–630
- Tebano MT, Martire A, Potenza RL, Gro C, Pepponi R, Armida M, Domenici MR, Schwarzschild MA, Chen JF, Popoli P (2008) Adenosine A(2A) receptors are required for normal BDNF levels and BDNF-induced potentiation of synaptic transmission in the mouse hippocampus. *J Neurochem* 104:279–286
- The Huntington's Disease Collaborative Research Group (THsDCR) (1993) A novel gene containing a trinucleotide repeat that is expanded and unstable on Huntington's disease chromosomes. The Huntington's Disease Collaborative Research Group. *Cell* 72:971–983
- Torvinen M, Marcellino D, Canals M, Agnati LF, Lluís C, Franco R, Fuxe K (2005) Adenosine A2A receptor and dopamine D3 receptor interactions: evidence of functional A2A/D3 heteromeric complexes. *Mol Pharmacol* 67:400–407
- Trevitt J, Vallance C, Harris A, Goode T (2009) Adenosine antagonists reverse the cataleptic effects of haloperidol: implications for the treatment of Parkinson's disease. *Pharmacol Biochem Behav* 92:521–527
- Tronci E, Simola N, Borsini F, Schintu N, Frau L, Carminati P, Morelli M (2007) Characterization of the antiparkinsonian effects of the new adenosine A2A receptor antagonist ST1535: acute and subchronic studies in rats. *Eur J Pharmacol* 566:94–102
- Tsutsui S, Vergote D, Shariat N, Warren K, Ferguson SS, Power C (2008) Glucocorticoids regulate innate immunity in a model of multiple sclerosis: reciprocal interactions between the A1 adenosine receptor and beta-arrestin-1 in monocytoid cells. *FASEB J* 22:786–796
- Tsutsui S, Schnermann J, Noorbakhsh F, Henry S, Yong VW, Winston BW, Warren K, Power C (2004) A1 adenosine receptor upregulation and activation attenuates neuroinflammation and demyelination in a model of multiple sclerosis. *J Neurosci* 24:1521–1529



- Turner CP, Seli M, Ment L, Stewart W, Yan H, Johansson B, Fredholm BB, Blackburn M, Rivkees SA (2003) A1 adenosine receptors mediate hypoxia-induced ventriculomegaly. *Proc Natl Acad Sci USA* 100:11718–11722
- Ulas J, Brunner LC, Nguyen L, Cotman CW (1993) Reduced density of adenosine A1 receptors and preserved coupling of adenosine A1 receptors to G proteins in Alzheimer hippocampus: a quantitative autoradiographic study. *Neuroscience* 52:843–854
- Valenza M, Carroll JB, Leoni V, Bertram LN, Bjorkhem I, Singaraja RR, Di Donato S, Lutjohann D, Hayden MR, Cattaneo E (2007) Cholesterol biosynthesis pathway is disturbed in YAC128 mice and is modulated by huntingtin mutation. *Hum Mol Genet* 16:2187–2198
- van Boxtel MP, Schmitt JA, Bosma H, Jolles J (2003) The effects of habitual caffeine use on cognitive change: a longitudinal perspective. *Pharmacol Biochem Behav* 75:921–927
- van Dam RM, Willett WC, Manson JE, Hu FB (2006) Coffee, caffeine, and risk of type 2 diabetes: a prospective cohort study in younger and middle-aged U.S. women. *Diabetes Care* 29:398–403
- Varani K, Rigamonti D, Sipione S, Camurri A, Borea PA, Cattabeni F, Abbraccio MP, Cattaneo E (2001) Aberrant amplification of A(2A) receptor signaling in striatal cells expressing mutant huntingtin. *FASEB J* 15:1245–1247
- Varani K, Abbraccio MP, Cannella M, Cislighi G, Giallonardo P, Mariotti C, Cattabriga E, Cattabeni F, Borea PA, Squitieri F, Cattaneo E (2003) Aberrant A2A receptor function in peripheral blood cells in Huntington's disease. *FASEB J* 17:2148–2150
- Varma MR, Dixon CE, Jackson EK, Peters GW, Melick JA, Griffith RP, Vagni VA, Clark RS, Jenkins LW, Kochanek PM (2002) Administration of adenosine receptor agonists or antagonists after controlled cortical impact in mice: effects on function and histopathology. *Brain Res* 951:191–201
- Varty GB, Hodgson RA, Pond AJ, Grzelak ME, Parker EM, Hunter JC (2008) The effects of adenosine A2A receptor antagonists on haloperidol-induced movement disorders in primates. *Psychopharmacology* 200:393–401
- Vidi PA, Chemel BR, Hu CD, Watts VJ (2008) Ligand-dependent oligomerization of dopamine D (2) and adenosine A(2A) receptors in living neuronal cells. *Mol Pharmacol* 74:544–551
- Von Lubitz DK, Lin RC, Jacobson KA (1995) Cerebral ischemia in gerbils: effects of acute and chronic treatment with adenosine A2A receptor agonist and antagonist. *Eur J Pharmacol* 287:295–302
- Von Lubitz DK, Lin RC, Melman N, Ji XD, Carter MF, Jacobson KA (1994) Chronic administration of selective adenosine A1 receptor agonist or antagonist in cerebral ischemia. *Eur J Pharmacol* 256:161–167
- Wang J, Wang C-E, Orr A, Tydlacka S, Li S-H, Li X-J (2008) Impaired ubiquitin-proteasome system activity in the synapses of Huntington's disease mice. *J Cell Biol* 180:1177–1189
- Washington CB, Jackson EK, Janesko KL, Vagni VA, Lefferis Z, Jenkins LW, Clark RS, Dixon CE, Kochanek PM (2005) Chronic caffeine administration reduces hippocampal neuronal cell death after experimental traumatic brain injury in mice. *J Neurotrauma* 22:366–370
- Wiese S, Jablonka S, Holtmann B, Orel N, Rajagopal R, Chao MV, Sendtner M (2007) Adenosine receptor A2A-R contributes to motoneuron survival by transactivating the tyrosine kinase receptor TrkB. *Proc Natl Acad Sci USA* 104:17210–17215
- Winkelmayer WC, Stampfer MJ, Willett WC, Curhan GC (2005) Habitual caffeine intake and the risk of hypertension in women. *JAMA* 294:2330–2335
- Winn HR, Morii S, Berne RM (1985) The role of adenosine in autoregulation of cerebral blood flow. *Ann Biomed Eng* 13:321–328
- Wirkner K, Assmann H, Koles L, Gerevich Z, Franke H, Norenberg W, Boehm R, Illes P (2000) Inhibition by adenosine A(2A) receptors of NMDA but not AMPA currents in rat neostriatal neurons. *Br J Pharmacol* 130:259–269
- Wyttenbach A, Swartz J, Kita H, Thykjaer T, Carmichael J, Bradley J, Brown R, Maxwell M, Schapira A, Orntoft TF, Kato K, Rubinsztein DC (2001) Polyglutamine expansions cause

- decreased CRE-mediated transcription and early gene expression changes prior to cell death in an inducible cell model of Huntington's disease. *Hum Mol Genet* 10:1829–1845
- Xiao D, Bastia E, Xu YH, Benn CL, Cha JH, Peterson TS, Chen JF, Schwarzschild MA (2006) Forebrain adenosine A2A receptors contribute to L-3,4-dihydroxyphenylalanine-induced dyskinesia in hemiparkinsonian mice. *J Neurosci* 26:13548–13555
- Xu K, Xu YH, Chen JF, Schwarzschild MA (2002) Caffeine's neuroprotection against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine toxicity shows no tolerance to chronic caffeine administration in mice. *Neurosci Lett* 322:13–16
- Yamamoto M, Schapira AH (2008) Dopamine agonists in Parkinson's disease. *Expert Rev Neurother* 8:671–677
- Yang JN, Chen JF, Fredholm BB (2009a) Physiological roles of A1 and A2A adenosine receptors in regulating heart rate, body temperature, and locomotion as revealed using knockout mice and caffeine. *Am J Physiol* 296:H1141–H1149
- Yang JN, Bjorklund O, Lindstrom-Tornqvist K, Lindgren E, Eriksson TM, Kahlstrom J, Chen JF, Schwarzschild MA, Tobler I, Fredholm BB (2009b) Mice heterozygous for both A1 and A(2A) adenosine receptor genes show similarities to mice given long-term caffeine. *J Appl Physiol* 106:631–639
- Yu L, Huang Z, Mariani J, Wang Y, Moskowitz M, Chen JF (2004) Selective inactivation or reconstitution of adenosine A2A receptors in bone marrow cells reveals their significant contribution to the development of ischemic brain injury. *Nat Med* 10:1081–1087
- Yu L, Coelho JE, Zhang X, Fu Y, Tillman A, Karaoz U, Fredholm BB, Weng Z, Chen JF (2009) Uncovering multiple molecular targets for caffeine using a drug target validation strategy combining A2A receptor knockout mice with microarray profiling. *Physiol Genomics* 37:199–210
- Yu L, Shen HY, Coelho JE, Araujo IM, Huang QY, Day YJ, Rebola N, Canas PM, Rapp EK, Ferrara J, Taylor D, Muller CE, Linden J, Cunha RA, Chen JF (2008) Adenosine A2A receptor antagonists exert motor and neuroprotective effects by distinct cellular mechanisms. *Ann Neurol* 63:338–346
- Zahniser NR, Simosky JK, Mayfield RD, Negri CA, Hanania T, Larson GA, Kelly MA, Grandy DK, Rubinstein M, Low MJ, Fredholm BB (2000) Functional uncoupling of adenosine A(2A) receptors and reduced response to caffeine in mice lacking dopamine D2 receptors. *J Neurosci* 20:5949–5957
- Zhou H, Cao F, Wang Z, Yu Z-X, Nguyen H-P, Evans J, Li S-H, Li X-J (2003) Huntingtin forms toxic NH2-terminal fragment complexes that are promoted by the age-dependent decrease in proteasome activity. *J Cell Biol* 163:109–118
- Zhou JG, Meno JR, Hsu SS, Winn HR (1994) Effects of theophylline and cyclohexyladenosine on brain injury following normo- and hyperglycemic ischemia: a histopathologic study in the rat. *J Cereb Blood Flow Metab* 14:166–173
- Zipser BD, Johanson CE, Gonzalez L, Berzin TM, Tavares R, Hulette CM, Vitek MP, Hovanesian V, Stopa EG (2007) Microvascular injury and blood-brain barrier leakage in Alzheimer's disease. *Neurobiol Aging* 28:977–986

# Methylxanthines and Pain

Jana Sawynok

## Contents

1	Introduction .....	312
2	Preclinical Studies with Methylxanthines .....	313
2.1	Intrinsic Antinociception by Caffeine .....	313
2.2	Caffeine and NSAIDs/Acetaminophen .....	316
2.3	Methylxanthines and Morphine .....	318
2.4	Methylxanthines and Other Analgesics .....	321
3	Clinical Studies with Caffeine .....	323
4	Summary and Conclusions .....	324
	References .....	325

**Abstract** Caffeine, an antagonist of adenosine A<sub>1</sub>, A<sub>2A</sub> and A<sub>2B</sub> receptors, is known as an adjuvant analgesic in combination with non-steroidal anti-inflammatory drugs (NSAIDs) and acetaminophen in humans. In preclinical studies, caffeine produces intrinsic antinociceptive effects in several rodent models, and augments the actions of NSAIDs and acetaminophen. Antagonism of adenosine A<sub>2A</sub> and A<sub>2B</sub> receptors, as well as inhibition of cyclooxygenase activity at some sites, may explain intrinsic antinociceptive and adjuvant actions. When combined with morphine, caffeine can augment, inhibit or have no effect depending on the dose, route of administration, nociceptive test and species; inhibition reflects spinal inhibition of adenosine A<sub>1</sub> receptors, while augmentation may reflect the intrinsic effects noted above. Low doses of caffeine given systemically inhibit antinociception by several analgesics (acetaminophen, amitriptyline, oxcarbazepine, cizolirtine), probably reflecting block of a component of action involving adenosine A<sub>1</sub> receptors. Clinical studies have demonstrated adjuvant analgesia, as well as some intrinsic analgesia, in the treatment of headache conditions, but not in the treatment of

---

J. Sawynok

Department of Pharmacology, Dalhousie University, Halifax, NS B3H 1X5, Canada  
e-mail: sawynok@dal.ca

postoperative pain. Caffeine clearly exhibits complex effects on pain transmission; knowledge of such effects is important for understanding adjuvant analgesia as well as considering situations in which dietary caffeine intake may have an impact on analgesic regimens.

**Keywords** Acetaminophen · Amitriptyline · Aspirin · Caffeine · Morphine · Non-steroidal anti-inflammatory drugs

## 1 Introduction

Caffeine has been a component of analgesic formulations containing aspirin (acetylsalicylic acid) for some time. Clinical studies published in the 1960s and 1970s indicated that caffeine-containing analgesics produced effects that were similar to those of the analgesics alone (Zhang 2001). However, in the mid 1980s, an analysis of 30 unpublished clinical studies over 20 years derived a relative potency of 1.4 for analgesics (aspirin, acetaminophen) containing caffeine compared with the analgesic alone (Laska et al. 1984). This analysis established caffeine's reputation as an adjuvant analgesic, an agent that augments the action of a known analgesic.

During the 1960s and early 1970s, with the discovery of cyclic AMP as a second messenger system and the appreciation that methylxanthines inhibit phosphodiesterase enzymes, the actions of caffeine were considered within the context of this system. In the late 1970s, the actions of caffeine and other methylxanthines as adenosine receptor antagonists came to be appreciated (Fredholm 1980), and, along with the recognition that adenosine receptors are involved in pain regulation, this added a further dimension to the interpretation of experimental observations with these agents. This shift in focus is clearly illustrated by studies that examined the effects of methylxanthines on morphine analgesia, tolerance and dependence over this interval (Ho et al. 1973; Ahljianian and Takemori 1985). When methods for spinal delivery of drugs were introduced in 1976, this allowed for the spinal pharmacology of pain transmission and its regulation by methylxanthine-sensitive adenosine receptors to be elaborated. Spinal administration of methylxanthines consistently inhibited spinal analgesia by morphine (Jurna 1984; DeLander and Hopkins 1986; Sweeney et al. 1987a), and this led to the adenosine hypothesis of opiate action within the spinal cord (Sawynok et al. 1989).

The main established targets for caffeine at concentrations normally achieved by human consumption are as an antagonist of adenosine  $A_1$ ,  $A_{2A}$  and  $A_{2B}$  receptors (Fredholm et al. 1999). The adjuvant and intrinsic analgesic properties of caffeine are largely considered in the context of such actions (Sawynok and Yaksh 1993). Other pharmacological actions (inhibition of phosphodiesterase,  $Ca^{2+}$  release, block of  $GABA_A$  receptors) are unlikely to manifest themselves in humans through any form of normal dietary use, but such actions potentially contribute in cases of unusually high doses of caffeine (Fredholm et al. 1999). Adenosine is known to be

involved in several aspects of pain regulation, with actions depending on the particular receptor subtype ( $A_1$ ,  $A_{2A}$ ,  $A_{2B}$ ,  $A_3$ ) and on the site of action (periphery, spinal cord, supraspinal sites, with systemic effects reflecting actions at several of these sites) (Sawynok 2006). Interest in exploring the adenosine system has been reflected in the potential development of novel analgesic agents (receptor agonists, antagonists or regulators, also indirectly acting agents such as inhibitors of adenosine kinase), as well as potential adjuvant agents that interact with other analgesics. Furthermore, because of the widespread dietary use of caffeine, it is important to know the potential impact that caffeine may have on a range of therapeutic agents, even procedures, used in pain management.

This chapter addresses several aspects of the effect of methylxanthines on pain. Initially, intrinsic antinociceptive properties of caffeine in a broad range of preclinical models of pain are considered. Subsequently, interactions with several different classes of analgesics (non-steroidal anti-inflammatory drugs, or NSAIDs, acetaminophen, opioids, antidepressants, anticonvulsants) in preclinical studies are reviewed. Finally, the actions of caffeine as an adjuvant analgesic in humans are addressed.

## 2 Preclinical Studies with Methylxanthines

### 2.1 *Intrinsic Antinociception by Caffeine*

There are several reports of intrinsic antinociceptive effects of systemically administered (by intraperitoneal injection) caffeine in the preclinical literature. The effect of caffeine depends on the nature of the test, on the dose of caffeine and on the species tested. Several generalities emerge.

#### 2.1.1 Caffeine Is Often Inactive at Doses up to $50 \text{ mg kg}^{-1}$

There are several reports that caffeine has no intrinsic effects in several different nociceptive tests in which a range of doses of caffeine have been evaluated. These include (1) tail immersion test, rats ( $25, 50 \text{ mg kg}^{-1}$ ) (Malec and Michalska 1988); (2) hot plate test, rats ( $25, 50 \text{ mg kg}^{-1}$ ) and mice ( $10, 50 \text{ mg kg}^{-1}$ ) (Malec and Michalska 1988); (3) hot plate test, mice ( $20, 40 \text{ mg kg}^{-1}$ ) (Engelhardt et al. 1997); (4) writhing test, mice ( $5\text{--}50 \text{ mg kg}^{-1}$ ) (Fialip et al. 1989; Gayawali et al. 1991); (5) pain-induced functional impairment test ( $10\text{--}56 \text{ mg kg}^{-1}$ ) (Granados-Soto et al. 1993; Flores-Acevedo et al. 1995; Díaz-Reval et al. 2001; López et al. 2006); (6) spinal nerve ligation, thermal hyperalgesia, rats ( $1.5\text{--}7.5 \text{ mg kg}^{-1}$ ) (Esser and Sawynok 2000); and (7) partial sciatic nerve ligation, mechanical allodynia, rats ( $10, 20 \text{ mg kg}^{-1}$ ) (Wu et al. 2006). Several works, in which the main goal was to examine the interaction of caffeine with another agent, have also evaluated effects of single doses of caffeine within this range and reported no effect for caffeine using these tests (noted in subsequent sections).

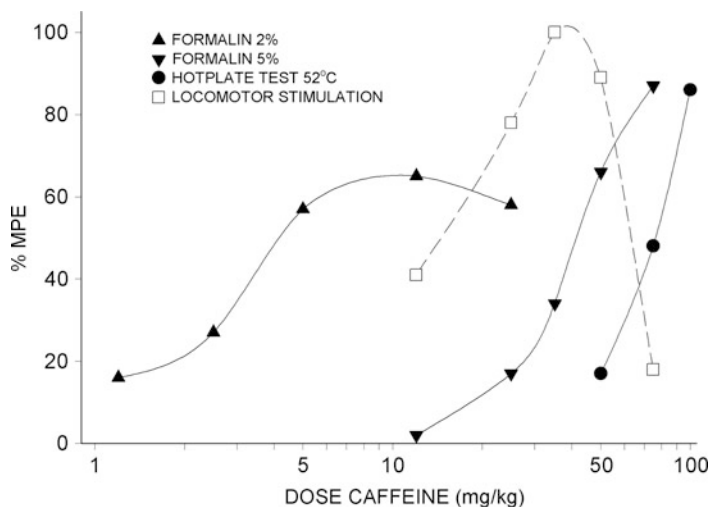
Curiously, some studies noted hyperalgesia with systemic doses of caffeine in this range. Thus, caffeine led to hyperalgesia in the tail immersion test in mice at 10 mg kg<sup>-1</sup> (Godfrey et al. 2006). The end point in this test is largely a spinal reflex (it is intact in spinalized animals), and hyperalgesia may reflect caffeine acting spinally to inhibit tonic activity at adenosine A<sub>1</sub> receptors. In rats, spinal (intrathecal) administration of the methylxanthines aminophylline (Jurna 1984), theophylline and 8-phenyltheophylline produced hyperalgesia in the tail flick test, particularly at a lower intensity of stimulation (Sawynok et al. 1986). In mice, intrathecal administration of caffeine and of theophylline were noted not to alter tail flick latencies, but no data were shown (Delander and Hopkins 1986).

### 2.1.2 Caffeine Produces Antinociception Generally at Doses of 35–100 mg kg<sup>-1</sup>

There have been reports of intrinsic antinociceptive properties for caffeine at doses generally between 35 and 100 mg kg<sup>-1</sup> in studies where several doses of caffeine were examined. These include (1) inflammatory hyperalgesia models, rats (50, 100 mg kg<sup>-1</sup>) (Sieggers 1973; Seegers et al. 1981); (2) tail compression test, rats (30, 100 mg kg<sup>-1</sup>) (Person et al. 1985); (3) hot plate test, mice (75, 100 mg kg<sup>-1</sup>) (Malec and Michalska 1988; Abo-Salem et al. 2004); (4) hot plate test, rats (50–100 mg kg<sup>-1</sup>) (Sawynok et al. 1995); (5) 5% formalin test, rats (35–75 mg kg<sup>-1</sup>) (Sawynok et al. 1995); and (6) partial sciatic nerve ligation, mechanical allodynia, rats (40, 80 mg kg<sup>-1</sup>) (Wu et al. 2006).

A few studies have not observed intrinsic antinociceptive actions at higher doses of caffeine. For example, Fialip et al. (1989) saw no effect of caffeine at several doses (40, 80, 100, 160, 200 mg kg<sup>-1</sup>) in the hot plate test; however, that study used a higher intensity of stimulation (56°C) than in many other studies. On occasion, studies that examined the effects of only a single high dose of caffeine also saw no effect (e.g. 50 mg kg<sup>-1</sup>, mouse hot plate test, Oliverio et al. 1983; 100 mg kg<sup>-1</sup>, rat tail flick test, Misra et al. 1985).

It is important to note that several studies have reported antinociceptive effects with low doses of caffeine. Antinociception occurs with 2.5–10 mg kg<sup>-1</sup> in the 2% formalin test in rats (Sawynok and Reid 1996a), with 1–5 mg kg<sup>-1</sup> in the hot plate and writhing tests in mice and in the paw pressure and tail flick tests in rats (Ghelardini et al. 1997), and with 10 mg kg<sup>-1</sup> in the hot plate test in mice (Abo-Salem et al. 2004). Stimulus intensity can be important in revealing antinociception with caffeine, as the ED<sub>40</sub> at 2% formalin is 10 times lower than at 5% formalin (Sawynok and Reid 1996a). There may also be species differences between mice and rats. For example, 10 mg kg<sup>-1</sup> caffeine produces antinociception in the 2% formalin test in rats (Sawynok and Reid 1996a) but not in mice (Sawynok et al. 2008). A further factor to note is that there may well be strain differences in expression of analgesia in mouse studies (Wilson and Mogil 2001); this is known to be important in general studies on analgesics, but no systematic studies are available for caffeine.



**Fig. 1** Antinociception by caffeine in several nociceptive tests in rats from a single laboratory. Shown are (1) antinociception from 35 to 100 mg kg<sup>-1</sup> in several tests, (2) the intensity dependence of antinociception in the same test (2 vs. 5% formalin) and (3) dissociation of antinociception from locomotor stimulation. (Data redrawn from Sawynok et al. 1995 and Sawynok and Reid 1996a)

Figure 1 is derived from data from a single laboratory using a single species and strain, and illustrates several aspects of the antinociceptive profile of caffeine. It shows (1) antinociceptive actions of caffeine at doses of 35–100 mg kg<sup>-1</sup> in multiple nociceptive tests (thermal threshold test and a tonic chemogenic model of pain); (2) the dependence of antinociception on stimulus intensity (2 vs. 5% formalin); and (3) dissociation of antinociception from motor effects. With respect to the latter issue, locomotor stimulation can be a concern with respect to confounding end points commonly interpreted as antinociception. However, in rats, locomotor stimulation is prominent up to 30–40 mg kg<sup>-1</sup>, with higher doses of 75–100 mg kg<sup>-1</sup> showing locomotor depression (Fredholm et al. 1999). Given that antinociception manifests itself in both motor stimulation and depression dose ranges, this suggests antinociception by caffeine is independent of motor effects. Locomotor stimulation by caffeine involves antagonism of adenosine A<sub>2A</sub> receptors (Ledent et al. 1997).

### 2.1.3 Mechanisms of Antinociception

Adenosine A<sub>1</sub>, A<sub>2A</sub> and A<sub>2B</sub> receptors are involved in antinociception, with the effects depending on the receptor subtype and the site of action (Sawynok 2006). Caffeine has a similar affinity as an antagonist of each of these receptors (Fredholm et al. 1999). Adenosine A<sub>1</sub> receptors at peripheral, spinal and supraspinal sites mediate antinociceptive actions, so the ability of caffeine to block these receptors cannot account for antinociception by caffeine. Adenosine A<sub>2A</sub> and A<sub>2B</sub> receptors

on peripheral nerve terminals and other peripheral sites are recognized to play a pronociceptive role; while central actions are less well characterized, they also may be involved in pain facilitation. The selective adenosine  $A_{2A}$  receptor antagonist SCH58261 produces antinociception in several models (mouse writhing, hot plate, formalin tests) (Bastia et al. 2002; Godfrey et al. 2006; Hussey et al. 2007), while adenosine  $A_{2A}$  receptor knockout mice exhibit decreased pain responses (hot plate, tail flick, formalin tests) (Ledent et al. 1997; Hussey et al. 2007). In another study, the effects of PSB1115, a selective  $A_{2B}$  receptor antagonist, together with a series of other selective antagonists, produced antinociception (mouse hot plate test) mimicking both the antinociceptive action of caffeine and the augmentation of morphine (Abo-Salem et al. 2004). Both intrinsic and adjuvant effects of caffeine were suggested to result from adenosine  $A_{2B}$  receptor blockade (Abo-Salem et al. 2004).

Antinociception by higher doses of caffeine, where antinociception is observed more reproducibly, involves central noradrenergic mechanisms. Antinociception is potentiated by phentolamine ( $\alpha$ -adrenergic receptor antagonist) in both the hot plate test and the formalin test, while depletion of spinal cord noradrenaline levels markedly reduced caffeine antinociception in the hot plate test (Sawynok et al. 1995). Depletion of central noradrenaline levels augmented antinociception by caffeine in the formalin test, but depletion of central 5-hydroxytryptamine levels exhibited no effect (Sawynok and Reid 1996b). Caffeine is known to alter the turnover of biogenic amines in several regions of the central nervous system (Nehlig et al. 1992), and these results indicate that central amine systems are important for antinociception by caffeine at medium (25–50 mg kg<sup>-1</sup>, formalin test) and high (75–100 mg kg<sup>-1</sup>, hot plate test) doses. However, there are some differences in effects between the two tests (e.g. depletion of spinal noradrenaline levels inhibits antinociception in one but not the other test), and a range of actions may be recruited.

There is one study that has demonstrated antinociception with low doses of caffeine (1–5 mg kg<sup>-1</sup>) in several models in mice and rats (Ghelardini et al. 1997). This low-dose antinociception in the hot plate and writhing tests in mice involved central cholinergic mechanisms, because antinociception was blocked by muscarinic receptor antagonists as well as depletion of central cholinergic levels with hemicholinium-3 (Ghelardini et al. 1997). Such antinociception did not involve opioid or GABA<sub>B</sub> receptors, or central amines (Ghelardini et al. 1997). The latter observation distinguishes low-dose caffeine antinociception (5 mg kg<sup>-1</sup>) from that seen at higher doses (35–100 mg kg<sup>-1</sup>), as central amine mechanisms are not involved in the former, but are involved in the latter (Sawynok et al. 1995, Sawynok and Reid 1996b).

## 2.2 Caffeine and NSAIDs/Acetaminophen

The effects of caffeine on antinociception by aspirin and acetaminophen have been examined for some time in order to explore the rationale for the historical combination of caffeine with these agents in analgesic formulations. Several earlier



preclinical studies demonstrated that caffeine augmented antinociception by both aspirin and acetaminophen in inflammatory hyperalgesia models in rats (Vinegar et al. 1976; Seegers et al. 1981; but see Engelhardt et al. 1997) and in the writhing model in mice (Gayawali et al. 1991). The doses of caffeine used in these studies were generally 10–50 mg kg<sup>-1</sup>. In one study that extended the dose to 100 mg kg<sup>-1</sup>, augmentation of antinociception was no longer observed at the highest dose (Gayawali et al. 1991).

The effects of caffeine on acetaminophen and several NSAIDs have been explored in a series of studies using the pain-induced functional impairment model in rats. In this model, uric acid is injected into the knee joint and walking activity on a revolving cylinder is examined; the test is sensitive to both NSAIDs and opioids. Caffeine, at 10–56 mg kg<sup>-1</sup>, was inactive alone in this model (Granados-Soto et al. 1993). Sixteen acetaminophen–caffeine dose combinations were examined (acetaminophen 100, 178, 316, 562 mg kg<sup>-1</sup> with caffeine 10, 18, 32, 56 mg kg<sup>-1</sup>) and augmentation of the action of acetaminophen was observed only at certain doses. The 316 mg kg<sup>-1</sup> dose of acetaminophen was augmented by all doses of caffeine, with a maximal effect at 32 mg kg<sup>-1</sup>. The maximally effective dose of caffeine did not change the plasma levels or kinetics of acetaminophen. The selective augmentation of a mid-range dose of acetaminophen by caffeine was explained by considering a three-zone model of action for a sigmoidal function whereby only the mid zone exhibits sensitivity (Granados-Soto and Castenada-Hernández 1999). This analysis emphasized the need for examining several dose combinations in order to more fully explore the potential for significant drug interactions.

In a subsequent series of studies using the pain-induced functional impairment model, caffeine was also shown to augment the action of aspirin (Castenada-Hernández et al. 1994), tolmetin (Flores-Acevedo et al. 1995), ketolorac (López-Munoz et al. 1996; Aguirre-Banuelos et al. 1999), ibuprofen (López et al. 2006) and ketoprofen (Díaz-Reval et al. 2001). The doses of caffeine that were most effective were 32 mg kg<sup>-1</sup> (aspirin), 32–56 mg kg<sup>-1</sup> (tolmetin), 18–32 mg kg<sup>-1</sup> (ketolorac), 10–18 mg kg<sup>-1</sup> (ketoprofen) and 18–32 mg kg<sup>-1</sup> (ibuprofen). Studies that examined the widest range of dose combinations (Díaz-Reval et al. 2001; López et al. 2006) also showed that only certain dose combinations showed augmented activity. These observations support the zone model previously proposed (Granados-Soto and Castenada-Hernández 1999).

With regard to the mechanism by which caffeine augments antinociception by acetaminophen and other NSAIDs, several studies determined the effect of caffeine on plasma levels and kinetics, and observed no effect on these parameters for acetaminophen (Granados-Soto et al. 1993), aspirin (Castenada-Hernández et al. 1994) or tolmetin (Flores-Acevedo et al. 1995). Functional changes, therefore, represent pharmacodynamic rather than pharmacokinetic interactions. Caffeine had no effect on cyclooxygenase (COX) activity alone and did not affect inhibition of COX activity in the brain by acetaminophen or aspirin (Engelhardt et al. 1997). However, when examined for activity in rat primary microglial cells, caffeine and acetaminophen both inhibited microglial prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) synthesis alone,

and augmented the ability of aspirin to inhibit lipopolysaccharide-induced PGE<sub>2</sub> synthesis (Fiebich et al. 2000). Caffeine also was shown to inhibit COX-2 protein synthesis (Fiebich et al. 2000). These actions of caffeine may result from inhibition of adenosine A<sub>2A</sub> receptors on microglia, as these receptors cause upregulation of the COX-2 gene and release of PGE<sub>2</sub> in microglia (Fiebich et al. 1996). It was proposed that inhibition of COX in microglial cells contributes to the adjuvant analgesic activity of caffeine (Fiebich et al. 2000). Additional mechanisms considered to be involved in the intrinsic antinociceptive properties of caffeine, including block of adenosine A<sub>2B</sub> receptors and recruitment of central noradrenergic mechanisms (Sect. 2.1), also may be involved in adjuvant analgesia by caffeine.

Some studies have noted that low doses of caffeine can inhibit antinociception by acetaminophen. Siegers (1973) reported that 10 mg kg<sup>-1</sup> caffeine inhibited antinociception by acetaminophen in an inflammatory hyperalgesia model in rats. Granados-Soto et al. (1993) observed that 10 mg kg<sup>-1</sup> caffeine appeared to inhibit the action of acetaminophen at doses of 100 and 562 mg kg<sup>-1</sup> in the pain-induced functional impairment model in rats, and while this was not significantly different, the observation was noteworthy enough to merit mention in a subsequent analysis of the same data (Granados-Soto and Castenada-Hernández 1999). More recently, Godfrey et al. (2006) reported that 10 mg kg<sup>-1</sup> caffeine inhibited the antinociceptive action of acetaminophen in the tail immersion and hot plate tests in mice; this effect was not mimicked by a selective adenosine A<sub>2A</sub> (SCH58261) or A<sub>2B</sub> (PSB1115) receptor antagonist. Curiously, this inhibitory effect of caffeine on acetaminophen mimics inhibitory effects reported with several different classes of analgesics; in those cases the inhibitory effect of caffeine is attributed to blockade of central adenosine A<sub>1</sub> receptors (Sect. 2.4).

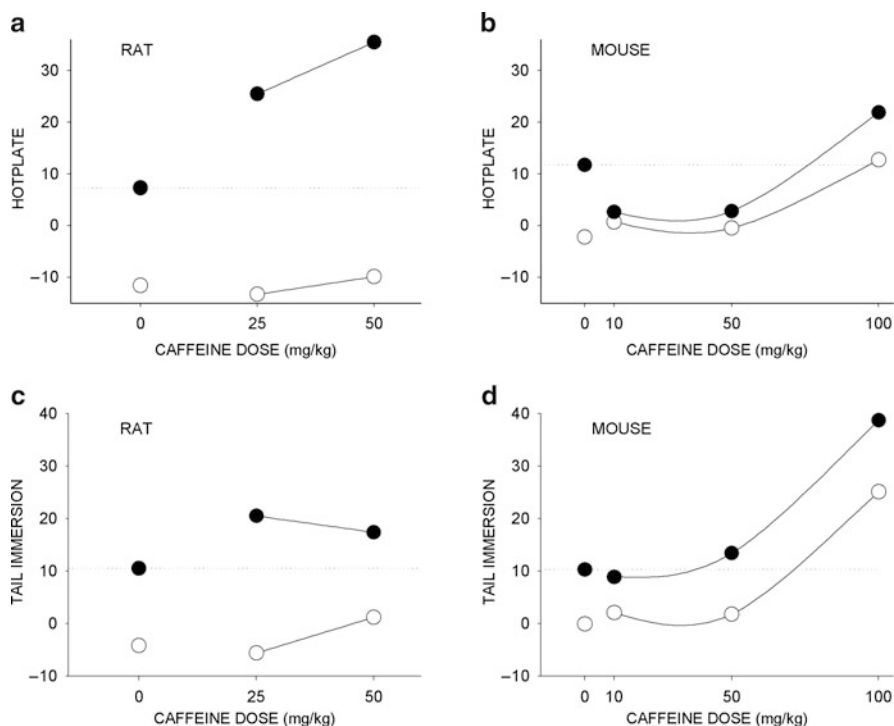
### 2.3 *Methylxanthines and Morphine*

Methylxanthine interactions with morphine need to be considered in two distinct contexts, that of systemic interactions, whereby drugs are distributed to several compartments and the behavioural effect reflects the integrated action of several systems, and local interactions, whereby drug actions represent an interaction within a specific compartment (spinal, supraspinal, peripheral). Systemic interactions were originally explored to determine aspects of the mechanism of action of morphine, but they are also relevant to interactions that might occur owing to dietary intake of caffeine when opioids are given, or to drug combinations in which caffeine is added to an analgesic formulation consisting of a combination of codeine and an NSAID or acetaminophen.

#### 2.3.1 Systemic Interactions

During the 1980s, caffeine was reported to potentiate antinociception by morphine (100 mg kg<sup>-1</sup>, tail flick test, rats, Misra et al. 1985; 30 mg kg<sup>-1</sup>, tail pressure test,

rats, Person et al. 1985; 75, 100 mg kg<sup>-1</sup>, hot plate test, mice, Malec and Michalska 1988), not to influence morphine (50 mg kg<sup>-1</sup>, hot plate test, mice, Oliverio et al. 1983) or to antagonize morphine (40 mg kg<sup>-1</sup>, tail flick test, mice, Ahlijanian and Takemori 1985; 25–50 mg kg<sup>-1</sup>, tail flick and hot plate tests, mice, Malec and Michalska 1988). The Malec and Michalska (1988) study seemed to resolve disparate observations by demonstrating dose dependence and species dependence of interactions within the same study – in mice, both inhibition (low dose) and augmentation (high dose) of morphine by caffeine were observed, while low doses of caffeine augmented the action of morphine in rats. Figure 2 presents a compilation of data from that study, illustrating the variety of effects that can occur with a single drug combination (caffeine – morphine). Similar biphasic effects also were seen with theophylline and morphine (Malec and Michalska 1988). More recently, 5 mg kg<sup>-1</sup> caffeine was shown to augment the action of morphine in the hot plate



**Fig. 2** Biphasic effects of caffeine on antinociception by morphine in (a, c) rats and (b, d) mice in two tests for nociception. Data points represent cumulative differences from the baseline; *hollow symbols* represent caffeine alone and *solid symbols* represent caffeine combined with 5 mg kg<sup>-1</sup> morphine; *the dashed line* represents antinociception by morphine alone. In mice, there is (1) inhibition of morphine antinociception by low doses of caffeine, (2) augmentation of such antinociception by high doses of caffeine and (3) a lack of effect of intermediate doses of caffeine. In rats, low doses of caffeine augment the action of morphine, illustrating a species difference between mice and rats. (Data redrawn from Malec and Michalska 1988)

test in mice (Abo-Salem et al. 2004). In that study, both intrinsic antinociception seen with caffeine (10, 75 mg kg<sup>-1</sup>; Sect. 2.1) and augmentation of the action of morphine (5 mg kg<sup>-1</sup>) were mimicked by several selective adenosine A<sub>2B</sub> receptor antagonists, and it was proposed that both actions of caffeine reflected adenosine A<sub>2B</sub> receptor blockade.

### 2.3.2 Spinal Interactions

In 1981, it was shown that intraperitoneal administration of aminophylline inhibited the antinociceptive action of intraperitoneally administered morphine in the tail flick test in both intact and spinalized rats, which clearly implicated the spinal cord as the locus of the interaction (Jurna 1981). Subsequent studies showed that intrathecal administration of aminophylline, theophylline (non-selective adenosine receptor antagonists) or 8-phenyltheophylline (selective adenosine A<sub>1</sub> receptor antagonist) inhibited the action of intrathecally administered morphine in the tail flick and hot plate tests in rats (Jurna 1984; Sweeney et al. 1987a) and in the tail flick test in mice (DeLander and Hopkins 1986). When morphine was shown to increase the release of endogenous adenosine from *in vitro* and *in vivo* spinal cord preparations (Sweeney et al. 1987a, b), the methylxanthine blockade was understood to reflect block of spinal adenosine A<sub>1</sub> receptors activated by increased endogenous levels of adenosine. The characteristics and mechanisms of opioid-induced release of adenosine have been reviewed comprehensively (Sawynok and Liu 2003). A recent study has demonstrated that antinociception following intrathecal administration of morphine was reduced in adenosine A<sub>1</sub> receptor knockout mice, confirming involvement of spinal adenosine and A<sub>1</sub> receptors in the local action of morphine (Wu et al. 2005). The adenosine component of spinal opioid action, via adenosine A<sub>1</sub> receptors, also manifests itself prominently in nerve injury models of pain (Zhang et al. 2005). Finally, it is interesting to note that intrathecal, but not intravenous, opioids have been demonstrated to lead to spinal release of adenosine in humans (Eisenach et al. 2004).

### 2.3.3 Supraspinally Administered Morphine, Spinally Administered Methylxanthines

When morphine is given supraspinally by intracerebroventricular injection, antinociception is blocked by intrathecal administration of caffeine and theophylline in mice (DeLander and Hopkins 1986) and rats (Sweeney et al. 1991). In rats, supraspinally administered morphine results in spinal release of adenosine which is secondary to activation of descending serotonergic pathways; the adenosine that is released subsequently activates spinal adenosine A<sub>1</sub> receptors (Sweeney et al. 1991). In mice, spinal adenosine A<sub>2</sub> receptors appear to contribute to antinociception by supraspinally administered morphine (Suh et al. 1997).

### 2.3.4 Supraspinally Administered Morphine, Supraspinally Administered Methylxanthines

When opioids and caffeine were both given supraspinally, caffeine (inactive alone) inhibited antinociception by a  $\mu$ -opioid and by a  $\delta$ -opioid receptor agonist, but not by a  $\kappa$ -opioid receptor agonist (tail pinch and hot plate tests, mice) (Pham et al. 2003). The  $\mu$ -opioid and  $\delta$ -opioid receptor agonists inhibited adenosine uptake into synaptosomes, and the resultant increase in extracellular adenosine levels at supraspinal sites, with subsequent activation of adenosine receptors, was proposed to account for the methylxanthine effect (Pham et al. 2003).

## 2.4 Methylxanthines and Other Analgesics

### 2.4.1 Antidepressants

Tricyclic antidepressants are widely used in the treatment of neuropathic pain (Dworkin et al. 2007). Both caffeine and theophylline at 50 mg kg<sup>-1</sup>, doses which were inactive alone, inhibited antinociception produced by systemic administration of the antidepressants amitriptyline and imipramine (tail flick test, mice) (Pareek et al. 1994). Aminophylline (10 mg kg<sup>-1</sup>), also inactive alone, inhibited antinociception by the antidepressants clomipramine, maprotiline, imipramine and zimelidine (writhing test, mice) (Sierralta et al. 1995). Caffeine, generally between 5 and 10 mg kg<sup>-1</sup>, inhibits antinociception by amitriptyline in several further models of pain – in the spinal nerve ligation model (thermal hyperalgesia, rats) (Esser and Sawynok 2000), in the streptozotocin-induced diabetic model (mechanical allodynia, rats) (Ulugol et al. 2002) and in the formalin model (2% formalin, mice) (Sawynok et al. 2008). Caffeine (5 mg kg<sup>-1</sup>) also inhibits the action of a newer antidepressant, venlafaxine (hot plate test, mice) (Yaba et al. 2006). Collectively, these studies provide a coherent body of information indicating block of antidepressant actions by low doses of caffeine.

In all of these studies, antagonism by caffeine was considered to indicate an involvement of endogenous adenosine and adenosine A<sub>1</sub> receptors in the actions of antidepressants. A recent study provides direct support for this concept. While 10 mg kg<sup>-1</sup> caffeine clearly blocked the ability of amitriptyline to produce antinociception in wild-type mice, this action was no longer observed in adenosine A<sub>1</sub> receptor knockout mice (Sawynok et al. 2008). In the latter situation, amitriptyline was still able to produce antinociception, and this was necessary in order to observe the differential effect of caffeine. Antidepressants are complex agents with several pharmacological actions, many of which contribute to antinociception (Micó et al. 2006), and this multiplicity of action may be what allows for intrinsic effects of amitriptyline to occur in A<sub>1</sub> receptor knockout mice.

Antidepressants also produce antinociception following local peripheral administration to the rodent hind paw, and this action is inhibited by local peripheral

administration of caffeine. Local administration of caffeine inhibits antinociception produced by peripherally administered amitriptyline in the formalin test in rats (as does 8-phenyltheophylline) (Sawynok et al. 1999), in the spinal nerve ligation model in rats (thermal hyperalgesia) (Esser and Sawynok 2000) and in the streptozotocin-induced diabetes model in rats (mechanical allodynia) (Ulugol et al. 2002). Caffeine was no longer able to inhibit the peripheral action of amitriptyline in adenosine A<sub>1</sub> receptor knockout mice (Sawynok et al. 2008). Peripherally administered amitriptyline has been shown to increase extracellular levels of adenosine using microdialysis, perhaps by inhibiting adenosine uptake, and this increased tissue availability of adenosine is believed to contribute to its actions (Sawynok et al. 2005). Such studies clearly implicate adenosine and A<sub>1</sub> receptors as one component of the peripheral action of amitriptyline.

#### 2.4.2 Carbamazepine/Oxcarbazepine

Carbamazepine is an anticonvulsant drug used to treat neuralgia and neuropathic pain, and oxcarbazepine is an analogue (10-keto derivative) better tolerated than the parent drug (Ambrósio et al. 2002). Caffeine (5 mg kg<sup>-1</sup>) inhibited antinociception by carbamazepine in stressed rats (tail flick test) (Mashimoto et al. 1998). Caffeine (5–20 mg kg<sup>-1</sup>; inactive alone) also inhibited the antinociceptive effects of both carbamazepine and oxcarbazepine administered systemically in an inflammatory hyperalgesia model in rats (Tomić et al. 2004). 1,3-Dipropyl-8-cyclopentylxanthine (DPCPX), a selective adenosine A<sub>1</sub> receptor antagonist, also inhibited the action of both drugs in a manner similar to caffeine (Tomić et al. 2004). Several reports indicate that carbamazepine and its derivatives can bind to adenosine A<sub>1</sub> and A<sub>2</sub> receptors, although it is not clear whether they function as agonists or antagonists (Ambrósio et al. 2002).

Peripheral administration of oxcarbazepine also produces antinociception in rats (inflammatory hyperalgesia), and this action is blocked by peripheral administration of caffeine (Tomić et al. 2006). Peripheral antagonism of oxcarbazepine by caffeine is mimicked by DPCPX, suggesting involvement of adenosine A<sub>1</sub> receptors in peripheral as well as systemic actions (Tomić et al. 2006). Agonist actions at adenosine A<sub>1</sub> receptors appear to contribute to peripheral, as well as systemic, actions of oxcarbazepine, although the nature of such involvement remains unclear.

#### 2.4.3 Cizolirtine

Cizolirtine is a novel analgesic drug with a wide profile of activity in several preclinical pain models (Alvarez et al. 2000). The mechanism by which this agent produces antinociception is unclear as it does not interact with several receptors or ion channels (Alvarez et al. 2000). Cizolirtine produces antinociception in a preclinical model of neuropathic pain (streptozotocin-induced diabetes, rats), and its actions are blocked by 5 mg kg<sup>-1</sup> caffeine (Aubel et al. 2007). Cizolirtine

does not bind to adenosine receptors, and it was suggested that this agent leads to increased extracellular levels of endogenous adenosine (Aubel et al. 2007).

#### 2.4.4 Summary of Analgesia-Modifying Properties of Caffeine

There are several general observations that can be made with respect to the ability of caffeine to modify antinociception by other analgesic agents. (1) *Low doses of caffeine (up to 10 mg kg<sup>-1</sup>) inhibit antinociception by several analgesics.* Such doses of caffeine inhibit antinociception by acetaminophen (Siegers 1973; Godfrey et al. 2006), amitriptyline (Esser and Sawynok 2000; Ulugol et al. 2002; Yaba et al. 2006, Sawynok et al. 2008), carbamazepine and oxcarbazepine (Tomić et al. 2004) and cizolirtine (Aubel et al. 2007). This antagonism may reflect block of adenosine A<sub>1</sub> receptors recruited by increased endogenous levels of adenosine. The ability of 5 mg kg<sup>-1</sup> caffeine to augment analgesia by morphine is proposed to be due to block of adenosine A<sub>2B</sub> receptors (Abo-Salem et al. 2004). (2) *Moderate doses of caffeine (10–35 mg kg<sup>-1</sup>) augment antinociception.* This is seen with acetaminophen (Gayawali et al. 1991; Granados-Soto et al. 1993) and several different NSAIDs (Gayawali et al. 1991; Castenada-Hernández et al. 1994; Flores-Acevedo et al. 1995; López-Munoz et al. 1996; Aguirre-Banuelos et al. 1999; Díaz-Reval et al. 2001; López et al. 2006). Augmentation is more readily observed within a medium-effect range (Granados-Soto and Castenada-Hernández 1999). Pharmacokinetic explanations for the positive interactions were excluded in several of these studies. Augmentation of antinociception may involve inhibition of COX in microglia (Fiebich et al. 2000). Mechanisms involving central amine systems may also be involved at moderate doses of caffeine (Sawynok et al. 1995). While some studies have reported potentiation of antinociception by morphine at such doses (Person et al. 1985), others have noted inhibition of antinociception by morphine (Malec and Michalska 1988; Ahlijanian and Takemori 1985); the latter interaction probably reflects actions at spinal sites. (3) *High doses of caffeine (75, 100 mg kg<sup>-1</sup>) often show intrinsic antinociceptive effects* (Sect. 2.1) and augment antinociception by morphine (Misra et al. 1985; Malec and Michalska 1988), perhaps simply reflecting this intrinsic activity. The effects of 50 mg kg<sup>-1</sup> caffeine can be variable, depending on the species and the test system, and may reflect biphasic effects by different doses of caffeine (Malec and Michalska 1988; Gayawali et al. 1991).

### 3 Clinical Studies with Caffeine

In the mid 1980s, an analysis of 30 trials indicated that caffeine (at least 65 mg) added to analgesia formulations containing aspirin and acetaminophen reduced the amount of analgesic needed by 40% (relative potency 1.4) (Laska et al. 1984). These adjuvant effects were confirmed in several subsequent trials (Schachtel et al. 1991; Migliardi et al. 1994; Lipton et al. 1998; Diener et al. 2005) and

extended to include combination with ibuprofen (Diamond et al. 2000). Further meta-analysis indicates that adding caffeine to analgesics increases the number of patients who experience pain relief from headache (rate ratio 1.36); it also leads to more patients with nervousness and dizziness (relative risk 1.60) (Zhang 2001). While there are some reports of adjuvant analgesic effects of caffeine in other pain conditions, meta-analysis shows no appreciable adjuvant effect with acetaminophen (Zhang and Po 1996), aspirin (Zhang and Po 1997) or ibuprofen (Po and Zhang 1998) for postoperative pain.

There are several issues to be considered with regard to caffeine and headaches. In some studies, caffeine also displayed intrinsic analgesic properties for treating tension-type headaches in doses of 130–200 mg caffeine (Ward et al. 1991; Diamond et al. 2000). Caffeine given orally at 300 mg relieved postdural puncture headache (Camann et al. 1990) but repeat oral doses of 75–125 mg caffeine (with acetaminophen) did not provide a prophylactic effect (Esmooglu et al. 2005); an intravenous dose of 500 mg caffeine, however, did provide significant pain relief from postdural puncture headaches (Yucel et al. 1999). With chronic administration, caffeine withdrawal can lead to headaches (onset within 24 h of interruption of daily caffeine intake), although there is variability in the dose/duration requirements (Shapiro 2008). Postoperative headaches may be a manifestation of caffeine withdrawal resulting from perisurgical fasting, and perisurgical administration of caffeine is useful for relieving postoperative headaches (Fennelly et al. 1991; Hampl et al. 1995; Weber et al. 1997). Vascular effects (vasoconstriction) are considered to account for some of the pain relief that occurs in headaches (Shapiro 2008). On the other hand, habitual caffeine consumption or analgesic overuse can also be associated with the development of headache, migraine and chronic daily headache, but there are geographic and cultural differences in the prevalence of this (Shapiro 2008). While consumption of caffeine in children may be less than the adult rate, a case study noted resolution of headaches in 33 of 36 children when caffeine-containing beverages were withdrawn (Hering-Hanit and Gadoth 2003).

## 4 Summary and Conclusions

Interest in methylxanthine effects on pain arose from the presence of caffeine in many analgesic formulations, particularly those containing acetaminophen and aspirin. Caffeine (and other methylxanthines) subsequently became an investigative tool to determine the potential involvement of adenosine receptors in the mechanism of action of other analgesics. In preclinical studies, caffeine has been shown to exhibit several effects on nociception, depending on dose, nociceptive test, stimulus intensity and species. At low doses (up to 10–12.5 mg kg<sup>-1</sup>) caffeine is usually inactive alone but inhibits the antinociceptive action of acetaminophen, amitriptyline, carbamazepine/oxcarbazepine and cizolirtine. A few studies have reported antinociception with caffeine at these low doses. At higher doses (approximately 15–45 mg kg<sup>-1</sup>), it produces intrinsic effects in some preclinical models of



antinociception, and augments antinociception by acetaminophen and NSAIDs even in tests where it lacks intrinsic effects. Doses in this range generally produce locomotor stimulation. At even higher doses (50–100 mg kg<sup>-1</sup>), caffeine leads to intrinsic antinociception in a wide range of tests, locomotor suppression and diverse effects on the action of morphine depending on the test and species. Mechanisms implicated in *antinociception and augmentation of antinociception* involve inhibition of central adenosine A<sub>2B</sub> (and possibly A<sub>2A</sub>) receptors, inhibition of COX in microglia, engagement of central cholinergic systems and involvement of central noradrenergic systems. The mechanisms implicated in *inhibition of antinociception* involve block of central adenosine A<sub>1</sub> receptors. Clinical studies with caffeine have indicated adjuvant analgesic actions in combination with acetaminophen and NSAIDs and intrinsic analgesia in several headache conditions. Efficacy in headache states may involve alterations in central blood flow. The consistent preclinical observation of blockade of antinociception by several different classes of analgesics may be relevant to dietary intake of caffeine in humans; there are, however, no clinical studies that have addressed this issue.

**Acknowledgement** I thank Allison Reid for editorial and graphical assistance.

## References

- Abo-Salem OM, Hayallah AM, Bilkei-Gorzo A et al (2004) Antinociceptive effects of novel A<sub>2B</sub> adenosine receptor antagonists. *J Pharmacol Exp Ther* 308:358–366
- Aguirre-Banuelos P, Castaneda-Hernández G, López-Munoz FJ et al (1999) Effect of coadministration of caffeine and either adenosine agonists or cyclic nucleotides on ketolorac analgesia. *Eur J Pharmacol* 377:175–182
- Ahlijanian MK, Takemori AE (1985) Effects of (-)-N<sup>6</sup>-(R-phenylisopropyl)-adenosine (PIA) and caffeine on nociception and morphine-induced analgesia, tolerance and dependence in mice. *Eur J Pharmacol* 112:171–179
- Alvarez I, Andreu F, Buxens J et al (2000) Pharmacology of cizolirtine: a new analgesic agent. *Methods Find Exp Clin Pharmacol* 22:211–221
- Ambrósio AF, Soares-da-Silva P, Carvalho CM et al (2002) Mechanisms of action of carbamazepine and its derivatives, oxcarbazepine, BIA 2-093, and BIA 2-024. *Neurochem Res* 27:121–130
- Aubel B, Kayser V, Farré A et al (2007) Evidence for adenosine- and serotonin-mediated antihyperalgesic effects of cizolirtine in rats suffering from diabetic neuropathy. *Neuropharmacology* 52:487–496
- Bastia E, Varani K, Monopoli A et al (2002) Effects of A(1) and A(2A) adenosine receptor ligands in mouse acute models of pain. *Neurosci Lett* 328:241–244
- Camann WR, Murray RS, Mushlin PS et al (1990) Effects of oral caffeine on postdural puncture headache. A double-blind, placebo-controlled trial. *Anesth Analg* 70:181–184
- Castaneda-Hernández G, Castillo-Méndez MS, López-Munoz FJ et al (1994) Potentiation by caffeine of the analgesic effect of aspirin in the pain-induced functional impairment model in the rat. *Can J Physiol Pharmacol* 72:1127–1131
- DeLander GE, Hopkins CJ (1986) Spinal adenosine modulates descending antinociceptive pathways stimulated by morphine. *J Pharmacol Exp Ther* 239:88–93
- Diamond S, Balm TK, Freitag FG (2000) Ibuprofen plus caffeine in the treatment of tension-type headache. *Clin Pharmacol Ther* 68:312–319

- Díaz-Reval MI, Ventura-Martínez R, Hernández-Delgadillo GP et al (2001) Effect of caffeine on antinociceptive action of ketoprofen in rats. *Arch Med Res* 32:13–20
- Diener HC, Pfaffenrath V, Pagler L et al (2005) The fixed combination of acetylsalicylic acid, paracetamol and caffeine is more effective than single substances and dual combination for the treatment of headache: a multicentre, randomized, double-blind, single-dose, placebo-controlled parallel group study. *Cephalalgia* 25:776–787
- Dworkin RH, O'Conner AB, Backonja M et al (2007) Pharmacological management of neuropathic pain: evidence-based recommendations. *Pain* 132:237–251
- Eisenach JC, Hood DD, Curry R et al (2004) Intrathecal but not intravenous opioids release adenosine from the spinal cord. *J Pain* 5:64–68
- Engelhardt G, Mauz AB, Pairet M (1997) Role of caffeine in combined analgesic drugs from the point of view of experimental pharmacology. *Arzneimittelforschung/Drug Res* 47:917–927
- Esmoğlu A, Akpınar H, Uğur F (2005) Oral multidose caffeine-paracetamol combination is not effective for the prophylaxis of postdural puncture headache. *J Clin Anesth* 17:58–61
- Esser MJ, Sawynok J (2000) Caffeine blockade of the thermal antihyperalgesic effect of acute amitriptyline in a rat model of neuropathic pain. *Eur J Pharmacol* 399:131–139
- Fennelly M, Galletly DC, Purdie GI (1991) Is caffeine withdrawal the mechanism of postoperative headache? *Anesth Analg* 72:449–453
- Fialip J, Porteix A, Marty H et al (1989) Lack of importance of caffeine as an analgesic adjuvant of dipyrone in mice. *Arch Int Pharmacodyn Ther* 302:86–95
- Fiebich BL, Biber K, Lieb K et al (1996) Cyclooxygenase-2 expression in rat microglia is induced by adenosine A<sub>2A</sub>-receptors. *Glia* 18:152–160
- Fiebich BL, Lieb K, Hüll M et al (2000) Effects of caffeine and paracetamol alone or in combination with acetylsalicylic acid on prostaglandin E<sub>2</sub> synthesis in rat microglial cells. *Neuropharmacology* 39:2205–2213
- Flores-Acevedo DM, Flores-Murrieta FJ, Castenada-Hernández G et al (1995) Potentiation of the analgesic effect of tolmetin, a potent non-steroidal anti-inflammatory drug, by caffeine in the rat. *Pharmaceut Sci* 1:441–444
- Fredholm BB (1980) Are methylxanthine effects due to antagonism of endogenous adenosine? *Trends Pharmacol Sci* 1:129–132
- Fredholm BB, Bättig K, Holmén J et al (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Gayawali K, Pandhi P, Sharma PL (1991) Determination of the optimal analgesia-potentiating dose of caffeine and a study of its effect on the pharmacokinetics of aspirin in mice. *Methods Find Clin Pharmacol* 13:529–533
- Ghelardini C, Gaelotti N, Bartolini A (1997) Caffeine induces central cholinergic analgesia. *Naunyn Schmiedebergs Arch Pharmacol* 356:590–595
- Godfrey L, Yan L, Clarke GD et al (2006) Modulation of paracetamol antinociception by caffeine and by selective adenosine A<sub>2</sub> receptor antagonists in mice. *Eur J Pharmacol* 531:80–86
- Granados-Soto V, Castenada-Hernández G (1999) A review of the pharmacokinetic and pharmacodynamic factors in the potentiation of the antinociceptive effect of nonsteroidal anti-inflammatory drugs by caffeine. *J Pharmacol Toxicol* 42:67–72
- Granados-Soto V, López-Munoz FJ, Castenada-Hernández G et al (1993) Characterization of the analgesic effects of paracetamol and caffeine combinations in the pain-induced functional impairment model in the rat. *J Pharm Pharmacol* 45:627–631
- Hampf KF, Schneider MC, Rüttimann U et al (1995) Perioperative administration of caffeine tablets for prevention of post-operative headaches. *Can J Anesth* 42:789–792
- Hering-Hanit R, Gadot N (2003) Caffeine-induced headache in children and adolescents. *Cephalalgia* 23:332–335
- Ho IK, Loh HH, Way EL (1973) Cyclic adenosine monophosphate antagonism of morphine analgesia. *J Pharmacol Exp Ther* 185:336–346
- Hussey MJ, Clarke CD, Ledent C et al (2007) Reduced response to the formalin test and lowered spinal NMDA glutamate receptor binding in adenosine A<sub>2A</sub> receptor knockout mice. *Pain* 129:287–294

- Jurna I (1981) Aminophylline differentiates between the depressant effects of morphine on the spinal reflex and on the spinal ascending activity evoked from afferent C fibres. *Eur J Pharmacol* 71:393–400
- Jurna I (1984) Cyclic nucleotides and aminophylline produce different effects on nociceptive motor and sensory responses in the rat spinal cord. *Naunyn Schmiedebergs Arch Pharmacol* 327:23–30
- Laska EM, Sunshine A, Mueller F et al (1984) Caffeine as an analgesic adjuvant. *JAMA* 251:1711–1718
- Ledent C, Vaugeois JM, Schiffman SN et al (1997) Aggressiveness, hypoalgesia and high blood pressure in mice lacking the adenosine  $A_{2A}$  receptor. *Nature* 388:674–678
- Lipton RB, Stewart WF, Ryan RE et al (1998) Efficacy and safety of acetaminophen, aspirin, and caffeine in alleviating migraine headache pain. *Arch Neurol* 55:210–217
- López JRM, Domínguez-Ramírez AM, Cook HG et al (2006) Enhancement of antinociception by co-administration of ibuprofen and caffeine in arthritic rats. *Eur J Pharmacol* 544:31–38
- López-Munoz FJ, Castenada-Hernández G, Flores-Mirrieta FJ et al (1996) Effect of caffeine coadministration and nitric oxide synthesis inhibition on the antinociceptive action of ketolorac. *Eur J Pharmacol* 308:275–277
- Malec D, Michalska E (1988) The effect of methylxanthines on morphine analgesia in mice and rats. *Pol J Pharmacol Pharm* 40:223–232
- Mashimoto S, Ushijima I, Suetsugi M et al (1998) Stress-dependent antinociceptive effects of carbamazepine: a study in stressed and non-stressed rats. *Prog Neuropsychopharmacol Biol Psychiatry* 22:159–168
- Micó JA, Ardid D, Berrococo E et al (2006) Antidepressants and pain. *Trends Pharmacol Sci* 27:348–354
- Migliardi JR, Armellino JJ, Friedman M et al (1994) Caffeine as an analgesic adjuvant in tension headache. *Clin Pharmacol Ther* 56:576–586
- Misra AL, Pontani RB, Vadlamani NL (1985) Potentiation of morphine analgesia by caffeine. *Br J Pharmacol* 84:789–791
- Nehlig A, Daval JL, Debry G (1992) Caffeine and the central nervous system: mechanisms of action, biochemical, metabolic and psychostimulant effects. *Brain Res Rev* 17:39–170
- Oliverio A, Castellano C, Pavone F et al (1983) Caffeine interferes with morphine-induced hyperactivity but not analgesia. *Pol J Pharmacol Pharm* 35:445–449
- Pareek SS, Chopde CT, Thakur Desai PA (1994) Adenosine enhances analgesic effect of tricyclic antidepressants. *Indian J Pharmacol* 26:159–161
- Person DL, Kissin I, Brown PT et al (1985) Morphine-caffeine analgesic interaction in rats. *Anesth Analg* 64:851–856
- Pham T, Carrega L, Sauze N et al (2003) Supraspinal antinociceptive effects of  $\mu$  and  $\delta$  agonists involve modulation of adenosine uptake. *Anesthesiology* 98:459–464
- Po ALW, Zhang WY (1998) Analgesic efficacy of ibuprofen alone and in combination with codeine or caffeine in post-surgical pain: a meta-analysis. *Eur J Clin Pharmacol* 53:303–311
- Sawynok J (2006) Adenosine and ATP receptors. In: Stein C (ed) *Handbook of experimental pharmacology*. Springer, Berlin, pp 301–320
- Sawynok J, Liu XJ (2003) Adenosine in the spinal cord and periphery: release and regulation of pain. *Prog Neurobiol* 69:313–340
- Sawynok J, Reid A (1996a) Caffeine antinociception: role of formalin concentration and adenosine A1 and A2 receptors. *Eur J Pharmacol* 298:105–111
- Sawynok J, Reid A (1996b) Neurotoxin-induced lesions to central serotonergic, noradrenergic and dopaminergic systems modify caffeine-induced antinociception in the formalin test and locomotor stimulation in rats. *J Pharmacol Exp Ther* 277:646–653
- Sawynok J, Yaksh TL (1993) Caffeine as an analgesic adjuvant: a review of pharmacology and mechanisms of action. *Pharmacol Rev* 45:43–85
- Sawynok J, Sweeney MI, White TD (1986) Classification of adenosine receptors mediating antinociception in the rat spinal cord. *Br J Pharmacol* 88:923–930

- Sawynok J, Sweeney MI, White TD (1989) Adenosine release may mediate spinal analgesia by morphine. *Trends Pharmacol Sci* 10:186–189
- Sawynok J, Reid A, Doak GJ (1995) Caffeine antinociception in the rat hot-plate and formalin tests and locomotor stimulation: involvement of noradrenergic mechanisms. *Pain* 61:203–213
- Sawynok J, Reid AR, Esser MJ (1999) Peripheral antinociceptive action of amitriptyline in the rat formalin test: involvement of adenosine. *Pain* 80:45–55
- Sawynok J, Reid AR, Liu XJ et al (2005) Amitriptyline enhances extracellular levels of adenosine in the rat hindpaw and inhibits adenosine uptake. *Eur J Pharmacol* 518:116–122
- Sawynok J, Reid A, Fredholm BB (2008) Caffeine reverses antinociception by amitriptyline in wild type mice but not in those lacking adenosine A<sub>1</sub> receptors. *Neurosci Lett* 440:181–184
- Schachtel BP, Thoden WR, Konerman JP et al (1991) Headache pain model for assessing and comparing the efficacy of over-the-counter analgesic agents. *Clin Pharmacol Ther* 50:322–329
- Seegers AJM, Jager LP, Zandberg P et al (1981) The anti-inflammatory, analgesic and antipyretic activities of non-narcotic analgesic drug mixtures in rats. *Arch Int Pharmacodyn Ther* 251:237–254
- Shapiro RE (2008) Caffeine and headaches. *Curr Pain Headache Rep* 12:311–315
- Siegers CP (1973) Effects of caffeine on the absorption and analgesic efficacy of paracetamol in rats. *Pharmacology* 10:19–27
- Sierralta F, Pinardi G, Mendez M et al (1995) Interaction of opioids with antidepressant antinociception. *Psychopharmacology* 122:374–378
- Suh HW, Song DK, Kim YH (1997) Differential effects of adenosine receptor antagonists injected intrathecally on antinociception by morphine and  $\beta$ -endorphin administered intracerebroventricularly in the mouse. *Neuropeptides* 31:339–344
- Sweeney MI, White TD, Sawynok J (1987a) Involvement of adenosine in the spinal antinociceptive effects of morphine and noradrenaline. *J Pharmacol Exp Ther* 243:657–665
- Sweeney MI, White TD, Sawynok J (1987b) Morphine releases endogenous adenosine from the spinal cord *in vivo*. *Eur J Pharmacol* 141:169–170
- Sweeney MI, White TD, Sawynok J (1991) Intracerebroventricular morphine releases cyclic AMP and adenosine from the spinal cord via a serotonergic mechanism. *J Pharmacol Exp Ther* 259:1013–1028
- Tomić MA, Vučković SM, Stepanović-Petrović RM et al (2004) The anti-hyperalgesic effects of carbamazepine and oxcarbazepine are attenuated by treatment with adenosine receptor antagonists. *Pain* 111:253–260
- Tomić MA, Vučković SM, Stepanović-Petrović RM et al (2006) Peripheral anti-hyperalgesia by oxcarbazepine: involvement of adenosine A<sub>1</sub> receptors. *Pharmazie* 61:566–568
- Ulugol A, Karadag HC, Tamer M et al (2002) Involvement of adenosine in the anti-allodynic effect of amitriptyline in streptozotocin-induced diabetic rats. *Neurosci Lett* 328:129–132
- Vinegar R, Traus JF, Selph JL et al (1976) Potentiation of the anti-inflammatory and analgesic activity of aspirin by caffeine in the rat. *Proc Soc Exp Biol Med* 151:556–560
- Ward N, Whitney C, Avery D et al (1991) The analgesic effects of caffeine in headache. *Pain* 44:151–155
- Weber JG, Klinderworth JT, Arnold JJ et al (1997) Prophylactic intravenous administration of caffeine and recovery after ambulatory surgical procedures. *Mayo Clin Proc* 72:621–626
- Wilson SG, Mogil JS (2001) Measuring pain in the (knockout) mouse: big challenges in a small mammal. *Behav Brain Res* 125:65–73
- Wu WP, Hao JX, Halldner L et al (2005) Increased nociceptive response in mice lacking the adenosine A<sub>1</sub> receptor. *Pain* 113:395–404
- Wu WP, Hao JX, Fredholm BB et al (2006) Effect of acute and chronic administration of caffeine on pain-like behaviors in rats with partial sciatic nerve injury. *Neurosci Lett* 402:164–166
- Yaba G, Sezer Z, Tekol Y (2006) Interaction between venlafaxine and caffeine on antinociception in mice. *Pharmazie* 61:60–62
- Yucel A, Ozyalcin S, Talu GK et al (1999) Intravenous administration of caffeine sodium benzoate for postdural puncture headache. *Reg Anesth Pain Med* 24:51–54

- Zhang WY (2001) A benefit-risk assessment of caffeine as an analgesic adjuvant. *Drug Saf* 24: 1127–1142
- Zhang WY, Po ALW (1996) Analgesic efficacy of paracetamol and its combination with codeine and caffeine in surgical pain – a meta-analysis. *J Clin Pharm Ther* 21:261–282
- Zhang WY, Po ALW (1997) Do codeine and caffeine enhance the analgesic effect of aspirin? – a systematic overview. *J Clin Pharm Ther* 22:79–97
- Zhang Y, Conklin DR, Li X et al (2005) Intrathecal morphine reduces allodynia after peripheral nerve injury in rats via activation of a spinal A<sub>1</sub> adenosine receptor. *Anesthesiology* 102: 416–420

# Methylxanthines and Sleep

Tarja Porkka-Heiskanen

## Contents

1	What Is Sleep? .....	332
2	Regulation of Sleep .....	334
2.1	Waking Mechanisms .....	334
2.2	Sleep Induction Mechanisms .....	335
3	Caffeine and Sleep .....	338
3.1	General Actions of Caffeine in the CNS with Potential Significance in the Regulation of Sleep .....	338
3.2	Mechanisms of Caffeine Action on Sleep .....	338
3.3	Effects of Caffeine on Sleep in Animals .....	341
3.4	Effects of Caffeine on Sleep in Humans .....	341
3.5	Caffeine and Performance .....	343
3.6	Individual Sensitivity to Caffeine .....	343
3.7	Sleep During Caffeine Withdrawal .....	344
3.8	Effects of Other Methylxanthines on (Human) Sleep .....	344
	References .....	345

**Abstract** Caffeine is widely used to promote wakefulness and counteract fatigue induced by restriction of sleep, but also to counteract the effects of caffeine abstinence. Adenosine is a physiological molecule, which in the central nervous system acts predominantly as an inhibitory neuromodulator. Adenosine is also a sleep-promoting molecule. Caffeine binds to adenosine receptors, and the antagonism of the adenosinergic system is believed to be the mechanism through which caffeine counteracts sleep in humans as well as in other species. The sensitivity for caffeine varies markedly among individuals. Recently, genetic variations in genes related to adenosine metabolism have provided at least a partial explanation for this variability. The main effects of caffeine on sleep are decreased sleep latency,

---

T. Porkka-Heiskanen  
Institute of Biomedicine/Physiology, University of Helsinki, P.O. Box 63, Haartmaninkatu 8,  
00014 Helsinki, Finland  
e-mail: porkka@cc.helsinki.fi

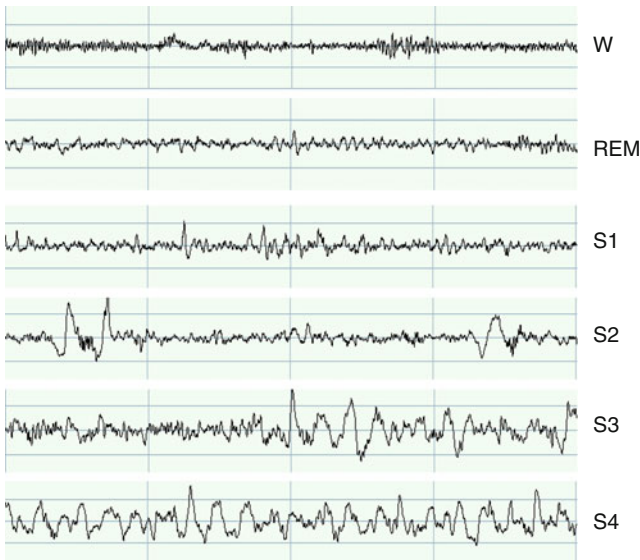
shortened total sleep time, decrease in power in the delta range, and sleep fragmentation. Caffeine may also decrease the accumulation of sleep propensity during waking, thus inducing long-term harmful effects on sleep quality.

**Keywords** Adenosine · Adenosine receptors · Caffeine · Sleep · Sleep deprivation

## 1 What Is Sleep?

In humans, sleep is characterized by lack of consciousness: we are no longer able to evaluate ourselves and we lose contact with the surrounding reality. An important feature in brain function during sleep is decreased cortical reactivity: we are unable to respond to external stimuli as we would during wakefulness.

In mammals, sleep is defined through an electroencephalogram (EEG): in waking, cortical activity is desynchronized, leading to high-frequency, low-amplitude waveforms. When sleep is initiated, the EEG becomes synchronized, expressed as low-frequency, high-amplitude waveforms (Fig. 1). Sleep is further divided into two main stages: non-REM sleep and REM sleep. In REM sleep cortical activity is as desynchronized as in waking, and only on the basis of the



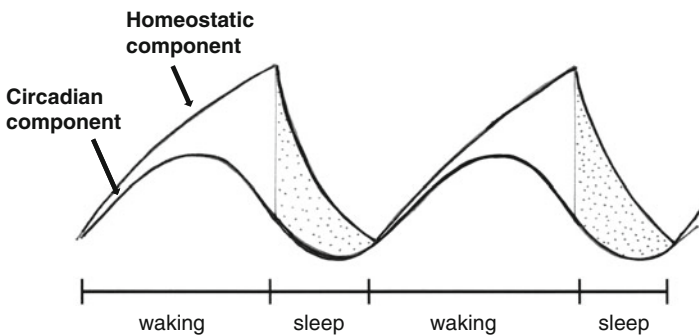
**Fig. 1** Human vigilance states defined by an electroencephalogram (EEG). Waking (W) is characterized with a high-frequency but low-amplitude waveform, similarly to REM sleep (REM). Non-REM sleep is divided into stages 1–4, with deepening of sleep from stage 1 to stage 4. Upon deepening of sleep, the EEG wave frequency decreases while the amplitude increases

EEG, these stages cannot be distinguished. Non-REM sleep is characterized by slowing down of the body and brain functions: synchronized cortical activity, decreased heart rate and blood pressure, as well as breathing. In REM sleep, heart rate, blood pressure, and breathing become irregular, with large and sudden variations. Muscle tone is almost completely lost in body muscles, but is maintained in the muscles of the head and in muscles used in breathing. However, the unconsciousness that is so characteristic to sleep is maintained.

The electric activity of the brain (cortex) can be measured using an electroencephalograph and can be either described visually as “brain waves” (Fig. 1) or quantized as a power spectrum over the frequencies of the EEG. A power spectrum is a mathematical measure of the intensity of brain electric activity, and it is calculated from the original EEG signal using Fourier transformation.

Low EEG frequencies are typical for sleep and high frequencies are typical for waking. The more power there is in the low-frequency range (typically 0.75–4 Hz, called delta range), the deeper the sleep is, while during waking, delta power is low. Powers in other frequency ranges also have significance for the vigilance state – in this chapter power in the theta range will be assessed in addition to that in the delta range. Theta activity is high during waking, and it increases during brain activation. In rodents, theta activity is characterized by exploratory behavior, meaning that the animals have motor activity (Vyazovskiy and Tobler 2005; Wigren et al. 2009). In humans, theta power increases in the course of waking, and it has been suggested to be a marker of sleep homeostasis during waking, while delta power is a marker of sleep homeostasis during sleep (Finelli et al. 2000).

An important feature of sleep is its homeostatic regulation, which means that a long waking period is followed by a correspondingly longer and deeper sleep period. This feature has been characterized from human (and animal) EEG power spectra and formalized mathematically in the two-process model of sleep regulation (Borbely 1982; see also Fig. 2). Sleep homeostasis is often described as accumulation of sleep propensity, or “sleep pressure” during the waking period, but what, at



**Fig. 2** The two-process model of sleep regulation. Sleep is regulated by two processes: the circadian component and the homeostatic component. During waking the homeostatic component increases until sleep is initiated. If waking is prolonged, the homeostatic component keeps increasing, while the circadian component continues its cycling variation. (Modified from Borbely 1982)



the molecular level, its accumulation is less clear. The accumulation of sleep pressure is necessary for initiation of sleep – if this mechanism is disturbed during the waking period, there will be difficulties in falling asleep and the sleep period may be terminated early. Also, the deep-sleep phases may be short, or absent.

Adenosine has been suggested to be one of the molecular correlates of sleep pressure and an inducer of sleepiness (Porkka-Heiskanen et al. 1997). Caffeine is an adenosine receptor blocker, counteracting sleepiness through this action.

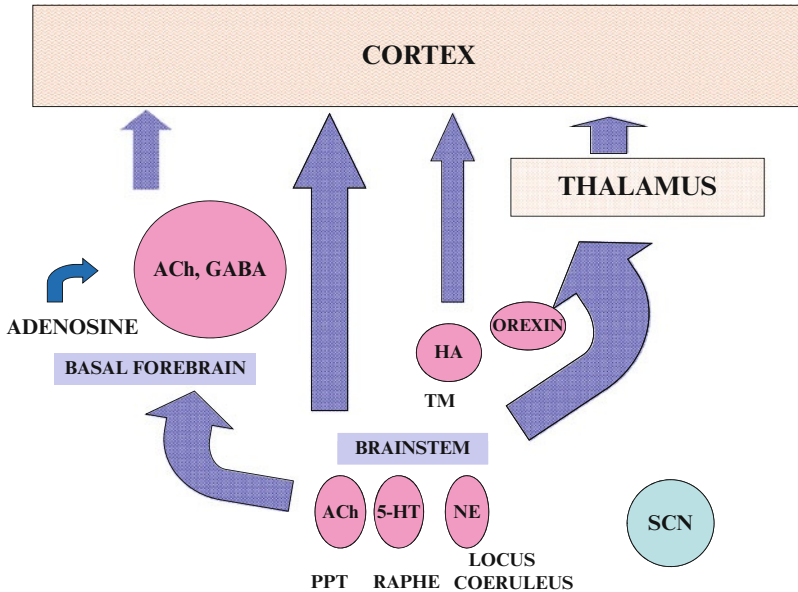
## 2 Regulation of Sleep

### 2.1 *Waking Mechanisms*

Sleep and wakefulness are mutually exclusive states of vigilance and thus need to be regulated in concert. As a consequence, we cannot understand the regulation of sleep without understanding the regulation of wakefulness.

Cortical excitability underlies all cortical activity, and its level is maintained and regulated by several, partly overlapping neuronal networks with very similar anatomical structure. The key elements of these networks are small cell groups, lying either in the brainstem or in the hypothalamus and sending widespread projections to the cortex and other brain areas. The level of cortical excitability is defined by the level of activity of these cell groups, which release neurotransmitters that regulate the reactivity of cortical neurons (predominantly glutamatergic): during wakefulness the activity in the cell groups is high and a lot of neurotransmitters are released into the cortex, while in sleep the activity of the cell groups decreases or ceases and only small amounts of neurotransmitters are released, leading to low cortical reactivity (for a review, see Stenberg, 2007). The waking-maintaining cell groups in the brainstem consist of serotonin neurons in the raphe nuclei, noradrenaline neurons in the locus coeruleus, and acetylcholine neurons in the laterodorsal tegmental and pedunculopontine tegmental nuclei. In addition, in the hypothalamus/basal forebrain (BF) area, histamine neurons in the tuberomammillary nuclei, orexin neurons in the lateral hypothalamic nucleus, and cholinergic neurons in the BF serve this function (for a summary, see Fig. 3). The activity of all these cells is high during waking and decreases upon falling asleep. Initiation of sleep is not possible if these neurons continue their activity. The activity of these cells continues to decrease during deep-sleep phases, but interestingly, during REM sleep one group of neurons, the cholinergic neurons, increase their activity to the level of waking, while other waking-maintaining cell groups virtually cease firing.

All these neurons have adenosine receptors on them, thus making them targets for caffeine and offering it an opportunity to modulate the vigilance state through these neurons.



**Fig. 3** Summary of the cell groups that regulate the vigilance state. All vigilance-regulating cell groups are located in the old parts of the brain: brainstem, basal forebrain and hypothalamus. The groups in the brainstem – pedunculopontine tegmental nucleus (PPT) with cholinergic cells (ACh), raphe nuclei with serotonergic cells (5-HT) and locus coeruleus with noradrenergic cells (NE) – project either directly or via the thalamus or via the basal forebrain to the cortex. In the hypothalamus tuberomammillary nuclei (TM) histaminergic cells (HA) and in the lateral hypothalamus orexinergic cells (OREXIN) send projections to the cortex, as do cholinergic cells (ACh) and GABAergic cells (GABA) in the basal forebrain. SCN suprachiasmatic nucleus – the site of the inner clock

## 2.2 Sleep Induction Mechanisms

So far only one area in the brain has been shown to contain neurons that are more active during sleep than during waking – sleep-active neurons: the preoptic area of the hypothalamus, particularly the ventrolateral preoptic area. The cells of this nucleus are GABAergic and send inhibitory projections to all waking nuclei. This system forms the core of the mutually exclusive vigilance state regulation: when sleep-active neurons are on, they send inhibitory signals to waking-active neurons, and vice versa (McGinty and Szymusiak, 2000; Saper et al. 2001).

What about induction of sleep homeostasis? Several molecules have been proposed to take part in this regulation, including adenosine (Porkka-Heiskanen et al. 1997), nitric oxide (Kalinchuk et al. 2006), prostaglandins (Hayashi 1988), and cytokines (Krueger and Fang 1997). This chapter concentrates on the role of adenosine in this regulation.

The adenosine concentration increases during prolonged wakefulness in the BF (Porkka-Heiskanen et al. 1997), and as adenosine is predominantly an inhibitory neuromodulator (through activation of  $A_1$  receptors), it will decrease neuronal activity. The inhibition of the BF wake-active cholinergic neurons will result in increased sleepiness and sleep propensity and initiation of recovery sleep. If the adenosine receptors are blocked during waking, also the accumulation of sleep pressure is attenuated (Gass et al. 2009; Landolt et al. 2004). This has relevance for the mechanisms by which caffeine reduces sleep: it can modulate the accumulation of sleep pressure.

### 2.2.1 Adenosine

#### The Effects of Adenosine in the Central Nervous System with Relevance for Regulation of the Vigilance State

Adenosine is predominantly an inhibitory neurotransmitter in the central nervous system (CNS), acting through  $A_1$  receptors. There is evidence that the release of excitatory transmitters is inhibited by adenosine more strongly than that of inhibitory neurotransmitters (Fredholm and Dunwiddie 1988). Adenosine inhibits neuronal activity through several mechanisms, e.g., through decreasing neurotransmitter release (Dunwiddie 1985) and inhibiting the firing rate of neurons (Phillis and Edstrom 1976). The BF cholinergic neurons, which are part of the waking-maintaining system, are tonically inhibited by adenosine (Rainnie et al. 1994), and adenosine inhibits the tuberomammillar histaminergic neurons via  $A_1$  receptors (Oishi et al. 2008), giving rise to the hypothesis that adenosine may promote sleepiness by inhibiting activity and neurotransmitter release of wakefulness-promoting neurons.

#### Adenosine Receptors in the CNS: Types and Distribution

There are four types of adenosine receptors in the CNS (for details, see Chap. 6, in this volume). Of the four receptor types,  $A_1$  and  $A_{2A}$  appear to be relevant in regulation of vigilance states.

$A_1$  receptors are located in all parts of the CNS and on both inhibitory and excitatory neurons, while the  $A_{2A}$  receptor location appears to be limited to the striatum (for details, see Chap. 6, in this volume). The result of  $A_1$  receptor stimulation is neuronal inhibition, and the functional outcome depends on whether inhibitory or excitatory neurons were inhibited. The result of  $A_{2A}$  receptor stimulation is neuronal activation.

#### Adenosine and Sleep

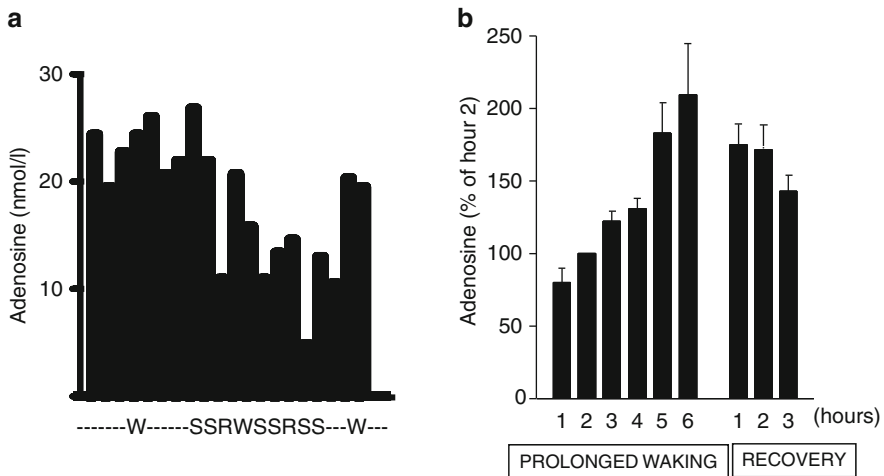
Circadian variation in the enzymes that metabolize adenosine suggests that also adenosine levels may undergo circadian- and/or vigilance-state-related variation (Chagoya de Sanchez et al. 1993). In rats, adenosine levels were high during the

active (dark) period of this species both in the cortical areas (Chagoya de Sanchez et al. 1993; Huston et al. 1996) and in the BF (Murillo-Rodriguez et al. 2004). In freely moving cats, adenosine levels were lower in samples that were collected during sleep than in those collected during wakefulness (Porkka-Heiskanen et al. 1997). Adenosine, acting on A<sub>1</sub> receptors, promotes non-REM sleep by inhibiting the histaminergic neurons in the tuberomammillary nucleus (Oishi et al. 2008) (Fig. 3).

Prolonged waking increased adenosine levels in the BF of cats (Fig. 4) and increasing adenosine concentration experimentally in the BF induced sleep (Porkka-Heiskanen et al. 1997), while blocking the A<sub>1</sub> receptors during prolonged wakefulness prevented recovery sleep (Gass et al. 2009). Taken together, these data imply that adenosine is a mediator of sleep homeostasis.

### Genetic Variations in the Human Adenosinergic System Relevant for Sleep

A functional genetic variation in adenosine deaminase (ADA) modulates sleep (Retey et al. 2005): both the duration and the intensity of slow-wave sleep were increased in individuals with ADA G/A genotype as compared with individuals with G/G genotype (Retey et al. 2005). The polymorphism (G to A transition at nucleotide 22) is functional, since the enzymatic activity of the G/A variant ADA is



**Fig. 4** (a) Variation in the basal forebrain adenosine concentration during a spontaneous sleep-wake cycle. Adenosine concentrations were measured during waking (W), sleep (S), and REM sleep (R). During waking the adenosine levels are high, decrease during sleep, and increase during REM sleep. (b) Adenosine concentration in the basal forebrain during prolonged wakefulness. Adenosine concentrations were measured during a 6-h wakefulness period from samples collected in the basal forebrain. The adenosine concentration increased steadily during the wakefulness period and started to decrease upon falling asleep (recovery). (Modified from Porkka-Heiskanen et al. 1997)

20–30% lower than that of the G/G variant (Battistuzzi et al. 1981), presumably leading to higher tissue adenosine levels.

### 3 Caffeine and Sleep

#### 3.1 *General Actions of Caffeine in the CNS with Potential Significance in the Regulation of Sleep*

In general, the inhibition of CNS A<sub>1</sub> receptors will increase neuronal firing, while inhibition of A<sub>2A</sub> receptors will decrease it.

In addition to the adenosine-receptor-mediated actions of caffeine (discussed in the next sections), caffeine can affect vigilance through other known mechanisms in the CNS. Caffeine increases the turnover of several monoamine neurotransmitters, including 5-hydroxytryptamine (5-HT) (Fernstrom et al. 1984), dopamine, and noradrenaline (Fredholm et al. 1984; Hadfield and Milio 1989) and it inhibits release of neurotransmitters (Fredholm and Dunwiddie 1988). High doses of caffeine can also activate phosphodiesterase, but this is usually not the case with normal human caffeine consumption (Fredholm et al. 1999).

The tonic inhibition of neurons by adenosine is inhibited by caffeine, which elevates the excitability of rat hippocampus slices (Dunwiddie et al. 1981; Greene et al. 1985) and activates the theta rhythm in rabbit hippocampus (Popoli et al. 1987). High caffeine doses (100 mg/kg or above) provoke seizurelike activity in the hippocampus.

#### 3.2 *Mechanisms of Caffeine Action on Sleep*

There is consensus that caffeine acts through adenosine receptors to reduce sleepiness/sleep, but the question of whether this action takes place through A<sub>1</sub> or A<sub>2A</sub> receptors is still debated (for a review, see Fredholm et al. 1999). This question has been intimately related to the question of whether adenosine acts on vigilance states through A<sub>1</sub> or A<sub>2A</sub> receptors (Fredholm et al. 1999; Basheer et al. 2004) but it should be appreciated that these are two related, but separate questions.

Probably, both adenosine A<sub>1</sub> and A<sub>2A</sub> receptors are involved in producing the sleep-promoting effects of adenosine, but these effects appear to be exerted in different parts of the brain. Consequently, several adenosine-receptor-mediated mechanisms may contribute to the sleep-reducing effects of caffeine, depending on the location of the receptors (whether they lie on waking-active or sleep-active cells) and whether they are predominantly of the inhibitory A<sub>1</sub> type or the activating A<sub>2A</sub> type. One hypothesis states that caffeine induces waking through inhibition of the cholinergic waking-active neurons (Rainnie et al. 1994). This view has recently

been challenged by results showing that caffeine does not affect vigilance in  $A_1$ -receptor-knockout mice, but significantly reduces sleep in  $A_{2A}$ -receptor-knockout mice (Huang et al. 2005), indicating that the effects of caffeine on the vigilance state are mediated through  $A_{2A}$  receptors.

In addition to this immediate arousing effect, caffeine may also inhibit the buildup of sleep homeostasis (Landolt et al. 2004). This effect is less well studied and generally neglected in discussions on the effect of caffeine on sleep, and particularly regarding its role in inducing sleeping problems (mainly prolonged sleep latency and/or premature awakenings). The mechanisms of this inhibition – whether  $A_1$  or  $A_{2A}$  receptors are involved and which neuronal groups are involved – remain to be determined.

### 3.2.1 $A_1$ Receptors

Studies indicating that adenosine acts through  $A_1$  receptors to reduce sleepiness/sleep are numerous. Early studies by Ticho and Radulovacki (1991) showed that local injections of adenosine  $A_1$  receptor agonists in the preoptic area of the rat induced sleep, while an  $A_{2A}$  receptor agonist did not. Also, systemic as well as intracerebroventricular administrations of  $A_1$  receptor agonists induced sleep (Benington and Heller 1995; Schwierin et al. 1996), while blocking the  $A_1$  receptors pharmacologically (Strecker et al. 2000) or by inactivating them by using an antisense oligonucleotide targeted at the  $A_1$  receptor (Thakkar et al. 2003) decreased sleep and increased waking. The administration of the adenosine  $A_1$ -receptor-selective agonist cyclopentyladenosine mimicked the EEG effects of sleep deprivation (Benington et al. 1995) and non-REM sleep (Schwierin et al. 1996).

The role of adenosine in induction of recovery sleep after sleep restriction has been convincingly shown in a series of experiments where an increase in BF adenosine concentration through either sleep restriction or pharmacological manipulation increased sleep (Porkka-Heiskanen et al. 1997; Porkka-Heiskanen et al. 2000), while blocking the  $A_1$  receptors during the deprivation, either pharmacologically (Gass et al. 2009) or using an  $A_1$ -specific antisense oligonucleotide (Thakkar et al. 2003), decreased it. Interestingly, the number of  $A_1$  receptors increased both in rat (Basheer et al. 2007) and in human (Elmenhorst et al. 2009) brain during sleep deprivation, further suggesting a role for  $A_1$  receptors in regulation of sleepiness.

The site(s) where adenosine increases sleep through  $A_1$  receptors is not known, but inhibition of wake-active neurons appears a plausible mechanism. The BF wake-active cholinergic neurons are tonically inhibited by adenosine through  $A_1$  receptors (Rainnie et al. 1994), and the removal of this inhibition by blocking these receptors would increase their firing and cortical arousal. There is evidence that the activity of the cholinergic neurons can be regulated by adenosinergic manipulations: both theophylline (Murray et al. 1982) and caffeine can modify acetylcholine levels and metabolism in the brain (Phillis and Wu 1981; Murray et al. 1982; Carter et al. 1995). Caffeine increases the level of cortical acetylcholine dose dependently

and even at doses that can be induced by ordinary caffeine consumption (Carter et al. 1995).

Evidently the induction of recovery sleep is connected with the BF cholinergic cells, since recovery sleep remained absent in animals that had been lesioned of these cells (Kalinchuk et al. 2008). In the lesioned animals, the increase in BF adenosine concentration during sleep restriction also remained absent.

### 3.2.2 A<sub>2</sub> Receptors

Several studies also suggest a role for A<sub>2A</sub> receptors in regulation of vigilance states, and that caffeine may act through these receptors when reducing sleep. The key structure in this action appears to be the subarachnoid space, below the rostral BF, where prostaglandin D<sub>2</sub> receptor activation releases adenosine, which induces sleep through activation of A<sub>2A</sub> receptors (for a review, see Urade and Hayaishi 1999; Hayaishi 2002).

Pharmacology studies showed that infusion of the A<sub>2A</sub> receptor agonist CGS 21680 into the subarachnoid space increased slow-wave sleep. This effect was blocked by the A<sub>2A</sub> receptor antagonist KF17837 (Satoh et al. 1996; Ram et al. 1997; Satoh et al. 1999). Injection of the selective adenosine A<sub>2A</sub> receptor agonist CGS 21680 into the subarachnoid space mimicked the sleep-promoting effects of prostaglandin D<sub>2</sub>, whereas an A<sub>1</sub> receptor agonist did not (Satoh et al. 1996). The authors stated that the adenosine A<sub>2A</sub> receptors in the tuberculum olfactorium/ventral nucleus accumbens are a likely site of action (Satoh et al. 1996).

The strongest evidence to support a role for A<sub>2A</sub> receptors in caffeine-mediated increase in waking comes from a study with A<sub>2A</sub>-receptor-knockout mice. In this study caffeine was administered to mice that had been genetically engineered so that they had either no A<sub>2A</sub> receptors or no A<sub>1</sub> receptors, as well as to normal mice with both receptors intact. Caffeine increased waking in the wild-type mice as well as in A<sub>1</sub>-receptor-knockout mice, but not in A<sub>2A</sub>-receptor-knockout mice, indicating that the effects of caffeine are mediated through A<sub>2A</sub> receptors (Huang et al. 2005). This view is supported by human data: A single c.1083T>C polymorphism in the A<sub>2A</sub> receptor gene (ADORA<sub>2A</sub>) modulates individual sensitivity to subjective and objective effects of caffeine on sleep (Retey et al. 2007).

Interestingly, a recent study conducted on fruit flies showed that chronic administration of caffeine reduces and fragments their sleep (Wu et al. 2009), while mutants lacking the fly adenosine receptor (with sequence similarity to mammalian A<sub>2A</sub> receptor) had normal amounts of baseline sleep, as well as normal homeostatic responses to sleep deprivation. Surprisingly, these mutants respond normally to caffeine. However, a phosphodiesterase inhibitor, 3-isobutyl-1-methylxanthine (IBMX), mimicked the effects of caffeine in the mutant flies, while the effects of caffeine on sleep were blocked in flies that had reduced neuronal phosphodiesterase (PKA) activity. The authors concluded that chronic administration of caffeine promotes wakefulness in *Drosophila*, at least in part by inhibiting cyclic AMP PKA activity, but not through adenosine receptors. Whether such mechanisms

contribute to wakefulness induced by chronic caffeine intake also in mammals remains to be clarified through further studies.

### 3.3 *Effects of Caffeine on Sleep in Animals*

Caffeine decreases sleep and increases wakefulness in all animal species tested so far, e.g., cats (Sinton and Petitjean 1989), (Yanik et al. 1987) and mice (Stenberg et al. 2003). In rats, the effects of caffeine on recovery sleep have also been addressed: caffeine reduced attempts to sleep during sleep deprivation and slowed the rate of recovery sleep, but did not prevent it (Wurts and Edgar 2000).

The effects of caffeine on the vigilance state are not restricted to mammals. A vigilance state that resembles sleep (characterized by immobility, increased arousal threshold, and signs of homeostatic regulation) can also be defined in species that do not have a CNS, which would allow EEG recording. Caffeine decreases sleep in the fruit fly (*Drosophila melanogaster*) (Shaw et al. 2000). A sleeplike state has also been characterized in zebra fish (*Danio rerio*) (Zhdanova et al. 2001; Yokogawa et al. 2007). Recently we showed that caffeine increases wakefulness and decreases “sleep” in this species (unpublished data).

### 3.4 *Effects of Caffeine on Sleep in Humans*

Sleep in humans is readily affected by caffeine. Epidemiology studies have reported an association between daily caffeine intake and sleep problems and daytime sleepiness (for a review, see Roehrs and Roth 2008).

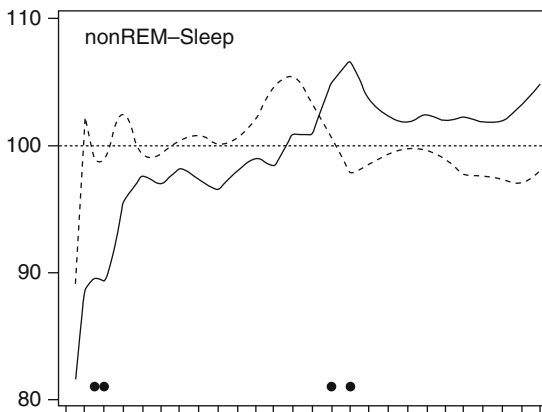
The most prominent effects of caffeine on sleep are prolonged sleep latency, shorter total sleep time, increases in light sleep, and decreases in deep sleep. REM sleep is less affected but the latency to the first REM period is shortened. Subjective sleep quality is also decreased reflecting the longer sleep latency, increased awakenings, and the shorter total sleep time. Remarkably, these effects are obvious not only when caffeine is ingested just before bedtime, but also when caffeine is ingested in the morning and the effects on sleep are similar on the following night.

When caffeine is ingested shortly before bedtime, experimental findings include a dose-dependent decrease in total sleep time, an increase in sleep latency and a decrease in deep sleep (Karacan et al. 1976; Nicholson and Stone 1980), impaired sleep efficacy and a decrease in slow-wave sleep (Landolt et al. 1995a, b), as well as decreased power in the delta frequency range, but an increase in power in the spindle frequency range (Landolt et al. 1995a, b, see Fig. 5).

Caffeine ingested even in the morning has prominent effects on sleep during the subsequent night: when modest coffee drinkers ingested 200 mg caffeine in the morning, their total sleep time was reduced by about 10 min, the latency to stage 2 sleep was prolonged, and sleep efficiency (ratio of time asleep to time in bed) was reduced by about 3% (Landolt et al. 1995a, b). The changes in the EEG



**Fig. 5** Effect of caffeine on human sleep EEG. Caffeine decreases the EEG power density at low frequencies and increases it at theta frequencies. *Solid line* EEG recorded after caffeine intake, *dashed line* before-caffeine EEG. The Y-axis is the EEG power density (%). (Modified from Landolt et al. 1995a, b)



were similar to those observed after night ingestion of caffeine. Caffeine had already metabolized, as evidenced by low saliva concentrations, but it is possible that the psychoactive metabolites induced some of the changes (Reitey et al. 2007; Landolt et al. 2004).

The caffeine-induced EEG changes in non-REM sleep are comparable in rested and sleep-deprived subjects. They are consistently found irrespective of whether the study participants maintain habitual morning caffeine consumption (Landolt et al. 1995a, b) or abstain from caffeine for days (Landolt et al. 1995a, b) (Landolt et al. 2004) and weeks (Reitey et al. 2007; Landolt et al. 2004) before the experiments. It is thus improbable that reversal of withdrawal syndromes would explain these changes (James and Keane 2007).

Studies that compare the effects of caffeine on sleep between high- and low-coffee consumers have shown that caffeine intake in low consumers will induce prolonged sleep latency, disturbances in different sleep phases, and shorter total sleep time (Curatolo and Robertson 1983), while the effects in habitual coffee drinkers are smaller (Colton et al. 1968).

Not only caffeine itself but also abstinence from caffeine will induce sleep difficulties (for a review, see James and Keane 2007). In a field study, regular caffeine drinkers were assigned to groups that received either regular coffee or decaffeinated coffee for 9 days, or to a group that at 2-day intervals was switched between regular and decaffeinated coffee. Caffeine abstinence resulted in increased heart rate, decreased motor activity, subjective wakefulness, and an increased number of headaches and use of analgesics. The subjective effects and headaches disappeared after a few days of abstinence and weakened over successive, separated abstinence periods. The intermittent onset of caffeine consumption resulted in increased wakefulness (Höfer and Bättig 1994).

Caffeine has even been used as a model of insomnia in different experimental designs (Okuma et al. 1982).

Insomnia was mimicked in volunteers by administering to them 400 mg caffeine three times a day for 7 days. The treatment decreased the total sleep time and increased

the sleep latency, while stage 4 sleep was reduced. Partial tolerance to the effects of caffeine developed in the course of the experiment (Bonnet and Arand 1992).

In summary, the sleep-disruptive effects of caffeine, even at doses equivalent to a single cup of coffee, have been well documented.

### ***3.5 Caffeine and Performance***

Caffeine restores performance and mood under sleep loss, when sleep has been previously restricted owing to, e.g., shift work or jet lag. Several studies, including laboratory and field studies, have documented the beneficial effect of caffeine on performance impairment associated with shift work (Bonnet et al. 2005; Schweitzer et al. 2006).

Naps are frequently used, and have been documented to be useful, in ameliorating the feeling of sleepiness and fatigue as well as a decrease of performance. The benefits of naps can be improved by combining them with caffeine intake (Bonnet 1991; Bonnet et al. 1995; Bonnet and Arand 1996).

However, does caffeine improve performance when ingested without a previous increase in sleep propensity? A study conducted on 36 healthy volunteers found no evidence of beneficial effects of caffeine on human performance (James 1998) although the subjects reported being more alert and feeling less tired when ingesting caffeine. The initially improved feeling of increased alertness disappeared when caffeine was used chronically (in the course of 6 days). It is possible that caffeine has only small, if any, beneficial effects on performance under sleep satiety but it is effective in improving performance under sleep loss. There are also methodological issues that cast doubts regarding the results on caffeine's effect on performance and mood (see Sect. 3.7 for details).

### ***3.6 Individual Sensitivity to Caffeine***

Human sensitivity to the effects of caffeine is variable. The basis of this variability has been widely debated. Physiological differences in the development of tolerance to the effects of caffeine may explain some of the differences (Colton et al. 1968; Curatolo and Robertson 1983; Bonnet and Arand 1992).

The plasma concentration of caffeine induced by ingestion of the same amount of caffeine by different people can vary between individuals by a factor of 15.9 (Birkett and Miners 1991), suggesting that slow metabolizers may be more sensitive to caffeine.

A genetic difference in caffeine sensitivity has been also suggested, and recently a distinct c.1083T > C polymorphism in the A<sub>2A</sub> receptor gene (ADORA<sub>2A</sub>) has been found to modulate individual sensitivity to subjective and objective effects of caffeine on sleep (Retey et al. 2007), giving a firm biological explanation at least partly for the differences in caffeine sensitivity (Landolt 2008). Furthermore,

another polymorphism in the same receptor gene, 1976T/T 2592 Tins/Tins polymorphism, induces differences in caffeine-induced anxiety: individuals with the polymorphism reported more anxiety after caffeine intake than individuals without the modification (Alsene et al. 2003).

There is also a large variability in how individuals respond to sleep deprivation: in some individuals a short deprivation period reduces performance, while others can maintain their performance level for prolonged periods of sleep loss (Haavisto et al. 2010). The relation of these individual differences to the adenosinergic system was assessed in a study where young volunteers stayed awake for one night. The subjects rated themselves as either caffeine-sensitive or caffeine-insensitive, and ingested either caffeine (200 mg) or a placebo twice during the waking period. In the placebo condition, those who had rated themselves as caffeine-sensitive showed a greater decrease in performance than those who had rated themselves as caffeine-insensitive. However, caffeine improved the performance particularly in caffeine-sensitive individuals (Hotta et al. 2009). The behavioral findings were supported by EEG findings: in those subjects in whom prolonged waking induced the largest increase in the frontooccipital power ratio, caffeine most potently reduced this ratio. In other words, there is negative association between the effects of sleep deprivation and caffeine. These findings suggest that adenosinergic mechanisms could at least partly explain differences in responses to sleep restriction.

### ***3.7 Sleep During Caffeine Withdrawal***

Caffeine induces dependency, even after a relatively short period of exposure with doses that are commonly used: withdrawal syndromes have been documented after three to seven daily uses of 100 mg caffeine (Juliano and Griffiths 2004). The withdrawal syndromes include headache, fatigue, drowsiness, decreased alertness, depressed mood, and irritability. Sleepiness commonly increases in the course of abstinence (Roehrs and Roth 2008).

The withdrawal syndrome introduces a serious confounding factor to human experiments with caffeine: most subjects habitually consume caffeine in variable daily amounts, but before the experiments they should stop caffeine use to ensure a zero level of caffeine in the blood. The half-life of caffeine is 3–7 h, while the withdrawal symptoms appear 12–24 h from the beginning of the abstinence. If overnight abstinence is used, the experiments start in the middle of the abstinence syndrome, including increased sleepiness. It has been argued that most of the reported effects of caffeine on mood and performance are the reversal of withdrawal symptoms (James and Keane 2007).

### ***3.8 Effects of Other Methylxantines on (Human) Sleep***

Methylxanthines include, in addition to caffeine, aminophylline, IBMX, paraxanthine, pentoxifylline, theobromine, and theophylline. The affinity of these substances

for adenosine receptors varies (see Chap. 6, in this volume), as well as their efficacy in promoting wakefulness. Common beverages, such as tea and chocolate, contain, in addition to caffeine, other methylxanthines. Tea contains caffeine at a level of about 3% of its dry weight and, in addition, small amounts of theobromine and theophylline. Dry tea has more caffeine by weight than coffee, but as more dry coffee is used than dry tea to prepare a beverage, a cup of brewed tea usually contains significantly less caffeine than a cup of coffee of the same size. Chocolate contains theobromine, which has physiological effects similar to those of caffeine.

## References

- Alsene K, Deckert J, Sand P, de Wit H (2003) Association between  $A_{2A}$  receptor gene polymorphism and caffeine-induced anxiety. *Neuropharmacology* 28:1694–1702
- Basheer R, Bauer A, Elmenhorst D, Ramesh V, McCarley RW (2007) Sleep deprivation upregulates  $A_1$  adenosine receptors in the rat basal forebrain. *Neuroreport* 18:1895–1859
- Basheer R, Strecker RE, Thakkar MM, McCarley RW (2004) Adenosine and sleep-wake regulation. *Prog Neurobiol* 73:379–396
- Battistuzzi G, Iudicone P, Santolamazza P, Petrucci R (1981) Activity of adenosine deaminase allelic forms in intact erythrocytes and in lymphocytes. *Ann Hum Genet* 45:15–19
- Benington JH, Heller HC (1995) Restoration of brain energy metabolism as the function of sleep. *Prog Neurobiol* 45:347–360
- Benington JH, Kodali SK, Heller HC (1995) Stimulation of  $A_1$  adenosine receptors mimics the electroencephalographic effects of sleep deprivation. *Brain Res* 692:79–85
- Birkett DJ, Miners JO (1991) Caffeine renal clearance and urine caffeine concentrations during steady state dosing. Implications for monitoring caffeine intake during sports events. *Br J Clin Pharmacol* 31:405–408
- Bonnet MH (1991) The effect of varying prophylactic naps on performance, alertness and mood throughout a 52-hour sustained operation. *Sleep* 14:307–315
- Bonnet MH, Arand DL (1992) Caffeine use as a model of acute and chronic insomnia. *Sleep* 15:526–536
- Bonnet MH, Arand DL (1996) Metabolic rate and the restorative function of sleep. *Physiol Behav* 59:777–782
- Bonnet MH, Balkin TJ, Dinges DF, Roehrs T, Rogers NL, Wesenten NJ (2005) The use of stimulants to modify performance during sleep loss: a review by the sleep deprivation and stimulant task force of the American Academy of Sleep Medicine. *Sleep* 28:1163–1187
- Bonnet MH, Gomez S, Wirth O, Arand DL (1995) The use of caffeine versus prophylactic naps in sustained performance. *Sleep* 18:97–104
- Borbely AA (1982) A two process model of sleep regulation. *Hum Neurobiol* 1:195–204
- Carter AJ, O'Connor CMJ, Ungerstedt U (1995) Caffeine enhances acetylcholine release in the hippocampus in vivo by a selective interaction with adenosine  $A_1$  receptors. *J Pharmacol Exp Ther* 273:637–642
- Chagoya de Sanchez V, Hernandez Munoz R, Suarez J, Vidrio S, Yanez L, Diaz Munoz M (1993) Day-night variations of adenosine and its metabolizing enzymes in the brain cortex of the rat—possible physiological significance for the energetic homeostasis and the sleep-wake cycle. *Brain Res* 612:115–121
- Colton T, Gosselin RE, Smith RP (1968) The tolerance of coffee drinkers to caffeine. *Clin Pharmacol Ther* 9:31–39
- Curatolo PW, Robertson D (1983) The health consequences of caffeine. *Ann Intern Med* 98:641–653

- Dunwiddie TV (1985) The physiological role of adenosine in the central nervous system. *Int Rev Neurobiol* 27:63–139
- Dunwiddie TV, Hoffer BJ, Fredholm BB (1981) Alkylxanthines elevate hippocampal excitability. Evident role of endogenous adenosine. *Naunyn Schmiedebergs Arch Pharmacol* 316:326–330
- Elmenhorst D, Meyer PT, Winz OH, Matusch A, Ermert J, Coenen HH, Basheer R, Haas HL, Zilles K, Bauer A (2009) Sleep deprivation increases A1 adenosine receptor binding in the human brain: a positron emission tomography study. *J Neurosci* 27:2410–2415
- Fernstrom MH, Bazil CW, Fernstrom JD (1984) Caffeine injection raises brain tryptophan level, but does not stimulate the rate of serotonin synthesis in rat brain. *Life Sci* 35:1241–1247
- Finelli LA, Baumann H, Borbely AA, Achermann P (2000) Dual electroencephalogram markers of human sleep homeostasis: correlation between theta activity in waking and slow-wave activity in sleep. *Neuroscience* 101:523–529
- Fredholm BB, Dunwiddie TV (1988) How does adenosine inhibit transmitter release? *Trends Pharmacol Sci* 9:130–134
- Fredholm BB, Bättig K, Holmén J, Nehling A, Zvartau EE (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fredholm BB, Jonzon B, Lindgren E (1984) Changes in noradrenaline release and in beta receptor number in rat hippocampus following long-term treatment with theophylline or L-phenylisopropyladenosine. *Acta Physiol Scand* 122:55–59
- Gass N, Porkka-Heiskanen T, Kalinchuk AV (2009) The role of the basal forebrain adenosine receptors in sleep homeostasis. *Neuroreport* 20:1013–1018
- Greene RW, Haas HL, Hermann A (1985) Effects of caffeine on hippocampal pyramidal cells in vitro. *Br J Pharmacol* 85:163–169
- Haavisto M-L, Porkka-Heiskanen T, Hublin C, Härmä M, Mutanen P, Muller K, Virkkala J, Sallinen M (2010) Sleep restriction for the duration of a work week impairs multitasking performance. *J Sleep Res* (epub ahead of printing)
- Hadfield MG, Milio C (1989) Caffeine and regional brain monoamine utilization in mice. *Life Sci* 45:2637–2644
- Hayaishi O (1988) Sleep-wake regulation of prostaglandin-D2 and prostaglandin-E2. *J Biol Chem* 263:19758
- Hayaishi O (2002) Molecular genetic studies on sleep-wake regulation, with special emphasis on the prostaglandin D2 system. *J Appl Physiol* 92:863–868
- Hotta H, Kagitani F, Kondo M, Uchida S (2009) Basal forebrain stimulation induces NGF secretion in ipsilateral parietal cortex via nicotinic receptor activation in adult, but not aged rats. *Neurosci Res* 63:122–8
- Huang ZL, Qu WM, Eguchi N, Chen JF, Schwarzschild MA, Fredholm BB, Urade Y, Hayaishi O (2005) Adenosine A<sub>2A</sub>, but not A<sub>1</sub>, receptors mediate the arousal effect of caffeine. *Nat Neurosci* 8:858–859
- Huston JP, Haas HL, Pfister M, Decking U, Schrader J, Schwarting RK (1996) Extracellular adenosine levels in neostriatum and hippocampus during rest and activity periods of rats. *Neuroscience* 73:99–107
- Höfer I, Bättig K (1994) Cardiovascular, behavioral, and subjective effects of caffeine under field conditions. *Pharmacol Biochem Behav* 48:899–908
- James JE (1998) Acute and chronic effects of caffeine on performance, mood, headache, and sleep. *Neuropsychobiology* 38:32–41
- James JE, Keane MA (2007) Caffeine, sleep and wakefulness: implications of new understanding about withdrawal reversal. *Hum Psychopharmacol* 22:549–558
- Juliano LM, Griffiths RR (2004) A critical review of caffeine withdrawal: empirical validation of symptoms and signs, incidence, severity and associated features. *Psychopharmacology* 176:1–29

- Kalinchuk AV, McCarley RW, Stenberg D, Porkka-Heiskanen T, Basheer R (2008) The role of cholinergic basal forebrain neurons in adenosine-mediated homeostatic control of sleep: lessons from 192 IgG-saporin lesions. *Neuroscience* 157:238–253
- Kalinchuk AV, Stenberg D, Rosenberg PA, Porkka-Heiskanen T (2006) Inducible and neuronal nitric oxide synthases (NOS) have complementary roles in recovery sleep induction. *Eur J Neurosci* 24:1–14
- Karacan I, Thornby JJ, Anch M, Booth GH, Williams RL, Salis PJ (1976) Dose-related sleep disturbances induced by coffee and caffeine. *Clin Pharm Ther* 20:682–689
- Krueger JM, Fang J (1997) Cytokines in sleep regulation. In: Hayaishi O, Inoue S (eds) *Sleep and sleep disorders: from molecule to behavior*. Academic/Harcourt Brace, Tokyo
- Landolt HP (2008) Sleep homeostasis: a role for adenosine in humans. *Biochem Pharmacol* 75:2070–2079
- Landolt HP, Dijk DJ, Gaus SE, Borbely AA (1995a) Caffeine reduces low-frequency delta activity in the human sleep EEG. *Neuropsychopharmacology* 12:229–238
- Landolt HP, Retey J, Tönz K, Gottselig JM, Khatami R, Buckelmüller I, Achermann P (2004) Caffeine attenuates waking and sleep electroencephalographic markers of sleep homeostasis in humans. *Neuropsychopharmacology* 29:1933–1939
- Landolt HP, Werth E, Borbely AA, Dijk DJ (1995b) Caffeine intake (200 mg) in the morning affects human sleep and EEG power spectra at night. *Brain Res* 675:67–74
- McGinty DJ, Szymusiak RS (2000) The sleep-wake switch: a neuronal alarm clock. *Nat Med* 6:510–511
- Murillo-Rodríguez E, Blanco-Centurion C et al (2004) The diurnal rhythm of adenosine levels in the basal forebrain of young and old rats. *Neuroscience* 123:361–370
- Murray TF, Blaker WD, Cheney DL, Costa E (1982) Inhibition of acetylcholine turnover rate in rat hippocampus and cortex by intraventricular injection of adenosine analogs. *J Pharmacol Exp Ther* 222:550–554
- Nicholson AN, Stone BM (1980) Heterocyclic amphetamine derivative and caffeine on sleep in man. *Br J Clin Pharmacol* 9:195–203
- Oishi Y, Huang ZL, Fredholm BB, Urade Y, Hayaishi O (2008) Adenosine in the tuberomammillary nucleus suppresses the histaminergic system via A<sub>1</sub> receptors and promotes non-rapid eye movement sleep. *Proc Natl Acad Sci USA* 105:19992–19997
- Okuma T, Matsuoka H, Yosihiko M, Toyomura K (1982) Model insomnia by methylphenidate and caffeine and use in the evaluation of temazepam. *Psychopharmacology* 76:201–208
- Phillis JW, Edstrom JP (1976) Effects of adenosine analogs on rat cerebral cortical neurons. *Life Sci* 19:1041–1053
- Phillis JW, Wu PH (1981) The role of adenosine and its nucleotides in central synaptic transmission. *Prog Neurobiol* 16:187–293
- Popoli P, Sagratella S, Scotti De Carolis A (1987) An EEG and behavioural study on the excitatory properties of caffeine in rabbits. *Arch Int Pharmacodyn Ther* 290:5–15
- Porkka-Heiskanen T, Strecker RE, Bjorkum AA, Thakkar M, Greene RW, McCarley RW (1997) Adenosine: a mediator of the sleep-inducing effects of prolonged wakefulness. *Science* 276:1265–1268
- Porkka-Heiskanen T, Strecker RE, McCarley RW (2000) Brain site specificity of extracellular concentration changes during sleep deprivation and spontaneous sleep: an in vivo microdialysis study. *Neuroscience* 99:507–517
- Rainnie DG, Grunze HC, McCarley RW, Greene RW (1994) Adenosine inhibition of mesopontine cholinergic neurons: implications for EEG arousal. *Science* 263:689–692
- Ram A, Pandey HP, Matsumura H, Kasahara-Orita K, Nakajima T, Takahata R, Satoh S, Terao A, Hayaishi O (1997) CSF levels of prostaglandins, especially the level of prostaglandin D<sub>2</sub>, are correlated with increasing propensity towards sleep in rats. *Brain Res* 751:81–89
- Retey JV, Adam M, Honegger E, Khatami R, Luhmann UFO, Jung HH, Landolt HP (2005) A functional genetic variation of adenosine deaminase affects the duration and intensity of deep sleep in humans. *Proc Natl Acad Sci USA* 102:15676–15681

- Retey JV, Adam M, Khatami R, Luhmann UFO, Jung HH, Berger W, Landolt HP (2007) A genetic variation in adenosine A<sub>2A</sub> receptor gene (ADORA<sub>2A</sub>) contributes to individual sensitivity to caffeine effects on sleep. *Clin Pharm Ther* 81:692–698
- Roehrs T, Roth T (2008) Caffeine: sleep and daytime sleepiness. *Sleep Med Rev* 12:153–162
- Saper CB, Chou TC, Scammell TE (2001) The sleep switch: hypothalamic control of sleep and wakefulness. *Trends Neurosci* 24:726–731
- Satoh S, Matsamura H, Suzuki F, Hayaishi O (1996) Promotion of sleep mediated by the A<sub>2a</sub>-adenosine receptor and possible involvement of this receptor in the sleep induced by prostaglandin D<sub>2</sub> in rats. *Proc Natl Acad Sci USA* 93:5980–5984
- Satoh S, Matsumura H, Koike N, Tokunaga Y, Maeda T, Hayaishi O (1999) Region dependent difference in the sleep-promoting potency of an adenosine A<sub>2A</sub> receptor agonist. *Eur J Neurosci* 11:1587–1597
- Schweitzer PK, Randazzo AC, Stone K, Erman M, Walsh JK (2006) Laboratory and field studies of naps and caffeine as practical countermeasures for sleep-wake problems associated with night work. *Sleep* 29:39–50
- Schwierin B, Borbely AA, Tobler I (1996) Effects of N<sup>6</sup>-cyclopentyladenosine and caffeine on sleep regulation in the rat. *Eur J Pharmacol* 300:163–171
- Shaw PJ, Cirelli C, Greenspan RJ, Tononi G (2000) Correlates of sleep and waking in *Drosophila melanogaster*. *Science* 287:1834–1837
- Sinton CM, Petitjean F (1989) The influence of chronic caffeine administration on sleep parameters in the cat. *Pharmacol Biochem Behav* 32:459–462
- Stenberg D (2007) Neuroanatomy and neurochemistry of sleep. *Cell Mol Life Sci* 64:1187–1204
- Stenberg D, Litonius E, Halldner L, Johansson B, Fredholm BB, Porkka-Heiskanen T (2003) Sleep and its homeostatic regulation in mice lacking adenosine A<sub>1</sub> receptor. *J Sleep Res* 12:283–290
- Strecker RE, Morairty SR, Thakkar MM, Porkka-Heiskanen T, Basheer R, Dauphin LJ, Rainnie DG, Greene RW, McCarley RW (2000) Adenosinergic modulation of basal forebrain and preoptic/anterior hypothalamic neuronal activity in the control of behavioral state. *Behav Brain Res* 115:183–204
- Thakkar MM, Winston S, McCarley RW (2003) A<sub>1</sub> receptor and adenosinergic homeostatic regulation of sleep-wakefulness: effects of antisense to the A<sub>1</sub> receptor in the cholinergic basal forebrain. *J Neurosci* 23:4278–4287
- Ticho SR, Radulovacki M (1991) Role of adenosine in sleep and temperature regulation in the preoptic area of rats. *Pharmacol Biochem Behav* 40:33–40
- Urade Y, Hayaishi O (1999) Prostaglandin D<sub>2</sub> and sleep regulation. *Biochim Biophys Acta* 1436:606–615
- Vyazovskiy VV, Tobler I (2005) Theta activity in the waking EEG is a marker of sleep propensity in the rat. *Brain Res* 1050:64–71
- Wigren H-K, Rytkönen K-M, Porkka-Heiskanen T (2009) Basal forebrain lactate release and promotion of cortical arousal during prolonged waking is attenuated in aging. *J Neurosci* 29:11698–11707
- Wu MN, Ho K, Crocker A, Yue Z, Seghal A (2009) The effects of caffeine on sleep in *Drosophila* require PKA activity, but not the adenosine receptor. *J Neurosci* 29:11029–11037
- Wurts SW, Edgar DM (2000) Caffeine during sleep deprivation: sleep tendency and dynamics of recovery sleep in rats. *Pharmacol Biochem Behav* 65:155–162
- Yanik G, Glaum S, Radulovacki M (1987) The dose-response effects of caffeine on sleep in rats. *Brain Res* 403:177–180
- Yokogawa T, Marin W, Faraco J, Pezeron G, Appelbaum L, Zhang J, Rosa F, Mourrain P, Mignot E (2007) Characterization of sleep in zebrafish and insomnia in hypocretin mutants. *PLoS Biol* 5:e277
- Zhdanova IV, Wang SY, Leclair OU, Danilova NP (2001) Melatonin promotes sleep-like state in zebrafish. *Brain Res* 903:263–268

# Methylxanthines and Reproduction

Alba Minelli and Ilaria Bellezza

## Contents

1	The Male Reproductive System .....	350
1.1	Spermatogenesis .....	351
1.2	Acquisition of Sperm Fertility .....	355
2	The Female Reproductive Tract .....	357
2.1	Oocyte Maturation .....	357
2.2	Caffeine and Female Fertility .....	361
3	Assisted Reproductive Techniques .....	364
3.1	Methylxanthines in Assisted Reproductive Techniques .....	364
3.2	Caffeine in Assisted Reproductive Techniques .....	365
4	Conclusion .....	366
	References .....	366

**Abstract** Reproduction is the process by which organisms create descendants. In human reproduction, two kinds of sex cells, or gametes, are involved. Sperm, the male gamete, and egg, or ovum, the female gamete, must meet in the female reproductive system to create a new individual and both the female and the male reproductive systems are essential to the occurrence of reproduction. Scientific reports dealing with the effects of methylxanthines on reproduction are mostly centred on the use of these compounds as phosphodiesterase inhibitors that, by maintaining high intracellular levels of cyclic AMP (cAMP), will affect the gametes differently. High cAMP levels will sustain sperm maturation while they hold the oocytes in mitotic arrest. Caffeine, being the methylxanthine most widely consumed by every segment of the population, has been the subject of greatest interest among health professionals and researchers. Conflicting results still seem to characterize the association between male/female caffeine consumption in adult life

---

A. Minelli (✉) and I. Bellezza

Dipartimento di Medicina Sperimentale e Scienze Biochimiche, Università degli Studi di Perugia,  
Via del Giochetto, 06123 Perugia, Italy  
e-mail: aminelli@unipg.it



and semen quality/fertility, although moderate daily caffeine consumption of levels up to 400–450 mg/day (5.7–6.4 mg/kg/day in a 70-kg adult) do not seem to be associated with adverse effects, i.e. general toxicity, effects on bone status and calcium balance, cardiovascular effects, behavioural changes, increased incidence of cancer, or effects on male fertility. A clear stimulation of egg-laying by the coffee leaf pest *Leucoptera coffeella* was recently reported, providing support for the hypothesis that caffeine, in a dose-dependent way, in insects stimulates egg-laying, thus leading to the death of coffee trees.

**Keywords** Acquisition of sperm fertility · Assisted reproductive techniques · Oocytes maturation · Spermatogenesis

## Abbreviation

AC	Adenylyl cyclase
AR	Acrosome reaction
cAMP	Cyclic AMP
Cdk1	Cyclin-dependant kinase
dbcAMP	Dibutyryl cyclic AMP
FSH	Follicle-stimulating hormone
GIFT	Gamete intra-Fallopian transfer
IBMX	3-Isobutyl-1-methylxanthine
IVF	In vitro fertilization
LH	Luteinizing hormone
LHR	Luteinizing hormone receptor
MI	Metaphase I
MPF	Maturation/meiosis or mitosis promoting factor
NOEL	No-observed-effect level
PDE	Phosphodiesterase
PKA	Protein kinase A
PKB	Protein kinase B
UPP	Ubiquitin-proteasome pathway

## 1 The Male Reproductive System

The male reproductive system consists of a number of sex organs that are part of the reproductive process. The testes are responsible for production of sperm and androgens, i.e. sex hormones essential to development and functional maintenance of the entire male reproductive tract. Each testis comprises two tissue compartments, which are functionally related but structurally separate: the seminiferous

tubule compartment, lined with a complex epithelium of highly specialized Sertoli cells and developing spermatogenic cells, and the interstitial tissue compartment, which contains the androgen-producing Leydig cells, as well as the testicular vasculature, lymphatic and immune cells. The seminiferous tubules are connected through a structure called the rete testis, via a series of efferent ducts, to the adjacent epididymis, which concentrates and facilitates the maturation of the sperm. At ejaculation, epididymal fluid and sperm are propelled along the muscular vas deferens to the urethra, where they are combined with the secretions of the accessory glands to form the seminal plasma.

### ***1.1 Spermatogenesis***

Spermatogenesis is the process by which male spermatogonia develop into mature spermatozoa in sexually reproducing organisms. In mammals this process occurs in the male testes and epididymis in a stepwise fashion, and for humans takes approximately 64 days (Heller and Clermont 1963). Starting at puberty, it usually continues uninterrupted until death, although a slight decrease can be discerned in the quantity of the sperm produced with increase in age. The entire process can be broken up into several distinct stages, i.e. spermatocytogenesis, spermatidogenesis, and spermiogenesis. The initial stages occur within the testes and progress to the epididymis, where the developing gametes mature and are stored until ejaculation. The seminiferous tubules of the testes are the starting point for the process, where stem cells adjacent to the inner tubule wall divide in a centripetal direction to produce immature sperm. Maturation occurs in the epididymis and involves the acquisition of a tail and hence motility.

In spermatocytogenesis, a diploid spermatogonium in the basal compartment of seminiferous tubules divides mitotically to produce two diploid intermediate cells called primary spermatocytes.

On the basis of the appearance of the nuclei, three functionally separate spermatogonial cell types are recognized: type A dark spermatogonia, type A pale spermatogonia, and type B spermatogonia. The population of spermatogonia is maintained by type A dark spermatogonia, which do not directly participate in producing sperm and simply ensure a supply of stem cells. Type A pale spermatogonia repeatedly divide mitotically to produce identical cell clones. When repeated division ceases, the cells differentiate into type B spermatogonia. This stage is referred to as the spermatogonial phase. Type B spermatogonia undergo mitosis to produce diploid primary spermatocytes.

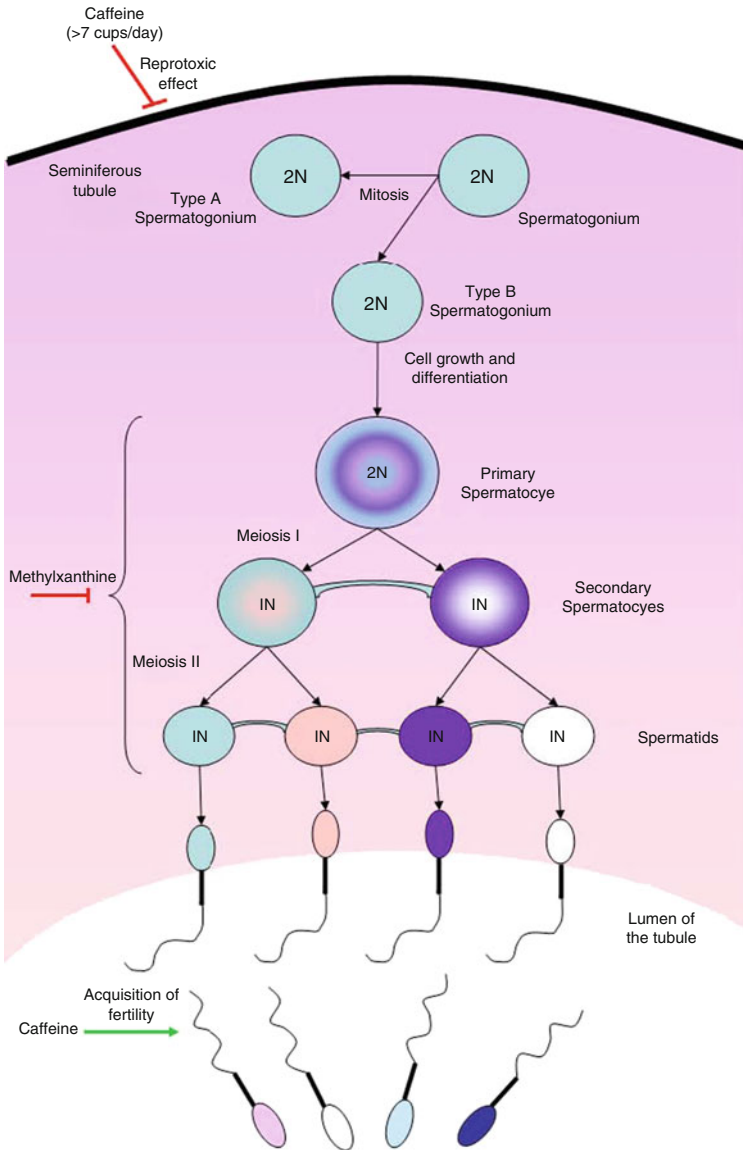
Each primary spermatocyte then moves into the adluminal compartment of the seminiferous tubules, duplicates its DNA and subsequently undergoes meiosis I to produce two secondary spermatocytes. This division implicates sources of genetic variation, such as random inclusion of either parental chromosome, and chromosomal crossover, to increase the genetic variability of the gamete.

Secondary spermatocytes rapidly enter meiosis II and divide to produce haploid spermatids in spermatidogenesis. Owing to the brevity of this stage, secondary spermatocytes are rarely seen in histological preparations. Each cell division from a spermatogonium to a spermatid is incomplete; the cells remain connected to one another by bridges of cytoplasm to allow synchronous development.

During spermiogenesis, the spermatids begin to grow a tail, and develop a thickened mid-piece, where the mitochondria gather and form an axoneme. Spermatid DNA also undergoes packaging, becoming highly condensed. The DNA is packaged firstly with specific nuclear basic proteins, which are subsequently replaced with protamines during spermatid elongation. The resultant tightly packed chromatin is transcriptionally inactive. The Golgi apparatus surrounds the now condensed nucleus, becoming the acrosome. One of the centrioles of the cell elongates to become the tail of the sperm. Maturation then takes place under the influence of testosterone, which is involved in the removal of the remaining unnecessary cytoplasm and organelles. The excess cytoplasm, known as residual bodies, is phagocytosed by surrounding Sertoli cells in the testes. The resulting spermatozoa are now mature but lack motility, rendering them sterile. The mature spermatozoa are released from the protective Sertoli cells into the lumen of the seminiferous tubule in a process called spermiation. The non-motile spermatozoa are transported to the epididymis in testicular fluid secreted by the Sertoli cells by peristaltic contraction. While residing in the epididymis, they acquire motility and become capable of fertilization. However, transport of the mature spermatozoa through the remainder of the male reproductive system is achieved via muscle contraction rather than the spermatozoon's recently acquired motility. During spermatogenesis, Sertoli cells support the developing gamete by maintaining the environment necessary for development and maturation via the blood–testis barrier, secreting the substances initiating meiosis, the androgen-binding protein, and inhibin, by phagocytosing residual cytoplasm left over from spermiogenesis (Fig. 1).

### 1.1.1 Methylxanthines and Spermatogenesis

Studies on the teratogenic or sperm-injuring potential of methylxanthine started nearly 40 years ago by investigating the effects of caffeine on spermatogenesis (Ax et al. 1976). It is to note that all animal studies have demonstrated that, depending on the method of administration and the species, the developmental no-observed-effect level (NOEL) is approximately 30 mg/kg per day, the teratogenic NOEL is 100 mg/kg per day, and the reproductive NOEL approximately 80–120 mg/kg per day (Christian and Brent 2001). Roosters, fed 0.1% caffeine mixed by weight into a standard ration (about 145 mg/day), after 14 days of treatment showed a significant decrease in fertility. Semen output and sperm concentration were markedly reduced after 17–21 days of treatment, and no semen could be collected from the roosters after a 30-day treatment. Testicular histological investigation showed interruption of spermatocyte divisions and abnormal spermiogenesis, but removal of dietary



**Fig. 1** Spermatogenesis and site of action of methylxanthines

caffeine resulted in resumption of semen production and a return of fertility to the control level. In rats fed caffeine, theobromine, or theophylline at a very high dietary level of 0.5% by weight into a standard ration ( $LD_{50} = 200$  mg/kg, Fredholm et al. 1999) for periods ranging from 14 to 75 weeks, a significant positive finding was the occurrence of severe bilateral testicular atrophy with aspermatogenesis or oligospermatogenesis in 85–100% of the rats (Weinberger et al. 1978;

Friedman et al. 1979). The relative testicular toxicity of the methylxanthines was reported as caffeine being the most potent, theobromine slightly less potent, and theophylline considerably less potent. Somewhat variable atrophic changes of the accessory sexual organs (epididymis, prostate, and seminal vesicles) accompanied the testicular changes. Cytogenetic analysis of testes from caffeine- or theophylline-fed rats revealed a significantly reduced number of mitotic cells in the caffeine-treated group. Plasma testosterone concentrations were significantly elevated in the theobromine group and were elevated in the caffeine-treated group; this correlated morphologically with an apparent hyperplasia of interstitial cells in severely atrophied testes in these groups. Plasma cholesterol concentrations were significantly increased in the caffeine and theobromine groups (Gans 1982; Ettlin et al. 1986; Ezzat and El-Gohary 1994; Funabashi et al. 2000). Studies of the toxicities of theobromine and cocoa extract on the reproductive tract of male rats showed that theobromine and high-dose cocoa extract caused vacuolation within the Sertoli cell, abnormally shaped spermatids, and failed release of late spermatids in treated animals. However, the frequencies of some parameters of testis alterations were significantly lower in the high-dose cocoa-extract-treated group compared with the theobromine-treated group, demonstrating the ability of a cocoa extract containing theobromine to alter testis structure in a similar pattern but with reduced intensity compared with that observed after oral exposure to pure theobromine (Wang et al. 1992; Wang and Waller 1994).

The effects of caffeine at a concentration of 0.5% and fed to male rats for 7 weeks were compared with those of 0.8% dietary theobromine. Both dietary methylated xanthines produced significant decreases in food consumption and body-weight gain in rats when compared with their respective control groups. The theobromine-fed rats showed severe testicular atrophy with extensive spermatogenic cell degeneration and necrosis, while the testes of rats fed caffeine showed only scattered vacuolar degeneration of spermatogenic cells. Caffeine appeared to be more potent than theobromine as an anorexic agent in rats, but to be equivalent to theobromine in its potential for inducing thymic atrophy and spermatogenic cell destruction with testicular atrophy (Gans 1984). Long-term intake of caffeine caused suppression of spermatogenesis mainly through inhibition of the release of follicle-stimulating hormone (FSH). Daily administration of caffeine (30 or 60 mg/kg) to mature male rabbits for four consecutive weeks caused an increase in the plasma FSH level and a decrease in the luteinizing hormone (LH) level. A light microscope study revealed reduced size of the seminiferous tubules, inhibition of spermatogenesis, fatty degeneration of the liver, and hepatic lesions, whereas the adrenal glands exhibited signs of stimulated steroidogenesis (Ezzat and El-Gohary 1994). Caffeine, when administered to the rat (30 mg/kg/day) during pregnancy, affected certain aspects of normal sexual differentiation of the fetal gonads (Pollard et al. 1990). In the male fetus, caffeine significantly inhibited differentiation of the interstitial tissue and Leydig cells, with a significant consequent reduction in testosterone biosynthesis in the fetal testes. Caffeine also had an effect on the earlier morphogenic organization of the seminiferous cords. In the female fetus, caffeine did not modify ovarian differentiation nor the morphology of

the ovaries, tissue arrangement, and overall appearance. These results indicating that caffeine, when administered during pregnancy, significantly inhibited the differentiation of the seminiferous cords and subsequent Leydig cell development in the interstitium, prompted the investigation of whether the observed effects were caused either by direct effects of caffeine or by intermediary secondary toxic effects of metabolites, i.e. theophylline and theobromine. Explants of 13-day-old fetal testis were cultured for 4 days *in vitro* in the presence of graded doses of caffeine, theophylline, or theobromine. Fetal testes exposed to caffeine or theobromine differentiated normally, developing seminiferous cords made up of Sertoli and germ cells, soon followed by the differentiation of functionally active Leydig cells appearing in the newly formed interstitium. In contrast, explants exposed to theophylline failed to develop seminiferous cords and, as a consequence, Leydig cells (Pollard et al. 2001). Recently, theophylline was shown to induce infertility by causing germ cell apoptosis in the testicular seminiferous epithelium. Theophylline exposure altered the expression of the genes within the ubiquitin–proteasome pathway (UPP), implicated in spermatogenesis and epididymal sperm quality control (Tengowski et al. 2007). Results suggest that the reprotoxic exposure alters the tissue-specific expression of UPP genes in the testis and epididymis, which may contribute to the aberrant spermatogenesis and epididymal processing of both normal and defective spermatozoa. Moreover, theophylline induced infertility by incapacitating the nurturing Sertoli cells, thus resulting in the premature release of late-differentiating spermatogenic cells, round spermatids (Weinberger et al. 1978). This leads to the depletion of spermatids and mature spermatozoa from the adluminal compartment of the seminiferous epithelium, ultimately causing testicular atrophy (Strandgaard and Miller 1998; Tengowski et al. 2005).

Other authors have suggest a beneficial effect of caffeine in the regulation of male gamete maturation since, acting as an agonist of ryanodine receptors, which induce release of  $\text{Ca}^{2+}$  from intracellular stores in spermatogonia, pachytene spermatocytes, and round spermatids, caffeine modulates calcium mobilization and plays a fundamental role in spermatozoa maturation (Chiarella et al. 2004). Conflicting results still seem to characterize the association between male caffeine consumption in adult life and semen quality, whereas the association between prenatal coffee consumption and semen quality and levels of reproductive hormones seems to be responsible for a small to moderate effect on semen volume and the levels of reproductive hormones (Ramlau-Hansen et al. 2008) .

## ***1.2 Acquisition of Sperm Fertility***

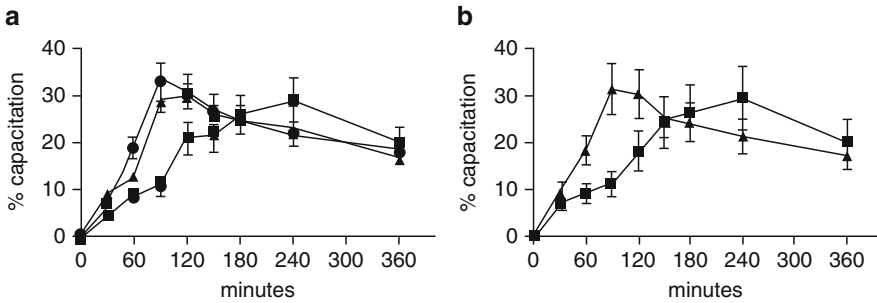
Mammalian spermatozoa emerging from the male reproductive tract are incapable of fertilizing eggs. They acquire this ability either during transit in the female reproductive tract (Yanagimachi 1994) or during incubation in suitable *in vitro* media (Allegrucci et al. 2001). Such conditioning, called capacitation, renders the spermatozoa capable interacting with the oocyte and thereby inducing the acrosome

reaction (AR). Capacitation and AR are related to many effectors and signal transduction pathways, but the molecular basis of these processes is still to be fully elucidated (Kopf and Wilde 1990; Florman et al. 1992; de Lamirande et al. 1997; Tulsiani et al. 1998, 2007; Thundathil et al. 2002; Morales et al. 2007; Salicioni et al. 2007; Aitken et al. 2007; Wassarman 2009; Abou-haila and Tulsiani 2009). Sperm capacitation is a multistep process that involves several biochemical and ultrastructural changes in the sperm membrane, ranging from modification of membrane lipid composition to an increased permeability to ions. The efflux of membrane cholesterol leads to bovine sperm capacitation (Visconti et al. 1999). Albumin, high-density lipoproteins, and follicular and oviductal lipoproteins are capacitation effectors of human and bovine spermatozoa (Moreau et al. 1998; Thérien et al. 2001). Capacitation is correlated with an increase of protein tyrosine phosphorylation (Visconti et al. 1995a, b, 1999; Aitken et al. 1998) and the signal transduction pathway leading to protein tyrosine phosphorylation is thought to be central to either the attainment of the capacitative state (Visconti and Kopf 1998) or the concomitant expression of hyperactivated motility (Mahony and Gwathmey 1999; Si and Okuno 1999). As capacitation proceeds, several proteins undergo serine/threonine phosphorylation or threonine/tyrosine double phosphorylation (Thundathil et al. 2002). The AR is an exocytotic process by which lytic enzymes are released from the sperm acrosome and digest the zona pellucida so that spermatozoa can reach and fertilize the oocyte (Yanagimachi 1994; de Lamirande et al. 1997). Sperm AR occurs within minutes, cannot be reversed once it is induced, and can be triggered *in vitro* by different inducers, such as zona pellucida (Yanagimachi 1994), progesterone (Harrison et al. 2000), calcium ionophores, lysophosphatidylcholine (de Lamirande et al. 1997), follicular fluid (De Jonge et al. 1993), and ATP (Luria et al. 2002). Sperm AR takes place after fusion between the acrosome and the overlying plasma membrane and involves calcium influx, actin polymerization, a rise in intracellular pH, and protein activation (phospholipases, kinases, G proteins, etc.) (Yanagimachi 1994; Baldi et al. 2000; Liguori et al. 2005; Abou-Haila and Tulsiani 2009).

### 1.2.1 Methylxanthines and Acquisition of Sperm Fertility

The greatest part of the data in the literature reports the use xanthines/methylxanthines as phosphodiesterase (PDE) inhibitors that maintain intracellular levels of cyclic AMP (cAMP), thereby acting as motility-enhancing agents or capacitating effectors (Hong et al. 1981; Jiang et al. 1984; Depeiges and Dacheux 1985; Galantino-Homer et al. 1997; Leclerc et al. 1998; Jaiswal and Majumder 1998; Harayama et al. 1998; Leclerc and Goupil 2002; Buffone et al. 2005; Lachance et al. 2007; Yeste et al. 2008).

In a study finalized to clarify the role of the adenosine A<sub>1</sub> receptor in the acquisition of fertilizing capacity, caffeine was used as an adenosine A<sub>1</sub> receptor antagonist at low concentrations that binds to and inhibits half of the adenosine receptors (Fredholm et al. 1999). This dose of caffeine had very little, if any, effect



**Fig. 2** **a** Percentage of capacitated cells as function of incubation time in  $A_1R^{+/+}$  (filled circle),  $A_1R^{-/-}$  (filled triangle), and  $A_1R^{-/-}$  (filled square) mouse spermatozoa (filled square). **b** Percentage of capacitated cells as function of incubation time of  $A_1R^{+/+}$  murine sperm in the presence of 15  $\mu M$  caffeine (filled triangle) and 100  $\mu M$  caffeine. (Adapted from Minelli et al. 2004)

on the acquisition of the capacitated status, whereas a very high dose of caffeine caused a significant reduction in sperm capacitation (Minelli et al. 2004) (Fig. 2). However, it is of note that high concentrations of caffeine are unlikely to be reached by caffeine consumers since strong side effects would preclude ingestion of this amount. Hence, the data suggest that regular caffeine consumption is unlikely to significantly affect spermatozoa function, thereby reassuring all coffee drinkers of the lack of negative effects of caffeine on male fertility. The scientific literature dealing with the AR contains references to the use of methylxanthines as PDE inhibitors (Kopf et al. 1983a, b; Carr and Acott 1990; Ain et al. 1999; Lachance et al. 2007) that maintain intracellular cAMP levels and induce the AR.

## 2 The Female Reproductive Tract

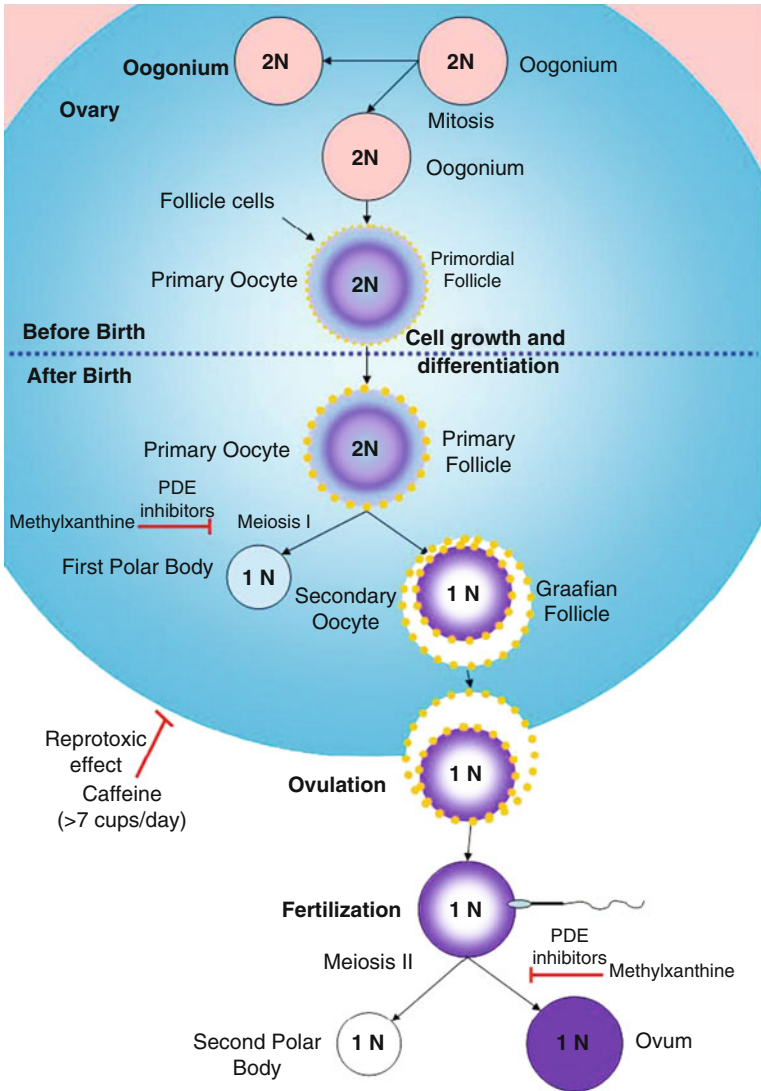
The female reproductive tract is a dynamic system cycling under the control of the key ovarian steroid hormones oestrogen and progesterone. It contains the uterus, which act as the receptacle for male sperm, and the ovaries, which produce the egg cells. The Fallopian tubes attach the uterus to the ovaries, which, at certain intervals, release an ovum, which passes through the Fallopian tube into the uterus. If, in this transit, the ovum meets with sperm, the sperm penetrate and merge with the egg, fertilizing it. The diploid zygote then implants itself in the wall of the uterus, where it begins the processes of embryogenesis and morphogenesis.

### 2.1 Oocyte Maturation

Despite the universal requirement of a haploid gamete for sexual reproduction, meiosis is regulated differently in oocytes and spermatocytes. As shown in Fig. 1, spermatocytes proceed through the meiotic divisions uninterrupted, whereas in



female mammals, meiosis occurs over a prolonged period of time (Eppig et al. 2004). Mammalian oocytes are engaged in a complex meiotic cell division, characterized by several “stops and starts”, and after resuming meiosis, they rely on maternal factors to sustain the subsequent developmental steps until the maternal-to-zygotic transition occurs. The process by which the oocyte completes the first meiotic division and undergoes other cytoplasmic changes and progresses to metaphase II is called oocyte maturation (Fig. 3). Because the mature, fertilizable oocyte



**Fig. 3** Oogenesis and site of action of methylxanthines

has a relatively short lifespan in the female reproductive tract, the timing of oocyte meiotic arrest, as well as maturation, is tightly regulated (Mehlmann 2005). The functional unit within the ovary is the follicle, formed during embryonic development, and it comprises one or more layers of granulosa cells surrounding the oocyte (Gougeon 1996; Zeleznik 2004). During follicular growth, the somatic cells divide to form several layers, the oocyte enlarges, and a fluid-filled antrum begins to form. Some follicles at the early antral stage are “recruited” to continue growing; this growth is dependent on the pituitary gonadotropin FSH (Gougeon 1996; Zeleznik 2004). During this phase, the antrum divides the granulosa cells into two separate compartments: mural granulosa cells form the outer layers, while the cumulus cells surround the oocyte. The oocyte grows to its full size (about 75- $\mu\text{m}$  diameter in the mouse, about 100  $\mu\text{m}$  in the human), but remains arrested in prophase I. If an oocyte is removed from an antral follicle, it spontaneously resumes meiosis and progresses to second metaphase (Pincus and Enzmann 1935). This indicates that the follicle cells hold the oocyte in prophase arrest. Meiosis resumes in response to a surge of LH from the pituitary gland during the oestrous or menstrual cycle, shortly before ovulation. LH receptors (LHRs) are located on the mural granulosa cells but not on the cumulus cells or the oocyte (Peng et al. 1991; Richards et al. 2002), so the mechanism(s) by which LH stimulates oocyte maturation is indirect. Prior to the midcycle surge of LH, the growing oocyte acquires the ability to undergo oocyte maturation. The acquisition of meiotic competence occurs around the time of antrum formation (Mehlmann et al. 2004) and corresponds to a point at which the oocyte achieves a threshold level of maturation-promoting proteins, such as cyclin-dependant kinase (Cdk1) and cyclin B (Kanatsu-Shinohara et al. 2000). Meiotic arrest is regulated by cAMP levels within the oocyte (Conti et al. 2002; Eppig et al. 2004) since the cAMP level affects the activity of the Cdk/cyclin B protein complex, also known as maturation/meiosis or mitosis promoting factor (MPF). High cAMP levels result in the phosphorylation of Cdk1 on Thr14 and Tyr15, rendering it inactive (Duckworth et al. 2002), while a decrease in cAMP levels leads to the dephosphorylation of Cdk1 on Thr14 and Tyr15, the MPF complex becomes active, and the oocyte can re-enter meiosis. A hypothesis for how high levels of cAMP are maintained in competent, fully grown oocytes is that the oocyte produces its own cAMP through a G-protein-linked receptor in the oocyte plasma membrane that stimulates  $G_s$  and, subsequently, adenylyl cyclase (AC). Direct evidence for an essential role of  $G_s$  in the maintenance of meiotic arrest was obtained by microinjecting either a function-blocking antibody or a dominant negative form of the  $\alpha$ -subunit of  $G_s$  into follicle-enclosed oocytes (Mehlmann et al. 2002; Kalinowski et al. 2003). This pathway was confirmed by the finding that oocytes from mice lacking the AC3 AC isoform, which is present in the oocyte, spontaneously undergo germinal vesicle breakdown within ovarian follicles (Horner et al. 2003).

Recently, it was shown that heat shock transcription factor 1, which triggers the transcription of several genes encoding heat shock proteins, is highly expressed in oocytes and plays an important role in normal progression of meiosis by directly regulating Hsp90 $\alpha$  expression (Metchat et al. 2009).

### 2.1.1 Methylxanthines and Oocyte Maturation

As for male gametes, the largest part of the data in the literature reports the use xanthines/methylxanthines as PDE inhibitors that maintain the intracellular levels of cAMP responsible for the meiotic arrest. Several groups (Cho et al. 1974; Dekel and Beers 1978; Schultz et al. 1983a, b; Vivarelli et al. 1983; Bornslaeger and Schultz 1985; Törnell et al. 1990; Haider and Chaube 1996; Webb et al. 2000; Conti et al. 2002) showed that spontaneous maturation of oocytes isolated from their follicles can be prevented by including membrane-permeant cAMP analogues or cAMP PDE inhibitors, such as hypoxanthine and 3-isobutyl-1-methylxanthine (IBMX), in the culture medium. The cAMP levels decrease in oocytes following their removal from their follicles as well as in isolated oocytes after removal of IBMX. The decrease in the level of oocyte cAMP occurs within 2 h after washing out IBMX, a time during which the oocyte becomes committed to resuming meiosis and cAMP levels increase in isolated oocytes. When transported into oocytes from the cumulus cells via gap junctions, cAMP also plays an important role in the regulation of meiotic progression beyond the meiosis I (MI) stage (Shimada et al. 2002). Using selective PDE inhibitors, such as milrinone (a PDE3 inhibitor), cilostamide (a PDE3 inhibitor), and rolipram (a PDE4 inhibitor), studies focused on the differential regulation of cAMP levels within the oocyte and somatic (cumulus) cell compartments of the follicle showed that specific PDE subtypes are differentially localized within the two compartments of the follicle, i.e. the type 3 PDE in the oocyte and the type 4 PDE in the granulosa cells (Thomas et al. 2002). Moreover, oocyte cAMP levels are primarily regulated in oocytes by its degradation by PDE, whereas granulosa cell cAMP levels are controlled mainly by active AC, with both sources able to participate in oocyte meiotic regulation. IBMX does not interfere with the expression of LHR in cumulus cells surrounding oocytes, whereas the binding of LH to its receptor induces a further increase in cAMP level, progesterone production, and acceleration of meiotic progression to the metaphase I stage. The role of cAMP in the oocyte meiotic arrest was further supported by Laforest et al. (2005), who showed a fundamental significance of cAMP pathways in controlling meiotic resumption in porcine oocytes. This control is at two levels, the ability to synthesize cAMP via active AC, where the cyclase of porcine oocyte is sensitive to forskolin, and the degradation of cAMP via cilostamide-sensitive PDE. A more detailed study of the effects of IBMX on the oocyte meiotic block (Barretto et al. 2007) showed that IBMX is able to prevent resumption of meiosis by maintaining elevated cAMP concentrations in the oocyte, whereas roscovitine, a purine known to specifically inhibit MPF kinase activity, maintains bovine oocytes at the germinal vesicle stage, indicating that the meiotic inhibitors delay the progression of nuclear maturation without affecting cytoplasmic maturation. It was proposed that the inhibitory cAMP is synthesized within oocytes via a stimulatory  $\alpha$ -subunit of G protein. After the presence of  $G_s$ - $\alpha$  molecules in porcine oocytes had been shown, an anti- $G_s$ - $\alpha$  antibody was injected into porcine immature oocytes and this inhibition of ooplasmic  $G_s$ - $\alpha$  functions significantly promoted germinal vesicle breakdown of the oocytes, whose spontaneous meiotic resumption was prevented by IBMX treatment. Moreover, although cyclin B synthesis and

MPF activation were largely prevented until 30 h of culture in IBMX-treated oocytes, injection of anti- $G_s$ - $\alpha$  antibody into these oocytes partially recovered cyclin B synthesis and activated MPF activity at 30 h, suggesting that meiotic resumption of porcine oocytes is prevented by ooplasmic  $G_s$ - $\alpha$ , which may stimulate cAMP synthesis within porcine oocytes, and that synthesized cAMP prevents meiotic resumption of oocytes through the signalling pathways involved in MPF activation (Morikawa et al. 2007). More recently, Ozawa et al. (2008) focused their attention on cAMP content, gap-junctional communication status, and LHR expression in porcine cumulus–oocyte complexes treated with IBMX or with FSH. They found that the inhibition of PDEs in porcine cumulus–oocyte complexes makes the oocyte ready for release from meiotic arrest, whereas the maintenance of a moderate cAMP content may prolong gap-junctional communications and stimulate LHR expression. A recent paper (Pirino et al. 2009) showed that meiotic resumption requires activation of the MPF. Protein kinase A (PKA) activity sustains the prophase arrest by inhibiting Cdk1. Therefore, the inhibition of the activity of the Cdc25 protein required for MPF activation results in mitotic arrest. Phosphorylation of a highly conserved serine 321 residue of Cdc25B 21 plays a key role in the negative regulation and localization of Cdc25B during prophase arrest, suggesting that Cdc25B is a direct target of PKA.

## 2.2 *Caffeine and Female Fertility*

Caffeine is a subject of interest among consumers and health professionals because it is widely consumed in the diet by most segments of the population and can exert several pharmacological effects (Dews 1982; Fredholm 1995; Christian and Brent 2001; Mandel 2002; Derbyshire and Abdula 2008; Yu et al. 2009).

The medical literature contains many varied references indicating that human adverse reproductive/developmental effects are produced by caffeine. Although it is difficult to compare doses of caffeine in animals and humans, the medical literature dealing with developmental and reproductive risks of caffeine underwent a thorough revision by evaluating the biological plausibility of the epidemiological and animal findings. When comparing effects among different species, one can only accomplish dose equivalence by considering the results of pharmacokinetics studies, metabolic studies, and dose-response investigations in the human and the species being studied. Moreover, the importance of dose within a species is of fundamental concern when determining developmental risks since most drugs/chemicals are potentially associated with developmental toxicity/teratogenicity only at some exposure level. The genetic constitution of an organism, i.e. both the maternal and the fetal genotypes, is also an important factor in the susceptibility of a species. Indeed, more than 30 disorders of increased sensitivity to drug toxicity or effects have been reported in the human owing to an inherited trait (McKusick 1988). Unlike human epidemiology studies, which are difficult to control and with multiple inherent flaws that prevent identification of causality, animal studies are conducted under conditions in which all the variables can be better controlled.

In addition, current non-clinical studies generally include identification of achieved blood levels/exposures in the maternal animal and the developing offspring, a critical factor because the severity of the effect is related to the ability of the conceptus to recover from the insult (Johnson and Christian 1984). Nevertheless, results of non-clinical animal studies provide excellent tools for predicting potential effects of caffeine on human reproduction and development. Indeed the LD<sub>50</sub> of caffeine is fairly consistent across species, including *Homo sapiens* (Dews, 1982). The plasma level resulting from 1.1 mg/kg caffeine (a single cup of coffee containing 80 mg of caffeine ingested by a 70-kg human) ranges from 0.5 to 1.5 mg/L. A similar dose–concentration relationship is found in many species, including rodents and primates (Hirsh 1984). It is generally assumed that 10 mg/kg in a rat represents about 250 mg of caffeine in a human weighing 70 kg (3.5 mg/kg), and that this would correspond to about two to three cups of coffee.

Nawrot et al. (2003) reviewed the effects of caffeine on human health and concluded that for the healthy adult population, moderate daily caffeine consumption at levels up to 400–450 mg/day (5.7–6.4 mg/kg/day in a 70-kg adult, equivalent to four to five cups per day) was not associated with adverse effects, which include general toxicity, effects on bone status and calcium balance, cardiovascular effects, behavioural changes, increased incidence of cancer, and effects on male fertility. However, the authors also reported that children and women of reproductive age were “at risk” subgroups who might require dietary advice to moderate their caffeine intake. High levels of caffeine intake may delay conception among fertile women (Bolúmar et al. 1997). The effects of caffeine consumption on delayed conception were evaluated in a European multicentre study on risk factors of infertility in a randomly selected sample of 3,187 women aged 25–44 years. A significantly increased odds ratio for subfecundity in the first pregnancy was observed for women drinking more than 500 mg of caffeine per day (more than six cups), the effect being relatively stronger in smokers than in non-smokers. Women with the highest level of consumption had an increase in the time leading to the first pregnancy. In addition, women whose caffeine consumption was high had less than a third of the risk for a long menses (8 days or more) compared with women who did not consume caffeine. Those whose caffeine consumption was high also had a doubled risk for a short cycle length (24 day or less); this association was also evident in those whose caffeine consumption was high but did not smoke. However, caffeine intake was not strongly related to an increased risk for anovulation, short luteal phase (10 days or less), long follicular phase (24 days or more), long cycle (36 days or more), or measures of within-woman cycle variability (Fenster et al. 1999). The mean birth weight was reduced by high reported caffeine consumption, but this small decrease in birth weight, observed for maternal caffeine consumption, is unlikely to be clinically important except for women consuming 600 mg of caffeine daily (more than 7.2 cups) (Bracken et al. 2003). More recently, in a study finalized to determine whether smoking, alcohol, and caffeine may be related to the four indicators of ovarian age, i.e. antral follicle count, FSH, inhibin B, and oestradiol, and therefore to fecundability and fertility (Kinney et al. 2007), 188 women, aged 22–49, were investigated and least-squares was regression used

to estimate differences in antral follicle count and hormone levels for women who smoke cigarettes or who drink alcohol or caffeine. Current smoking is related to elevated FSH levels, but not to the antral follicle count, inhibin B, or oestradiol. Neither alcohol nor caffeine was found to be related to any ovarian age indicator, suggesting that caffeine, at the dosage of 156 mg/day (1.9 cups), does not affect ovarian age indicators. On the basis of data from a large retrospective epidemiology study and from a large retrospective case-control study in humans, it appears that use of caffeine does not impair ovulation to the point of decreasing fertility and during pregnancy has little, if any, effect on the outcome of pregnancy. Nevertheless, although caffeine use during pregnancy does not appear to be associated with substantial risk and the association between soft drinks and ovulatory disorder infertility does not seem to be attributable to their caffeine content, most clinicians recommend that pregnant women limit their consumption of foods, beverages, and drugs containing caffeine, since caffeine crosses the placenta (Care Study Group 2008; Chavarro et al. 2009). Recent research (Björklund et al. 2008) has confirmed the concerns about caffeine consumption during pregnancy or the early postnatal period because there may be long-lasting behavioural changes after caffeine exposure early in life. Indeed, pregnant wild-type mice, given modest doses of caffeine (0.3 g/L in drinking water), gave birth to offspring that as adults exhibited increased locomotor activity in an open field. The offspring also responded to cocaine challenge with greater locomotor activity than mice not perinatally exposed to caffeine. The same behavioural experiments on mice heterozygous for adenosine A<sub>1</sub> receptor gene, where signalling via adenosine A<sub>1</sub> receptors is reduced to about the same degree as after modest consumption of caffeine, showed a behavioural profile similar to that of wild-type mice perinatally exposed to caffeine. It appeared that the mother's genotype was critical for behavioural changes in adult offspring, suggesting that perinatal caffeine, by acting on adenosine A<sub>1</sub> receptors in the

**Table 1** Effects of male genotype

Parameter	A <sub>1</sub> R +/+	A <sub>1</sub> R +/-	A <sub>1</sub> R -/-
In vivo fertility <sup>a</sup>			
Average number of pups	8 ± 2.1	8 ± 1.9	5 ± 1.7
Birthweight (g)	2.07 ± 0.8	2.05 ± 0.6	2.08 ± 0.4
Days between litters	45 ± 8	47 ± 8	53 ± 15
Reproductive parameters <sup>b</sup>			
Number of spermatozoa	13 × 10 <sup>6</sup> ± 2 × 10 <sup>6</sup>	11 × 10 <sup>6</sup> ± 3 × 10 <sup>6</sup>	12 × 10 <sup>6</sup> ± 1 × 10 <sup>6</sup>
Viability (%)	80 ± 10	76 ± 7	77 ± 9
Motility (%)	75 ± 7	72 ± 9	71 ± 11
Phenotype <sup>c</sup>			
Weight of adult animal (g)	33 ± 5	31 ± 7	32 ± 4
Weight of testis (mg)	132 ± 21	121 ± 15	122 ± 18

Adapted from Minelli et al. (2004)

<sup>a</sup>The values are the means ± the standard error of the mean (SEM) of 40 litters.

<sup>b</sup>The values are the means ± SEM of 20 male mice, *P* < 0.05.

<sup>c</sup>The values are the means ± SEM of 20 male mice, *P* < 0.05.

mother, causes long-lasting behavioural changes in the offspring that even manifest themselves in the second generation. Mice homozygous for genetic deletion of the adenosine A<sub>1</sub> receptor showed a reduction in number of offspring and increased time between litters (Table 1) (Minelli et al. 2004). Interestingly, caffeine and some of its derivatives present in coffee leaves affect egg-laying by the coffee leaf miner *Leucoptera coffeella*, one of the main coffee pests in the Neotropical region. In fact, increased leaf levels of caffeine favour egg-laying by the coffee leaf miner with a significant concentration–response relationship, providing support for the hypothesis that caffeine stimulates egg-laying by the coffee leaf miner in coffee leaves (Magalhães et al. 2008).

### 3 Assisted Reproductive Techniques

Since the birth of the first baby conceived with in vitro fertilization (IVF) and embryo transfer, assisted reproductive technology, an extremely successful form of therapy for many infertile couples, is currently practised all over the world.

#### 3.1 *Methylxanthines in Assisted Reproductive Techniques*

The effects of AC, IBMX, and dibutyryl cAMP (dbcAMP) on porcine oocyte in vitro maturation, IVF, and subsequent embryonic development were investigated by Somfai et al. (2003). They showed that a change in the intracellular level of cAMP during oocyte collection does not affect the maturational and developmental competence of the oocytes and that synchronization of meiotic maturation using dbcAMP enhances the meiotic potential of oocytes by promoting the MI to metaphase II transition and results in high developmental competence by monospermic fertilization. In IVF experiments, IBMX in association with FSH and LH, was used to synchronize the oocytes. At 6 days after IVF, the blastocyst rate in oocytes matured under these conditions was significantly higher than that for oocytes cultured in the absence of LH. The results suggest that the treatment of oocytes with FSH and IBMX causes the expression of LHR in cumulus cells, holds the oocytes at the germinal vesicle II stage, and can be considered as a beneficial procedure to obtain in-vitro-matured oocytes with high developmental competence (Shimada et al. 2003a, b). Besides PKA, several protein kinases are involved in oocyte maturation, and studies of the mechanisms of protein kinase B (PKB) activation and its role in cumulus cells during in vitro meiotic resumption of oocytes showed that the addition of PDE inhibitors maintained the level of PKB activity in cumulus cells at levels comparable with those in cumulus cells just after collection from their follicles and that the inhibitory effect of hypoxanthine on spontaneous meiotic resumption was overcome by addition of a PKB inhibitor.



### 3.2 Caffeine in Assisted Reproductive Techniques

Recently, Maalouf et al. (2009) reported the effects of cumulus cell removal and caffeine treatment on the development of in-vitro-matured ovine oocytes. Whereas removal of cumulus cells and aging increases polyspermy, caffeine was effective in reducing this phenomenon, showing that caffeine treatment statistically increases the development to blastocyst and lowers the frequency of polyspermy.

Caffeine increases MPF and mitogen-activated protein kinase activities in ovine oocytes, prevents age-related changes, and increases cell numbers in blastocysts produced by somatic cell nuclear transfer (Lee and Campbell 2006, 2008). Used in experiments of nuclear remodelling of somatic cell nuclear transfer embryos and subsequent development and DNA methylation patterns, caffeine induces premature chromosome condensation at a high rate, a high blastocyst formation rate, and lowers the apoptotic cell index, suggesting that the nuclear remodelling type controlled by caffeine treatment can affect in vitro development and the methylation status of nuclear transfer in relation to nuclear reprogramming (Kwon et al. 2008). These results confirmed previous observations that showed that caffeine treatment promotes nuclear remodelling although it does not prevent the decrease in the developmental ability of cloned embryos caused by oocyte aging (Iwamoto et al. 2005; Kawahara et al. 2005). On the other hand, with use of a mouse model, it was shown that caffeine had no effect on the quality of oocytes matured in vivo, whereas it was detrimental to the quality of oocytes matured in vitro (Miao et al. 2007). However, in vivo studies with female mice administered 150 mg/kg caffeine at various times prior to metaphase I, showed non-significant differences in the frequencies of hyperploid, MI, diploid, premature centromere separation, single chromatids, and structural chromosome aberrations between the controls and each of the caffeine groups (Mailhes et al. 1996; Jaakma et al. 1997).

Studies of the effects of caffeine on male gametes showed stimulation of sperm capacitation and spontaneous AR (Funahashi 2003, 2005); therefore, during IVF procedures, supplementation with  $\beta$ -mercaptoethanol, which neutralizes the stimulatory effect of caffeine, has a beneficial effect in maintaining the function of gametes, the incidence of normal fertilization, and, consequently, the quality of IVF embryos. However, a limited exposure of gametes to caffeine significantly reduced the mean number of sperm cells that penetrated into the oocyte and asynchrony in the morphology of sperm nuclei in polyspermic oocytes (Funahashi and Romar 2004). Other epidemiology studies evaluated the timing and amount of caffeine intake by women and men undergoing IVF and gamete intra-Fallopian transfer (GIFT) on oocyte retrieval, sperm parameters, fertilization, multiple gestations, miscarriage, and live births. A prospective study of 221 couples was conducted between 1993 and 1998. "Usual" caffeine intake during the lifetime and 1 year prior to the study, caffeine intake during the week of the initial clinic visit, as well as caffeine intake during the week of the procedure were evaluated for beverages (coffee, soda, and tea) and chocolates. Not achieving a live birth was significantly associated with "usual" female caffeine consumption for an intake of



more than 50 mg/day and consumption of 0–2 mg/day during the week of the initial visit. Infant gestational age decreased by 3.8 or 3.5 weeks for women who consumed more than 50 mg/day of caffeine “usually” or during the week of the initial visit. The odds of having multiple gestations increased by 2.2 and 3.0 for men who increased their “usual” intake or intake during the week of the initial visit by an extra 100 mg/day. Caffeine intake was not significantly associated with other outcomes. This was the first IVF/GIFT study to report any effect of caffeine on live births, gestational age, and multiple gestations and if these findings are replicated, caffeine use should be minimized prior to and while undergoing IVF/GIFT (Klonoff-Cohen et al. 2002).

## 4 Conclusion

Reports on methylxanthines and their effects on reproduction have mainly focused on their use as *in vitro* PDE inhibitors that, by maintaining high intracellular levels of cAMP, differently affect female and male gametes. Animal studies have largely shown that methylxanthines have toxic effects on gonads and gametogenesis of both sexes, although comparing doses of caffeine in animals and humans is for many reasons not an easy task. Caffeine is present in many beverages (coffee, tea, colas, and chocolate) and in over-the-counter medications. The medical literature contains many varied references that appear to indicate that human adverse reproductive/developmental effects are produced by caffeine. However, if caffeine causes such effects, the reproductive consequences could be very serious because caffeine-containing foods and beverages are consumed by most of the human populations of the world, and, as world “coffee culture” continues to grow, world caffeine intakes continue to increase. After revising the medical literature dealing with developmental and reproductive risks of caffeine on the basis of the biological plausibility of the epidemiological and animal findings and the methods and conclusions of previous investigators, clinical counsellors can inform pre-pregnant/pregnant women who do not smoke or drink alcohol and who consume moderate amounts of caffeine (5–6 mg/kg per day, five cups) that they do not have an increase in reproductive risks or adverse effects.

## References

- Abou-haila A, Tulsiani DR (2009) Signal transduction pathways that regulate sperm capacitation and the acrosome reaction. *Arch Biochem Biophys* 485:72–81
- Ain R, Uma Devi K, Shivaji S, Seshagiri PB (1999) Pentoxifylline-stimulated capacitation and acrosome reaction in hamster spermatozoa: involvement of intracellular signalling molecules. *Mol Hum Reprod* 5:618–626

- Aitken RJ, Harkiss D, Knox W, Paterson M, Irvine DS (1998) A novel signal transduction cascade in capacitating human spermatozoa characterised by a redox-regulated, cAMP-mediated induction of tyrosine phosphorylation. *J Cell Sci* 111:645–656
- Aitken RJ, Nixon B, Lin M, Koppers AJ, Lee YH, Baker MA (2007) Proteomic changes in mammalian spermatozoa during epididymal maturation. *Asian J Androl* 9:554–564
- Ax RL, Collier RJ, Lodge JR (1976) Effects of dietary caffeine on the testis of the domestic fowl, *Gallus domesticus*. *J Reprod Fertil* 47:235–238
- Allegretti C, Liguori L, Minelli A (2001) Stimulation by N6-cyclopentyladenosine of A1 adenosine receptors, coupled to G $\alpha$ i2 protein subunit, has a capacitative effect on human spermatozoa. *Biol Reprod* 64:1653–1659
- Baldi E, Luconi M, Bonaccorsi L, Muratori M, Forti G (2000) Intracellular events and signaling pathways involved in sperm acquisition of fertilizing capacity and acrosome reaction. *Front Biosci* 5:110–123
- Barretto LS, Caiado Castro VS, Garcia JM, Mingoti GZ (2007) Role of roscovitine and IBMX on kinetics of nuclear and cytoplasmic maturation of bovine oocytes in vitro. *Anim Reprod Sci* 99:202–207
- Björklund O, Kahlström J, Salmi P, Fredholm BB (2008) Perinatal caffeine, acting on maternal adenosine A(1) receptors, causes long-lasting behavioral changes in mouse offspring. *PLoS ONE* 3:e3977
- Bolúmar F, Olsen J, Rebagliato M, Bisanti L (1997) Caffeine intake and delayed conception: a European multicenter study on infertility and subfecundity. European study group on infertility subfecundity. *Am J Epidemiol* 145:324–334
- Bornslaeger EA, Schultz RM (1985) Regulation of mouse oocyte maturation: effect of elevating cumulus cell cAMP on oocyte cAMP levels. *Biol Reprod* 33:698–704
- Bracken MB, Triche EW, Belanger K, Hellenbrand K, Leaderer B (2003) Association of maternal caffeine consumption with decrements in fetal growth. *Am J Epidemiol* 157:456–466
- Buffone MG, Calamera JC, Verstraeten SV, Doncel GF (2005) Capacitation-associated protein tyrosine phosphorylation and membrane fluidity changes are impaired in the spermatozoa of asthenozoospermic patients. *Reproduction* 129:697–705
- CARE Study Group (2008) Maternal caffeine intake during pregnancy and risk of fetal growth restriction: a large prospective observational study. *BMJ* 337:a2332. doi:10.1136/bmj.a2332
- Carr DW, Acott TS (1990) The phosphorylation of a putative sperm microtubule-associated protein 2 (MAP2) is uniquely sensitive to regulation. *Biol Reprod* 43:795–805
- Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC (2009) Caffeinated and alcoholic beverage intake in relation to ovulatory disorder infertility. *Epidemiology* 20:374–381
- Chiarella P, Puglisi R, Sorrentino V, Boitani C, Stefanini M (2004) Ryanodine receptors are expressed and functionally active in mouse spermatogenic cells and their inhibition interferes with spermatogonial differentiation. *J Cell Sci* 117:4127–4134
- Cho WK, Stern S, Biggers JD (1974) Inhibitory effect of dibutyryl cAMP on mouse oocyte maturation in vitro. *J Exp Zool* 187:383–386
- Christian MS, Brent RL (2001) Teratogen update: evaluation of the reproductive and developmental risks of caffeine. *Teratology* 64:51–78
- Conti M, Andersen CB, Richard F, Mehats C, Chun SY, Horner K, Jin C, Tsafirri A (2002) Role of cyclic nucleotide signaling in oocyte maturation. *Mol Cell Endocrinol* 187:153–159
- de Lamirande E, Leclerc P, Gagnon C (1997) Capacitation as a regulatory event that primes spermatozoa for the acrosome reaction and fertilization. *Mol Hum Reprod* 3:175–194
- Dekel N, Beers WH (1978) Rat oocyte maturation in vitro: relief of cyclic AMP inhibition by gonadotropins. *Proc Natl Acad Sci USA* 75:4369–4373
- De Jonge CJ, Barratt CLR, Radwanska EWA, Cooke ID (1993) The acrosome reaction-inducing effect of human follicular and oviductal fluid. *J Androl* 14:359–365
- Depeiges A, Dacheux JL (1985) Acquisition of sperm motility and its maintenance during storage in the lizard, *Lacerta vivipara*. *J Reprod Fertil* 74:23–27

- Derbyshire E, Abdula S (2008) Habitual caffeine intake in women of childbearing age. *J Hum Nutr Diet* 21:159–164
- Dews PB (1982) Caffeine. *Annu Rev Nutr* 2:323–241
- Duckworth BC, Weaver JS, Ruderman JV (2002) G2 arrest in *Xenopus* oocytes depends on phosphorylation of cdc25 by protein kinase A. *Proc Natl Acad Sci USA* 99:16794–16799
- Eppig JJ, Vivieros MM, Marin-Bivens C, De La Fuente R (2004) Regulation of mammalian oocyte maturation. In: Leung PCK, Adashi EY (eds) *The ovary*. Elsevier, Amsterdam
- Ettlin RA, Armstrong JM, Buser S, Hennes U (1986) Retardation of spermiation following short-term treatment of rats with theobromine. *Arch Toxicol Suppl* 9:441–446
- Ezzat AR, Gohary ZM (1994) Hormonal and histological effects of chronic caffeine administration on the pituitary-gonadal and pituitary-adrenocortical axes in male rabbits. *Funct Dev Morphol* 4:45–50
- Fenster L, Quale C, Waller K, Windham GC, Elkin EP, Benowitz N, Swan SH (1999) Caffeine consumption and menstrual function. *Am J Epidemiol* 149:550–557
- Florman HM, Corron ME, Kim TD, Babcock DF (1992) Activation of voltage-dependent calcium channels of mammalian sperm is required for zona pellucida-induced acrosomal exocytosis. *Dev Biol* 152:304–314
- Fredholm BB (1995) Astra award lecture. Adenosine, adenosine receptors and the actions of caffeine. *Pharmacol Toxicol* 76:93–101
- Fredholm B, Battig K, Holmen J, Nehlig A, Zvartau E (1999) Action of caffeine in the brain with special reference to factors that contribute to its wide spread use. *Pharmacol Rev* 51:83–133
- Friedman L, Weinberger MA, Farber TM, Moreland FM, Peters EL, Gilmore CE, Khan MA (1979) Testicular atrophy and impaired spermatogenesis in rats fed high levels of the methylxanthines caffeine, theobromine, or theophylline. *J Environ Pathol Toxicol* 2:687–706
- Funabashi H, Fujioka M, Kohchi M, Tateishi Y, Matsuoka N (2000) Collaborative work to evaluate toxicity on male reproductive organs by repeated dose studies in rats 22). Effects of 2- and 4-week administration of theobromine on the testis. *J Toxicol Sci* 25:211–221
- Funahashi H (2003) Polyspermic penetration in porcine IVM-IVF systems. *Reprod Fertil Dev* 15:167–177
- Funahashi H, Romar R (2004) Reduction of the incidence of polyspermic penetration into porcine oocytes by pretreatment of fresh spermatozoa with adenosine and a transient co-incubation of the gametes with caffeine. *Reproduction* 128:789–800
- Funahashi H (2005) Effect of beta-mercaptoethanol during in vitro fertilization procedures on sperm penetration into porcine oocytes and the early development in vitro. *Reproduction* 130:889–898
- Galantino-Homer HL, Visconti PE, Kopf GS (1997) Regulation of protein tyrosine phosphorylation during bovine sperm capacitation by a cyclic adenosine 3′5′-monophosphate-dependent pathway. *Biol Reprod* 56:707–719
- Gans JH (1982) Dietary influences on theobromine-induced toxicity in rats. *Toxicol Appl Pharmacol* 63:312–320
- Gans JH (1984) Comparative toxicities of dietary caffeine and theobromine in the rat. *Food Chem Toxicol* 22:365–369
- Gougeon A (1996) Regulation of ovarian follicular development in primates: facts and hypotheses. *Endocr Rev* 17:121–155
- Haider S, Chaube SK (1996) The in vitro effects of forskolin, IBMX and cyanoketone on meiotic maturation in follicle-enclosed catfish (*Clarias batrachus*) oocytes. *Comp Biochem Physiol C Pharmacol Toxicol Endocrinol* 115:117–123
- Harayama H, Miyake M, Shidara O, Iwamoto E, Kato S (1998) Effects of calcium and bicarbonate on head-to-head agglutination in ejaculated boar spermatozoa. *Reprod Fertil Dev* 10:445–450
- Harrison DA, Carr DW, Meizel S (2000) Involvement of protein kinase A and A kinase anchoring protein in the progesterone-initiated human sperm acrosome reaction. *Biol Reprod* 62:811–820
- Heller CG, Clermont Y (1963) Spermatogenesis in man: an estimate of its duration. *Science* 140:184–186

- Hirsh K (1984) Central nervous system pharmacology of the methylxanthines. In: Spiller GA (ed) The methylxanthine beverages and foods: chemistry, composition and health effects. Liss, New York
- Hong CY, Chaput De Saintonge DM, Turner P (1981) The inhibitory action of procaine, (+) propranolol and ( $\pm$ ) propranolol on human sperm motility: antagonism by caffeine. *Br J Clin Pharmacol* 12:751–753
- Horner K, Livera G, Hinckley M, Trinh K, Storm D, Conti M (2003) Rodent oocytes express an active adenylyl cyclase required for meiotic arrest. *Dev Biol* 258:385–396
- Iwamoto M, Onishi A, Fuchimoto D, Somfai T, Suzuki S, Yazaki S, Hashimoto M, Takeda K, Tagami T, Hanada H, Noguchi J, Kaneko H, Nagai T, Kikuchi K (2005) Effects of caffeine treatment on aged porcine oocytes: parthenogenetic activation ability, chromosome condensation and development to the blastocyst stage after somatic cell nuclear transfer. *Zygote* 13:335–345
- Jaakma U, Zhang BR, Larsson B, Niwa K, Rodriguez-Martinez H (1997) Effects of sperm treatments on the in vitro development of bovine oocytes in semidefined and defined media. *Theriogenology* 48:711–720
- Jiang CS, Kilfeather SA, Pearson RM, Turner P (1984) The stimulatory effects of caffeine, theophylline, lysine-theophylline and 3-isobutyl-1-methylxanthine on human sperm motility. *Br J Clin Pharmacol* 18:258–262
- Jaiswal BS, Majumder GC (1998) Biochemical parameters regulating forward motility initiation in vitro in goat immature epididymal spermatozoa. *Reprod Fertil Dev* 10:299–307
- Johnson EM, Christian MS (1984) When is a teratology study not an evaluation of teratogenicity? *J Am Coll Toxicol* 3:431–434
- Kalinowski RR, Jaffe LA, Foltz KR, Giusti AF (2003) A receptor linked to a Gi-family G-protein functions in initiating oocyte maturation in starfish but not frogs. *Dev Biol* 253:139–149
- Kanatsu-Shinohara M, Schultz RM, Kopf GS (2000) Acquisition of meiotic competence in mouse oocytes: absolute amounts of p34cdc2, cyclin B1, cdc25C, and wee1 in meiotically incompetent and competent oocytes. *Biol Reprod* 63:1610–1616
- Kawahara M, Wakai T, Yamanaka K, Kobayashi J, Sugimura S, Shimizu T, Matsumoto H, Kim JH, Sasada H, Sato E (2005) Caffeine promotes premature chromosome condensation formation and in vitro development in porcine reconstructed embryos via a high level of maturation promoting factor activity during nuclear transfer. *Reproduction* 130:351–357
- Kinney A, Kline J, Kelly A, Reuss ML, Levin B (2007) Smoking, alcohol and caffeine in relation to ovarian age during the reproductive years. *Hum Reprod* 22:1175–1185
- Klonoff-Cohen H, Bleha J, Lam-Kruglick P (2002) A prospective study of the effects of female and male caffeine consumption on the reproductive endpoints of IVF and gamete intra-Fallopian transfer. *Hum Reprod* 17:1746–1754
- Kopf GS, Lewis CA, Vacquier VD (1983a) Methylxanthines stimulate calcium transport and inhibit cyclic nucleotide phosphodiesterases in abalone sperm. *Dev Biol* 99:115–120
- Kopf GS, Lewis CA, Vacquier VD (1983b) Regulation of abalone sperm cyclic AMP concentrations and the acrosome reaction by calcium and methylxanthines. *Dev Biol* 98:28–36
- Kopf GS, Wilde MW (1990) Signal transduction processes leading to acrosomal exocytosis in mammalian spermatozoa. *Trends Endocrinol Metab* 1:362–368
- Kwon DJ, Park CK, Yang BK, Cheong HT (2008) Control of nuclear remodelling and subsequent in vitro development and methylation status of porcine nuclear transfer embryos. *Reproduction* 135:649–656
- Lachance C, Bailey JL, Leclerc P (2007) Expression of Hsp60 and Grp78 in the human endometrium and oviduct, and their effect on sperm functions. *Hum Reprod* 22:2606–2614
- Laforest MF, Pouliot E, Guéguen L, Richard FJ (2005) Fundamental significance of specific phosphodiesterases in the control of spontaneous meiotic resumption in porcine oocytes. *Mol Reprod Dev* 70:361–372

- Leclerc P, de Lamirande E, Gagnon C (1998) Interaction between  $\text{Ca}^{2+}$ , cyclic 3',5' adenosine monophosphate, the superoxide anion, and tyrosine phosphorylation pathways in the regulation of human sperm capacitation. *J Androl* 19:434–443
- Leclerc P, Goupil S (2002) Regulation of the human sperm tyrosine kinase c-yes. Activation by cyclic adenosine 3',5'-monophosphate and inhibition by  $\text{Ca}(2+)$ . *Biol Reprod* 67:301–307
- Lee JH, Campbell KH (2006) Effects of enucleation and caffeine on maturation-promoting factor (MPF) and mitogen-activated protein kinase (MAPK) activities in ovine oocytes used as recipient cytoplasts for nuclear transfer. *Biol Reprod* 74:691–698
- Lee JH, Campbell KH (2008) Caffeine treatment prevents age-related changes in ovine oocytes and increases cell numbers in blastocysts produced by somatic cell nuclear transfer. *Cloning Stem Cells* 10:381–390
- Liguori L, de Lamirande E, Minelli A, Gagnon C (2005) Various protein kinases regulate human sperm acrosome reaction and the associated phosphorylation of Tyr residues and of the Thr-Glu-Tyr motif. *Mol Hum Reprod* 11:211–221
- Luria Y, Rubinstein S, Lax Y, Breitbart H (2002) Extracellular adenosine triphosphate stimulates acrosomal exocytosis in bovine spermatozoa via P2 purinoceptor. *Biol Reprod* 66:429–437
- Maalouf WE, Lee JH, Campbell KH (2009) Effects of caffeine, cumulus cell removal and aging on polyspermy and embryo development on in vitro matured and fertilized ovine oocytes. *Theriogenology* 71:1083–1092
- Magalhães ST, Guedes RN, Demuner AJ, Lima ER (2008) Effect of coffee alkaloids and phenolics on egg-laying by the coffee leaf miner *Leucoptera coffeella*. *Bull Entomol Res* 98:483–489
- Mailhes JB, Young D, London SN (1996) Cytogenetic effects of caffeine during in vivo mouse oocyte maturation. *Mutagenesis* 11:395–399
- Mahony MC, Gwathmey T (1999) Protein tyrosine phosphorylation during hyperactivated motility of cynomolgus monkey (*Macaca fascicularis*) spermatozoa. *Biol Reprod* 60:1239–1243
- Mandel HG (2002) Update on caffeine consumption, disposition and action. *Food Chem Toxicol* 40:1231–1234
- McKusick VA (1988) Mendelian inheritance in man: catalogs of autosomal dominant, autosomal recessive, and x-linked phenotypes, 8th edn. Johns Hopkins University Press, Baltimore
- Mehlmann LM, Jones TL, Jaffe LA (2002) Meiotic arrest in the mouse follicle maintained by a Gs protein in the oocyte. *Science* 297:1343–1345
- Mehlmann LM, Saeki Y, Tanaka S, Brennan TJ, Evsikov AV, Pendola FL, Knowles BB, Eppig JJ, Jaffe LA (2004) The Gs-linked receptor GPR3 maintains meiotic arrest in mammalian oocytes. *Science* 306:1947–1950
- Mehlmann LM (2005) Stops and starts in mammalian oocytes: recent advances in understanding the regulation of meiotic arrest and oocyte maturation. *Reproduction* 130:791–799
- Metchat A, Akerfelt M, Bierkamp C, Delsinne V, Sistonen L, Alexandre H, Christians ES (2009) Mammalian heat shock factor 1 is essential for oocyte meiosis and directly regulates Hsp90-alpha expression. *J Biol Chem* 284:9521–9528
- Miao YL, Shi LH, Lei ZL, Huang JC, Yang JW, Ouyang YC, Sun QY, Chen DY (2007) Effects of caffeine on in vivo and in vitro oocyte maturation in mice. *Theriogenology* 68:640–645
- Minelli A, Liguori L, Bellazza I, Mannucci R, Johansson B, Fredholm BB (2004) Involvement of A1 adenosine receptors in the acquisition of fertilizing capacity. *J Androl* 25:286–292
- Morales P, Díaz ES, Kong M (2007) Proteasome activity and its relationship with protein phosphorylation during capacitation and acrosome reaction in human spermatozoa. *Soc Reprod Fertil* 65:269–273
- Moreau R, Thérien I, Lazure C, Manjunath P (1998) Type II domains of BSP-A1/A2 proteins: binding properties, lipid efflux, and sperm capacitation potential. *Biochem Biophys Res Commun* 246:148–154
- Morikawa M, Seki M, Kume S, Endo T, Nishimura Y, Kano K, Naito K (2007) Meiotic resumption of porcine immature oocytes is prevented by ooplasmic Gsalpha functions. *J Reprod Dev* 53:1151–1157
- Nawrot P, Jordan S, Eastwood J, Rotstein J, Hughenholtz A, Feeley M (2003) Effects of caffeine on human health. *Food Addit Contam* 20:1–30

- Ozawa M, Nagai T, Somfai T, Nakai M, Maedomari N, Fahrudin M, Karja NW, Kaneko H, Noguchi J, Ohnuma K, Yoshimi N, Miyazaki H, Kikuchi K (2008) Comparison between effects of 3-isobutyl-1-methylxanthine and FSH on gap junctional communication, LH-receptor expression, and meiotic maturation of cumulus-oocyte complexes in pigs. *Mol Reprod Dev* 75:857–866
- Peng XR, Hsueh AJ, LaPolt PS, Bjersing L, Ny T (1991) Localization of luteinizing hormone receptor messenger ribonucleic acid expression in ovarian cell types during follicle development and ovulation. *Endocrinology* 129:3200–3207
- Pincus G, Enzmann EV (1935) The comparative behaviour of mammalian eggs in vivo and in vitro: I. The activation of ovarian eggs. *J Exp Med* 62:665–675
- Pirino G, Wescott MP, Donovan PJ (2009) Protein kinase A regulates resumption of meiosis by phosphorylation of Cdc25B in mammalian oocytes. *Cell Cycle* 8:665–670
- Pollard I, Williamson S, Magre S (1990) Influence of caffeine administered during pregnancy on the early differentiation of foetal rat ovaries and testes. *J Dev Physiol* 13:59–65
- Pollard I, Locquet O, Solvar A, Magre S (2001) Effects of caffeine and its reactive metabolites theophylline and theobromine on the differentiating testis. *Reprod Fertil Dev* 13:435–441
- Ramlau-Hansen CH, Thulstrup AM, Bonde JP, Olsen J, Bech BH (2008) Semen quality according to prenatal coffee and present caffeine exposure: two decades of follow-up of a pregnancy cohort. *Hum Reprod* 23:2799–2805
- Richards JS, Russell DL, Ochsner S, Espey LL (2002) Ovulation: new dimensions and new regulators of the inflammatory-like response. *Annu Rev Physiol* 64:69–92
- Salicioni AM, Platt MD, Wertheimer EV, Arcelay E, Allaire A, Sosnik J, Visconti PE (2007) Signalling pathways involved in sperm capacitation. *Soc Reprod Fertil Suppl* 65:245–259
- Schultz RM, Montgomery RR, Belanoff JR (1983a) Regulation of mouse oocyte meiotic maturation: implication of a decrease in oocyte cAMP and protein dephosphorylation in commitment to resume meiosis. *Dev Biol* 97:264–273
- Schultz RM, Montgomery RR, Ward-Bailey PF, Eppig JJ (1983b) Regulation of oocyte maturation in the mouse: possible roles of intercellular communication, cAMP, and testosterone. *Dev Biol* 95:294–304
- Shimada M, Kawano N, Terada T (2002) Delay of nuclear maturation and reduction in developmental competence of pig oocytes after mineral oil overlay of in vitro maturation media. *Reproduction* 124:557–564
- Shimada M, Nishibori M, Isobe N, Kawano N, Terada T (2003a) Luteinizing hormone receptor formation in cumulus cells surrounding porcine oocytes and its role during meiotic maturation of porcine oocytes. *Biol Reprod* 68:1142–1149
- Shimada M, Ito J, Yamashita Y, Okazaki T, Isobe N (2003b) Phosphatidylinositol 3-kinase in cumulus cells is responsible for both suppression of spontaneous maturation and induction of gonadotropin-stimulated maturation of porcine oocytes. *J Endocrinol* 179:25–34
- Si Y, Okuno M (1999) Role of tyrosine phosphorylation of flagellar proteins in hamster sperm hyperactivation. *Biol Reprod* 61:240–246
- Somfai T, Kikuchi K, Onishi A, Iwamoto M, Fuchimoto D, Papp AB, Sato E, Nagai T (2003) Meiotic arrest maintained by cAMP during the initiation of maturation enhances meiotic potential and developmental competence and reduces polyspermy of IVM/IVF porcine oocytes. *Zygote* 11:199–206
- Strandgaard C, Miller MG (1998) Germ cell apoptosis in rat testis after administration of 1,3-dinitrobenzene. *Reprod Toxicol* 12:97–103
- Tengowski MW, Sutovsky P, Hedlund LW, Guyot DJ, Burkhardt JE, Thompson WE, Sutovsky M, Johnson GA (2005) Reproductive cytotoxicity is predicted by magnetic resonance microscopy and confirmed by ubiquitin-proteasome immunohistochemistry in a theophylline-induced model of rat testicular and epididymal toxicity. *Microsc Microanal* 11:300–312
- Tengowski MW, Feng D, Sutovsky M, Sutovsky P (2007) Differential expression of genes encoding constitutive and inducible 20S proteasomal core subunits in the testis and epididymis of theophylline- or 1,3-dinitrobenzene-exposed rats. *Biol Reprod* 76:149–163

- Thérien I, Bousquet D, Manjunath P (2001) Effect of seminal phospholipid-binding proteins and follicular fluid on bovine sperm capacitation. *Biol Reprod* 65:41–51
- Thomas RE, Armstrong DT, Gilchrist RB (2002) Differential effects of specific phosphodiesterase isoenzyme inhibitors on bovine oocyte meiotic maturation. *Dev Biol* 244:215–225
- Törnell J, Billig H, Hillensjö T (1990) Resumption of rat oocyte meiosis is paralleled by a decrease in guanosine 3',5'-cyclic monophosphate (cGMP) and is inhibited by microinjection of cGMP. *Acta Physiol Scand* 139:511–517
- Tulsiani DR, Abou-Haila A, Loeser CR, Pereira BM (1998) The biological and functional significance of the sperm acrosome and acrosomal enzymes in mammalian fertilization. *Exp Cell Res* 240:151–164
- Tulsiani DR, Zeng HT, Abou-Haila A (2007) Biology of sperm capacitation: evidence for multiple signalling pathways. *Soc Reprod Fertil Suppl* 63:257–272
- Thundathil J, de Lamirande E, Gagnon C (2002) Different signal pathways are involved during human sperm capacitation induced by biological and pharmacological agents. *Mol Hum Reprod* 8:811–816
- Vivarelli E, Conti M, De Felici M, Siracusa G (1983) Meiotic resumption and intracellular cAMP levels in mouse oocytes treated with compounds which act on cAMP metabolism. *Cell Differ* 12:271–276
- Visconti PE, Bailey JL, Moore GD, Pan D, Olds-Clarke P, Kopf GS (1995a) Capacitation of mouse spermatozoa: I. Correlation between the capacitation state and the protein tyrosine phosphorylation. *Development* 121:1129–1137
- Visconti PE, Moore GD, Bailey JL, Leclerc P, Connors SA, Pan D, Olds-Clarke P, Kopf GS (1995b) Capacitation of mouse spermatozoa: II. Protein tyrosine phosphorylation and capacitation are regulated by a cAMP-dependent pathway. *Development* 121:1139–1150
- Visconti PE, Kopf GS (1998) Regulation of protein phosphorylation during sperm capacitation. *Biol Reprod* 59:1–6
- Visconti PE, Galantino-Homer H, Ning X, Moore GD, Valenzuela JP, Jorgez CJ, Alvarez JG, Kopf GS (1999) Cholesterol efflux-mediated signal transduction in mammalian sperm. beta-cyclodextrins initiate transmembrane signaling leading to an increase in protein tyrosine phosphorylation and capacitation. *J Biol Chem* 274:3235–3242
- Wang Y, Waller DP, Hikim AP, Russell LD (1992) Reproductive toxicity of theobromine and cocoa extract in male rats. *Reprod Toxicol* 6:347–353
- Wang Y, Waller DP (1994) Theobromine toxicity on Sertoli cells and comparison with cocoa extract in male rats. *Toxicol Lett* 70:155–164
- Wassarman PM (2009) Mammalian fertilization: the strange case of sperm protein 56. *Bioessays* 31:153–158
- Webb RJ, Marshall F, Swann K, Carroll J (2000) Follicle-stimulating hormone induces a gap junction-dependent dynamic change in [cAMP] and protein kinase A in mammalian oocytes. *Dev Biol* 246:441–454
- Weinberger MA, Friedman L, Farber TM, Moreland FM, Peters EL, Gilmore CE, Khan MA (1978) Testicular atrophy and impaired spermatogenesis in rats fed high levels of the methylxanthines caffeine, theobromine, or theophylline. *J Environ Pathol Toxicol* 1:669–688
- Yanagimachi R (1994) Mammalian fertilization. In: Knobil E, Neill JD (eds) *The physiology of reproduction*. Raven, New York
- Yeste M, Briz M, Pinart E, Sancho S, Garcia-Gil N, Badia E, Bassols J, Pruneda A, Bussalleu E, Casas I, Bonet S (2008) Hyaluronic acid delays boar sperm capacitation after 3 days of storage at 15 degrees C. *Anim Reprod Sci* 109:236–250
- Yu L, Coelho J, Zhang X, Fu Y, Tillman A, Karaoz U, Fredholm BB, Weng Z, Chen JF (2009) Uncovering multiple molecular targets for caffeine by a drug target validation strategy of combined A2A receptor knockouts and microarray profiling. *Physiol Genomics* 37:199–210
- Zeleznik AJ (2004) Dynamics of primate follicular growth: a physiological perspective. In: Leung PCK, Adashi EY (eds) *The ovary*, 2nd edn. Elsevier, Amsterdam

# Methylxanthines During Pregnancy and Early Postnatal Life

Ulrika Ådén

## Contents

1	Introduction .....	374
2	Mechanisms of Action in the Neonate. Effector Systems and Their Maturation .....	375
2.1	Mechanisms of Action at Clinically Relevant Concentrations .....	375
2.2	Maturation of Adenosine and GABA <sub>A</sub> Receptor Systems .....	375
3	Exposure to Methylxanthines During Fetal Life .....	377
3.1	Metabolism of Methylxanthines in the Pregnant Woman and in the Fetus .....	377
3.2	Exposure to Methylxanthines in Fetal Life and Perinatal Outcomes .....	378
4	Exposure to Methylxanthines in Neonates .....	379
4.1	Absorption, Metabolism, and Elimination .....	379
4.2	Methylxanthines and Breast Feeding .....	379
4.3	Methylxanthines for Apnea of Prematurity .....	380
5	Short-Term Effects of Methylxanthines in the Infant .....	382
5.1	Physiological Effects .....	382
5.2	Withdrawal Symptoms .....	382
6	Long-Term Effects of Methylxanthines for the Developing Organism .....	383
6.1	CNS Function .....	383
6.2	Respiration and Cardiovascular Function .....	383
7	Conclusions .....	384
	References .....	385

**Abstract** World-wide, many fetuses and infants are exposed to methylxanthines via maternal consumption of coffee and other beverages containing these substances. Methylxanthines (caffeine, theophylline and aminophylline) are also commonly used as a medication for apnea of prematurity.

The metabolism of methylxanthines is impaired in pregnant women, fetuses and neonates, leading to accumulating levels thereof. Methylxanthines readily passes

---

U. Ådén

Department of Woman and Child Health, Karolinska Institute, 171 77 Stockholm, Sweden  
e-mail: [ulrika.aden@ki.se](mailto:ulrika.aden@ki.se)



the placenta barrier and enters all tissues and thus may affect the fetus/newborn at any time during pregnancy or postnatal life, given that the effector systems are mature.

At clinically relevant doses, the major effector system for methylxanthines is adenosine receptors. Animal studies suggest that adenosine receptors in the cardiovascular, respiratory and immune system are developed at birth, but that cerebral adenosine receptors are not fully functional. Furthermore animal studies have shown protective positive effects of methylxanthines in situations of hypoxia/ischemia in neonates. Similarly, a positive long-term effect on lung function and CNS development was found in human preterm infants treated with high doses of caffeine for apneas. There is now evidence that the overall benefits from methylxanthine therapy for apnea of prematurity outweigh potential short-term risks.

On the other hand it is important to note that experimental studies have indicated that long-term effects of caffeine during pregnancy and postnatally may include altered behavior and altered respiratory control in the offspring, although there is currently no human data to support this.

Some epidemiology studies have reported negative effects on pregnancy and perinatal outcomes related to maternal ingestion of high doses of caffeine, but the results are inconclusive. The evidence base for adverse effects of caffeine in first third of pregnancy are stronger than for later parts of pregnancy and there is currently insufficient evidence to advise women to restrict caffeine intake after the first trimester.

**Keywords** Caffeine · Fetus · Methylxanthines · Neonatal · Newborn · Pregnancy · Theophylline

## 1 Introduction

The majority of expecting mothers in the Western world drink beverages containing methylxanthines during pregnancy and they continue their consumption during lactation. Many fetuses and infants are thus exposed to these substances. In addition, a large number of premature infants need pharmacological treatment for apnea of prematurity with methylxanthines.

There has been considerable concern over fetuses and infants being subjected to methylxanthines, with reported negative effects on pregnancy, perinatal, and long-term outcomes.

This chapter discusses how fetuses and newborns are subjected to methylxanthines, how the effects are mediated, and what information the current literature provides on the long-term effects of exposure to methylxanthines at clinically relevant doses.

## 2 Mechanisms of Action in the Neonate. Effector Systems and Their Maturation

### 2.1 Mechanisms of Action at Clinically Relevant Concentrations

The only known effect of caffeine at concentrations relevant to daily intake of coffee is blockade of adenosine  $A_1$  and  $A_{2A}$  receptors that occurs at serum concentrations of 0.2–2 mg/L (0.001–0.01 mM; Fredholm et al. 1999). When pregnant women drink beverages containing caffeine, serum levels of caffeine soon become similar in the fetus and in the mother. When preterm infants are treated with caffeine for apnea of prematurity, more than 10 times higher serum levels are reached and then, in addition to adenosine  $A_1$  and  $A_{2A}$  receptor blockade, an inhibitory effect on adenosine  $A_3$  receptors, and a minimal blockade of phosphodiesterases,  $GABA_A$  receptors, and  $Ca^{++}$  release can occur. The maturation of these effector systems may affect the way the fetus and the infant react to caffeine (see below).

It is now less common that fetuses and infants are exposed to theophylline than caffeine, since theophylline has been replaced in modern asthma therapy and for treatment of apneas in preterm infants, and caffeine is usually preferred to theophylline/aminophylline (see below). Theophylline is structurally related to caffeine and is thus a potent inhibitor of adenosine  $A_1$ ,  $A_{2A}$ , and to some extent  $A_3$  receptors at therapeutically relevant doses. The degree of inhibition of phosphodiesterases is minimal at these concentrations.

Methylxanthines have both pro- and anti-inflammatory properties. At therapeutic doses, most effects result from adenosine receptor antagonism and the proinflammatory actions may be more relevant (Haskó and Cronstein 2010; Ohta and Sitkovsky 2010), at least in adults. However, a recent study in umbilical cord blood monocytes indicated that caffeine inhibited TNF- $\alpha$  production in neonates, possibly via adenosine  $A_1$  receptors (Chavez-Valdez et al. 2009).

Thus, some of the immunomodulatory mechanisms of methylxanthines are functioning at an early developmental stage, whereas other effects do not mature until later in infancy.

### 2.2 Maturation of Adenosine and $GABA_A$ Receptor Systems

As mentioned, the major effect of the methylxanthines caffeine and theophylline/aminophylline at therapeutic doses is antagonism of adenosine  $A_1$  and  $A_{2A}$  receptors. The maturation of these receptor systems affects how the fetus and child react to methylxanthines. Data on the development of adenosine receptors are obtained from mice and rats, but there are no available data from human fetuses or infants to our knowledge.

Adenosine A<sub>1</sub> receptors are mainly present in the heart (conductive system and myocytes), brain (predominantly in cortex, hippocampus, cerebellum, pons, and medulla oblongata) kidney, testis, and adipose tissue. Adenosine-A<sub>1</sub>-receptor-knockout mice appear to develop normally (Johansson et al. 2001), indicating that this receptor has no main effect on fetal development, but recent studies have pointed out differences in heart rate, body temperature, and locomotion in adult life of these mice (Yang et al. 2007). Adenosine A<sub>1</sub> receptors are present in a small amount early in brain development (Rivkees 1995; Aden et al. 2000), but are not functionally coupled to G proteins in the brain until adolescence (2–3 weeks of age in a mouse) (Aden et al. 2001). An important exception is the medulla oblongata, where functional coupling to G proteins was confirmed in rats just before birth using guanylyl-5'-O-(γ-[<sup>35</sup>S]thio)-triphosphate binding (Herlenius et al. 2002). To this end, it is known that adenosine and adenosine A<sub>1</sub> receptor agonists depress respiratory rhythmogenesis *in vivo* and *in vitro* (Lagercrantz et al. 1984; Eldridge et al. 1985). Accordingly, methylxanthines exert profound stimulatory effects on respiration in newborns by antagonism of adenosine A<sub>1</sub> receptors in the pons and medulla oblongata (Lagercrantz et al. 1984; Tilley 2010).

An early functional coupling of adenosine A<sub>1</sub> receptors to G proteins was also reported in the heart (Aden et al. 2001), which is in agreement with the clinical observation that tachycardia is a frequently seen side effect of methylxanthines in premature infants. There are data indicating that adenosine A<sub>1</sub> receptors function in umbilical cord blood monocytes (Chavez-Valdez et al. 2009) Adenosine A<sub>2A</sub> receptors are present in the brain and (predominantly in the basal ganglia), at endothelial cells, platelets, and on inflammatory cells (neutrophils, platelets, macrophages/microglial cells, and T cells), where they exhibit important anti-inflammatory properties (Haskó and Cronstein 2010; Ohta and Sitkovsky 2010, 2001). The first targeted disruption of the adenosine A<sub>2A</sub> receptor gene in mice was reported in 1997 (Ledent et al. 1997) and in agreement with previous pharmacology studies, these animals had increased blood pressure and platelet aggregation. There were no major malformations in adenosine-A<sub>2A</sub>-receptor-knockout mice, indicating that the stimulation of this receptor is not necessary for normal fetal development. The major development of cerebral adenosine A<sub>2A</sub> receptors takes place after birth, as pointed out by rat studies (Rivkees 1995; Aden et al. 2000). It is not known whether adenosine A<sub>2A</sub> receptors on inflammatory cells are functional early in life.

Adenosine A<sub>3</sub> receptors have a low affinity for caffeine and are therefore not considered as its primary target when ingested in beverages. However, therapeutic levels of methylxanthines given as therapy for preterm apneas may reach concentrations where adenosine A<sub>3</sub> receptors are blocked. Adenosine A<sub>3</sub> receptors are very scarce in the brain, but can be detected at low levels in several regions in the adult rodent brain. The functionality of these receptors in neurons remains unclear. It is, however, now evident that functional adenosine A<sub>3</sub> receptors are present on microglial cells and astrocytes (Hammarberg et al. 2003; Abbracchio et al. 2001; see Haskó and Cronstein 2010), but developmental data are lacking. Recent data show that deletion of adenosine A<sub>3</sub> receptors

(adenosine- $A_3$ -receptor-knockout mice) during brain development leads to a reduced response to caffeine in adult life (Bjorklund et al. 2008a). Although the mechanisms have not been elucidated, a developmental role of adenosine  $A_3$  receptors might therefore be indicated, in particular during circumstances when there is fetal exposure to caffeine.

GABA<sub>A</sub> receptors may be a target for high doses of methylxanthines, for example, when given postnatally for the treatment of apnea of prematurity. This receptor system develops in the brain early during fetal life (Aden et al. 2000), but whereas activation of GABA<sub>A</sub> receptors in mature neurons results in membrane hyperpolarization, GABA<sub>A</sub> receptor activation during early stages of brain development (up until the first postnatal week in rodents, corresponding to term age in a human baby) causes depolarization of the postsynaptic membrane. The GABA-mediated depolarization is thought to regulate neurogenesis, synaptogenesis, and final neuron number by regulating second-messenger systems (Varani et al. 2005) and modulating DNA synthesis (Leinekugel et al. 1997). Blockade of GABA<sub>A</sub> receptors with methylxanthines has, therefore, a theoretic potential to affect brain development. However, it is completely unknown whether the net effect on outcome would be positive or negative.

### 3 Exposure to Methylxanthines During Fetal Life

As mentioned, it is nowadays uncommon that fetuses are exposed to theophylline, but many fetuses are subjected to caffeine because the majority (75%) of expecting mothers drink beverages containing caffeine (Eskenazi 1999).

#### 3.1 *Metabolism of Methylxanthines in the Pregnant Woman and in the Fetus*

Methylxanthines are rapidly and completely absorbed from the gastrointestinal tract. There is only a minimal first-pass effect and once absorbed, methylxanthines pass blood–brain and placenta barriers, entering all tissues (Arnaud 1987). The half life of caffeine in pregnant women varies between 2 and 4.5 h during the first trimester, which is similar to that in nonpregnant women, but increases to 10 h at 17 weeks gestation and up to 18 h in the end of pregnancy, leading to an accumulation of caffeine in the mother and the fetus (Aldridge et al. 1981).

The main enzyme controlling caffeine metabolism is cytochrome P450 1A2 (CYP1A2). There are interindividual differences due to polymorphisms of this enzyme (Grosso and Bracken 2005). Another enzyme that regulates the metabolism to a lesser extent is *N*-acetyltransferase (Fenster et al. 1998). The activities of both of these enzymes are reduced during pregnancy (Tsutsumi et al. 2001), resulting in

gradually increasing plasma concentrations (to about twice the prepregnancy levels) of caffeine during pregnancy, despite little change in reported consumption (Cook et al. 1996). Both the fetus and the placenta lack the enzymes needed to metabolize methylxanthines (Kalow and Tang 1991) and therefore elimination in the fetus is almost entirely dependent upon renal excretion. The urinary excretion rate increases with dose in both mothers and fetuses.

### ***3.2 Exposure to Methylxanthines in Fetal Life and Perinatal Outcomes***

Since caffeine and its metabolites can pass the placenta barrier, maternal coffee consumption may affect the fetus at any time throughout pregnancy, given that the effector systems are mature (see earlier).

Some studies have raised concern about fetuses being exposed to caffeine during pregnancy, but others have failed to find any associations between maternal caffeine intake and adverse perinatal outcomes.

There is some evidence from animal studies that high doses of caffeine may result in malformations of the fetus, including cleft palate and cardiovascular malformations (for a review, see Nehlig and Debry 1994). Epidemiology studies, though, have not been able to detect significant risks for teratogenic effects of caffeine exposure in humans (Browne 2006). Some studies have shown that excessive maternal consumption of caffeine in humans (more than 300 mg/day) may be related to reduced fertility (Marie-Soleil and Graham 2010) and was associated with increased rate of spontaneous abortions (Godel et al. 1992; Klebanoff et al. 1999; Cnattingius et al. 2000; Wen et al. 2001), intrauterine growth restriction (Bracken et al. 2003a; Klebanoff et al. 2002; Vlajinac et al. 1997; CARE Study Group 2008), and still birth (Wisborg et al. 2003). One theoretically possible explanation for some of these effects may be that caffeine increases the levels of catecholamines in the mother and in the fetus, which may induce uteroplacental vasoconstriction (Kirkinen et al. 1983).

On the other hand, several prospective epidemiology studies have shown no major effects of caffeine on ovulation (Chavarro et al. 2009; Chap. 21), intrauterine growth restriction, low birth weight, or preterm delivery (Bracken et al. 2003a).

The major limitations of the epidemiology studies in this field lie in their retrospective nature and the data on caffeine exposure are thus subject to recall and other types of bias and that some studies lack information on confounding variables such as smoking. Confounding due to pregnancy symptoms is another important issue that complicates the relation between caffeine consumption and spontaneous abortion. Nausea may influence the amount of caffeine consumed in early pregnancy and it is also a marker of fetal viability.

Only one randomized controlled study investigated the effect of caffeine versus restricted caffeine intake on pregnancy outcome (Bech et al. 2007). Caffeinated instant coffee was compared with decaffeinated coffee during the second and the

third trimester. A moderate caffeine reduction of 182 mg/day did not affect birth weight or length of gestation. Thus, there is currently insufficient evidence for advising mothers to avoid caffeine during the last two thirds of pregnancy.

The few studies that have examined theophylline in pregnancy in relation to perinatal outcomes have found no association between theophylline and low birth weight (Schatz et al. 2004) or small size for gestational age (Schatz et al. 2004; Stenius-Aarniala et al. 1995; Bracken et al. 2003b).

## 4 Exposure to Methylxanthines in Neonates

### 4.1 Absorption, Metabolism, and Elimination

Methylxanthines, from breast milk or given orally to infants for treatment of apneas, are rapidly and completely absorbed from the gastrointestinal tract of the neonate with a minimal first-pass effect (Arnaud 1987). Once absorbed, they freely enter all body tissues, including the brain and gonads (Arnaud 1987).

Pharmacokinetic studies have shown that newborn infants (up to about term-equivalent age) have a prolonged half life of caffeine of around 100 h (see Table 1), which decreases with gestational age (Aranda et al. 1979), reflecting the immaturity of the hepatic CYP1A2 enzyme system. The enzyme capacity then gradually improves and reaches adult function at about 3 months (Aranda et al. 1979) after birth.

Like caffeine, theophylline is readily absorbed orally (Ogilvie 1978) and no dose adjustment is needed when switching from intravenous to oral administration. Theophylline is, like caffeine, metabolized by CYP1A2, but has variable pharmacokinetics during the first few days in newborns, which is why monitoring of the plasma concentrations is required. Although there is a faster metabolic clearance of theophylline relative to caffeine in neonates, because theophylline can be back-methylated to caffeine, the net effect of methylxanthines may be the same since caffeine and theophylline exert similar biological actions. The half life in premature infants is 20–30 h.

Even though caffeine and theophylline are excreted in the urine mainly unchanged in the neonate, significant methylation of theophylline to caffeine occurs and the latter may exert additional pharmacological effects (Dani et al. 2000).

### 4.2 Methylxanthines and Breast Feeding

As mentioned already, although caffeine is excreted to a limited extent in breast milk, the immature metabolism of methylxanthines in neonates and, in particular, in preterm infants (Aranda et al. 1979) makes them at risk of accumulating

methylxanthines. Theophylline (e.g., given to the mother for asthma) and theobromine (from chocolate) are also present in breast milk after administration (Yurchak and Jusko 1976) and accumulate in the neonate (Aranda et al. 1976).

While some studies have suggested possible risk effects of methylxanthines in pregnancy (see Sect. 3.2), a growth-promoting effect has been demonstrated in breast-feeding rat pups (Hart and Grimble 1990), which was shown to be due to increased milk volume. By contrast, there are a few animal studies showing that theophylline (Milsap et al. 1980; Carnielli et al. 2000) and caffeine (Bauer et al. 2001) increase the metabolic rate, which would infer a negative effect on growth. Similarly, a minimal decrease in fetal growth was observed in humans after excessive maternal caffeine drinking (more than 600 mg/day, about six cups of coffee) (Bracken et al. 2003a).

There has also been concern over the use of caffeine in mothers at the particular time point around delivery. Adenosine acting via adenosine receptors has endogenous neuroprotective effects in the adult brain (Fredholm 2007; Müller and Jacobson 2010). Although results in the neonate are complex (for a review, see Millar and Schmidt 2004), it was anticipated that the presence of the antagonist caffeine would possibly not be beneficial in a situation of birth asphyxia or postnatal apnea (with subsequent hypoxia). It was therefore surprising that when rat dams were given caffeine in their drinking water during pregnancy and lactation, in a dose that produced plasma concentrations similar to those achieved after 300–400 mg/day in humans, hypoxic ischemic brain damage was reduced by about 30% (Bona et al. 1995). No major effects on adenosine receptors or GABA<sub>A</sub> receptors were found (Aden et al. 2000). A similar perinatal exposure to caffeine prevented periventricular white matter damage in mice reared in hypoxia by enhancing myelination (Back et al. 2006). Further studies in adenosine-A<sub>1</sub>-receptor-knockout mice (Turner et al. 2003) indicated that blockade of this receptor might contribute to the protective effect of caffeine in the immature brain. It is also possible that the anti-inflammatory effects of caffeine (Haskó and Cronstein 2010; Ohta and Sitkovsky 2010) may be partly instrumental for the beneficial effects of caffeine in the developing brain.

### 4.3 *Methylxanthines for Apnea of Prematurity*

More than three decades ago, it was demonstrated that methylxanthines can reduce the frequency of apneic episodes in premature infants (Kuzemko 1973; Aranda et al. 1977). Since then, methylxanthines have become part of the routine clinical management of apnea of prematurity. Apnea commonly occurs in premature infants below 34 weeks of gestational age and is a cessation of breathing due to immaturity of the respiratory drive, followed by decreased oxygen saturation in the blood and bradycardia. Recommended doses and desired plasma concentrations for

**Table 1** Pharmacokinetics of methylxanthines in neonates. Data from Dani et al. (2000) and Aranda et al. (1976)

	Theophylline <sup>a</sup>	Caffeine
Plasma levels in neonates after moderate maternal coffee intake	No data available	0.2–1 mg/L (Neims and von Borstel 1983)
Route of administration	Intravenous, per os	Intravenous, per os
Dose (mg/kg)		
Loading	4–6	20
Maintenance	1.5–3 every 8–12 hr	5 (–10) every 24 hr
Plasma half life (h, range)	20–30	100 (40–230)
Therapeutic window (mg/L)	6–12	5–25
Adverse effects	Commonly include tachycardia, vomiting, hyperglycemia, irritability, sleeplessness	Usually mild, but includes tachycardia, vomiting, restlessness

<sup>a</sup>Theophylline is often administered as the salt aminophylline, which consists of approximately 80% theophylline. If changing from intravenous administration of aminophylline to oral administration, the dose needs to be increased by 20%.

therapeutic use of caffeine and theophylline in premature neonates are shown in Table 1. It is important to note that the plasma concentration of caffeine used to treat this condition is 10–100 times higher than the plasma concentrations reached if the mother drinks moderate doses of coffee and the child is exposed via breast milk (Table 1). The methylxanthine therapy often goes on for several weeks until the premature child has matured.

Caffeine and theophylline exert similar effects, but have differences in pharmacokinetic properties (Table 1). In clinical practice, caffeine citrate has now become the methylxanthine of choice because it has a wider therapeutic range, has essentially complete full bioavailability, and can thus be given orally and routine measurements of blood concentrations are not needed.

As mentioned, given the neuroprotective effects of endogenous adenosine, there has been concern over the effects of high doses of methylxanthines in neonates at risk of apneas and postnatal hypoxic ischemia. It was therefore surprising that when rat pups were given theophylline in therapeutically relevant doses just before the induction of a hypoxia ischemia, the brain damage was reduced by almost 50% (Bona et al. 1997). The authors speculated that the mechanism for this protection might include anti-inflammatory effects of theophylline.

To this end, in a study where premature infants were randomized to either caffeine or a placebo for apnea of prematurity, it was shown that caffeine improved the rate of survival, decreased the number of children with severe respiratory sequelae (bronchopulmonary dysplasia), and reduced the incidences of cerebral palsy and cognitive delay at 18 months of age (Schmidt et al. 2007). With use of post hoc analysis, about half of the protective effect of caffeine was attributed to positive effects on the respiration, but half of the effect remained unexplained. These results indicate that caffeine has inherent direct or indirect neuroprotective effects.



## 5 Short-Term Effects of Methylxanthines in the Infant

### 5.1 *Physiological Effects*

Methylxanthines exert profound stimulatory effects on respiratory drive in neonates by antagonism of adenosine A<sub>1</sub> receptors in the pons and medulla oblongata (Lagercrantz et al. 1984; Tilley 2010), improved chemoreceptor sensitivity to CO<sub>2</sub> (Davi et al. 1978), mainly via blockade of adenosine A<sub>2A</sub> receptors (Conde et al. 2006), increased oxygen consumption (Bauer et al. 2001), and increased cardiac output (Walther et al. 1990). As mentioned already, experimental data show that adenosine A<sub>1</sub> receptors are clearly G-protein-coupled (Aden et al. 2001) and capable of function at birth in the heart. Accordingly, a commonly seen clinical effect/side effect of methylxanthines is tachycardia.

Adverse effects are similar for theophylline and caffeine (Table 1) but are milder and occur less often for caffeine. One specific adverse effect of theophylline is that cerebral blood hemodynamics are transiently affected (Dani et al. 2000), possibly via blockade of cerebrovascular adenosine A<sub>2A</sub> receptors

Renal effects of methylxanthines include increased diuresis (via tubular adenosine A<sub>1</sub> receptors) and increased calcium excretion (Bauer et al. 2001; McPhee and Whiting 1989; Rieg et al. 2005).

Unfavorable symptoms can also occur in neonates if a chronic administration of methylxanthines during pregnancy suddenly ceases, i.e., if breast feeding is not established early after birth.

### 5.2 *Withdrawal Symptoms*

There are case reports in the literature on transient withdrawal symptoms in full-term neonates exposed to high concentrations of caffeine after excessive maternal ingestion of coffee, mate, or cola drinks (450–1,800 mg/day). These symptoms include jitteriness, high-pitched cry, hypertonia in the limbs, brisk tendon reflexes, and vomiting and resolved spontaneously within 1–2 days (Khanna and Somani 1984; McGowan et al. 1988). In these cases the withdrawal symptoms were preceded by lack of breast feeding.

It is also possible that some apneic episodes encountered in neonates who were exposed to caffeine during pregnancy but then were not breast-fed, may be due to a central withdrawal effect of caffeine at the pontomedullary level. This speculation was supported by experimental data from neonatal rat pups where caffeine was given in the drinking water to pregnant and lactating dams. If caffeine was withdrawn at birth, apneas could be induced and subsequent caffeine exposure during breast feeding was able to prevent apneas in the rat pups (Bodineau et al. 2006).

## 6 Long-Term Effects of Methylxanthines for the Developing Organism

Even though the short-term effects of caffeine, in relevant doses, seem to be largely beneficial, there has been considerable concern over the long-term effects of methylxanthines in the developing brain and other organs.

### 6.1 CNS Function

Exposure to psychostimulant drugs during brain development may lead to long-term effects beyond the time point when the drug exposure is withdrawn (Andersen 2005). There is some experimental evidence that high doses of methylxanthines induce long-lasting behavioral changes in the offspring (Nehlig and Debray 1994; Henderson et al. 1991; Nakamoto et al. 1991). Also, when a modest dose of caffeine (similar to plasma levels achieved in moderate maternal coffee drinking) was given in the drinking water to mice throughout pregnancy and lactation, the adult offspring exhibited increased locomotor activity in an open field (Nehlig and Debray 1994). A similar behavioral profile was found in mice heterozygous for the adenosine A<sub>1</sub> receptor gene, where signaling via adenosine A<sub>1</sub> receptors was reduced to about the same degree as after modest consumption of caffeine (Bjorklund et al. 2008b). Furthermore, it appeared that the mother's genotype, not the offspring's, was critical for behavioral changes in adult offspring, thus indicating that perinatal caffeine acting on adenosine A<sub>1</sub> receptors in the mother caused a long-term effect in the offspring (Bjorklund et al. 2008b). Interestingly, these effects even manifested themselves in the second generation (Bjorklund et al. 2008b).

However, in humans, perinatal exposure to coffee per se was not related to any increased risk of having ADHD or hyperkinetic diagnosis (Linnet et al. 2009). Similarly, in a follow-up study of 500 pregnant women and their offspring, there was no association between perinatal caffeine exposure and IQ and attention tests at 7 years of age (Barr and Streissguth 1991). Furthermore in a large randomized controlled trial on caffeine (given postnatally in high doses) for apnea of prematurity, a reduced incidence of cerebral palsy and cognitive delay at 18 months of age was shown (Schmidt et al. 2007).

Although no detrimental long-term effects from perinatal methylxanthine exposure have been demonstrated in humans so far, studies on development of CNS functions require a very long follow-up period and complex neuropsychological testing; therefore, effects may nevertheless be present but difficult to detect.

### 6.2 Respiration and Cardiovascular Function

There is some experimental evidence that neonatal caffeine treatment with plasma levels comparable to these achieved with treatment for apnea of prematurity alters

the ventilatory response to hypercapnia and hypoxia in adolescence and adulthood (Montandon et al. 2008). Mainly adenosine A<sub>1</sub> receptors were found to be involved in these plasticity changes, which speculatively might have implications for diseases implied in respiratory control dysfunction such as SIDS and sleep apnea (Montandon et al. 2008).

It is also conceivable that perinatal exposure to caffeine may have persisting effects on cardiovascular function, since adenosine is an important regulator thereof (Riksen et al. 2010). Effector systems for adenosine receptors actually mature earlier in the cardiovascular system than in the CNS, at least in rats (Aden et al. 2001), and there is some evidence that early caffeine exposure can alter gene expression of adenosine and dopamine receptors and tyrosine hydroxylase in the carotid body and adrenal glands of rats (Montandon et al. 2008). Another study showed adverse effects of a dose of caffeine relevant to daily intake of coffee on embryonic arteries with transiently decreased blood flow. These effects were adenosine A<sub>2A</sub> dependent (Momoi et al. 2008). It is thus possible that early exposure to caffeine might have long-lasting cardiovascular effects, but there is currently a lack of long-term follow-up data of cardiorespiratory function and human data.

As mentioned, an important positive influence of perinatal caffeine on lung function was seen in a randomized controlled study of apnea of prematurity, where caffeine decreased the number of children with a chronic respiratory disease (bronchopulmonary dysplasia) at 18 months of age (Schmidt et al. 2007). The major protective effect of caffeine in this context was that ventilator-induced lung injury was avoided to a large extent, because the caffeine-treated infants came off the ventilator earlier than the controls (Schmidt et al. 2007). Another possibly contributing effect might be that caffeine and other methylxanthines have inherent immunomodulatory mechanisms (Haskó and Cronstein 2010; Ohta and Sitkovsky 2010) that might protect both from lung and brain injury.

## 7 Conclusions

Many fetuses and neonates are exposed to low levels of methylxanthines owing to maternal drinking of coffee. Much higher doses of caffeine and theophylline are used in long-term pharmacological treatment of apneas in preterm infants. Pregnant women, fetuses, and neonates have an inability to detoxify methylxanthines, rendering the developing organism with accumulating levels of methylxanthines, which implies that there is a risk for adverse effects. Brain development is of particular concern.

Animal studies, however, suggest that the major effector system for methylxanthines, adenosine receptors, is not fully developed in the brain at birth. Furthermore, animal studies have shown protective positive effects of methylxanthines in situations of hypoxia/ischemia in neonates. Similarly, a positive long-term effect on lung function and CNS development was found in human preterm infants treated

with high doses of caffeine for apneas. There is now evidence that the overall benefits from methylxanthine therapy for apnea of prematurity outweigh potential risks in the short term.

Experimental studies, however, have indicated that long-term effects of low-dose caffeine during pregnancy and lactation may include altered behavior in the offspring, but there are currently no human data to support this.

Negative effects on pregnancy and perinatal outcomes have been reported in epidemiology studies, but the results are inconclusive. The evidence base for adverse effects of caffeine in first third of pregnancy is stronger than for later parts of pregnancy. Whereas it may be prudent for women in early pregnancy to limit caffeine intake (less than 300 mg/day), there is currently insufficient scientific evidence to advise mothers to avoid consuming caffeine during later parts of pregnancy.

## References

- Abbracchio MP, Camurri A, Ceruti S, Cattabeni F, Falzano L, Giammarioli AM, Jacobson KA, Trincavelli L, Martini C, Malorni W, Fiorentini C (2001) The A3 adenosine receptor induces cytoskeleton rearrangement in human astrocytoma cells via a specific action on rho proteins. *Ann N Y Acad Sci* 939:63–73
- Aden U, Herlenius E, Tang LQ, Fredholm BB (2000) Maternal caffeine intake has minor effects on adenosine receptor ontogeny in the rat brain. *Pediatr Res* 48:177–183
- Aden U, Leverin AL, Hagberg H, Fredholm BB (2001) Adenosine A(1) receptor agonism in the immature rat brain and heart. *Eur J Pharmacol* 426:185–192
- Aldridge A, Bailey J, Neims AH (1981) The disposition of caffeine during and after pregnancy. *Semin Perinatol* 5:310–314
- Andersen SL (2005) Stimulants and the developing brain. *Trends Pharmacol Sci* 26:237–243
- Aranda JV, Sitar DS, Parsons WD, Loughnan PM, Neims AH (1976) Pharmacokinetic aspects of theophylline in premature newborns. *N Engl J Med* 295:413–416
- Aranda JV, Gorman W, Bergsteinsson H, Gunn T (1977) Efficacy of caffeine in treatment of apnea in the low-birth-weight infant. *J Pediatr* 90:467–472
- Aranda JV, Collinge JM, Zinman R, Watters G (1979) Maturation of caffeine elimination in infancy. *Arch Dis Child* 54:946–949
- Arnaud MJ (1987) The pharmacology of caffeine. *Prog Drug Res* 31:273–313
- Back SA, Craig A, Luo NL, Ren J, Akundi RS, Ribeiro I, Rivkees SA (2006) Protective effects of caffeine on chronic hypoxia-induced perinatal white matter injury. *Ann Neurol* 60:696–705
- Barr HM, Streissguth AP (1991) Caffeine use during pregnancy and child outcome: a 7-year prospective study. *Neurotoxicol Teratol* 13:441–448
- Bauer J, Maier K, Linderkamp O, Hentschel R (2001) Effect of caffeine on oxygen consumption and metabolic rate in very low birth weight infants with idiopathic apnea. *Pediatrics* 107:660–663
- Bech BH, Obel C, Henriksen TB, Olsen J (2007) Effect of reducing caffeine intake on birth weight and length of gestation: randomised controlled trial. *BMJ* 334:409
- Bjorklund O, Halldner-Henriksson L, Yang J, Eriksson TM, Jacobson MA, Dare E, Fredholm BB (2008a) Decreased behavioral activation following caffeine, amphetamine and darkness in A3 adenosine receptor knock-out mice. *Physiol Behav* 95:668–676
- Bjorklund O, Kahlstrom J, Salmi P, Fredholm BB (2008b) Perinatal caffeine, acting on maternal adenosine A(1) receptors, causes long-lasting behavioral changes in mouse offspring. *PLoS ONE* 3:e3977

- Bodineau L, Saadani-Makki F, Jullien H, Frugiere A (2006) Caffeine in the milk prevents respiratory disorders caused by in utero caffeine exposure in rats. *Respir Physiol Neurobiol* 150:94–98
- Bona E, Aden U, Fredholm BB, Hagberg H (1995) The effect of long term caffeine treatment on hypoxic-ischemic brain damage in the neonate. *Pediatr Res* 38:312–318
- Bona E, Aden U, Gilland E, Fredholm BB, Hagberg H (1997) Neonatal cerebral hypoxia-ischemia: the effect of adenosine receptor antagonists. *Neuropharmacology* 36:1327–1338
- Bracken MB, Triche EW, Belanger K, Hellenbrand K, Leaderer BP (2003a) Association of maternal caffeine consumption with decrements in fetal growth. *Am J Epidemiol* 157:456–466
- Bracken MB, Triche EW, Belanger K, Saftlas A, Beckett WS, Leaderer BP (2003b) Asthma symptoms, severity, and drug therapy: a prospective study of effects on 2205 pregnancies. *Obstet Gynecol* 102:739–752
- Browne ML (2006) Maternal exposure to caffeine and risk of congenital anomalies: a systematic review. *Epidemiology* 17:324–331
- CARE Study Group (2008) Maternal caffeine intake during pregnancy and risk of fetal growth restriction: a large prospective observational study. *BMJ* 337:a2332
- Carnielli VP, Verlato G, Benini F, Rossi K, Cavedagni M, Filippone M, Baraldi E, Zacchello F (2000) Metabolic and respiratory effects of theophylline in the preterm infant. *Arch Dis Child Fetal Neonatal Ed* 83:F39–F43
- Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC (2009) Caffeinated and alcoholic beverage intake in relation to ovulatory disorder infertility. *Epidemiology* 20:374–381
- Chavez-Valdez R, Wills-Karp M, Ahlawat R, Cristofalo EA, Nathan A, Gauda EB (2009) Caffeine modulates tnfr-alpha production by cord blood monocytes: the role of adenosine receptors. *Pediatr Res* 65:203–208
- Cnattingius S, Signorello LB, Anneren G, Clausson B, Ekblom A, Ljunger E, Blot WJ, McLaughlin JK, Petersson G, Rane A, Granath F (2000) Caffeine intake and the risk of first-trimester spontaneous abortion. *N Engl J Med* 343:1839–1845
- Conde SV, Obeso A, Vicario I, Rigual R, Rocher A, Gonzalez C (2006) Caffeine inhibition of rat carotid body chemoreceptors is mediated by A2A and A2B adenosine receptors. *J Neurochem* 98:616–628
- Cook DG, Peacock JL, Feyerabend C, Carey IM, Jarvis MJ, Anderson HR, Bland JM (1996) Relation of caffeine intake and blood caffeine concentrations during pregnancy to fetal growth: prospective population based study. *BMJ* 313:1358–1362
- Cosio BG, Tsaprouni L, Ito K, Jazrawi E, Adcock IM, Barnes PJ (2004) Theophylline restores histone deacetylase activity and steroid responses in COPD macrophages. *J Exp Med* 200:689–695
- Dani C, Bertini G, Reali MF, Tronchin M, Wiechmann L, Martelli E, Rubaltelli FF (2000) Brain hemodynamic changes in preterm infants after maintenance dose caffeine and aminophylline treatment. *Biol Neonate* 78:27–32
- Davi MJ, Sankaran K, Simons KJ, Simons FE, Seshia MM, Rigatto H (1978) Physiologic changes induced by theophylline in the treatment of apnea in preterm infants. *J Pediatr* 92:91–95
- Eldridge FL, Millhorn DE, Kiley JP (1985) Antagonism by theophylline of respiratory inhibition induced by adenosine. *J Appl Physiol* 59:1428–1433
- Eskenazi B (1999) Caffeine—filtering the facts. *N Engl J Med* 341:1688–1689
- Fenster L, Quale C, Hiatt RA, Wilson M, Windham GC, Benowitz NL (1998) Rate of caffeine metabolism and risk of spontaneous abortion. *Am J Epidemiol* 147:503–510
- Fredholm BB, Battig K, Holmen J, Nehlig A, Zvartau EE (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fredholm BB (2007) Adenosine, an endogenous distress signal, modulates tissue damage and repair. *Cell Death Differ* 14:1315–1323
- Godel JC, Pabst HF, Hodges PE, Johnson KE, Froese GJ, Joffres MR (1992) Smoking and caffeine and alcohol intake during pregnancy in a northern population: effect on fetal growth. *CMAJ* 147:181–188

- Grosso LM, Bracken MB (2005) Caffeine metabolism, genetics, and perinatal outcomes: a review of exposure assessment considerations during pregnancy. *Ann Epidemiol* 15: 460–466
- Hammarberg C, Schulte G, Fredholm BB (2003) Evidence for functional adenosine A3 receptors in microglia cells. *J Neurochem* 86:1051–1054
- Hart AD, Grimble RF (1990) The effect of methylxanthines on milk volume and composition, and growth of rat pups. *Br J Nutr* 64:339–350
- Haskó G, Cronstein B (2010) Methylxanthines and inflammatory cells. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Henderson MG, McConnaughey MM, McMillen BA (1991) Long-term consequences of prenatal exposure to cocaine or related drugs: effects on rat brain monoaminergic receptors. *Brain Res Bull* 26:941–945
- Herlenius E, Aden U, Tang LQ, Lagercrantz H (2002) Perinatal respiratory control and its modulation by adenosine and caffeine in the rat. *Pediatr Res* 51:4–12
- Ito K, Lim S, Caramori G, Cosio B, Chung KF, Adcock IM, Barnes PJ (2002) A molecular mechanism of action of theophylline: induction of histone deacetylase activity to decrease inflammatory gene expression. *Proc Natl Acad Sci USA* 99:8921–8926
- Johansson B, Halldner L, Dunwiddie TV, Masino SA, Poelchen W, Gimenez-Llort L, Escorihuela RM, Fernandez-Teruel A, Wiesenfeld-Hallin Z, Xu XJ, Hardemark A, Betsholtz C, Herlenius E, Fredholm BB (2001) Hyperalgesia, anxiety, and decreased hypoxic neuroprotection in mice lacking the adenosine A1 receptor. *Proc Natl Acad Sci USA* 98:9407–9412
- Kalow W, Tang BK (1991) Use of caffeine metabolite ratios to explore CYP1A2 and xanthine oxidase activities. *Clin Pharmacol Ther* 50:508–519
- Khanna NN, Somani SM (1984) Maternal coffee drinking and unusually high concentrations of caffeine in the newborn. *J Toxicol Clin Toxicol* 22:473–483
- Kirkinen P, Jouppila P, Koivula A, Vuori J, Puukka M (1983) The effect of caffeine on placental and fetal blood flow in human pregnancy. *Am J Obstet Gynecol* 147:939–942
- Klebanoff MA, Levine RJ, DerSimonian R, Clemens JD, Wilkins DG (1999) Maternal serum paraxanthine, a caffeine metabolite, and the risk of spontaneous abortion. *N Engl J Med* 341:1639–1644
- Klebanoff MA, Levine RJ, Clemens JD, Wilkins DG (2002) Maternal serum caffeine metabolites and small-for-gestational age birth. *Am J Epidemiol* 155:32–37
- Kuzemko JA (1973) Aminophylline in apnoeic attacks of newborn. *Lancet* 1:1509
- Lagercrantz H, Yamamoto Y, Fredholm BB, Prabhakar NR, von Euler C (1984) Adenosine analogues depress ventilation in rabbit neonates. Theophylline stimulation of respiration via adenosine receptors? *Pediatr Res* 18:387–390
- Ledent C, Vaugeois JM, Schiffmann SN, Pedrazzini T, El Yacoubi M, Vanderhaeghen JJ, Costentin J, Heath JK, Vassart G, Parmentier M (1997) Aggressiveness, hypoalgesia and high blood pressure in mice lacking the adenosine A2A receptor. *Nature* 388:674–678
- Leinekugel X, Medina I, Khalilov I, Ben-Ari Y, Khazipov R (1997) Ca<sup>2+</sup> oscillations mediated by the synergistic excitatory actions of GABA(A) and NMDA receptors in the neonatal hippocampus. *Neuron* 18:243–255
- Linet KM, Wisborg K, Secher NJ, Thomsen PH, Obel C, Dalsgaard S, Henriksen TB (2009) Coffee consumption during pregnancy and the risk of hyperkinetic disorder and ADHD: a prospective cohort study. *Acta Paediatr* 98:173–179
- Marie-Soleil B, Graham TE (2010) Methylxanthines and human health. Epidemiological and experimental evidence. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- McGowan JD, Altman RE, Kanto WP Jr (1988) Neonatal withdrawal symptoms after chronic maternal ingestion of caffeine. *South Med J* 81:1092–1094
- McPhee MD, Whiting SJ (1989) The effect of adenosine and adenosine analogues on methylxanthine-induced hypercalciuria in the rat. *Can J Physiol Pharmacol* 67:1278–1282
- Millar D, Schmidt B (2004) Controversies surrounding xanthine therapy. *Semin Neonatol* 9:239–244

- Milsap RL, Krauss AN, Auld PA (1980) Oxygen consumption in apneic premature infants after low-dose theophylline. *Clin Pharmacol Ther* 28:536–540
- Momoi N, Tinney JP, Liu LJ, Elshershari H, Hoffmann PJ, Ralphe JC, Keller BB, Tobita K (2008) Modest maternal caffeine exposure affects developing embryonic cardiovascular function and growth. *Am J Physiol Heart Circ Physiol* 294:H2248–H2256
- Montandon G, Kinkead R, Bairam A (2008) Adenosinergic modulation of respiratory activity: developmental plasticity induced by perinatal caffeine administration. *Respir Physiol Neurobiol* 164:87–95
- Müller C, Jacobson KA (2010) Xanthines as adenosine receptor antagonists. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Nakamoto T, Roy G, Gottschalk SB, Yazdani M, Rossowska M (1991) Lasting effects of early chronic caffeine feeding on rats' behavior and brain in later life. *Physiol Behav* 49:721–727
- Nehlig A, Debry G (1994) Potential teratogenic and neurodevelopmental consequences of coffee and caffeine exposure: a review on human and animal data. *Neurotoxicol Teratol* 16:531–543
- Neims AH, von Borstel RW (1983) Caffeine: Metabolism and biochemical mechanisms of action. In: Wurtman RJ, Wurtman JJ (Eds) *Nutrition and the Brain*, Vol. 6. Raven Press, New York, pp 1–30
- Ogilvie RI (1978) Clinical pharmacokinetics of theophylline. *Clin Pharmacokinet* 3:267–293
- Ohta A, Sitkovsky M (2001) Role of g-protein-coupled adenosine receptors in downregulation of inflammation and protection from tissue damage. *Nature* 414:916–920
- Ohta A, Sitkovsky M (2010) Methylxanthines, inflammation and cancer: fundamental mechanisms. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Rieg T, Steigele H, Schnermann J, Richter K, Osswald H, Vallon V (2005) Requirement of intact adenosine A1 receptors for the diuretic and natriuretic action of the methylxanthines theophylline and caffeine. *J Pharmacol Exp Ther* 313:403–409
- Riksen NP, Smits P, Rongen GA (2010) The cardiovascular effects of methylxanthines. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Rivkees SA (1995) The ontogeny of cardiac and neural A1 adenosine receptor expression in rats. *Brain Res Dev Brain Res* 89:202–213
- Schatz M, Dombrowski MP, Wise R, Momirova V, Landon M, Mabie W, Newman RB, Hauth JC, Lindheimer M, Caritis SN, Leveno KJ, Meis P, Miodovnik M, Wapner RJ, Paul RH, Varner MW, O'Sullivan MJ, Thurnau GR, Conway DL (2004) The relationship of asthma medication use to perinatal outcomes. *J Allergy Clin Immunol* 113:1040–1045
- Schmidt B, Roberts RS, Davis P, Doyle LW, Barrington KJ, Ohlsson A, Solimano A, Tin W (2007) Long-term effects of caffeine therapy for apnea of prematurity. *N Engl J Med* 357:1893–1902
- Stenius-Aarniala B, Riikonen S, Teramo K (1995) Slow-release theophylline in pregnant asthmatics. *Chest* 107:642–647
- Tilley SL (2010) Methylxanthines in asthma. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Tsutsumi K, Kotegawa T, Matsuki S, Tanaka Y, Ishii Y, Kodama Y, Kuranari M, Miyakawa I, Nakano S (2001) The effect of pregnancy on cytochrome P4501A2, xanthine oxidase, and N-acetyltransferase activities in humans. *Clin Pharmacol Ther* 70:121–125
- Turner CP, Seli M, Ment L, Stewart W, Yan H, Johansson B, Fredholm BB, Blackburn M, Rivkees SA (2003) A1 adenosine receptors mediate hypoxia-induced ventriculomegaly. *Proc Natl Acad Sci USA* 100:11718–11722
- Varani K, Portaluppi F, Gessi S, Merighi S, Vincenzi F, Cattabriga E, Dalpiaz A, Bortolotti F, Belardinelli L, Borea PA (2005) Caffeine intake induces an alteration in human neutrophil A2A adenosine receptors. *Cell Mol Life Sci* 62:2350–2358
- Vlajinac HD, Petrovic RR, Marinkovic JM, Sipetic SB, Adanja BJ (1997) Effect of caffeine intake during pregnancy on birth weight. *Am J Epidemiol* 145:335–338
- Walther FJ, Erickson R, Sims ME (1990) Cardiovascular effects of caffeine therapy in preterm infants. *Am J Dis Child* 144:1164–1166

- Wen W, Shu XO, Jacobs DR Jr, Brown JE (2001) The associations of maternal caffeine consumption and nausea with spontaneous abortion. *Epidemiology* 12:38–42
- Wisborg K, Kesmodel U, Bech BH, Hedegaard M, Henriksen TB (2003) Maternal consumption of coffee during pregnancy and stillbirth and infant death in first year of life: prospective study. *BMJ* 326:420
- Yang JN, Tiselius C, Dare E, Johansson B, Valen G, Fredholm BB (2007) Sex differences in mouse heart rate and body temperature and in their regulation by adenosine A1 receptors. *Acta Physiol (Oxf)* 190:63–75
- Yurchak AM, Jusko WJ (1976) Theophylline secretion into breast milk. *Pediatrics* 57:518–520



# Methylxanthines and the Kidney

Hartmut Osswald and Jürgen Schnermann

## Contents

1	Introduction .....	392
2	Diuresis .....	393
3	Natriuresis .....	394
4	Hemodynamics .....	396
5	Renin Secretion .....	398
6	Disease and Therapeutic Aspects .....	399
6.1	Polycystic Kidney Disease .....	399
6.2	Nephropathies .....	399
6.3	Radiocontrast Nephropathy .....	400
6.4	Calcineurin Inhibitors .....	402
6.5	Cisplatin .....	402
6.6	Glycerol .....	403
6.7	Ischemia–Reperfusion .....	403
	References .....	404

**Abstract** This chapter describes the effects of the natural methylxanthines caffeine and theophylline on kidney function. Theophylline in particular was used traditionally to increase urine output until more potent diuretics became available in the middle of the last century. The mildly diuretic actions of both methylxanthines are mainly the result of inhibition of tubular fluid reabsorption along the renal proximal

---

H. Osswald

Department of Pharmacology and Toxicology, University of Tübingen, Wilhelmstrasse 56, 72074 Tübingen, Germany

J. Schnermann (✉)

National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Building 10, Room 4D51, 10 Center Drive MSC 1370, Bethesda, MD 20892, USA

e-mail: jurgens@intra.niddk.nih.gov

tubule. Based upon the use of specific adenosine receptor antagonists and the observation of a complete loss of diuresis in mice with targeted deletion of the A1AR gene, transport inhibition by methylxanthines is mediated mainly by antagonism of adenosine A1 receptors (A1AR) in the proximal tubule. Methylxanthines are weak renal vasodilators, and they act as competitive antagonists against adenosine-induced preglomerular vasoconstriction. Caffeine and theophylline stimulate the secretion of renin by inhibition of adenosine receptors and removal of the general inhibitory brake function of endogenous adenosine. Since enhanced intrarenal adenosine levels lead to reduced glomerular filtration rate in several pathological conditions theophylline has been tested for its therapeutic potential in the renal impairment following administration of nephrotoxic substances such as radiocontrast media, cisplatin, calcineurin inhibitors or following ischemia-reperfusion injury. In experimental animals functional improvements have been observed in all of these conditions, but available clinical data in humans are insufficient to affirm a definite therapeutic efficacy of methylxanthines in the prevention of nephrotoxic or postischemic renal injury.

**Keywords** Adenosine · Caffeine · Diuresis · Natriuresis · Nephropathy · Renin · Theophylline

## 1 Introduction

The plant constituents caffeine and theophylline have been known to alter kidney function since the demonstration in 1864 that caffeine can increase urine production in patients with congestive heart failure and edema (Koschlakoff 1864). Interest in the renal actions of methylxanthines has remained acute although the clinical use of theophylline is largely restricted to the treatment of extra-renal diseases such as asthma. The focus of this chapter is a description of the renal effects of the natural methylxanthines, mainly of caffeine and theophylline, and a discussion of current understanding of the mechanisms underlying these effects. Important progress in mechanistic thinking has been made by using modified methylxanthines, and it will therefore sometimes be necessary to include data obtained with synthetic xanthine derivatives when this permits discrimination between the different targets of the natural compounds. This is especially relevant in the case of methylxanthines as antagonists of adenosine receptors, a predominant mechanism in many renal actions of methylxanthines. Nevertheless, a broad discussion of adenosine and its interaction with adenosine receptors is not the goal of this review, and adenosine will only be discussed in the context of understanding methylxanthine actions. We also will make reference to nonnatural xanthine compounds in cases where their actions are in conflict with those of caffeine or theophylline or where they shed light on likely mechanisms of action.

## 2 Diuresis

The natural methylxanthines caffeine and theophylline were used traditionally to increase urine output until more potent diuretics became available in the middle of the last century. In numerous studies in animal and humans the order of diuretic potency of the natural methylxanthines was found to be theophylline > caffeine > paraxanthine > theobromine. The actions of methylxanthines as diuretics have been extensively reviewed in an excellent chapter in the *Handbook of Experimental Pharmacology* in which the literature prior to 1970 was summarized and discussed in full detail (Fulgraff 1969). In the period since then, it has been confirmed repeatedly that caffeine and other methylxanthines can induce an increase in urine flow in humans and experimental animals. The dose of caffeine that elicits a significant acute diuresis has been reported to be on the order of 300 mg, the equivalent of about four to five cups of coffee (Grandjean et al. 2000; Passmore et al. 1987; Riesenhuber et al. 2006). As determined by impedance analysis, an acute intake of 642 mg of caffeine without a change in total fluid intake caused a measurable decrease in body weight corresponding to a 2.7% reduction of total body water (Neuhauser et al. 1997). Furthermore, a positive fluid balance prior to methylxanthine administration enhanced the diuretic efficacy of methylxanthines, while a negative fluid balance reduced the diuretic response (Fulgraff 1969). The diuretic potency of caffeine appears to be also modulated by age and habituation, with old age and previous exposure to caffeine causing further decreases in the diuretic effectiveness of caffeine (Izzo et al. 1983).

Overall, the relatively modest potency of caffeine to enhance water excretion and cause net water loss is consistent with studies in which caffeine (up to 6 mg/kg) given for 11 days did not affect 24-h urine volume and was not associated with symptoms of negative fluid balance (Armstrong et al. 2005). The same conclusion was drawn in a study where the diuretic effects of caffeinated and noncaffeinated electrolyte drinks were compared in individuals at rest and during moderate exercise (Wemple et al. 1997). While the ingestion of caffeine (25 mg/dl at 35 ml/kg) caused a higher urine flow in individuals at rest compared with ingestion of caffeine-free fluid, urine flow was reduced to the same level during exercise and this was associated with identical increments of plasma catecholamine concentrations. In a smaller study, consumption of tea as the major fluid source did not cause noticeable differences in hydration status compared with nontea fluid intake in a group of mountaineers at high altitude (Scott et al. 2004). Thus, the general advice against using caffeinated drinks for volume replacement may need to be qualified in that beverages containing moderate amounts of caffeine do not appear to cause significant fluid losses (Armstrong 2002).

Caffeine in high concentration supplied in a calcium-free medium has been shown to prevent the increase in calcium level caused by vasopressin in rat renal papillary collecting duct cells and this may blunt the increase of cyclic AMP (cAMP) level and the effect of vasopressin on water permeability (Ishikawa et al. 1992). Caffeine appears to exert this effect by depletion of endoplasmic calcium

stores, suggesting that its action may be caused by interaction with ryanodine-sensitive calcium-release channels. Nevertheless, there is no evidence in support of the notion that methylxanthines in lower concentrations inhibit solute-free water absorption in the distal part of the nephron. In fact, theophylline has been observed to mimic the effect of vasopressin on water permeability in isolated perfused collecting ducts and in the bladder of the toad (Grantham and Orloff 1968; Orloff and Handler 1962). Both vasopressin and theophylline increased cellular cAMP levels, indicating that the theophylline effects are mediated by inhibition of phosphodiesterase (PDE) (Handler et al. 1965). Caffeine is unlikely to reduce collecting duct water reabsorption through inhibition of adenosine receptors since in perfused and nonperfused collecting ducts adenosine has been shown to inhibit AVP-stimulated water permeability through activation of A<sub>1</sub> adenosine receptors (Edwards and Spielman 1994; Yagil 1990). Since A<sub>2</sub>-adenosine-receptor-mediated effects have not been identified (Edwards and Spielman 1994; Yagil 1990), caffeine would thus be expected to enhance, not inhibit, collecting duct water transport, just as inhibition of cAMP degradation by possible methylxanthine effects on PDE would not be predicted to inhibit water transport. Furthermore, there does not seem to be a direct effect of caffeine on vasopressin secretion since plasma vasopressin levels have been reported to be unaltered after caffeine ingestion (Izzo et al. 1983; Nussberger et al. 1990). One would conclude that the type of diuresis caused by methylxanthines is mostly or exclusively a solute diuresis.

### 3 Natriuresis

Increased urine flow caused by methylxanthines is accompanied by increased excretion of sodium, chloride, calcium, phosphate, magnesium, and other urinary solutes. Although methylxanthines have in some studies been found to increase the tubular sodium load, significant natriuresis can occur without changes in glomerular filtration rate (GFR) or renal blood flow (RBF), clearly indicating that the natriuresis caused by methylxanthines is predominantly the result of inhibition of tubular salt transport (Davis and Shock 1949; Ludens et al. 1970; Shirley et al. 2002). Natriuresis without hemodynamic changes was also caused by methylxanthines in premature infants and newborn rabbits (Gouyon and Guignard 1987; Mazkereth et al. 1997). The increased excretion of calcium caused by caffeine may have implications for calcium homeostasis. Abstinence from moderate daily caffeine consumption (200 mg or less) has been noted to significantly increase plasma concentrations of ionized calcium and to reduce PTH levels in women with a relatively low dietary calcium intake (Massey et al. 1994; Wise et al. 1996).

In addition to causing natriuresis and diuresis, the administration of theophylline to conscious rats (10–50 mg/kg oral) or anesthetized rabbits (15 mg/kg intravenously) was accompanied by increased urinary excretion of prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) and cAMP (Baer et al. 1983; Oliw et al. 1977). Pretreatment with indomethacin or meclofenamate prevented the natriuretic action of theophylline, and

conversely, theophylline caused a transient reversal of the antinatriuresis and antidiuresis elicited by indomethacin in patients with rheumatic diseases (Oliw et al. 1977; Seideman et al. 1987). The possibility that some effects of methylxanthines on urine excretion could be indirect is also supported by the observation that plasma and kidney atrial natriuretic factor (ANF) activity increased following prolonged caffeine ingestion (Eggertsen et al. 1993; Lee et al. 2002). Nevertheless, no change in plasma ANF was seen 2 h after an oral intake of a single 250 mg dose of caffeine (Nussberger et al. 1990). That the natriuretic effect of methylxanthines is probably not strictly dependent on ANF is also suggested by the observation that prior exposure to theophylline did not modify the natriuretic action of ANF (Beutler et al. 1990).

There is considerable evidence to indicate that methylxanthine-induced natriuresis is predominantly a consequence of inhibition of salt transport along the proximal convoluted tubule. Administration of 400 mg of caffeine to healthy human subjects caused an about 1.5-fold increase in sodium excretion and this was associated with an increase in the clearance of lithium (Shirley et al. 2002). A reduction of proximal solute reabsorption in humans as assessed by lithium clearance was also caused by theophylline and aminophylline (Beutler et al. 1990; Brater et al. 1983). At the level of the single tubule, systemic administration of theophylline (20 mg/kg subcutaneously) caused an about 20% reduction in proximal tubular reabsorptive capacity as determined with the split-droplet technique in the rat (Fulgraff 1969). Tubular microperfusion of the loop of Henle with solutions containing theophylline or 3-isobutyl-1-methylxanthine did not significantly alter chloride reabsorption, indicating that methylxanthines do not affect salt absorption along proximal straight tubules and thick ascending limbs (Schnermann et al. 1977). The effect of methylxanthines on sodium transport in tubular segments beyond the proximal tubule and the loop of Henle has not been explored in detail, although on the basis of indirect evidence an inhibitory action in more distal parts of the tubule has been proposed (Brater et al. 1983; Shirley et al. 2002).

The mechanism by which methylxanthines inhibit proximal NaCl reabsorption is related to their properties as antagonists of adenosine receptors. Xanthine derivatives such as doxofylline and enprofylline with low affinity for adenosine receptors, but similar ability to inhibit PDE, have been noted to exert only marginal effects on natriuresis compared with aminophylline or theophylline, suggesting that inhibition of adenosine receptors is critical for the natriuretic action (Andersson et al. 1984; Cirillo et al. 1989; Franzone et al. 1988). Strong experimental evidence indicates that it is the A<sub>1</sub> adenosine receptor subtype whose inhibition results in natriuresis. In mice with targeted deletion of A<sub>1</sub> adenosine receptors, the diuretic and natriuretic effect of caffeine (45 mg/kg) or theophylline (45 mg/kg) was entirely absent (Rieg et al. 2005). The natriuresis caused by systemic administration of xanthine derivatives designed to selectively inhibit A<sub>1</sub> adenosine receptors such as CVT-124, 1,3-dipropyl-8-cyclopentylxanthine (DPCPX), and KW-3902 has been shown by lithium clearance and renal tubular micropuncture approaches to be accompanied by inhibition of proximal tubular fluid reabsorption (Knight et al. 1993; Kost et al. 2000; Mizumoto and Karasawa 1993; Wilcox et al. 1999).

DPCPX-induced natriuresis can be prevented by pertussis toxin, consistent with an involvement of the Gi-coupled A<sub>1</sub> adenosine receptors (Kost et al. 2000).

In view of the dominant role of Na/H exchange for sodium reabsorption in the proximal tubule, it is not surprising that modulation of NHE3 has been implicated in adenosine-dependent modulation of sodium reabsorption. In opossum kidney cells, low concentrations of an A<sub>1</sub> adenosine receptor agonist (less than 10<sup>-8</sup> M) do in fact activate NHE3, an effect that is blocked by A<sub>1</sub> adenosine receptor antagonists and apparently mediated by inactivation of adenylyl cyclase (Di Sole et al. 2003). Downregulation of NHE3 and of  $\alpha_1/\beta_1$ -NaKATPase protein expression was observed following a 1-day treatment with caffeine in rats (Lee et al. 2002). On the other hand, theophylline (1 mM) did not affect HCO<sub>3</sub> flux as assessed from the pH recovery in stationary microperfusion studies in the rat (Bailey 2004). In renal proximal tubular cell cultures and opossum kidney cells, A<sub>1</sub> adenosine receptor activation stimulated and inhibition of A<sub>1</sub> adenosine receptors by DPCPX or KW-3902 inhibited apical sodium/phosphate (Cai et al. 1994, 1995; Coulson et al. 1991) and Na/glucose cotransport (Coulson et al. 1991, 1996). Inhibition of phosphate uptake by A<sub>1</sub> adenosine receptor antagonists was associated with a dose-dependent increase of cellular cAMP production as well as an increase in protein kinase C activity (Cai et al. 1995; Coulson et al. 1991, 1996). Theophylline and A<sub>1</sub>-adenosine-receptor-selective xanthine derivatives inhibited basolateral HCO<sub>3</sub> conductance in microperfused rabbit proximal convoluted tubules and this effect was mimicked by forskolin and chlorophenylthio-cAMP, suggesting that methylxanthines inhibit Na/HCO<sub>3</sub> cotransport activity by increasing intracellular cAMP levels (Takeda et al. 1993). To the extent that caffeine causes an increase in arterial blood pressure, a potential direct role of blood pressure in inhibiting tubular reabsorption and altering NHE3 distribution needs to be considered (Nussberger et al. 1990; Rachima-Maoz et al. 1998; Rakic et al. 1999). Finally, it should be pointed out that administration of methylxanthines only explores the impact of a reduction in adenosine-mediated effects, and that the consistent stimulatory effect of adenosine suggested by this intervention is therefore not in conflict with the possibility that high concentrations of adenosine may elicit inhibition of proximal tubular transport (Di Sole 2008; Di Sole et al. 2003).

## 4 Hemodynamics

Several studies in anesthetized dogs agree that the intrarenal infusion of methylxanthines does not affect RBF significantly, although renal vascular tone may decrease slightly because of small blood pressure reductions (Ibarrola et al. 1991; Osswald 1975; Premen et al. 1985). Furthermore, theophylline or caffeine do not alter renal plasma flow in humans to an extent that could be detected by clearance techniques (Beutler et al. 1990; Brater et al. 1983; Brown et al. 1993; Passmore et al. 1987). Changes of GFR in response to methylxanthines are sometimes more pronounced than those of RBF causing increases of filtration fraction (Fulgraff

1969). On the other hand, theophylline consistently inhibited adenosine-induced reductions of GFR and RBF in dogs and rats (Osswald 1975; Osswald et al. 1977; Pawlowska et al. 1987; Spielman 1984). Similarly, the vasodilator response of medullary blood flow to adenosine was blocked by 8-phenyltheophylline (Dinour and Brezis 1991). Thus, methylxanthines can affect renal hemodynamics by blocking the vascular actions of adenosine, at least in the range of supranormal adenosine levels. The absence of major effects of methylxanthines on renal hemodynamics could be due to low resting levels of adenosine. However, this seems unlikely since renal interstitial adenosine levels as determined by microdialysis are on the order of 50–200 nM, a range in which both  $A_1$  and  $A_{2a}$  adenosine receptors should be partially occupied (Baranowski and Westenfelder 1994; Nishiyama et al. 2001; Siragy and Linden 1996; Zou et al. 1999). Thus, renal vascular tone under basal conditions appears to represent a state of balanced activation of  $A_1$  and  $A_{2a}$  adenosine receptors. Vascular actions of theophylline could also result from a reduction of the inhibitory effect of adenosine on catecholamine release from renal sympathetic nerve terminals (Hedqvist and Fredholm 1976). In isolated perfused rabbit kidneys, theophylline did not affect norepinephrine release or RBF under basal conditions, presumably a reflection of the absence of a basal sympathetic tone (Hedqvist et al. 1978). However, the increased norepinephrine release following renal nerve stimulation was slightly potentiated by theophylline, and this was accompanied by a decrease in the vasoconstrictor response to nerve stimulation (Hedqvist et al. 1978). Similarly, theophylline attenuated the vasoconstrictor response to exogenous norepinephrine (Hedqvist et al. 1978; Yoneda et al. 1990). Finally, the reduction of GFR following dipyridamole and indomethacin administration in rheumatic patients was fully reversed by theophylline, suggesting that some of the vascular effects of indomethacin may be adenosine-mediated (Seideman et al. 1987).

In contrast to the negligible effects of methylxanthines on global renal vascular tone, theophylline has been found to fully inhibit the local tubuloglomerular feedback (TGF) response to changes of NaCl concentration in individual nephrons. Intratubular and intravenous administration of theophylline or 1,3-dipropyl-8-*p*-sulfophenylxanthine caused a dose-dependent blockade of the afferent arteriolar constriction induced by increases in NaCl concentration in the tubular fluid passing the macula densa segment (Franco et al. 1989; Osswald et al. 1980; Schnermann et al. 1977). This effect is the result of inhibition of  $A_1$  adenosine receptors since the effect of theophylline on TGF was fully mimicked by subtype-specific adenosine receptor antagonists (Kawabata et al. 1998; Ren et al. 2002; Schnermann et al. 1990; Thomson et al. 2000; Wilcox et al. 1999). Infusion of hypertonic saline into the renal artery of anesthetized dogs has been shown to cause sustained vasoconstriction, and this has been considered a model of “whole kidney TGF” (Gerken et al. 1983). Like single nephron TGF, vasoconstriction caused by hypertonic saline was inhibited by theophylline or aminophylline (Gerber and Nies 1986; Gerken et al. 1983). Extracellular conversion of released cAMP by ectophosphodiesterases and 5'-nucleotidase has been suggested to constitute a significant source of adenosine in the kidney (Jackson et al. 1997). Thus, the PDE-inhibitory effects of

xanthines may contribute to inhibition of TGF by reducing the interstitial adenosine levels (Mi et al. 1994).

## 5 Renin Secretion

It is remarkable that methylxanthines are able to cause natriuresis despite the fact that they also stimulate the strongly antinatriuretic renin–angiotensin system. Theophylline increased plasma renin activity in dogs, and this increase was shown to occur without changes in blood pressure or in the plasma levels of epinephrine and norepinephrine, indicating that it was not mediated by the renal baroreceptor mechanism or by adrenergic receptors (Reid et al. 1972). In fact, theophylline stimulated renin release even in dogs treated with propranolol. Likewise, oral administration of caffeine in rats for 10 days was associated with a marked rise of renin secretion (Tofovic and Jackson 1999). Even though it was thought that the theophylline effect may be mediated by inhibition of PDE and the resulting increase in cellular cAMP levels (Reid et al. 1972), it now seems likely that stimulation of renin by methylxanthines is at least in part, but probably predominantly, a consequence of inhibition of adenosine receptors. Inhibition of A<sub>1</sub> adenosine receptors by the selective antagonist FK-453 caused a significant increase of plasma renin levels (Balakrishnan et al. 1993), and DPCPX partially inhibited the stimulation of renin release caused by low NaCl concentration in a microperfused juxtaglomerular apparatus preparation (Weihprecht et al. 1990). Furthermore, infusion of caffeine or theophylline at a dose that did not change renal cortical cAMP levels has been reported to abolish the inhibitory effect of intrarenally infused adenosine on renin secretion, and this effect could be dissociated from hemodynamic changes (Arend et al. 1987; Choi et al. 1993; Spielman 1984). In support of the notion that endogenous adenosine exerts a general inhibitory “brake” function in renin release, caffeine or theophylline as well as specific A<sub>1</sub> adenosine receptor antagonistic xanthines have been reported to augment the renin-stimulatory effects of a low renal artery pressure, a low-sodium diet, furosemide, isoproterenol, and vasodilators such as diazoxide and hydralazine (Brown et al. 1991; Langard et al. 1983; Paul et al. 1989; Pfeifer et al. 1995; Tofovic et al. 1991; Tseng et al. 1993). Despite the convincing effect of methylxanthines in experimental animals, studies in normotensive human subjects examining the effect of caffeine (250 mg) or coffee drinking have reported plasma renin concentration to increase, decrease, or remain unchanged, indicating that caffeine in moderate doses may not consistently induce the plasma caffeine levels needed to stimulate renin release (Nussberger et al. 1990; Robertson et al. 1978; Smits et al. 1983). Similarly, no increases of plasma renin concentration were observed in response to caffeine or coffee drinking in hypertensive patients (Eggertsen et al. 1993; Palatini et al. 1996; Robertson et al. 1984) or in patients with autonomic failure (Onrot et al. 1985).



## 6 Disease and Therapeutic Aspects

### 6.1 Polycystic Kidney Disease

Generation of cAMP has been shown to play a role in the secretion of fluid that is thought to be partly responsible for the accumulation of fluid in renal cystic disease (Belibi et al. 2002). Thus, by augmenting cAMP levels, methylxanthines could contribute to the progression of cyst formation. In fact, in primary cultures of renal cysts from patients with autosomal dominant polycystic kidney disease (ADPKD), caffeine (10–1,000  $\mu\text{M}$ ) increased levels of cellular cAMP and this was associated with an increase of transepithelial chloride secretion. Furthermore, caffeine greatly potentiated the augmentation of cAMP induced by Gs-coupled agonists such as vasopressin and PGE<sub>2</sub>. The increase of cAMP levels by caffeine was in part mediated by inhibition of PDE since rolipram, a PDE inhibitor that does not interact with adenosine receptors, also caused a marked elevation of cAMP levels. However, an elevation in cAMP level was also seen with adenosine (10 mM), and this effect was attenuated, but not fully blocked by caffeine, suggesting that stimulation of A<sub>2</sub> adenosine receptors contributed to the accumulation of cAMP in cyst cells (Belibi et al. 2002). Cyst formation has also been related to impaired mechanosensation by primary cilia since both polycystin 1 (PC1) and polycystin 2 (PC2) are found in association with cilia. In fact, the increase in cytosolic calcium concentration caused by a flow stimulus in renal cells of wild-type mice was not seen in cells from PC1- or PC2-deficient mice or after treatment of wild-type cells with blocking antibodies against PC2 (Nauli et al. 2003). The increase in cytosolic calcium concentration by flow was blocked by high concentrations of caffeine, suggesting that it was caused by release of stored calcium across ryanodine-sensitive receptors in response to an initial calcium entry through PC2 cation channels. Nevertheless, caffeine ingestion did not accelerate cyst formation in the Han–Sprague–Dawley rat model of ADPKD although it was associated with the generation of hypertension (Tanner and Tanner 2001).

### 6.2 Nephropathies

In several experimental disease models, chronic caffeine administration has been found to exacerbate the development of hypertension and renal disease, perhaps through the effect of caffeine on renin secretion. For example, the presence of caffeine (0.1%) in the drinking water of rats augmented the blood pressure increase caused by renal arterial constriction and this was associated with a greater increase of plasma renin concentration (Choi et al. 1993; Kost et al. 1994; Ohnishi et al. 1986). Similarly, a 10-day caffeine exposure enhanced renin secretion to a markedly greater extent in spontaneously hypertensive heart failure (SHHF/Mcc-fa) rats than in control rats (Tofovic et al. 1999). In association with the increase of renin

secretion, prolonged caffeine ingestion caused a faster decline of renal function and a significant enhancement of urinary protein excretion (Tofovic and Jackson 1999). An adverse effect of chronic caffeine intake on renal function was also observed in puromycin aminonucleoside induced nephrosis in rats. Both the decline of renal function assessed as creatinine clearance and the increase of renin secretion were enhanced in puromycin-treated rats receiving caffeine in comparison with nephrotic rats receiving tap water (Tofovic et al. 2000). In addition, caffeine potentiated the development of interstitial fibrosis and glomerulosclerosis caused by puromycin (Tofovic et al. 2000). Finally, long-term treatment with caffeine reduced renal function and augmented proteinuria in obese diabetic ZSF1 rats despite improving glucose tolerance. Caffeine-induced renal deterioration was paralleled by enhanced fibrosis, proliferation, and inflammation (Tofovic et al. 2002, 2007). In addition to stimulating the release of renin, the effects of caffeine may be mediated through interference with the direct anti-inflammatory effects of adenosine (Tofovic et al. 2007).

In view of this solid body of evidence in support of caffeine as a risk factor in renal disease it is unexpected that the methylxanthine pentoxifylline has been found to produce exactly opposite outcomes. Pentoxifylline has only a low affinity for adenosine receptors and is generally considered a PDE inhibitor with some selectivity for PDE4 (Daly 2007). In animal studies, pentoxifylline markedly reduced the functional decline, proteinuria, fibrosis, and inflammation in rats following 5/6 nephrectomy or treatment with anti-GBM antiserum, and this was associated with an attenuation of the stimulated expression of mitogenic and profibrotic gene products (Chen et al. 2004; Lin et al. 2002). This effect was probably independent of the renin–angiotensin system since a combination of pentoxifylline with an ACE inhibitor further diminished disease progression (Lin et al. 2002), and since the plasma renin level was found to be unchanged in a human study (Chen et al. 2006). Furthermore, pentoxifylline protected against endotoxin-induced renal failure in mice and reduced plasma levels of TNF- $\alpha$ , IL-1 $\beta$ , and nitric oxide (Wang et al. 2006). In relatively small human trials, pentoxifylline reduced proteinuria and slowed the GFR decline in patients with chronic renal failure (Lin et al. 2008; Perkins et al. 2009), and it reduced proteinuria in patients with primary glomerular diseases in association with a reduction of urinary MCP-1 excretion (Chen et al. 2006). In nonhypertensive type 2 diabetic subjects pentoxifylline was as effective in reducing microalbuminuria as the ACE inhibitor captopril (Rodriguez-Moran and Guerrero-Romero 2005). A meta-analysis of ten randomized controlled studies in adult patients with diabetic kidney disease suggested comparable efficacies of pentoxifylline and captopril in reducing proteinuria (McCormick et al. 2008). The mechanism of action of the protective effects of pentoxifylline or its metabolite lisofylline is unclear.

### **6.3 Radiocontrast Nephropathy**

Iodinated radiocontrast agents used in a number of radiological imaging procedures can cause acute renal failure. The incidence is very low in the absence of

complicating factors, but it increases considerably in patients with pre-existing renal conditions or in other circumstances that represent a risk factor for developing acute renal failure in general, such as dehydration or low cardiac output. Since radiocontrast-induced renal failure is accompanied by a reduction of RBF and GFR, methylxanthines have been among a number of vasodilator agents that have been assessed in regard to their preventive potential. In sodium-depleted dogs, radiocontrast agents reduced RBF and GFR, and both of these effects were attenuated by prior administration of theophylline (Arend et al. 1987; Deray et al. 1990). Furthermore, theophylline partially prevented the reduction of medullary blood flow induced by iodixanol (Lancelot et al. 2002). Thus, theophylline appears to act by antagonizing vasoconstriction mediated by A<sub>1</sub> adenosine receptor activation, and this hypothesis was corroborated in studies in which the A<sub>1</sub>-adenosine-receptor-selective antagonist KW-3902 was even more effective than theophylline in attenuating iohexol-induced renal functional impairment in dogs with pre-existing renal insufficiency (Arakawa et al. 1996). Acute renal failure and cytotoxicity following iohexol administration were found to be more pronounced in wild-type than in A<sub>1</sub>-adenosine-receptor-deficient mice, and a similar protective effect could be seen when wild-type mice were pretreated with the A<sub>1</sub>-adenosine-receptor-selective antagonist DPCPX (Lee et al. 2006). In rats pretreated chronically with N<sup>ω</sup>-nitro-L-arginine methyl ester, but not in control rats, sodium diatrizoate caused a decrease of GFR and RBF that could be fully prevented by pretreatment with theophylline, DPCPX, or KW-3902 (Erley et al. 1997; Yao et al. 2001). It could also be prevented by extracellular volume expansion, the standard preventive strategy (Yao et al. 2001). In the majority of studies, prophylactic administration of theophylline or aminophylline has also been reported to provide protection against radiocontrast renal failure in humans (Erley et al. 1994; Huber et al. 2001, 2003; Kapoor et al. 2002; Katholi et al. 1995; Kolonko et al. 1998). Protection by theophylline was similar to that afforded by oral or intravenous hydration (Erley et al. 1999). Two recent meta-analyses, one including 480 patients and the other including 585 patients, concluded that theophylline or aminophylline appear to attenuate the radiocontrast-induced decline of renal function (Bagshaw and Ghali 2005; Ix et al. 2004). On the other hand, an analysis of 41 studies that used radiocontrast agents in combination with theophylline, N-acetylcysteine, fenoldopam, dopamine, iloprost, statins, furosemide or mannitol showed that only N-acetylcysteine provided significant renoprotection, whereas the risk reduction provided by theophylline was not significant (Kelly et al. 2008). Greater protection by N-acetylcysteine than by theophylline was also observed in a study in dehydrated rats (Efrati et al. 2009). Exactly how methylxanthines exert their limited protective effect is unclear in view of the well-recognized multifactorial pathophysiological mechanisms of radiocontrast-induced nephropathy (Cox and Tsikouris 2004; Persson et al. 2005). Because of similar effects of other vasoactive agents such as ANP, dopamine, endothelin antagonists, calcium channel blockers, and PGE<sub>2</sub>, nonspecific vasodilatation of the renal vascular bed is likely to play a major contributory role. Nevertheless, despite some promising results, the overall clinical experience does not support the

use of methylxanthines as a first-line defense against the induction of contrast nephropathy (Lin and Bonventre 2005).

## 6.4 *Calcineurin Inhibitors*

The main complication of immunosuppression by calcineurin inhibitors is nephrotoxicity manifesting itself as a decline of renal function associated with vasoconstriction and a reduction of RBF. In early studies in rats, theophylline failed to ameliorate cyclosporine-induced renal vasoconstriction, indicating that it was not caused by adenosine (Churchill et al. 1990). Subsequently however, caffeine, theophylline, and pentoxifylline were observed to reduce the acute contractile response to cyclosporine of isolated glomeruli and mesangial cells in culture (Potier et al. 1997). Furthermore, the acute reduction of GFR and RBF caused in rats by a single dose of tacrolimus was completely reversed by theophylline given 1 h after the drug (McLaughlin et al. 2003a). Concomitant chronic administration of theophylline together with tacrolimus prevented the decrease of creatinine clearance that was caused by tacrolimus in the control group (McLaughlin et al. 2003b). In contrast, chronic administration of theophylline did not protect against cyclosporine-induced renal failure in rabbits and even enhanced its cytotoxic effects, whereas functional recovery was seen when theophylline was given as a single dose following a 5-day treatment with cyclosporine (Prevot et al. 2002). Follow-up studies that would shed light on the reasons for these divergent observations are not available. In children with nonrenal transplants who showed signs of tacrolimus nephrotoxicity, such as an increase in serum creatinine levels and oliguria despite treatment with loop diuretics, a single dose of aminophylline (5 mg/kg) caused a doubling of urine flow rate and osmolar clearance as well as a moderate increase of renal function (McLaughlin and Abitbol 2005).

## 6.5 *Cisplatin*

Cisplatin-based chemotherapy is another treatment modality that is often associated with nephrotoxicity. In a placebo-controlled trial on 36 patients, the administration of theophylline before and for 5 days following cisplatin treatment completely prevented the fall of inulin clearance that was seen in the placebo group, in which GFR fell by 21% (Benoehr et al. 2005). This study in humans is thus consistent with observations in rats in which aminophylline ameliorated cisplatin-induced renal failure when given in the maintenance phase although it did not prevent the decline of renal function when administered prophylactically (Heidemann et al. 1989). Since enprofylline did not mimic the protective action of aminophylline, adenosine receptor activation is the likely cause for the decline of renal function (Heidemann et al. 1989). Specifically, the protective action seems related to inhibition of A<sub>1</sub> adenosine receptors because the A<sub>1</sub>-adenosine-receptor-specific antagonists

DPCPX and KW-3902 were effective both in preventing and in treating the nephrotoxic effects of cisplatin (Knight et al. 1991; Nagashima et al. 1995). A higher dose of cisplatin in vivo as well as exposure of LLC-PK1 cells to cisplatin caused upregulation of A<sub>1</sub> adenosine receptor expression, and this was associated with cytoprotection on the basis of the finding that nonselective and A<sub>1</sub>-adenosine-receptor-selective antagonists exacerbated cisplatin-induced nephrotoxicity (Bhat et al. 2002; Pingle et al. 2004; Saad et al. 2004). Thus, the potential of methylxanthines to exert both protective and injurious effects may be a reflection of the wide spectrum of adenosine actions in fundamental processes such as tissue oxygen supply and inflammation.

## 6.6 Glycerol

Theophylline and other methylxanthines have been consistently found to ameliorate the experimental acute renal failure caused by intramuscular injection of glycerol when treatment was started at the time of injury (Bidani and Churchill 1983; Bowmer et al. 1986, 1988). The protective effect appears to be a consequence of inhibition of A<sub>1</sub> adenosine receptors since subtype-specific antagonists mimic the action of the natural methylxanthines (Ishikawa et al. 1993; Panjehshahin et al. 1992; Suzuki et al. 1992) and since their effect can be seen in a dose range that has no discernible effect on renal PDE activity (Panjehshahin et al. 1992). In contrast to the protective action of methylxanthines in myoglobinuric acute renal failure, theophylline did not improve renal function in the renal failure caused by mercury chloride or gentamicin (Kellett et al. 1988; Rossi et al. 1990).

## 6.7 Ischemia–Reperfusion

Interest in the potential of methylxanthines to improve renal function following postschismic acute renal failure has arisen from the fact that ischemia is associated with increases of adenosine tissue content in the kidney as well as in other organs (Osswald et al. 1977). Thus, it was conceivable that methylxanthines may exert beneficial effects by preventing the renal vasoconstriction caused by excess adenosine. In rats, theophylline, administered as a single dose of 100 μmol/kg 10 min before the release of a 1-h renal artery occlusion, increased GFR and electrolyte excretion threefold to sixfold within 3 h of the postschismic period compared with vehicle-treated animals (Osswald et al. 1979), an observation that was later confirmed in both rats and rabbits (Gouyon and Guignard 1988; Lin et al. 1986). The protective mechanism of methylxanthines in the initiation phase of renal injury following ischemia–reperfusion is likely related at least in part to inhibition of vasoconstrictive adenosine receptors. Support for this notion comes from a recent study in which the A<sub>1</sub> adenosine receptor antagonist DPCPX infused prior to and following a 30-min period of bilateral renal artery obstruction was observed to

significantly improve creatinine clearance over the initial 4 h following reperfusion (Moosavi et al. 2009). Furthermore, the immediate postischemic reduction of GFR was enhanced by dipyridamole, an inhibitor of adenosine uptake through equilibrative nucleoside transporters, and this effect was abrogated by theophylline (Lin et al. 1987). In clinical studies, a single dose of theophylline given early after birth in asphyxiated full-term infants elicited beneficial effects by reducing the renal involvement and fall in GFR as determined over the first 5 days (Bakr 2005; Bhat et al. 2006; Eslami et al. 2009; Jenik et al. 2000).

The role of methylxanthines in the maintenance phase following renal ischemia is an area of considerable controversy. Early studies have shown that pretreatment of rats with a single dose of theophylline during a 30-min renal artery occlusion was associated with higher RBF and GFR during the maintenance phase of acute renal failure after 5 days, suggesting that theophylline administration in the acute phase affected the severity of renal failure in the maintenance phase (Lin et al. 1988). Furthermore, theophylline administered 5 days after ischemia acutely increased RBF and GFR in previously untreated rats (Lin et al. 1988). Theophylline also caused an increase of GFR measured 5 days after experimental renal transplantation in rats without affecting the inflammatory response (Grenz et al. 2006). On the other hand, in a small study with limited statistical power, theophylline was not found to afford protection against acute renal failure during cardiac surgery (Kramer et al. 2002). It is uncertain whether theophylline exerts these effects by antagonizing vasoconstrictor effects of adenosine. In fact, adenosine itself given immediately after a renal ischemia of 45 min provided renoprotection after 24 h, an effect mimicked by CGS-21680 and therefore apparently mediated by activation of  $A_{2a}$  adenosine receptors (Lee and Emala 2001). Similarly, the rise of serum creatinine level assessed 1 and 2 days following renal ischemia was found to be reduced by chronic administration of the selective  $A_{2a}$  agonist DWH-146 and enhanced in  $A_{2a}$ -adenosine-receptor-deficient mice (Day et al. 2003; Okusa et al. 1999). Relative renoprotection 24 h following renal ischemia was also provided by  $A_1$  adenosine receptor agonists, and a worsening of the outcome was observed in  $A_1$ -adenosine-receptor-deficient mice (Kim et al. 2009; Lee et al. 2004a, b). Comparable renoprotective effects of  $A_{2a}$  and  $A_1$  adenosine receptor activation suggest that the functional improvement after extended reperfusion is unrelated to the vascular actions of adenosine since the vascular effects of activating  $A_{2a}$  or  $A_1$  adenosine receptors are opposite. The common denominator may be a dominant anti-inflammatory action of adenosine that is exerted by both receptor subtypes.

## References

- Andersson KE, Johannesson N, Karlberg B, Persson CG (1984) Increase in plasma free fatty acids and natriuresis by xanthines may reflect adenosine antagonism. *Eur J Clin Pharmacol* 26:33–8
- Arakawa K, Suzuki H, Naitoh M, Matsumoto A, Hayashi K, Matsuda H, Ichihara A, Kubota E, Saruta T (1996) Role of adenosine in the renal responses to contrast medium. *Kidney Int* 49:1199–206

- Arend LJ, Bakris GL, Burnett JC Jr, Megerian C, Spielman WS (1987) Role for intrarenal adenosine in the renal hemodynamic response to contrast media. *J Lab Clin Med* 110:406–11
- Armstrong LE (2002) Caffeine, body fluid-electrolyte balance, and exercise performance. *Int J Sport Nutr Exerc Metab* 12:189–206
- Armstrong LE, Pumerantz AC, Roti MW, Judelson DA, Watson G, Dias JC, Sokmen B, Casa DJ, Maresh CM, Lieberman H, Kellogg M (2005) Fluid, electrolyte, and renal indices of hydration during 11 days of controlled caffeine consumption. *Int J Sport Nutr Exerc Metab* 15:252–65
- Baer PG, Armstrong EL, Cagen LM (1983) Dissociation of effects of xanthine analogs on renal prostaglandins and renal excretory function in the awake rat. *J Pharmacol Exp Ther* 227:600–4
- Bagshaw SM, Ghali WA (2005) Theophylline for prevention of contrast-induced nephropathy: a systematic review and meta-analysis. *Arch Intern Med* 165:1087–93
- Bailey MA (2004) Inhibition of bicarbonate reabsorption in the rat proximal tubule by activation of luminal P2Y1 receptors. *Am J Physiol Renal Physiol* 287:F789–96
- Bakr AF (2005) Prophylactic theophylline to prevent renal dysfunction in newborns exposed to perinatal asphyxia—a study in a developing country. *Pediatr Nephrol* 20:1249–52
- Balakrishnan VS, Coles GA, Williams JD (1993) A potential role for endogenous adenosine in control of human glomerular and tubular function. *Am J Physiol* 265:F504–10
- Baranowski RL, Westenfelder C (1994) Estimation of renal interstitial adenosine and purine metabolites by microdialysis. *Am J Physiol* 267:F174–82
- Belibi FA, Wallace DP, Yamaguchi T, Christensen M, Reif G, Grantham JJ (2002) The effect of caffeine on renal epithelial cells from patients with autosomal dominant polycystic kidney disease. *J Am Soc Nephrol* 13:2723–9
- Benoehr P, Krueth P, Bokemeyer C, Grenz A, Osswald H, Hartmann JT (2005) Nephroprotection by theophylline in patients with cisplatin chemotherapy: a randomized, single-blinded, placebo-controlled trial. *J Am Soc Nephrol* 16:452–8
- Beutler JJ, Koomans HA, Bijlsma JA, Dorhout Mees EJ (1990) Renal actions of theophylline and atrial natriuretic peptide in humans: a comparison by means of clearance studies. *J Pharmacol Exp Ther* 255:1314–9
- Bhat MA, Shah ZA, Makhdoomi MS, Mufti MH (2006) Theophylline for renal function in term neonates with perinatal asphyxia: a randomized, placebo-controlled trial. *J Pediatr* 149:180–4
- Bhat SG, Mishra S, Mei Y, Nie Z, Whitworth CA, Rybak LP, Ramkumar V (2002) Cisplatin up-regulates the adenosine A(1) receptor in the rat kidney. *Eur J Pharmacol* 442:251–64
- Bidani AK, Churchill PC (1983) Aminophylline ameliorates glycerol-induced acute renal failure in rats. *Can J Physiol Pharmacol* 61:567–71
- Bowmer CJ, Collis MG, Yates MS (1986) Effect of the adenosine antagonist 8-phenyltheophylline on glycerol-induced acute renal failure in the rat. *Br J Pharmacol* 88:205–12
- Bowmer CJ, Collis MG, Yates MS (1988) Amelioration of glycerol-induced acute renal failure in the rat with 8-phenyltheophylline: timing of intervention. *J Pharm Pharmacol* 40:733–5
- Brater DC, Kaojarern S, Chennavasin P (1983) Pharmacodynamics of the diuretic effects of aminophylline and acetazolamide alone and combined with furosemide in normal subjects. *J Pharmacol Exp Ther* 227:92–7
- Brown NJ, Porter J, Ryder D, Branch RA (1991) Caffeine potentiates the renin response to diazoxide in man. Evidence for a regulatory role of endogenous adenosine. *J Pharmacol Exp Ther* 256:56–61
- Brown NJ, Ryder D, Nadeau J (1993) Caffeine attenuates the renal vascular response to angiotensin II infusion. *Hypertension* 22:847–52
- Cai H, Batuman V, Puschett DB, Puschett JB (1994) Effect of KW-3902, a novel adenosine A1 receptor antagonist, on sodium-dependent phosphate and glucose transport by the rat renal proximal tubular cell. *Life Sci* 55:839–45
- Cai H, Puschett DB, Guan S, Batuman V, Puschett JB (1995) Phosphate transport inhibition by KW-3902, an adenosine A1 receptor antagonist, is mediated by cyclic adenosine monophosphate. *Am J Kidney Dis* 26:825–30

- Chen YM, Lin SL, Chiang WC, Wu KD, Tsai TJ (2006) Pentoxifylline ameliorates proteinuria through suppression of renal monocyte chemoattractant protein-1 in patients with proteinuric primary glomerular diseases. *Kidney Int* 69:1410–5
- Chen YM, Ng YY, Lin SL, Chiang WC, Lan HY, Tsai TJ (2004) Pentoxifylline suppresses renal tumour necrosis factor-alpha and ameliorates experimental crescentic glomerulonephritis in rats. *Nephrol Dial Transplant* 19:1106–15
- Choi KC, Lee J, Moon KH, Park KK, Kim SW, Kim NH (1993) Chronic caffeine ingestion exacerbates 2-kidney, 1-clip hypertension and ameliorates deoxycorticosterone acetate-salt hypertension in rats. *Nephron* 65:619–22
- Churchill PC, Rossi NF, Churchill MC, Bidani AK, McDonald FD (1990) Acute cyclosporine-induced renal vasoconstriction: lack of effect of theophylline. *Am J Physiol* 258:F41–5
- Cirillo R, Grossi E, Franzone JS (1989) Doxofylline, an adenosine-nonblocking xanthine, does not induce cardiostimulant effects. *Res Commun Chem Pathol Pharmacol* 65:21–34
- Coulson R, Johnson RA, Olsson RA, Cooper DR, Scheinman SJ (1991) Adenosine stimulates phosphate and glucose transport in opossum kidney epithelial cells. *Am J Physiol* 260:F921–8
- Coulson R, Proch PS, Olsson RA, Chalfant CE, Cooper DR (1996) Upregulated renal adenosine A1 receptors augment PKC and glucose transport but inhibit proliferation. *Am J Physiol* 270:F263–74
- Cox CD, Tsikouris JP (2004) Preventing contrast nephropathy: what is the best strategy? A review of the literature. *J Clin Pharmacol* 44:327–37
- Daly JW (2007) Caffeine analogs: biomedical impact. *Cell Mol Life Sci* 64:2153–69
- Davis JO, Shock NW (1949) The effect of theophylline ethylene diamine on renal function in control subjects and in patients with congestive heart failure. *J Clin Invest* 28:1459–68
- Day YJ, Huang L, McDuffie MJ, Rosin DL, Ye H, Chen JF, Schwarzschild MA, Fink JS, Linden J, Okusa MD (2003) Renal protection from ischemia mediated by A2A adenosine receptors on bone marrow-derived cells. *J Clin Invest* 112:883–91
- Deray G, Martinez F, Cacoub P, Baumelou B, Baumelou A, Jacobs C (1990) A role for adenosine calcium and ischemia in radiocontrast-induced intrarenal vasoconstriction. *Am J Nephrol* 10:316–22
- Di Sole F (2008) Adenosine and renal tubular function. *Curr Opin Nephrol Hypertens* 17:399–407
- Di Sole F, Cerull R, Petzke S, Casavola V, Burckhardt G, Helmle-Kolb C (2003) Bimodal acute effects of A1 adenosine receptor activation on Na<sup>+</sup>/H<sup>+</sup> exchanger 3 in opossum kidney cells. *J Am Soc Nephrol* 14:1720–30
- Dinour D, Brezis M (1991) Effects of adenosine on intrarenal oxygenation. *Am J Physiol* 261:F787–91
- Edwards RM, Spielman WS (1994) Adenosine A1 receptor-mediated inhibition of vasopressin action in inner medullary collecting duct. *Am J Physiol* 266:F791–6
- Efrati S, Berman S, Ilgiyev I, Siman-Tov Y, Averbukh Z, Weissgarten J (2009) Differential effects of N-acetylcysteine, theophylline or bicarbonate on contrast-induced rat renal vasoconstriction. *Am J Nephrol* 29:181–91
- Eggertsen R, Andreasson A, Hedner T, Karlberg BE, Hansson L (1993) Effect of coffee on ambulatory blood pressure in patients with treated hypertension. *J Intern Med* 233:351–5
- Erley CM, Duda SH, Rehfuß D, Scholtes B, Bock J, Muller C, Osswald H, Risler T (1999) Prevention of radiocontrast-media-induced nephropathy in patients with pre-existing renal insufficiency by hydration in combination with the adenosine antagonist theophylline. *Nephrol Dial Transplant* 14:1146–9
- Erley CM, Duda SH, Schlepckow S, Koehler J, Huppert PE, Strohmaier WL, Bohle A, Risler T, Osswald H (1994) Adenosine antagonist theophylline prevents the reduction of glomerular filtration rate after contrast media application. *Kidney Int* 45:1425–31
- Erley CM, Heyne N, Burgert K, Langanke J, Risler T, Osswald H (1997) Prevention of radiocontrast-induced nephropathy by adenosine antagonists in rats with chronic nitric oxide deficiency. *J Am Soc Nephrol* 8:1125–32



- Eslami Z, Shajari A, Kheirandish M, Heidary A (2009) Theophylline for prevention of kidney dysfunction in neonates with severe asphyxia. *Iran J Kidney Dis* 3:222–6
- Franco M, Bell PD, Navar LG (1989) Effect of adenosine A1 analogue on tubuloglomerular feedback mechanism. *Am J Physiol Renal Physiol* 257:F231–6
- Franzone JS, Cirillo R, Barone D (1988) Doxofylline and theophylline are xanthines with partly different mechanisms of action in animals. *Drugs Exp Clin Res* 14:479–89
- Fulgraff G (1969) Xanthinderivate als diuretika. In: Herken H (ed) *Handbuch der experimentellen Pharmakologie*, vol XXIV. Springer, Berlin, pp 596–640
- Gerber JG, Nies AS (1986) Renal vasoconstrictor response to hypertonic saline in the dog: effects of prostaglandins, indomethacin and theophylline. *J Physiol (Lond)* 380:35–43
- Gerkens JF, Heidemann HT, Jackson EK, Branch RA (1983) Aminophylline inhibits renal vasoconstriction produced by intrarenal hypertonic saline. *J Pharmacol Exp Ther* 225:611–5
- Gouyon JB, Guignard JP (1987) Renal effects of theophylline and caffeine in newborn rabbits. *Pediatr Res* 21:615–8
- Gouyon JB, Guignard JP (1988) Theophylline prevents the hypoxemia-induced renal hemodynamic changes in rabbits. *Kidney Int* 33:1078–83
- Grandjean AC, Reimers KJ, Bannick KE, Haven MC (2000) The effect of caffeinated, non-caffeinated, caloric and non-caloric beverages on hydration. *J Am Coll Nutr* 19:591–600
- Grantham JJ, Orloff J (1968) Effect of prostaglandin E1 on the permeability response of the isolated collecting tubule to vasopressin, adenosine 3',5'-monophosphate, and theophylline. *J Clin Invest* 47:1154–61
- Grenz A, Baier D, Petroktistis F, Wehrmann M, Kohle C, Schenk M, Sessler M, Gleiter CH, Fandrich F, Osswald H (2006) Theophylline improves early allograft function in rat kidney transplantation. *J Pharmacol Exp Ther* 317:473–9
- Handler JS, Butcher RW, Sutherland EW, Orloff J (1965) The effect of vasopressin and of theophylline on the concentration of adenosine 3',5'-phosphate in the urinary bladder of the toad. *J Biol Chem* 240:4524–6
- Hedqvist P, Fredholm BB (1976) Effects of adenosine on adrenergic neurotransmission; prejunctional inhibition and postjunctional enhancement. *Naunyn Schmiedebergs Arch Pharmacol* 293:217–23
- Hedqvist P, Fredholm BB, Olundh S (1978) Antagonistic effects of theophylline and adenosine on adrenergic neuroeffector transmission in the rabbit kidney. *Circ Res* 43:592–8
- Heidemann HT, Muller S, Mertins L, Stepan G, Hoffmann K, Ohnhaus EE (1989) Effect of aminophylline on cisplatin nephrotoxicity in the rat. *Br J Pharmacol* 97:313–8
- Huber W, Jeschke B, Page M, Weiss W, Salmhofer H, Schweigart U, Ilgmann K, Reichenberger J, Neu B, Classen M (2001) Reduced incidence of radiocontrast-induced nephropathy in ICU patients under theophylline prophylaxis: a prospective comparison to series of patients at similar risk. *Intensive Care Med* 27:1200–9
- Huber W, Schiepek C, Ilgmann K, Page M, Hennig M, Wacker A, Schweigart U, Lutlisky L, Valina C, Seyfarth M, Schomig A, Classen M (2003) Effectiveness of theophylline prophylaxis of renal impairment after coronary angiography in patients with chronic renal insufficiency. *Am J Cardiol* 91:1157–62
- Ibarrola AM, Inscho EW, Vari RC, Navar LG (1991) Influence of adenosine receptor blockade on renal function and renal autoregulation. *J Am Soc Nephrol* 2:991–9
- Ishikawa I, Shikura N, Takada K (1993) Amelioration of glycerol-induced acute renal failure in rats by an adenosine A1 receptor antagonist (FR-113453). *Ren Fail* 15:1–5
- Ishikawa SE, Okada K, Saito T (1992) Modulation by intracellular calcium pool of arginine vasopressin-induced cellular cyclic AMP production in rat renal papillary collecting tubule cells in culture. *J Pharmacol Exp Ther* 263:1050–5
- Ix JH, McCulloch CE, Chertow GM (2004) Theophylline for the prevention of radiocontrast nephropathy: a meta-analysis. *Nephrol Dial Transplant* 19:2747–53
- Izzo JL Jr, Ghosal A, Kwong T, Freeman RB, Jaenike JR (1983) Age and prior caffeine use alter the cardiovascular and adrenomedullary responses to oral caffeine. *Am J Cardiol* 52:769–73

- Jackson EK, Mi Z, Gillespie DG, Dubey RK (1997) Metabolism of cAMP to adenosine in the renal vasculature. *J Pharmacol Exp Ther* 283:177–82
- Jenik AG, Ceriani Cernadas JM, Gorenstein A, Ramirez JA, Vain N, Armadans M, Ferraris JR (2000) A randomized, double-blind, placebo-controlled trial of the effects of prophylactic theophylline on renal function in term neonates with perinatal asphyxia. *Pediatrics* 105: E45
- Kapoor A, Kumar S, Gulati S, Gambhir S, Sethi RS, Sinha N (2002) The role of theophylline in contrast-induced nephropathy: a case-control study. *Nephrol Dial Transplant* 17:1936–41
- Katholi RE, Taylor GJ, McCann WP, Woods WT Jr, Womack KA, McCoy CD, Katholi CR, Moses HW, Mishkel GJ, Lucore CL et al (1995) Nephrotoxicity from contrast media: attenuation with theophylline. *Radiology* 195:17–22
- Kawabata M, Ogawa T, Takabatake T (1998) Control of rat glomerular microcirculation by juxtaglomerular adenosine A1 receptors. *Kidney Int Suppl* 67:S228–30
- Kellelt R, Bowmer CJ, Collis MG, Yates MS (1988) Effect of alkylxanthines on gentamicin-induced acute renal failure in the rat. *J Pharm Pharmacol* 40:849–54
- Kelly AM, Dwamena B, Cronin P, Bernstein SJ, Carlos RC (2008) Meta-analysis: effectiveness of drugs for preventing contrast-induced nephropathy. *Ann Intern Med* 148:284–94
- Kim M, Chen SW, Park SW, Kim M, D'Agati VD, Yang J, Lee HT (2009) Kidney-specific reconstitution of the A1 adenosine receptor in A1 adenosine receptor knockout mice reduces renal ischemia-reperfusion injury. *Kidney Int* 75:809–23
- Knight RJ, Bowmer CJ, Yates MS (1993) The diuretic action of 8-cyclopentyl-1,3-dipropylxanthine, a selective A1 adenosine receptor antagonist. *Br J Pharmacol* 109:271–7
- Knight RJ, Collis MG, Yates MS, Bowmer CJ (1991) Amelioration of cisplatin-induced acute renal failure with 8-cyclopentyl-1,3-dipropylxanthine. *Br J Pharmacol* 104:1062–8
- Kolonko A, Wiecek A, Kokot F (1998) The nonselective adenosine antagonist theophylline does prevent renal dysfunction induced by radiographic contrast agents. *J Nephrol* 11:151–6
- Koschlakoff DJ (1864) Beobachtungen ueber die Wirkung des citronensauren Coffeins. *Virchows Arch Pathol Anat* 31:436–445
- Kost CK Jr, Herzer WA, Rominski BR, Mi Z, Jackson EK (2000) Diuretic response to adenosine A1 receptor blockade in normotensive and spontaneously hypertensive rats: role of pertussis toxin-sensitive G-proteins. *J Pharmacol Exp Ther* 292:752–60
- Kost CK Jr, Li P, Pfeifer CA, Jackson EK (1994) Telemetric blood pressure monitoring in benign 2-kidney, 1-clip renovascular hypertension: effect of chronic caffeine ingestion. *J Pharmacol Exp Ther* 270:1063–70
- Kramer BK, Preuner J, Ebenburger A, Kaiser M, Bergner U, Eilles C, Kammerl MC, Riegger GA, Birnbaum DE (2002) Lack of renoprotective effect of theophylline during aortocoronary bypass surgery. *Nephrol Dial Transplant* 17:910–5
- Lancelot E, Idee JM, Laclede C, Santus R, Corot C (2002) Effects of two dimeric iodinated contrast media on renal medullary blood perfusion and oxygenation in dogs. *Invest Radiol* 37:368–75
- Langard O, Holdaas H, Eide I, Kiil F (1983) Conditions for augmentation of renin release by theophylline. *Scand J Clin Lab Invest* 43:9–14
- Lee HT, Emala CW (2001) Systemic adenosine given after ischemia protects renal function via A2a adenosine receptor activation. *Am J Kidney Dis* 38:610–8
- Lee HT, Gallos G, Nasr SH, Emala CW (2004a) A1 adenosine receptor activation inhibits inflammation, necrosis, and apoptosis after renal ischemia-reperfusion injury in mice. *J Am Soc Nephrol* 15:102–11
- Lee HT, Jan M, Bae SC, Joo JD, Goubaeva FR, Yang J, Kim M (2006) A1 adenosine receptor knockout mice are protected against acute radiocontrast nephropathy in vivo. *Am J Physiol Renal Physiol* 290:F1367–75
- Lee HT, Xu H, Nasr SH, Schnermann J, Emala CW (2004b) A1 adenosine receptor knockout mice exhibit increased renal injury following ischemia and reperfusion. *Am J Physiol Renal Physiol* 286:F298–306

- Lee J, Ha JH, Kim S, Oh Y, Kim SW (2002) Caffeine decreases the expression of Na<sup>+</sup>/K<sup>+</sup>-ATPase and the type 3 Na<sup>+</sup>/H<sup>+</sup> exchanger in rat kidney. *Clin Exp Pharmacol Physiol* 29:559–63
- Lin J, Bonventre JV (2005) Prevention of radiocontrast nephropathy. *Curr Opin Nephrol Hypertens* 14:105–10
- Lin JJ, Churchill PC, Bidani AK (1986) Effect of theophylline on the initiation phase of post-ischemic acute renal failure in rats. *J Lab Clin Med* 108:150–4
- Lin JJ, Churchill PC, Bidani AK (1987) The effect of dipyridamole on the initiation phase of postischemic acute renal failure in rats. *Can J Physiol Pharmacol* 65:1491–5
- Lin JJ, Churchill PC, Bidani AK (1988) Theophylline in rats during maintenance phase of post-ischemic acute renal failure. *Kidney Int* 33:24–8
- Lin SL, Chen YM, Chiang WC, Wu KD, Tsai TJ (2008) Effect of pentoxifylline in addition to losartan on proteinuria and GFR in CKD: a 12-month randomized trial. *Am J Kidney Dis* 52:464–74
- Lin SL, Chen YM, Chien CT, Chiang WC, Tsai CC, Tsai TJ (2002) Pentoxifylline attenuated the renal disease progression in rats with remnant kidney. *J Am Soc Nephrol* 13:2916–29
- Ludens JH, Willis LR, Williamson HE (1970) The effect of aminophylline on renal hemodynamics and sodium excretion. *Arch Int Pharmacodyn Théor* 185:274–86
- Massey LK, Bergman EA, Wise KJ, Sherrard DJ (1994) Interactions between dietary caffeine and calcium on calcium and bone metabolism in older women. *J Am Coll Nutr* 13:592–6
- Mazkereth R, Laufer J, Jordan S, Pomerance JJ, Boichis H, Reichman B (1997) Effects of theophylline on renal function in premature infants. *Am J Perinatol* 14:45–9
- McCormick BB, Sydor A, Akbari A, Fergusson D, Doucette S, Knoll G (2008) The effect of pentoxifylline on proteinuria in diabetic kidney disease: a meta-analysis. *Am J Kidney Dis* 52:454–63
- McLaughlin GE, Abitbol CL (2005) Reversal of oliguric tacrolimus nephrotoxicity in children. *Nephrol Dial Transplant* 20:1471–5
- McLaughlin GE, Kashimawo LA, Steele BW, Kuluz JW (2003a) Reversal of acute tacrolimus-induced renal vasoconstriction by theophylline in rats. *Pediatr Crit Care Med* 4:358–62
- McLaughlin GE, Schober M, Perez M, Ruiz P, Steele BW, Abitbol C (2003b) Benefit of theophylline administration in tacrolimus-induced nephrotoxicity in rats. *Pediatr Nephrol* 18:860–4
- Mi Z, Herzer WA, Zhang Y, Jackson EK (1994) 3-Isobutyl-1-methylxanthine decreases renal cortical interstitial levels of adenosine and inosine. *Life Sci* 54:277–82
- Mizumoto H, Karasawa A (1993) Renal tubular site of action of KW-3902, a novel adenosine A<sub>1</sub>-receptor antagonist, in anesthetized rats. *Jpn J Pharmacol* 61:251–3
- Moosavi SM, Bayat G, Owji SM, Panjehshahin MR (2009) Early renal post-ischaemic tissue damage and dysfunction with contribution of A<sub>1</sub>-adenosine receptor activation in rat. *Nephrology (Carlton)* 14:179–88
- Nagashima K, Kusaka H, Karasawa A (1995) Protective effects of KW-3902, an adenosine A<sub>1</sub>-receptor antagonist, against cisplatin-induced acute renal failure in rats. *Jpn J Pharmacol* 67:349–57
- Nauli SM, Alenghat FJ, Luo Y, Williams E, Vassilev P, Li X, Elia AE, Lu W, Brown EM, Quinn SJ, Ingber DE, Zhou J (2003) Polycystins 1 and 2 mediate mechanosensation in the primary cilium of kidney cells. *Nat Genet* 33:129–37
- Neuhauser B, Beine S, Verwied SC, Luhrmann PM (1997) Coffee consumption and total body water homeostasis as measured by fluid balance and bioelectrical impedance analysis. *Ann Nutr Metab* 41:29–36
- Nishiyama A, Kimura S, He H, Miura K, Rahman M, Fujisawa Y, Fukui T, Abe Y (2001) Renal interstitial adenosine metabolism during ischemia in dogs. *Am J Physiol Renal Physiol* 280:F231–8
- Nussberger J, Mooser V, Maridor G, Juillerat L, Waeber B, Brunner HR (1990) Caffeine-induced diuresis and atrial natriuretic peptides. *J Cardiovasc Pharmacol* 15:685–91

- Ohnishi A, Branch RA, Jackson K, Hamilton R, Biaggioni I, Deray G, Jackson EK (1986) Chronic caffeine administration exacerbates renovascular, but not genetic, hypertension in rats. *J Clin Invest* 78:1045–50
- Okusa MD, Linden J, Macdonald T, Huang L (1999) Selective A2A adenosine receptor activation reduces ischemia-reperfusion injury in rat kidney. *Am J Physiol* 277:F404–12
- Oliw E, Anggard E, Fredholm BB (1977) Effect of indomethacin on the renal actions of theophylline. *Eur J Pharmacol* 43:9–16
- Onrot J, Goldberg MR, Biaggioni I, Hollister AS, Kingaid D, Robertson D (1985) Hemodynamic and humoral effects of caffeine in autonomic failure. Therapeutic implications for postprandial hypotension. *N Engl J Med* 313:549–54
- Orloff J, Handler JS (1962) The similarity of effects of vasopressin, adenosine-3',5'-phosphate (cyclic AMP) and theophylline on the toad bladder. *J Clin Invest* 41:702–9
- Osswald H (1975) Renal effects of adenosine and their inhibition by theophylline in dogs. *Naunyn Schmiedebergs Arch Pharmacol* 288:79–86
- Osswald H, Helmlinger J, Jendralski A, Abrar B (1979) Improvement of renal function by theophylline in acute renal failure of the rat. *Naunyn Schmiedebergs Arch Pharmacol* 307 (Suppl):R47 (abstract)
- Osswald H, Nabakowski G, Hermes H (1980) Adenosine as a possible mediator of metabolic control of glomerular filtration rate. *Int J Biochem* 12:263–7
- Osswald H, Schmitz HJ, Kemper R (1977) Tissue content of adenosine, inosine and hypoxanthine in the rat kidney after ischemia and postischemic recirculation. *Pflugers Arch* 371:45–9
- Palatini P, Canali C, Graniero GR, Rossi G, de Toni R, Santonastaso M, dal Follo M, Zanata G, Ferrarese E, Mormino P, Pessina AC (1996) Relationship of plasma renin activity with caffeine intake and physical training in mild hypertensive men. HARVEST Study Group. *Eur J Epidemiol* 12:485–91
- Panjehshahin MR, Munsey TS, Collis MG, Bowmer CJ, Yates MS (1992) Further characterization of the protective effect of 8-cyclopentyl-1,3-dipropylxanthine on glycerol-induced acute renal failure in the rat. *J Pharm Pharmacol* 44:109–13
- Passmore AP, Kondowe GB, Johnston GD (1987) Renal and cardiovascular effects of caffeine: a dose-response study. *Clin Sci (Lond)* 72:749–56
- Paul S, Jackson EK, Robertson D, Branch RA, Biaggioni I (1989) Caffeine potentiates the renin response to furosemide in rats. Evidence for a regulatory role of endogenous adenosine. *J Pharmacol Exp Ther* 251:183–7
- Pawlowska D, Granger JP, Knox FG (1987) Effects of adenosine infusion into renal interstitium on renal hemodynamics. *Am J Physiol* 252:F678–82
- Perkins RM, Aboudara MC, Uy AL, Olson SW, Cushner HM, Yuan CM (2009) Effect of pentoxifylline on GFR decline in CKD: a pilot, double-blind, randomized, placebo-controlled trial. *Am J Kidney Dis* 53:606–16
- Persson PB, Hansell P, Liss P (2005) Pathophysiology of contrast medium-induced nephropathy. *Kidney Int* 68:14–22
- Pfeifer CA, Suzuki F, Jackson EK (1995) Selective A1 adenosine receptor antagonism augments beta-adrenergic-induced renin release in vivo. *Am J Physiol* 269:F469–79
- Pingle SC, Mishra S, Marcuzzi A, Bhat SG, Sekino Y, Rybak LP, Ramkumar V (2004) Osmotic diuretics induce adenosine A1 receptor expression and protect renal proximal tubular epithelial cells against cisplatin-mediated apoptosis. *J Biol Chem* 279:43157–67
- Potter M, Aparicio M, Cambar J (1997) Protective effect of three xanthine derivatives (theophylline, caffeine and pentoxifylline) against the cyclosporin A-induced glomerular contraction in isolated glomeruli and cultured mesangial cells. *Nephron* 77:427–34
- Premen AJ, Hall JE, Mizelle HL, Cornell JE (1985) Maintenance of renal autoregulation during infusion of aminophylline or adenosine. *Am J Physiol* 248:F366–73
- Prevot A, Liet JM, Semama DS, Justabo E, Guignard JP, Gouyon JB (2002) Disparate effects of chronic and acute theophylline on cyclosporine A nephrotoxicity. *Pediatr Nephrol* 17:418–24

- Rachima-Maoz C, Peleg E, Rosenthal T (1998) The effect of caffeine on ambulatory blood pressure in hypertensive patients. *Am J Hypertens* 11:1426–32
- Rakic V, Burke V, Beilin LJ (1999) Effects of coffee on ambulatory blood pressure in older men and women: a randomized controlled trial. *Hypertension* 33:869–73
- Reid IA, Stockigt JR, Goldfien A, Ganong WF (1972) Stimulation of renin secretion in dogs by theophylline. *Eur J Pharmacol* 17:325–32
- Ren Y, Arima S, Carretero OA, Ito S (2002) Possible role of adenosine in macula densa control of glomerular hemodynamics. *Kidney Int* 61:169–76
- Rieg T, Steigele H, Schnermann J, Richter K, Osswald H, Vallon V (2005) Requirement of intact adenosine A1 receptors for the diuretic and natriuretic action of the methylxanthines theophylline and caffeine. *J Pharmacol Exp Ther* 313:403–9
- Riesenhuber A, Boehm M, Posch M, Aufricht C (2006) Diuretic potential of energy drinks. *Amino Acids* 31:81–3
- Robertson D, Frolich JC, Carr RK, Watson JT, Hollifield JW, Shand DG, Oates JA (1978) Effects of caffeine on plasma renin activity, catecholamines and blood pressure. *N Engl J Med* 298:181–6
- Robertson D, Hollister AS, Kincaid D, Workman R, Goldberg MR, Tung CS, Smith B (1984) Caffeine and hypertension. *Am J Med* 77:54–60
- Rodriguez-Moran M, Guerrero-Romero F (2005) Pentoxifylline is as effective as captopril in the reduction of microalbuminuria in non-hypertensive type 2 diabetic patients—a randomized, equivalent trial. *Clin Nephrol* 64:91–7
- Rossi N, Ellis V, Kontry T, Gunther S, Churchill P, Bidani A (1990) The role of adenosine in HgCl<sub>2</sub>-induced acute renal failure in rats. *Am J Physiol* 258:F1554–60
- Saad SY, Najjar TA, Alashari M (2004) Role of non-selective adenosine receptor blockade and phosphodiesterase inhibition in cisplatin-induced nephrogonadal toxicity in rats. *Clin Exp Pharmacol Physiol* 31:862–7
- Schnermann J, Osswald H, Hermle M (1977) Inhibitory effect of methylxanthines on feedback control of glomerular filtration rate in the rat. *Pflugers Arch* 369:39–48
- Schnermann J, Weihprecht H, Briggs JP (1990) Inhibition of tubuloglomerular feedback during adenosine 1 receptor blockade. *Am J Physiol Renal Physiol* 258:F553–61
- Scott D, Rycroft JA, Aspen J, Chapman C, Brown B (2004) The effect of drinking tea at high altitude on hydration status and mood. *Eur J Appl Physiol* 91:493–8
- Seideman P, Sollevi A, Fredholm BB (1987) Additive renal effects of indomethacin and dipyrindamole in man. *Br J Clin Pharmacol* 23:323–30
- Shirley DG, Walter SJ, Noormohamed FH (2002) Natriuretic effect of caffeine: assessment of segmental sodium reabsorption in humans. *Clin Sci (Lond)* 103:461–6
- Siragy HM, Linden J (1996) Sodium intake markedly alters renal interstitial fluid adenosine. *Hypertension* 27:404–7
- Smits P, Hoffmann H, Thien T, Houben H, Van't Laar A (1983) Hemodynamic and humoral effects of coffee after beta 1-selective and nonselective beta-blockade. *Clin Pharmacol Ther* 34:153–8
- Spielman WS (1984) Antagonistic effect of theophylline on the adenosine-induced decreased in renin release. *Am J Physiol* 247:F246–51
- Suzuki F, Shimada J, Mizumoto H, Karasawa A, Kubo K, Nonaka H, Ishii A, Kawakita T (1992) Adenosine A1 antagonists. 2. Structure-activity relationships on diuretic activities and protective effects against acute renal failure. *J Med Chem* 35:3066–75
- Takeda M, Yoshitomi K, Imai M (1993) Regulation of Na(+)-3HCO<sub>3</sub>- cotransport in rabbit proximal convoluted tubule via adenosine A1 receptor. *Am J Physiol* 265:F511–9
- Tanner GA, Tanner JA (2001) Chronic caffeine consumption exacerbates hypertension in rats with polycystic kidney disease. *Am J Kidney Dis* 38:1089–95
- Thomson S, Bao D, Deng A, Vallon V (2000) Adenosine formed by 5'-nucleotidase mediates tubuloglomerular feedback. *J Clin Invest* 106:289–98

- Tofovic SP, Branch KR, Oliver RD, Magee WD, Jackson EK (1991) Caffeine potentiates vasodilator-induced renin release. *J Pharmacol Exp Ther* 256:850–60
- Tofovic SP, Jackson EK (1999) Effects of long-term caffeine consumption on renal function in spontaneously hypertensive heart failure prone rats. *J Cardiovasc Pharmacol* 33:360–6
- Tofovic SP, Kost CK Jr, Jackson EK, Bastacky SI (2002) Long-term caffeine consumption exacerbates renal failure in obese, diabetic, ZSF1 (fa-fa(cp)) rats. *Kidney Int* 61: 1433–44
- Tofovic SP, Kusaka H, Rominski B, Jackson EK (1999) Caffeine increases renal renin secretion in a rat model of genetic heart failure. *J Cardiovasc Pharmacol* 33:440–50
- Tofovic SP, Rominski BR, Bastacky S, Jackso EK, Kost CK Jr (2000) Caffeine augments proteinuria in puromycin-aminonucleoside nephrotic rats. *Ren Fail* 22:159–79
- Tofovic SP, Salah EM, Jackson EK, Melhem M (2007) Early renal injury induced by caffeine consumption in obese, diabetic ZSF1 rats. *Ren Fail* 29:891–902
- Tseng CJ, Kuan CJ, Chu H, Tung CS (1993) Effect of caffeine treatment on plasma renin activity and angiotensin I concentrations in rats on a low sodium diet. *Life Sci* 52:883–90
- Wang W, Zolty E, Falk S, Basava V, Reznikov L, Schrier R (2006) Pentoxifylline protects against endotoxin-induced acute renal failure in mice. *Am J Physiol Renal Physiol* 291:F1090–5
- Weihprecht H, Lorenz JN, Schnermann J, Skott O, Briggs JP (1990) Effect of adenosine1-receptor blockade on renin release from rabbit isolated perfused juxtaglomerular apparatus. *J Clin Invest* 85:1622–8
- Wemple RD, Lamb DR, McKeever KH (1997) Caffeine vs caffeine-free sports drinks: effects on urine production at rest and during prolonged exercise. *Int J Sports Med* 18:40–6
- Wilcox CS, Welch WJ, Schreiner GF, Belardinelli L (1999) Natriuretic and diuretic actions of a highly selective adenosine A1 receptor antagonist. *J Am Soc Nephrol* 10:714–720
- Wise KJ, Bergman EA, Sherrard DJ, Massey LK (1996) Interactions between dietary calcium and caffeine consumption on calcium metabolism in hypertensive humans. *Am J Hypertens* 9:223–9
- Yagil Y (1990) Interaction of adenosine with vasopressin in the inner medullary collecting duct. *Am J Physiol* 259:F679–87
- Yao K, Heyne N, Erley CM, Risler T, Osswald H (2001) The selective adenosine A1 receptor antagonist KW-3902 prevents radiocontrast media-induced nephropathy in rats with chronic nitric oxide deficiency. *Eur J Pharmacol* 414:99–104
- Yoneda H, Hisa H, Satoh S (1990) Effects of adenosine on adrenergically induced renal vasoconstriction in dogs. *Eur J Pharmacol* 176:109–16
- Zou AP, Wu F, Li PL, Cowley AW Jr (1999) Effect of chronic salt loading on adenosine metabolism and receptor expression in renal cortex and medulla in rats. *Hypertension* 33:511–6

# The Cardiovascular Effects of Methylxanthines

Niels P. Riksen, Paul Smits, and Gerard A. Rongen

## Contents

1	Introduction .....	414
2	The Molecular Targets of the Methylxanthines .....	415
3	The Cardiovascular Effects of Adenosine Receptor Stimulation .....	416
4	The Cardiovascular Effects of Methylxanthines .....	418
4.1	Overview of the Cardiovascular Actions of Methylxanthines .....	418
4.2	Effect of Methylxanthines on Blood Vessels and Organ Perfusion .....	419
4.3	Actions of Methylxanthines on the Heart .....	421
4.4	Action of Methylxanthines on the Autonomic Nervous System and the Renin–Angiotensin System .....	422
5	The Cardiovascular Effects of Dietary Caffeine .....	424
5.1	Tolerance to the Cardiovascular Effects of Caffeine .....	424
5.2	The Effects of Long-Term Coffee Consumption on Cardiovascular Risk Factors .....	425
5.3	Association Between Coffee Consumption and Coronary Heart Disease .....	427
	References .....	430

**Abstract** In the concentration range that is normally achieved in humans, e.g., after the drinking of coffee or in patients treated with theophylline, the cardiovascular effects of methylxanthines are primarily due to antagonism of adenosine A<sub>1</sub> and A<sub>2</sub> receptors. Inhibition of phosphodiesterases or mobilization of intracellular calcium requires much higher concentrations. In conscious humans, acute

---

G.A.R. is a Clinical Established Investigator of the Netherlands Heart Foundation (2006 T035).

N.P. Riksen (✉), P. Smits and G.A. Rongen

Departments of Pharmacology-Toxicology and Internal Medicine, Radboud University Nijmegen Medical Centre, PO Box 9101, 6500 HB Nijmegen, The Netherlands

e-mail: n.riksen@aig.umcn.nl

exposure to caffeine results in an increase in blood pressure by an increased total peripheral resistance, and a slight decrease in heart rate. This overall hemodynamic response is composed of direct effects of caffeine on vascular tone, on myocardial contractility and conduction, and on the sympathetic nervous system. Caffeine is the most widely consumed methylxanthine, mainly derived from coffee intake. Regular coffee consumption can affect various traditional cardiovascular risk factors, including a slight increase in blood pressure, an increase in plasma cholesterol and homocysteine levels, and a reduced incidence of type 2 diabetes mellitus. Although most prospective studies have not reported an association between coffee consumption and coronary heart disease, these findings do not exclude that the acute hemodynamic and neurohumoral effects of coffee consumption could have an adverse effect in selected patient groups who are more vulnerable for these effects, based on their genetic profile or medication use.

**Keywords** Adenosine · Blood pressure · Caffeine · Cardiovascular effects

## 1 Introduction

Systemic administration of methylxanthines can profoundly affect hemodynamic parameters, such as blood pressure and heart rate. These hemodynamic changes result from direct effects on myocardial contractility and conduction, on vascular tone, and on the sympathoadrenal system. The net effect on blood pressure and heart rate can differ substantially between the various methylxanthines. Thorough knowledge of these cardiovascular effects of methylxanthines is important for three reasons. Firstly, methylxanthine derivatives have been used in patients for their distinct cardiovascular effects, such as pentoxifylline in patients with peripheral artery disease. Secondly, these cardiovascular effects can explain some of the side effects of methylxanthine derivatives that are used in other patient group, e.g., as bronchodilators in patients with obstructive pulmonary disease. But most importantly, the cardiovascular effects of caffeine, which is the most widely consumed methylxanthine derivative, can have important effects on cardiovascular morbidity and mortality in the general population (Riksen et al. 2009).

In this chapter, we will discuss in detail the separate effects of methylxanthines on the heart, the blood vessels, the sympathoadrenal and renin–angiotensin system, and the integrative hemodynamic effects. We will discuss effects observed in *in vitro* experiments, and in animal experiments, but our main focus is on the effects observed in humans *in vivo* in concentrations that are reached in daily clinical practice. We will start with a brief summary of the molecular targets of the methylxanthines, which are relevant to their cardiovascular effects. Finally, we will discuss the potential impact of the cardiovascular effects of dietary caffeine, mainly derived from coffee consumption, on cardiovascular morbidity and mortality in the general population.



## 2 The Molecular Targets of the Methylxanthines

The molecular mechanisms of methylxanthines are diverse, and include inhibition of cyclic nucleotide phosphodiesterases, mobilization of intracellular calcium, and adenosine receptor antagonism. Francis et al. (2010), Guerreiro et al. (2010), and Müller and Jacobson (2010) are devoted to describing these mechanisms in detail. Here, we will briefly review the molecular targets that are relevant to the cardiovascular effects of the methylxanthines, particularly when they are given in concentrations within the therapeutic range.

Caffeine can increase the free intracellular calcium concentration of muscle cells by activation of ryanodine receptors, which are calcium-release channels located in the endoplasmic reticulum, but only in concentrations of more than 1 mM, which is much higher than the concentration reached after coffee consumption (Fredholm et al. 1999; McPherson et al. 1991). Butcher and Sutherland (1962) reported in the early 1960s that theophylline, caffeine, and theobromine inhibited bovine cyclic adenosine monophosphate (cAMP) phosphodiesterase with  $EC_{50}$  values of approximately 200–300  $\mu$ M for theophylline, and 1,000  $\mu$ M for caffeine and theobromine. These values are in the same range as those obtained in more recent studies (Fredholm 1984). Therefore, at the therapeutic range of 20–80  $\mu$ M for theophylline there is relatively little phosphodiesterase inhibition. It is even more obvious for caffeine that the concentration required to inhibit phosphodiesterase is by far higher than the concentration reached after regular coffee consumption (Fredholm 1984).

As mobilization of intracellular calcium stores and inhibition of phosphodiesterase activity require millimolar and high micromolar concentrations of caffeine, these molecular targets do not explain the cardiovascular effects of caffeine in humans *in vivo*. In contrast, caffeine effectively antagonizes adenosine receptors in the concentration range reached after regular coffee consumption. Also, other methylxanthines, such as theophylline, have been shown to antagonize adenosine receptors in the therapeutic concentration range (Sattin and Rall 1970). Currently, four adenosine receptor subtypes have been identified, designated as adenosine  $A_1$ ,  $A_{2A}$ ,  $A_{2B}$ , and  $A_3$  receptors (Fredholm et al. 2001a). Traditionally, the adenosine receptors are classified on the basis of their ability to decrease or increase intracellular cAMP concentration: adenosine  $A_1$  and  $A_3$  receptors are coupled to  $G_i$  proteins and stimulation will decrease the intracellular cAMP level. In contrast, adenosine  $A_{2A}$  and  $A_{2B}$  receptor stimulation increases cAMP levels via  $G_s$  proteins (Fredholm et al. 2001a). Functional studies on human adenosine receptors using intracellular cAMP concentrations have shown that caffeine blocks adenosine receptors with  $K_b$  values of approximately 30, 10, 15, and more than 100  $\mu$ M for the adenosine  $A_1$ ,  $A_{2A}$ ,  $A_{2B}$ , and  $A_3$  receptors, respectively, which are similar to the potencies reported earlier in studies using binding assays (Fredholm et al. 2001b). The effect of competitive antagonists is dependent on the occupancy of the receptors with the endogenous ligand. The levels of adenosine under baseline normoxic conditions are sufficient to activate the adenosine  $A_1$ ,  $A_{2A}$ , and  $A_3$  receptors. Therefore, caffeine is generally believed to act on the adenosine  $A_1$  and  $A_{2A}$

receptors to exert most of its effects. Theophylline and paraxanthine have slightly higher affinities than caffeine for the adenosine A<sub>1</sub>, A<sub>2A</sub>, and A<sub>2B</sub> receptors and are also weak antagonists for the adenosine A<sub>3</sub> receptor (Fredholm et al. 2001b; Klotz et al. 1998). The affinities of other methylxanthines, including theobromine and pentoxifylline, are lower than that of caffeine (Fredholm and Persson 1982; Schwabe et al. 1985). Enprofylline has an affinity profile that differs from that of the other methylxanthines in that it has very low affinities for adenosine A<sub>1</sub>, A<sub>2A</sub>, and A<sub>3</sub> receptors, whereas it is as potent as theophylline in blocking the adenosine A<sub>2B</sub> receptor (Feoktistov and Biaggioni 1995). Additional evidence that the hemodynamic effects of caffeine are indeed due to adenosine receptor antagonism has recently been obtained in a study in mice heterozygous for both the adenosine A<sub>1</sub> receptor gene and the adenosine A<sub>2A</sub> receptor gene. In female control mice, 7 days of caffeine administration reduced the heart rate. In A<sub>1</sub>/A<sub>2A</sub> heterozygous mice, however, the baseline heart rate was lower, and was not affected by caffeine administration (Yang et al. 2009).

Two additional molecular targets of methylxanthines related to purine pharmacology have been reported, but, given the high concentrations required to activate these targets, both mechanisms are unlikely to contribute significantly to any cardiovascular effect of methylxanthines *in vivo*. At a concentration of 0.5 mM, theophylline, caffeine, and theobromide inhibit 5'-nucleotidase, isolated from rabbit kidney (Fredholm et al. 1978) and rat brain (Tsuzuki and Newburgh 1975). In addition, theophylline and caffeine can inhibit the rat nucleoside transporter with IC<sub>50</sub> values of approximately 1 and 3 mM (Plagemann and Wohlhueter 1984).

On the basis of these pharmacology studies, it has been argued that adenosine receptor antagonism is the single most important mechanism underlying the cardiovascular effects of caffeine and theophylline in humans *in vivo* (Fredholm 1980; Fredholm et al. 2001a). On the other hand, other methylxanthines, which are more potent phosphodiesterase inhibitors, might act primarily through this mechanism (Kamphuis et al. 1994). Indeed, the hemodynamic effects of adenosine in humans *in vivo* are blunted by concomitant administration of caffeine (Smits et al. 1987, 1989) and theophylline (Biaggioni et al. 1991; Smits et al. 1990), but not enprofylline (Smits et al. 1989).

### 3 The Cardiovascular Effects of Adenosine Receptor Stimulation

To facilitate accurate prediction of the cardiovascular effects of methylxanthines, the effects of adenosine receptor stimulation by its endogenous ligand, and the potential effects of methylxanthines on adenosine receptor expression and function will be briefly discussed in this section. A series of experiments on human adenosine receptors using changes in intracellular cAMP levels revealed EC<sub>50</sub> values for adenosine of approximately 0.3, 0.7, 24, and 0.3 μM for the adenosine A<sub>1</sub>, A<sub>2A</sub>,

$A_{2B}$ , and  $A_3$  receptors, respectively (Fredholm et al. 2001b). As the endogenous adenosine concentration during normoxia is in the range 20–300 nM (Ramakers et al. 2008), the adenosine  $A_1$  and  $A_3$  and to a lesser extent the  $A_{2A}$  receptor can be activated by physiological concentrations of adenosine, but this is also dependent on the receptor density of the target cells (Fredholm et al. 2001b). In contrast, the low-affinity adenosine  $A_{2B}$  receptor is only activated in situations in which the adenosine concentration is increased, such as during ischemia or inflammation.

Interestingly, in addition to direct adenosine receptor antagonism, methylxanthines have also been reported to affect adenosine receptor density and the endogenous adenosine concentration. In humans, administration of caffeine (750 mg per day for 1 week) induced an upregulation of the adenosine  $A_{2A}$  receptor on thrombocytes accompanied by an increased potency of an adenosine  $A_{2A}$  receptor agonist to increase intracellular cAMP levels (Varani et al. 1999). In rats, administration of caffeine and 8-sulphophenyltheophylline was reported to increase the circulating adenosine concentration, but this finding has not been confirmed since then (Conlay et al. 1997).

It has been well established that systemic administration of adenosine in conscious humans elicits a typical hemodynamic response, consisting of an increase in heart rate and systolic blood pressure, and a slight drop in diastolic blood pressure (Biaggioni et al. 1986; Smits et al. 1987). This response is the overall result of various separate effects of adenosine on the myocardium, the vascular tone, and the activity of the sympathetic nervous system (Riksen et al. 2008; Rongen et al. 1997). In isolated heart preparations, adenosine has a negative inotropic, dromotropic, and chronotropic effect by activation of adenosine  $A_1$  receptors (Belardinelli et al. 1989). In addition, adenosine can attenuate ischemia-, and catecholamine-induced cardiac arrhythmias, probably by preventing catecholamine-induced release of calcium from the sarcoplasmic reticulum via adenosine  $A_1$  receptor stimulation (Song et al. 2001). With regard to vascular tone, adenosine induces vasodilation in most vascular beds via activation of the adenosine  $A_{2A}$  receptor (Belardinelli et al. 1998). Also in humans, selective activation of the adenosine  $A_{2A}$  receptor increases coronary blood flow (Lieu et al. 2007). Adenosine  $A_{2B}$  receptors can also contribute to the vasodilator effect of adenosine in humans (Kemp and Cocks 1999). In addition, experimental animal studies have provided evidence that adenosine  $A_1$  receptor stimulation can attenuate the vasodilator effect of adenosine  $A_{2A}$  receptor activation (Tawfik et al. 2005) and that the adenosine  $A_{2B}$  and  $A_3$  receptors can also contribute to coronary vasodilation in rats (Hinschen et al. 2003).

In addition to direct effects on the heart and vascular tone, adenosine can modulate the activity of the sympathetic nervous system. In conscious humans, adenosine has two distinct and opposite actions. Firstly, adenosine induces a reflex activation of the sympathetic nervous system by stimulation of adenosine-sensitive chemoreceptors located in the carotid body (Biaggioni et al. 1987; Timmers et al. 2004), and sympathetic afferents from the heart (Cox et al. 1989), kidney, and forearm skeletal muscle (Costa and Biaggioni 1994). Secondly, stimulation of presynaptic adenosine receptors can inhibit norepinephrine release from sympathetic nerve terminals (Rongen et al. 1996). *In vivo* studies have suggested that the adenosine  $A_{2A}$  and  $A_3$

receptors are responsible for this prejunctional effect (Barraco et al. 1995; Donoso et al. 2006). The augmentation of sympathetic nervous system traffic by stimulation of the carotid chemoreceptors appears to explain most of the hemodynamic effects of systemic administration of adenosine in conscious humans: administration of adenosine into the aorta proximal to the carotid chemoreceptors increases blood pressure and heart rate, whereas injection distal to these receptors decreases blood pressure (by direct unopposed vasodilation) (Biaggioni et al. 1987). Also, systemic administration of adenosine in patients after bilateral carotid body tumor resection decreases blood pressure and does not affect muscle sympathetic nerve activity, in contrast to the effects in healthy volunteers (Timmers et al. 2004).

In addition to these hemodynamic effects, adenosine receptor stimulation affects the function of platelets and inflammatory cells, and modulates intrinsic tolerance against ischemia and reperfusion. Activation of adenosine  $A_{2A}$  receptors on human platelets inhibits their aggregation (Varani et al. 2000). The adenosine  $A_{2A}$  receptor is also the primary and dominant adenosine receptor subtype responsible for the potent anti-inflammatory effects of adenosine (Hasko et al. 2008). Finally, endogenous adenosine is an important mediator of myocardial ischemic pre- and post-conditioning, which is defined as a reduction of lethal ischemia–reperfusion injury by a brief sublethal period of ischemia and reperfusion immediately before or after the lethal insult (Hausenloy and Yellon 2007). This protective effect is initiated by adenosine  $A_1$  and  $A_3$  receptor stimulation (Carr et al. 1997). In addition, stimulation of the adenosine  $A_{2A}$  receptor at the moment of myocardial reperfusion can also limit infarct size.

## 4 The Cardiovascular Effects of Methylxanthines

### 4.1 Overview of the Cardiovascular Actions of Methylxanthines

In healthy volunteers, acute exposure to caffeine in a dose that is regularly encountered after intake of two to three cups of coffee increases blood pressure by increasing total peripheral resistance without relevantly affecting cardiac output (Casiglia et al. 1991; Farag et al. 2005; Pincomb et al. 1985), although the heart rate often decreases slightly (Rongen et al. 1995). Exposure to extremely high plasma concentrations of methylxanthines, as occurs in theophylline intoxication, induces tachyarrhythmias and hypotension (Woo et al. 1984).

Discussion of the mechanisms involved in these cardiovascular actions of methylxanthines should take into account the various targets of these compounds that are concentration-dependently engaged in the overall effect of methylxanthine exposure. Furthermore, a distinction between *direct actions* on the heart, blood vessels, and regulatory systems (in particular, the autonomic nervous system, adrenal glands and renin–angiotensin system) and *indirect or secondary effects* that may result, for example, from metabolic vasodilation in the coronary system in

response to increased afterload or from baroreflex modulation of the autonomic nervous system or the renin–angiotensin system in response to blood pressure changes should be appreciated. Finally, since the adenosine receptors are an important target of methylxanthines at relevant concentrations that occur during daily life, the availability of endogenous extracellular adenosine, which may differ between organs, the conditions studied, and the experimental setups used should be taken into account.

To disentangle all these different factors that could potentially modulate the cardiovascular response to a methylxanthine, we will first describe the action of methylxanthines in isolated organs. Subsequently, we will discuss the actions of methylxanthines on the autonomic nervous system and the renin–angiotensin system.

#### ***4.2 Effect of Methylxanthines on Blood Vessels and Organ Perfusion***

Most *in vitro* studies on the direct vasomotor action of xanthine derivatives in isolated vessel preparations have revealed a vasodilator action (Bardou et al. 2002; Brodmann et al. 2003; Grossmann et al. 1998; Harada et al. 1995; Lo et al. 2005; Sekiguchi et al. 2002). The explanation for the vasodilator response to methylxanthines in these isolated vessels is probably twofold: firstly, the concentrations used in these studies are relatively high, resulting in significant phosphodiesterase inhibition as reflected by increases in cAMP and cyclic guanosine monophosphate, two important vasodilating second messengers; secondly, in these denervated nonperfused vessels with low metabolic activity, extracellular adenosine formation is probably low, resulting in a reduced baseline contribution of adenosine receptor stimulation to vascular tone.

In humans, the vasomotor action of methylxanthines has been studied in the forearm model, a method to explore direct actions of drugs on vasomotor tone without relevant systemic exposure that could otherwise bias results by triggering central actions or reflex (counter) regulations. In these studies, both caffeine and theophylline induce a vasodilator response at relatively high doses without any effect on vascular tone at lower doses. Although both methylxanthines antagonize adenosine-induced vasodilation in this model, the vasodilator effect of methylxanthines on baseline tone cannot be explained by adenosine-receptor antagonism. Therefore, it is generally assumed that this action is mediated by phosphodiesterase inhibition. Alternatively, caffeine may augment endothelial nitric oxide release, resulting in vasodilation. However, although this phenomenon may occur in response to acetylcholine, an effect of caffeine on basal nitric oxide release in the forearm vasculature has not been demonstrated yet (Umemura et al. 2006). Furthermore, this observation contrasts with observations from others who reported a decrease in acetylcholine-mediated vasodilation in response to acute caffeine

exposure (Papamichael et al. 2005). The lack of a vasoconstrictor response in the forearm suggests that in this particular vascular bed adenosine does not importantly contribute to basal vascular tone. Most likely, this is explained by the low metabolic rate of resting muscle, although there is evidence for extracellular adenosine formation in the forearm as shown by the vasodilator response to inhibition of the equilibrative nucleoside transporter by either draflazine or dipyridamole (Bijlstra et al. 2004; Rongen et al. 1996). The extracellular adenosine concentration is mainly determined, however, by the rapid uptake of extracellular adenosine by neighboring cells. As such, extracellular adenosine concentrations can still be very low despite significant extracellular adenosine formation. Caffeine inhibited vasodilation in response to exercise without affecting baseline blood flow to muscle, supporting the concept of a differential contribution of adenosine to vascular tone in resting and exercising muscle (Daniels et al. 1998).

In the heart, the metabolic rate is much higher than in resting skeletal muscle. In the human coronary circulation, theophylline reduces basal coronary blood flow, which has been taken as evidence for a role for adenosine in the basal regulation of coronary vascular tone (Edlund et al. 1995; Edlund and Sollevi 1995). However, to our knowledge, these observations have not been confirmed by other research groups. Furthermore, evidence in animals suggests that methylxanthines only reduce coronary flow in conditions of increased metabolic demand or reduced oxygen supply (Ishibashi et al. 1998; Melchert et al. 1999; Phillis et al. 1998), although others have found a vasoconstrictor response to aminophylline in coronary arteries *in vivo* in baseline conditions at low doses that did not affect myocardial contractility (Paoloni and Wilcken 1975).

In the kidney of hypertensive patients, caffeine inhibits adenosine-induced vasodilation but has no effect on basal renal flow. In anaesthetized rabbits, injection of the adenosine receptor antagonist 8-sulfophenyltheophylline into the renal artery did not affect baseline renal flow either (Eppel et al. 2006). Adenosine has a complex action in the renal circulation to serve the tubuloglomerular feedback: in the afferent arterioles adenosine induces vasoconstriction (adenosine- $A_1$ -receptor-mediated), whereas in the efferent arterioles adenosine induces vasodilation by activation of adenosine  $A_2$  receptors. Therefore, caffeine may not affect overall renal blood flow but may still affect glomerular circulation, resulting in disruption of the tubuloglomerular feedback system and an increase in the glomerular filtration fraction (Persson 2001). This action of caffeine and theophylline may contribute to their mild diuretic action.

Thus, despite an increase in total peripheral resistance after acute exposure to caffeine, investigations of the effect of methylxanthines on isolated vessels or organ perfusion have not revealed a consistent vasoconstrictor effect. An explanation for this apparent discrepancy may involve the lack of sensitivity to detect minimal vasoconstriction in individual vascular beds, or a significant vasoconstrictor effect in a vascular bed in which the effect of methylxanthines has not been extensively studied, such as the mesenteric circulation (Rutherford et al. 1981).

Apart from an effect on vascular tone in resistance vessels, caffeine has also been shown to affect conduit vessels: it adversely affects arterial stiffness (Karatzis

et al. 2005; Mahmud and Feely 2001; Vlachopoulos et al. 2003) and reduces flow-mediated dilation in the brachial artery (Papamichael et al. 2005). Aortic pulse wave velocity, a measure of aortic stiffness, was associated with a higher incidence of coronary heart disease and cardiovascular mortality in a prospective study among healthy older subjects (Sutton-Tyrrell et al. 2005). The drinking of caffeinated coffee, but not decaffeinated coffee, acutely increased pulse wave velocity in healthy subjects (Mahmud and Feely 2001). Moreover, coffee drinking resulted in an increased augmentation index of the aortic pressure waveform, indicating increased wave reflection (Karatzis et al. 2005; Mahmud and Feely 2001). The intake of 250 mg of caffeine has been demonstrated to induce similar effects on arterial stiffness (Vlachopoulos et al. 2003), suggesting that caffeine is the compound in coffee responsible for the observed effects.

### ***4.3 Actions of Methylxanthines on the Heart***

Caffeine and theophylline antagonize the effects of endogenous adenosine on cardiac rhythm and conduction. Thus, in theory, these substances could increase the firing rate in the sinus node and increase atrioventricular nodal conduction velocity. At higher concentrations, methylxanthines also inhibit phosphodiesterases, resulting in augmentation of the  $\beta$ -adrenergic rise in intracellular cAMP levels. On top of that, through interactions with adenosine receptors in the autonomic nervous system, norepinephrine release in the heart may be increased (see Sect. 4.4). Therefore, it does not come as a surprise that abundant use of methylxanthines has been associated with cardiac arrhythmias (Agwunobi et al. 1996; Cannon et al. 2001; Ishida et al. 1996), in particular when combined with cocaine or amphetamines (McNamara et al. 2007; Mehta et al. 2004). However, in an unselected population the regular use of caffeine did not significantly modulate the risk for arrhythmias such as atrial fibrillation (Frost and Vestergaard 2005). Methylxanthines have been used successfully in the treatment of bradyarrhythmias, in particular in settings of increased extracellular adenosine appearance (Cawley et al. 2001; DeLago et al. 2008).

Aminophylline, caffeine, and theophylline have a direct positive inotropic action on myocardial cells (Dimarco et al. 1985; Paoloni and Wilcken 1975; Rutherford et al. 1981). The mechanism of this action depends on the concentration of the methylxanthine and the location (atrial versus ventricular tissue). In atrial tissue, adenosine  $A_1$  receptor stimulation results in sarcolemmal hyperpolarization, which prevents calcium influx and subsequent contractility. This response is independent of sympathetic tone (Urquhart and Broadley 1992a, b). Blockade of these adenosine receptors with methylxanthines in the presence of endogenous adenosine will result in increased contractility. In ventricular myocardial cells, adenosine  $A_1$  receptor stimulation only reduces contractility in the presence of  $\beta$ -adrenergic receptor stimulation by inhibition of  $\beta$ -adrenoceptor-induced activation of adenylate cyclase (Belardinelli et al. 1995). Therefore, blockade of these ventricular adenosine



receptors by methylxanthines will only result in increased contractility in the presence of sympathetic tone. At higher methylxanthine concentrations, this positive inotropic action of methylxanthines will be augmented by inhibition of phosphodiesterase, resulting in a further increase in cAMP levels. When the concentrations are raised to high (toxic) levels, caffeine and theophylline increase cellular calcium influx by activating the ryanodine receptor (Kong et al. 2008), with a positive effect on contractility (Rasmussen et al. 1987).

Only recently, the effect of coffee and caffeine on ischemia–reperfusion injury of the myocardium has been investigated. It has consistently been shown in experimental studies that a brief period of ischemia increases the tolerance against a subsequent prolonged ischemic insult, a phenomenon which has been termed “ischemic preconditioning” (Yellon and Downey 2003). This mechanism has been implied to explain the observation that the infarct size is smaller in patients who had experienced angina in the hours preceding the acute myocardial infarction (Yellon and Downey 2003). It transpired that several drugs, including hydroxymethylglutaryl-CoA reductase inhibitors (statins), could mimic this infarct size-limiting effect. Indeed, it has recently been demonstrated that treatment with atorvastatin improved the outcome in patients undergoing primary percutaneous intervention for an acute myocardial infarction (Patti et al. 2007). Interestingly, the cardioprotective effect of both ischemic preconditioning and statins is mediated by adenosine receptor stimulation (Sanada et al. 2004; Yellon and Downey 2003). Consequently, consumption of caffeinated coffee, but not decaffeinated coffee, appeared to abolish the cardioprotective effect of atorvastatin in a rat model of myocardial infarction (Ye et al. 2008). Recently, similar results were obtained with rosuvastatin and intravenous caffeine administration in a human forearm model of ischemia–reperfusion injury (Meijer et al. 2009). Likewise, in two human experimental models of ischemia–reperfusion injury, the intravenous administration of a single dose of caffeine (4 mg/kg) completely blocked the protective effect of ischemic preconditioning (Riksen et al. 2006).

#### ***4.4 Action of Methylxanthines on the Autonomic Nervous System and the Renin–Angiotensin System***

Acute exposure of humans to caffeine results in a doubling of the plasma epinephrine concentration without significantly affecting the plasma norepinephrine concentration (Smits et al. 1986), although a relatively small rise in plasma norepinephrine concentration has also been observed (Robertson et al. 1978). Comparison of the hemodynamic response to caffeine between healthy controls and bilaterally adrenalectomized patients (in whom plasma epinephrine remained undetectable) revealed that the adrenal release of epinephrine is only partially involved in the pressor response to caffeine (Smits et al. 1986).

Intravenous administration of caffeine reduces muscle sympathetic nerve activity in the human peroneal nerve of healthy volunteers, probably due to a baroreflex



inhibition of sympathetic outflow in response to the increase in blood pressure (Notarius et al. 2001). Taken together, these results suggest that caffeine facilitates norepinephrine release from sympathetic nerve endings. However, it should be realized that plasma norepinephrine concentrations do not directly reflect norepinephrine release from nerve endings. Owing to an active norepinephrine reuptake mechanism and metabolism, only a small fraction of the released norepinephrine appears in the circulation. Nevertheless, this proposed action of caffeine is supported by the observation that both endogenous as well as exogenous adenosine reduce forearm norepinephrine release in humans as measured with a tracer technique that partially circumvents the problems in the interpretation of plasma norepinephrine levels (Rongen et al. 1998a, 1996). Furthermore, a wealth of preclinical evidence indicates that adenosine inhibits norepinephrine release from sympathetic nerve endings by activation of a presynaptic adenosine receptor (see Sect. 3). By blocking these receptors, caffeine could facilitate norepinephrine release, thus preventing a baroreflex reduction in plasma norepinephrine concentration. Presynaptic facilitation of norepinephrine release by a caffeine-induced release of epinephrine provides an alternative explanation for the unaffected or even increased plasma norepinephrine concentration in the setting of a baroreflex-mediated inhibition of sympathetic nerve traffic (Newton et al. 1999). This is supported by the observation in adrenalectomized patients that the plasma norepinephrine concentration decreased in response to caffeine, whereas it remained unaffected in healthy controls (Smits et al. 1986).

In patients with heart failure, a condition that increases the endogenous adenosine formation, methylxanthines either do not affect baseline muscle sympathetic nerve activity or increase this measure of postganglionic sympathetic activity (Andreas et al. 2004; Notarius et al. 2001), an observation, however, that has been disputed by Notarius et al. (2003). Interestingly, the exercise pressor reflex in patients with heart failure is strongly increased. Caffeine inhibited this exaggerated response in patients with heart failure, suggesting that in patients with heart failure, caffeine may prevent some adverse actions of endogenous adenosine, namely, the activation of metaboreceptors in exercising muscle (Notarius et al. 2001).

The reported effect of caffeine on the plasma renin activity varies from a fall in plasma renin activity (Smits et al. 1986) to an increase (Robertson et al. 1978). Direct infusion of caffeine into the renal artery did not affect plasma renin concentration (Wierema et al. 2005), suggesting that the previous observations after systemic exposure were confounded by the blood pressure response to caffeine, which could reduce renal renin release, by the diuretic action of caffeine on renal tubules or by caffeine-induced epinephrine release, which could potentially increase renal renin release.

In theory, methylxanthines could modulate the vascular release of or response to angiotensin II by blocking adenosine receptors (Lai et al. 2006; Taddei et al. 1991). Whether these actions have any relevance for the cardiovascular effects of systemic exposure to methylxanthines remains to be established. In the human forearm, angiotensin II receptor blockade did not affect the vasomotor response to adenosine, suggesting that adenosine–angiotensin II interactions are not relevant in

peripheral vessels in humans (Rongen et al. 1998a). However, in the human renal circulation, the vasoconstrictor response to angiotensin II was reduced by oral caffeine administration (Brown et al. 1993), which is in agreement with observations in animals using 8-cyclopentyltheophylline, a methylxanthine with selectivity for the adenosine A<sub>1</sub> receptor (Lai et al. 2006).

## 5 The Cardiovascular Effects of Dietary Caffeine

Caffeine is by far the most widely consumed methylxanthine. Although caffeine is also present in tea, soft drinks, energy drinks, and chocolate, coffee is the most important source of dietary caffeine in adults. Experimental studies in healthy volunteers have demonstrated that the drinking of two cups of regular coffee acutely raises systolic and diastolic blood pressure and slightly lowers heart rate (Nurminen et al. 1999). It appears that caffeine is responsible for this pressor effect, as the same response is observed after administration of caffeine (Robertson et al. 1978), but not after the administration of decaffeinated coffee (Smits et al. 1985a).

### 5.1 *Tolerance to the Cardiovascular Effects of Caffeine*

When extrapolating these acute hemodynamic effects of coffee to the effects of its sustained consumption in daily life, one has to take into account tolerance to these acute effects. The pressor response and the increase in (nor)epinephrine levels in response to the administration of 250 mg of caffeine in healthy subjects was no longer present after 3 days of daily caffeine administration (Robertson et al. 1981). In addition, the daily drinking of caffeinated coffee acutely raised the blood pressure only in the first 5 days of the experiment (Ammon et al. 1983). More recent studies, however, have reported that only approximately half of all subjects showed complete tolerance to the acute pressor effect of caffeine, and that the others continued to show a significant pressor response with repeated dosing (Frag et al. 2005; Lovallo et al. 2004). In addition, after 1 week of regular coffee consumption (more than two cups per day), the pressor response to the administration of adenosine was still significantly attenuated 6 h after the last coffee intake (Rongen et al. 1998b). Furthermore, caffeinated coffee has been reported to consistently induce an acute increase in blood pressure in healthy regular coffee consumers, when administered after four to five half lives of caffeine in each subject (Smits et al. 1985b). Moreover, in this study, the increase in blood pressure appeared to be inversely related to the basal plasma caffeine concentration. These findings suggest that differences in the half life of caffeine could account for the observed inter-individual variation in the development of tolerance to the acute effects of caffeine, although in another study the baseline saliva caffeine concentrations did not differ between the tolerant and nontolerant groups (Lovallo et al. 2004).

## 5.2 The Effects of Long-Term Coffee Consumption on Cardiovascular Risk Factors

### 5.2.1 Long-Term Effect on Blood Pressure

Evidence for a sustained effect of coffee consumption on blood pressure can be derived from randomized controlled trials, usually performed in small groups of healthy subjects for a maximum of approximately 14 weeks (Jee et al. 1999; Noordzij et al. 2005), or from cross-sectional or longitudinal observational studies in large populations. Two meta-analyses of these randomized controlled trials have concluded that regular coffee intake slightly increases blood pressure by 1.2/0.5 mmHg (Noordzij et al. 2005) or 2.4/1.2 mmHg (Jee et al. 1999) for systolic/diastolic blood pressure, respectively. Table 1 summarizes the randomized controlled studies on coffee intake versus no coffee intake on blood pressure which were included in these meta-analyses. Interestingly, the blood pressure elevations were larger in the studies using caffeine tablets than in the studies on coffee consumption, suggesting that other compounds in the coffee could potentially counterbalance the pressor effect of caffeine. Cross-sectional epidemiology studies have provided conflicting results, but these studies are notoriously prone to confounding as patients with hypertension may have been advised to moderate their coffee intake. Four longitudinal cohort studies have investigated the impact of coffee consumption on blood pressure and the incidence of hypertension (Hu et al. 2007; Klag et al. 2002; Uiterwaal et al. 2007; Winkelmayr et al. 2005). Klag et al. (2002) have reported that consumption of one cup of coffee is associated with a blood pressure increase of 0.19/0.27 mmHg; however, after adjustment for confounding factors, coffee consumption was not associated with incident hypertension in this cohort. In 155,594 women participating in the Nurses' Health Study, coffee consumption was not associated with the incidence of hypertension

**Table 1** Summary of randomized controlled studies on the effect of long-term coffee intake versus no coffee intake on blood pressure

Reference	No. of subjects	Duration (days)	Type of coffee	Coffee (mL)	Caffeine (mg)	SBP change (mmHg)	DBP change (mmHg)
Bak and Grobbee (1990)	66	63	Filtered	700	469	6.1 (2.27)	3.0 (1.56)
Bak and Grobbee (1990)	62	63	Boiled	700	441	6.0 (2.17)	2.8 (1.77)
Burr et al. (1989)	54	28	Instant	1,235	741	2.9 (1.4)	-0.9 (1.20)
van Dusseldorp et al. (1991)	43	79	Boiled	900	774	3.5 (1.18)	0.9 (0.92)
van Dusseldorp et al. (1991)	42	79	Filtered	900	774	0.4 (0.98)	0.4 (0.90)
MacDonald et al. (1991) <sup>a</sup>	50	14	Instant	450	225	-0.7 (1.45)	0.1 (0.88)
Rakic et al. (1999) <sup>b</sup>	27	14	Instant	750	300	3.6 (1.60)	4.7 (1.20)
Rakic et al. (1999)	21	14	Instant	750	300	-1.6 (6.87)	-0.2 (4.18)
Rosmarin et al. (1990)	21	56	Filtered	540	270	2.1 (2.15)	-2.4 (2.45)
Superko et al. (1991)	120	56	Filtered	1,067	615	1.3 (1.57)	0.2 (1.18)
Superko et al. (1994)	99	56	Filtered	1,067	615	1.4 (1.51)	0.7 (1.29)

SBP systolic blood pressure, DBP diastolic blood pressure, Numbers in parenthesis indicate standard error values

<sup>a</sup>In patients with mild/moderate untreated hypertension

<sup>b</sup>In treated hypertensive patients

(Winkelmayer et al. 2005). More recently, Hu et al. (2007) have shown in a Finnish population that moderate coffee consumption (two to seven cups per day), but not heavy coffee consumption (8 cups or more per day), was associated with an increased risk for the initiation of antihypertensive drug treatment. In a Dutch population, coffee abstainers had a lower risk of hypertension than those with a coffee intake of more than zero to three cups per day (adjusted odds ratio 0.54, 95% confidence interval 0.31–0.92) (Uiterwaal et al. 2007).

### 5.2.2 Long-Term Effect on Plasma Cholesterol

In 1966 a significant correlation between coffee consumption and serum lipid concentrations was reported in men with coronary heart disease (Little et al. 1966). This possible association was subsequently studied in many cross-sectional observational studies (Thelle et al. 1987). The brewing method appeared to be a crucial factor in the cholesterol-raising effect of coffee. The diterpenes cafestol and kahweol, which are present in nonfiltered coffee, including boiled coffee, cafetière coffee, and Turkish coffee, but which are largely removed by filtering the coffee, appeared to be responsible for the increase in plasma cholesterol concentration (Urgert and Katan 1997). Indeed, only studies performed in populations drinking mainly boiled coffee or Turkish coffee showed an association between coffee consumption and the plasma cholesterol concentration (Jansen et al. 1995; Lindahl et al. 1991; Pietinen et al. 1990). A meta-analysis of randomized controlled trials in healthy subjects showed an average increase in total cholesterol of 0.31 mmol/L, particularly with consumption of six or more cups per day (Jee et al. 2001). Also in these studies, filtered coffee only slightly increased total cholesterol concentration, and did not increase LDL cholesterol concentration, in contrast to the studies using nonfiltered coffee. More recently, however, the drinking of 600 mL of filtered coffee daily for 4 weeks was shown to also significantly increase total cholesterol concentration (Strandhagen and Thelle 2003) and coffee abstinence for 6 weeks lowered total cholesterol concentration in healthy subjects normally consuming more than four cups of filtered coffee daily (Christensen et al. 2001), indicating that a cholesterol-raising effect is not absolutely restricted to unfiltered coffee.

### 5.2.3 Long-Term Effect on Plasma Homocysteine

Several cross-sectional studies have reported a positive dose-dependent relation between coffee consumption and the total plasma homocysteine concentration, with an increase of approximately 20% in the subjects with the highest coffee consumption (Nygard et al. 1997; Panagiotakos et al. 2004; Stolzenberg-Solomon et al. 1999). In addition, coffee abstinence for 6 weeks decreased the total plasma homocysteine concentration by approximately 1.5  $\mu\text{mol/L}$  in subjects who were used to drinking four or more cups per day, indicating a causal relationship (Christensen et al. 2001). Subsequent randomized controlled trials in healthy volunteers have obtained some insight into the possible mechanism of the

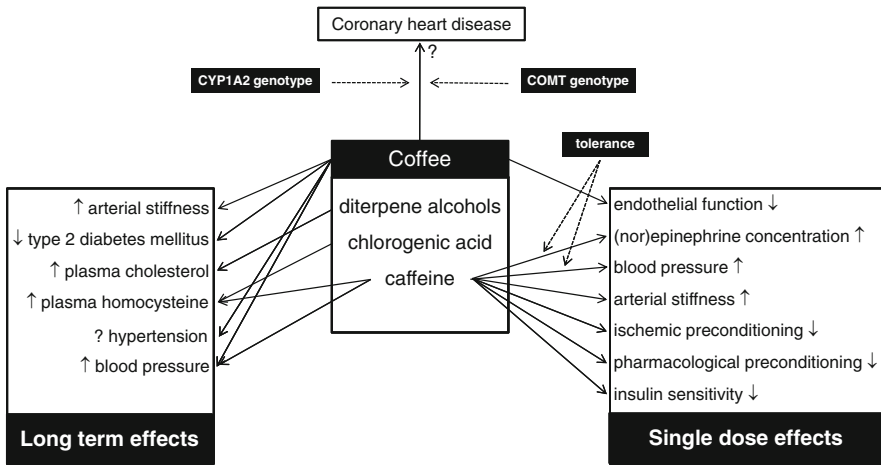
homocysteine-raising effect of coffee. Consumption of both filtered coffee (Verhoef et al. 2002) and unfiltered coffee (Grubben et al. 2000) for 2 weeks significantly increased plasma homocysteine levels. The observation that taking caffeine-filled capsules for 2 weeks also increased plasma homocysteine levels, but to a lesser extent than by drinking coffee, suggests that caffeine is partly, but not solely, responsible for this effect (Verhoef et al. 2002). Indeed, Olthof et al. (2001) demonstrated that a 7-day treatment with chlorogenic acid, one of the other components of coffee, also increased plasma homocysteine levels.

#### **5.2.4 Long-Term Effect on Type 2 Diabetes Mellitus**

In contrast to the potentially deleterious effect of coffee on plasma homocysteine and cholesterol levels, coffee consumption can also have beneficial effects with regard to cardiovascular health by reducing the incidence of type 2 diabetes mellitus. It was reported that subjects consuming seven or more cups of coffee per day had a relative risk of 0.50 (95% confidence interval 0.35–0.72) of developing type 2 diabetes mellitus compared with individuals drinking two or fewer cups per day (van Dam and Feskens 2002). A systematic review of nine prospective and seven cross-sectional cohort studies also showed a dose-dependent risk reduction for the development of type 2 diabetes, with a relative risk of 0.65 (95% confidence interval 0.54–0.78) and 0.72 (95% confidence interval 0.62–0.83) for drinking seven or more or five to six cups of coffee per day compared with two or fewer cups (van Dam and Hu 2005). More recent prospective studies have confirmed this beneficial effect of coffee intake on the incidence of type 2 diabetes mellitus (Paynter et al. 2006; Pereira et al. 2006; van Dam et al. 2006). Interestingly, in these latter studies, consumption of decaffeinated coffee protected to a similar extent against the development of diabetes as caffeinated coffee, which suggests that caffeine is not responsible for this observed effect (Pereira et al. 2006; van Dam et al. 2006). It has been speculated that chlorogenic acid and antioxidants present in coffee could mediate this protective effect (Riksen et al. 2009).

### **5.3 Association Between Coffee Consumption and Coronary Heart Disease**

As described in Sect. 4, administration of a single dose of caffeine induces various hemodynamic effects (Fig. 1). In addition, long-term caffeine administration can also affect hemodynamic parameters, as well as other independent cardiovascular risk factors (Fig. 1). As coffee is one of the most widely consumed beverages in the world, any possible association with the development of coronary heart disease might have a tremendous impact on the overall burden of disease. An impressive number of epidemiology studies have examined coffee as a potential risk factor for coronary heart disease. Four comprehensive meta-analyses of these trials have



**Fig. 1** Summary of the individual effects of coffee on cardiovascular parameters and risk factors, which could all contribute to an association between coffee consumption and cardiovascular disease. The effects of administration of a single dose of coffee are depicted on the *right*, whereas the effects of long-term coffee consumption are depicted on the *left*. It is indicated with *arrows* when an effect is attributed to one single component of coffee. The *text in the filled boxes* and the *dotted arrows* indicate effect modulators of specific effects of coffee (see the text). (Reproduced with permission from Riksen et al. 2009)

reported a positive association between coffee consumption and coronary heart disease in case-control studies, whereas there appeared to be no association, or a weaker association, in prospective cohort studies (Greenland 1993; Kawachi et al. 1994; Myers and Basinski 1992; Sofi et al. 2007). More recently, three large prospective studies concluded that coffee consumption does not increase the risk of an acute myocardial events and cardiovascular mortality (Lopez-Garcia et al. 2008; Rosner et al. 2007; Zhang et al. 2009). In contrast, case-control studies did show an association between caffeine intake and nonfatal myocardial infarction, with an estimated population attributable risk of 12.8% (95% confidence interval 5.9–25.7%) (Kabagambe et al. 2007; Klatsky et al. 2008).

In conclusion, prospective studies have not shown a consistent positive association between coffee intake and coronary heart disease, whereas retrospective studies in general do report such an association. This apparent discrepancy has often been explained in terms of the retrospective studies being more likely to suffer from bias and confounding, in particular recall bias (Kawachi et al. 1994; Sofi et al. 2007). However, this discrepancy could also be interpreted in favor of an acute rather than a chronic adverse effect of coffee on coronary heart disease: obviously, retrospective studies might provide a more accurate assessment of coffee intake in the period immediately before the coronary event, in contrast to the prospective studies, in which coffee consumption is generally assessed years before the event. Therefore, the discrepancy between the results of the case-control and prospective

studies might also be compatible with the hypothesis that coffee has an acute adverse effect (i.e., triggering a coronary event) rather than a long-term adverse effect (i.e., promoting the development of atherosclerosis). The recent observation, in a cohort study of older subjects, that there is an inverse correlation, particularly in women, between coffee consumption and coronary calcifications, which are a marker for atherosclerosis in coronary arteries, also suggests that chronic coffee consumption does not adversely affect the development of atherosclerosis (van Woudenberg et al. 2008). In contrast, experimental studies have consistently shown that coffee or caffeine acutely raises blood pressure, circulating levels of (nor)epinephrine, and arterial stiffness, and impairs endothelium-dependent vasodilation and ischemic and pharmacological preconditioning. It has generally been appreciated that acute coronary events can be triggered by physical and emotional stressors, which cause similar acute physiological hemodynamic and neurohumoral changes as coffee drinking. As such, coffee consumption could act as a trigger for coronary events. The recent finding, in a retrospective study, that there was an increased risk for acute myocardial infarction in the first hour after coffee consumption favors this hypothesis (Baylin et al. 2006).

Another factor that has to be taken into account when studying the association between coffee drinking and coronary heart disease is that several studies have suggested that there is a marked interindividual variation in the susceptibility to the adverse effects of coffee. In experimental studies, only half of all subjects show a complete tolerance for the acute hemodynamic and neurohumoral effects of coffee. Therefore, in the setting of chronic daily coffee consumption, some subjects could be more susceptible to the acute effects of coffee consumption than others. Interestingly, in a case-control study, coffee consumption was only associated with an increased risk of acute myocardial infarction in patients with the cytochrome P450 1A2 genotype, which predicts slow hepatic metabolism of caffeine, suggesting that caffeine plays a role in this association (Cornelis et al. 2006). Previous studies have suggested also that the effect of coffee on the plasma cholesterol and homocysteine concentrations shows interindividual variation, which is genetically determined (Strandhagen et al. 2004; Weggemans et al. 2001a, b). Finally, it has been examined whether the relation between coffee intake and coronary heart disease is dependent on the metabolism of circulating catecholamines. In a prospective study, the risk of an acute myocardial infarction in heavy coffee consumers with a low activity of catechol *O*-methyltransferase was higher than in participants with a high catechol *O*-methyltransferase activity (odds ratio 3.2, 90% confidence interval 1.2–8.4) (Happonen et al. 2006), suggesting that the acute effect of coffee on circulating catecholamines might be involved in the adverse cardiovascular effect of coffee.

In conclusion, a wealth of evidence demonstrate that chronic coffee consumption does not increase the risk of coronary heart disease in the general population. These findings do not exclude, however, that the acute hemodynamic and neurohumoral effects of coffee consumption could have an adverse effect in selected patient groups who are more vulnerable for these effects, based on their genetic profile or medication use (Riksen et al. 2009).

## References

- Agwunobi J, Abedin M, Young M, Beeram M, Sinkford S (1996) Impact of theophylline use in Wolff-Parkinson-White syndrome. *J Natl Med Assoc* 88:450–452
- Ammon HP, Bieck PR, Mandalaz D, Verspohl EJ (1983) Adaptation of blood pressure to continuous heavy coffee drinking in young volunteers. A double-blind crossover study. *Br J Clin Pharmacol* 15:701–706
- Andreas S, Reiter H, Luthje L et al (2004) Differential effects of theophylline on sympathetic excitation, hemodynamics, and breathing in congestive heart failure. *Circulation* 110:2157–2162
- Bak AA, Grobbee DE (1990) A randomized study on coffee and blood pressure. *J Hum Hypertens* 4:259–264
- Bardou M, Goirand F, Bernard A et al (2002) Relaxant effects of selective phosphodiesterase inhibitors on U46619 precontracted human intralobar pulmonary arteries and role of potassium channels. *J Cardiovasc Pharmacol* 40:153–161
- Barraco RA, Clough-Helfman C, Goodwin BP, Anderson GF (1995) Evidence for presynaptic adenosine A<sub>2A</sub> receptors associated with norepinephrine release and their desensitization in the rat nucleus tractus solitarius. *J Neurochem* 65:1604–1611
- Baylin A, Hernandez-Diaz S, Kabagambe EK, Siles X, Campos H (2006) Transient exposure to coffee as a trigger of a first nonfatal myocardial infarction. *Epidemiology* 17:506–511
- Belardinelli L, Linden J, Berne RM (1989) The cardiac effects of adenosine. *Prog Cardiovasc Dis* 32:73–97
- Belardinelli L, Shryock JC, Snowdy S et al (1998) The A<sub>2A</sub> adenosine receptor mediates coronary vasodilation. *J Pharmacol Exp Ther* 284:1066–1073
- Belardinelli L, Shryock JC, Song Y, Wang D, Srinivas M (1995) Ionic basis of the electrophysiological actions of adenosine on cardiomyocytes. *FASEB J* 9:359–365
- Biaggioni I, Olafsson B, Robertson RM, Hollister AS, Robertson D (1987) Cardiovascular and respiratory effects of adenosine in conscious man. Evidence for chemoreceptor activation. *Circ Res* 61:779–786
- Biaggioni I, Onrot J, Hollister AS, Robertson D (1986) Cardiovascular effects of adenosine infusion in man and their modulation by dipyridamole. *Life Sci* 39:2229–2236
- Biaggioni I, Paul S, Puckett A, Arzubiaga C (1991) Caffeine and theophylline as adenosine receptor antagonists in humans. *J Pharmacol Exp Ther* 258:588–593
- Bijlstra P, van Ginneken EE, Huls M, van Dijk R, Smits P, Rongen GA (2004) Glyburide inhibits dipyridamole-induced forearm vasodilation but not adenosine-induced forearm vasodilation. *Clin Pharmacol Ther* 75:147–156
- Brodmann M, Lischnig U, Lueger A, Pilger E, Stark G (2003) The effect of caffeine on peripheral vascular resistance in isolated perfused guinea pig hind limbs. *J Cardiovasc Pharmacol* 42:506–510
- Brown NJ, Ryder D, Nadeau J (1993) Caffeine attenuates the renal vascular response to angiotensin II infusion. *Hypertension* 22:847–852
- Burr ML, Gallacher JE, Butland BK, Bolton CH, Downs LG (1989) Coffee, blood pressure and plasma lipids: a randomized controlled trial. *Eur J Clin Nutr* 43:477–483
- Butcher RW, Sutherland EW (1962) Adenosine 3',5'-phosphate in biological materials. I. Purification and properties of cyclic 3',5'-nucleotide phosphodiesterase and use of this enzyme to characterize adenosine 3',5'-phosphate in human urine. *J Biol Chem* 237:1244–1250
- Cannon ME, Cooke CT, McCarthy JS (2001) Caffeine-induced cardiac arrhythmia: an unrecognized danger of healthfood products. *Med J Aust* 174:520–521
- Carr CS, Hill RJ, Masamune H et al (1997) Evidence for a role for both the adenosine A<sub>1</sub> and A<sub>3</sub> receptors in protection of isolated human atrial muscle against simulated ischaemia. *Cardiovasc Res* 36:52–59
- Casiglia E, Bongiovi S, Paleari CD et al (1991) Haemodynamic effects of coffee and caffeine in normal volunteers: a placebo-controlled clinical study. *J Intern Med* 229:501–504



- Cawley MJ, Al-Jazairi AS, Stone EA (2001) Intravenous theophylline—an alternative to temporary pacing in the management of bradycardia secondary to AV nodal block. *Ann Pharmacother* 35:303–307
- Christensen B, Mosdol A, Retterstol L, Landaas S, Thelle DS (2001) Abstention from filtered coffee reduces the concentrations of plasma homocysteine and serum cholesterol: a randomized controlled trial. *Am J Clin Nutr* 74:302–307
- Conlay LA, Conant JA, deBros F, Wurtman R (1997) Caffeine alters plasma adenosine levels. *Nature* 389:136
- Cornelis MC, El-Sohemy A, Kabagambe EK, Campos H (2006) Coffee, CYP1A2 genotype, and risk of myocardial infarction. *JAMA* 295:1135–1141
- Costa F, Biaggioni I (1994) Role of adenosine in the sympathetic activation produced by isometric exercise in humans. *J Clin Invest* 93:1654–1660
- Cox DA, Vita JA, Treasure CB, Fish RD, Selwyn AP, Ganz P (1989) Reflex increase in blood pressure during the intracoronary administration of adenosine in man. *J Clin Invest* 84:592–596
- Daniels JW, Mole PA, Shaffrath JD, Stebbins CL (1998) Effects of caffeine on blood pressure, heart rate, and forearm blood flow during dynamic leg exercise. *J Appl Physiol* 85:154–159
- DeLago A, El-Hajjar M, Kirnus M (2008) Aminophylline for prevention of bradyarrhythmias induced by rheolytic thrombectomy. *J Invasive Cardiol* 20:9A–11A
- Dimarco AF, Nochomovitz M, DiMarco MS, Altose MD, Kelsen SG (1985) Comparative effects of aminophylline on diaphragm and cardiac contractility. *Am Rev Respir Dis* 132:800–805
- Donoso MV, Aedo F, Huidobro-Toro JP (2006) The role of adenosine A<sub>2A</sub> and A<sub>3</sub> receptors on the differential modulation of norepinephrine and neuropeptide Y release from peripheral sympathetic nerve terminals. *J Neurochem* 96:1680–1695
- Edlund A, Conradsson T, Sollevi A (1995) A role for adenosine in coronary vasoregulation in man. Effects of theophylline and enprofylline. *Clin Physiol* 15:623–636
- Edlund A, Sollevi A (1995) Theophylline increases coronary vascular tone in humans: evidence for a role of endogenous adenosine in flow regulation. *Acta Physiol Scand* 155:303–311
- Eppel GA, Ventura S, Evans RG (2006) Regional vascular responses to ATP and ATP analogues in the rabbit kidney *in vivo*: roles for adenosine receptors and prostanoids. *Br J Pharmacol* 149:523–531
- Farag NH, Vincent AS, McKey BS, Whitsett TL, Lovallo WR (2005) Hemodynamic mechanisms underlying the incomplete tolerance to caffeine's pressor effects. *Am J Cardiol* 95:1389–1392
- Feoktistov I, Biaggioni I (1995) Adenosine A<sub>2B</sub> receptors evoke interleukin-8 secretion in human mast cells. An enprofylline-sensitive mechanism with implications for asthma. *J Clin Invest* 96:1979–1986
- Francis SH, Sekhar KR, Ke H, Corbin JD (2010) Inhibition of cyclic nucleotide phosphodiesterases by methylxanthines and related compounds. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Fredholm BB (1980) Are methylxanthine effects due to antagonism of endogenous adenosine? *Trends Pharmacol Sci* 1:129–132
- Fredholm BB (1984) Cardiovascular and renal actions of methylxanthines. *Prog Clin Biol Res* 158:303–330
- Fredholm BB, Battig K, Holmen J, Nehlig A, Zvartau EE (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fredholm BB, Hedqvist P, Vernet L (1978) Effect of theophylline and other drugs on rabbit renal cyclic nucleotide phosphodiesterase, 5'-nucleotidase and adenosine deaminase. *Biochem Pharmacol* 27:2845–2850
- Fredholm BB, IJzerman AP, Jacobson KA, Klotz KN, Linden J (2001a) International union of pharmacology. XXV. Nomenclature and classification of adenosine receptors. *Pharmacol Rev* 53:527–552

- Fredholm BB, Irenius E, Kull B, Schulte G (2001b) Comparison of the potency of adenosine as an agonist at human adenosine receptors expressed in Chinese hamster ovary cells. *Biochem Pharmacol* 61:443–448
- Fredholm BB, Persson CG (1982) Xanthine derivatives as adenosine receptor antagonists. *Eur J Pharmacol* 81:673–676
- Frost L, Vestergaard P (2005) Caffeine and risk of atrial fibrillation or flutter: the Danish diet, cancer, and health study. *Am J Clin Nutr* 81:578–582
- Greenland S (1993) A meta-analysis of coffee, myocardial infarction, and coronary death. *Epidemiology* 4:366–374
- Grossmann M, Braune J, Ebert U, Kirch W (1998) Dilatory effects of phosphodiesterase inhibitors on human hand veins *in vivo*. *Eur J Clin Pharmacol* 54:35–39
- Grubben MJ, Boers GH, Blom HJ et al (2000) Unfiltered coffee increases plasma homocysteine concentrations in healthy volunteers: a randomized trial. *Am J Clin Nutr* 71:480–484
- Guerreiro S, Marien M, Michel PP (2010) Methylxanthines and ryanodine receptor channels. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Happonen P, Voutilainen S, Tuomainen TP, Salonen JT (2006) Catechol-O-methyltransferase gene polymorphism modifies the effect of coffee intake on incidence of acute coronary events. *PLoS ONE* 1:e117
- Harada K, Ohashi K, Kumagai Y, Fujimura A (1995) Comparison of venodilatory effect of amrinone and theophylline in human subjects. *J Clin Pharmacol* 35:1067–1070
- Hasko G, Linden J, Cronstein B, Pacher P (2008) Adenosine receptors: therapeutic aspects for inflammatory and immune diseases. *Nat Rev Drug Discov* 7:759–770
- Hausenloy DJ, Yellon DM (2007) Preconditioning and postconditioning: united at reperfusion. *Pharmacol Ther* 116:173–191
- Hinschen AK, Rose-Meyer RB, Headrick JP (2003) Adenosine receptor subtypes mediating coronary vasodilation in rat hearts. *J Cardiovasc Pharmacol* 41:73–80
- Hu G, Jousilahti P, Nissinen A, Bidel S, Antikainen R, Tuomilehto J (2007) Coffee consumption and the incidence of antihypertensive drug treatment in Finnish men and women. *Am J Clin Nutr* 86:457–464
- Ishibashi Y, Duncker DJ, Zhang J, Bache RJ (1998) ATP-sensitive K<sup>+</sup> channels, adenosine, and nitric oxide-mediated mechanisms account for coronary vasodilation during exercise. *Circ Res* 82:346–359
- Ishida S, Ito M, Takahashi N, Fujino T, Akimitsu T, Saikawa T (1996) Caffeine induces ventricular tachyarrhythmias possibly due to triggered activity in rabbits *in vivo*. *Jpn Circ J* 60:157–165
- Jansen DF, Nedeljkovic S, Feskens EJ et al (1995) Coffee consumption, alcohol use, and cigarette smoking as determinants of serum total and HDL cholesterol in two Serbian cohorts of the Seven Countries Study. *Arterioscler Thromb Vasc Biol* 15:1793–1797
- Jee SH, He J, Appel LJ, Whelton PK, Suh I, Klag MJ (2001) Coffee consumption and serum lipids: a meta-analysis of randomized controlled clinical trials. *Am J Epidemiol* 153:353–362
- Jee SH, He J, Whelton PK, Suh I, Klag MJ (1999) The effect of chronic coffee drinking on blood pressure: a meta-analysis of controlled clinical trials. *Hypertension* 33:647–652
- Kabagambe EK, Baylin A, Campos H (2007) Nonfatal acute myocardial infarction in Costa Rica: modifiable risk factors, population-attributable risks, and adherence to dietary guidelines. *Circulation* 115:1075–1081
- Kamphuis J, Smits P, Thien T (1994) Vascular effects of pentoxifylline in humans. *J Cardiovasc Pharmacol* 24:648–654
- Karatzis E, Papaioannou TG, Aznaouridis K et al (2005) Acute effects of caffeine on blood pressure and wave reflections in healthy subjects: should we consider monitoring central blood pressure? *Int J Cardiol* 98:425–430
- Kawachi I, Colditz GA, Stone CB (1994) Does coffee drinking increase the risk of coronary heart disease? Results from a meta-analysis. *Br Heart J* 72:269–275

- Kemp BK, Cocks TM (1999) Adenosine mediates relaxation of human small resistance-like coronary arteries via A<sub>2B</sub> receptors. *Br J Pharmacol* 126:1796–1800
- Klag MJ, Wang NY, Meoni LA et al (2002) Coffee intake and risk of hypertension: the Johns Hopkins precursors study. *Arch Intern Med* 162:657–662
- Klatsky AL, Koplik S, Kipp H, Friedman GD (2008) The confounded relation of coffee drinking to coronary artery disease. *Am J Cardiol* 101:825–827
- Klotz KN, Hessling J, Hegler J et al (1998) Comparative pharmacology of human adenosine receptor subtypes – characterization of stably transfected receptors in CHO cells. *Naunyn Schmiedebergs Arch Pharmacol* 357:1–9
- Kong H, Jones PP, Koop A, Zhang L, Duff HJ, Chen SR (2008) Caffeine induces Ca<sup>2+</sup> release by reducing the threshold for luminal Ca<sup>2+</sup> activation of the ryanodine receptor. *Biochem J* 414:441–452
- Lai EY, Patzak A, Steege A et al (2006) Contribution of adenosine receptors in the control of arteriolar tone and adenosine-angiotensin II interaction. *Kidney Int* 70:690–698
- Lieu HD, Shryock JC, von Mering GO et al (2007) Regadenoson, a selective A<sub>2A</sub> adenosine receptor agonist, causes dose-dependent increases in coronary blood flow velocity in humans. *J Nucl Cardiol* 14:514–520
- Lindahl B, Johansson I, Huhtasaari F, Hallmans G, Asplund K (1991) Coffee drinking and blood cholesterol: effects of brewing method, food intake and life style. *J Intern Med* 230:299–305
- Little JA, Shanoff HM, Csima A, Toronto MA, Yano R (1966) Coffee and serum-lipids in coronary heart-disease. *Lancet* 1:732–734
- Lo YC, Tsou HH, Lin RJ et al (2005) Endothelium-dependent and -independent vasorelaxation by a theophylline derivative MCPT: roles of cyclic nucleotides, potassium channel opening and phosphodiesterase inhibition. *Life Sci* 76:931–944
- Lopez-Garcia E, van Dam RM, Li TY, Rodriguez-Artalejo F, Hu FB (2008) The relationship of coffee consumption with mortality. *Ann Intern Med* 148:904–914
- Lovallo WR, Wilson MF, Vincent AS, Sung BH, McKey BS, Whitsett TL (2004) Blood pressure response to caffeine shows incomplete tolerance after short-term regular consumption. *Hypertension* 43:760–765
- MacDonald TM, Sharpe K, Fowler G et al (1991) Caffeine restriction: effect on mild hypertension. *BMJ* 303:1235–1238
- Mahmud A, Feely J (2001) Acute effect of caffeine on arterial stiffness and aortic pressure waveform. *Hypertension* 38:227–231
- McNamara R, Maginn M, Harkin A (2007) Caffeine induces a profound and persistent tachycardia in response to MDMA (“Ecstasy”) administration. *Eur J Pharmacol* 555:194–198
- McPherson PS, Kim YK, Valdivia H et al (1991) The brain ryanodine receptor: a caffeine-sensitive calcium release channel. *Neuron* 7:17–25
- Mehta MC, Jain AC, Billie M (2004) Effects of cocaine and caffeine alone and in combination on cardiovascular performance: an experimental hemodynamic and coronary flow reserve study in a canine model. *Int J Cardiol* 97:225–232
- Meijer P, Oyen WJ, Dekker D et al (2009) Rosuvastatin increases extracellular adenosine formation in humans *in vivo*: a new perspective on cardiovascular protection. *Arterioscler Thromb Vasc Biol* 29:963–968
- Melchert PJ, Duncker DJ, Traverse JH, Bache RJ (1999) Role of K(+)(ATP) channels and adenosine in regulation of coronary blood flow in the hypertrophied left ventricle. *Am J Physiol* 277:H617–H625
- Müller C, Jacobson KA (2010) Xanthines as adenosine receptor antagonists. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Myers MG, Basinski A (1992) Coffee and coronary heart disease. *Arch Intern Med* 152:1767–1772
- Newton GE, Azevedo ER, Parker JD (1999) Inotropic and sympathetic responses to the intracoronary infusion of a beta2-receptor agonist: a human *in vivo* study. *Circulation* 99:2402–2407

- Noordzij M, Uiterwaal CS, Arends LR, Kok FJ, Grobbee DE, Geleijnse JM (2005) Blood pressure response to chronic intake of coffee and caffeine: a meta-analysis of randomized controlled trials. *J Hypertens* 23:921–928
- Notarius CF, Atchison DJ, Rongen GA, Floras JS (2001) Effect of adenosine receptor blockade with caffeine on sympathetic response to handgrip exercise in heart failure. *Am J Physiol Heart Circ Physiol* 281:H1312–H1318
- Notarius CF, Rongen GA, Floras JS (2003) Caffeine and coffee tolerance. *Circulation* 108:e38–e40
- Nurminen ML, Niittynen L, Korpela R, Vapaatalo H (1999) Coffee, caffeine and blood pressure: a critical review. *Eur J Clin Nutr* 53:831–839
- Nygard O, Nordrehaug JE, Refsum H, Ueland PM, Farstad M, Vollset SE (1997) Plasma homocysteine levels and mortality in patients with coronary artery disease. *N Engl J Med* 337:230–236
- Olthof MR, Hollman PC, Zock PL, Katan MB (2001) Consumption of high doses of chlorogenic acid, present in coffee, or of black tea increases plasma total homocysteine concentrations in humans. *Am J Clin Nutr* 73:532–538
- Panagiotakos DB, Pitsavos C, Zampelas A et al (2004) The association between coffee consumption and plasma total homocysteine levels: the “ATTICA” study. *Heart Vessels* 19:280–286
- Paoloni HJ, Wilcken DE (1975) The action of aminophylline on the acutely transplanted dog heart: effect of alpha- and beta-adrenoceptor blockade. *Br J Pharmacol* 53:163–171
- Papamichael CM, Aznaouridis KA, Karatzis EN et al (2005) Effect of coffee on endothelial function in healthy subjects. The role of caffeine. *Clin Sci (Lond)* 109:55–60
- Patti G, Pasceri V, Colonna G et al (2007) Atorvastatin pretreatment improves outcomes in patients with acute coronary syndromes undergoing early percutaneous coronary intervention: results of the ARMYDA-ACS randomized trial. *J Am Coll Cardiol* 49:1272–1278
- Paynter NP, Yeh HC, Voutilainen S et al (2006) Coffee and sweetened beverage consumption and the risk of type 2 diabetes mellitus: the atherosclerosis risk in communities study. *Am J Epidemiol* 164:1075–1084
- Pereira MA, Parker ED, Folsom AR (2006) Coffee consumption and risk of type 2 diabetes mellitus: an 11-year prospective study of 28 812 postmenopausal women. *Arch Intern Med* 166:1311–1316
- Persson PB (2001) Tubuloglomerular feedback in adenosine A<sub>1</sub> receptor-deficient mice. *Am J Physiol Regul Integr Comp Physiol* 281:R1361
- Phillis JW, Song D, O'Regan MH (1998) The role of adenosine in rat coronary flow regulation during respiratory and metabolic acidosis. *Eur J Pharmacol* 356:199–206
- Pietinen P, Aro A, Tuomilehto J, Uusitalo U, Korhonen H (1990) Consumption of boiled coffee is correlated with serum cholesterol in Finland. *Int J Epidemiol* 19:586–590
- Pincomb GA, Lovallo WR, Passey RB, Whitsett TL, Silverstein SM, Wilson MF (1985) Effects of caffeine on vascular resistance, cardiac output and myocardial contractility in young men. *Am J Cardiol* 56:119–122
- Plagemann PG, Wohlhueter RM (1984) Inhibition of the transport of adenosine, other nucleosides and hypoxanthine in Novikoff rat hepatoma cells by methylxanthines, papaverine, N<sub>6</sub>-cyclohexyladenosine and N<sub>6</sub>-phenylisopropyladenosine. *Biochem Pharmacol* 33:1783–1788
- Rakic V, Burke V, Beilin LJ (1999) Effects of coffee on ambulatory blood pressure in older men and women: a randomized controlled trial. *Hypertension* 33:869–873
- Ramakers BP, Pickkers P, Deussen A et al (2008) Measurement of the endogenous adenosine concentration in humans *in vivo*: methodological considerations. *Curr Drug Metab* 9:679–685
- Rasmussen CA Jr, Sutko JL, Barry WH (1987) Effects of ryanodine and caffeine on contractility, membrane voltage, and calcium exchange in cultured heart cells. *Circ Res* 60:495–504
- Riksen NP, Rongen GA, Smits P (2009) Acute and long-term cardiovascular effects of coffee: implications for coronary heart disease. *Pharmacol Ther* 121:185–191
- Riksen NP, Rongen GA, Yellon D, Smits P (2008) Human *in vivo* research on the vascular effects of adenosine. *Eur J Pharmacol* 585:220–227

- Riksen NP, Zhou Z, Oyen WJ et al (2006) Caffeine prevents protection in two human models of ischemic preconditioning. *J Am Coll Cardiol* 48:700–707
- Robertson D, Frolich JC, Carr RK et al (1978) Effects of caffeine on plasma renin activity, catecholamines and blood pressure. *N Engl J Med* 298:181–186
- Robertson D, Wade D, Workman R, Woosley RL, Oates JA (1981) Tolerance to the humoral and hemodynamic effects of caffeine in man. *J Clin Invest* 67:1111–1117
- Rongen GA, Brooks SC, Ando S, Abramson BL, Floras JS (1998a) Angiotensin AT1 receptor blockade abolishes the reflex sympatho-excitatory response to adenosine. *J Clin Invest* 101:769–776
- Rongen GA, Brooks SC, Ando S, Notarius CF, Floras JS (1998b) Caffeine abstinence augments the systolic blood pressure response to adenosine in humans. *Am J Cardiol* 81:1382–1385
- Rongen GA, Floras JS, Lenders JW, Thien T, Smits P (1997) Cardiovascular pharmacology of purines. *Clin Sci (Lond)* 92:13–24
- Rongen GA, Lenders JW, Lambrou J et al (1996) Presynaptic inhibition of norepinephrine release from sympathetic nerve endings by endogenous adenosine. *Hypertension* 27:933–938
- Rongen GA, Smits P, Verdonck K et al (1995) Hemodynamic and neurohumoral effects of various grades of selective adenosine transport inhibition in humans. Implications for its future role in cardioprotection. *J Clin Invest* 95:658–668
- Rosmarin PC, Applegate WB, Somes GW (1990) Coffee consumption and blood pressure: a randomized, crossover clinical trial. *J Gen Intern Med* 5:211–213
- Rosner SA, Akesson A, Stampfer MJ, Wolk A (2007) Coffee consumption and risk of myocardial infarction among older Swedish women. *Am J Epidemiol* 165:288–293
- Rutherford JD, Vatner SF, Braunwald E (1981) Effects and mechanism of action of aminophylline on cardiac function and regional blood flow distribution in conscious dogs. *Circulation* 63:378–387
- Sanada S, Asanuma H, Minamino T et al (2004) Optimal windows of statin use for immediate infarct limitation: 5'-nucleotidase as another downstream molecule of phosphatidylinositol 3-kinase. *Circulation* 110:2143–2149
- Sattin A, Rall TW (1970) The effect of adenosine and adenine nucleotides on the cyclic adenosine 3',5'-phosphate content of guinea pig cerebral cortex slices. *Mol Pharmacol* 6:13–23
- Schwabe U, Ukena D, Lohse MJ (1985) Xanthine derivatives as antagonists at A<sub>1</sub> and A<sub>2</sub> adenosine receptors. *Naunyn Schmiedebergs Arch Pharmacol* 330:212–221
- Sekiguchi F, Miyake Y, Kashimoto T, Sunano S (2002) Unaltered caffeine-induced relaxation in the aorta of stroke-prone spontaneously hypertensive rats (SHRSP). *J Smooth Muscle Res* 38:11–22
- Smits P, Boekema P, De Abreu R, Thien T, van 't Laar A (1987) Evidence for an antagonism between caffeine and adenosine in the human cardiovascular system. *J Cardiovasc Pharmacol* 10:136–143
- Smits P, Lenders JW, Thien T (1990) Caffeine and theophylline attenuate adenosine-induced vasodilation in humans. *Clin Pharmacol Ther* 48:410–418
- Smits P, Pieters G, Thien T (1986) The role of epinephrine in the circulatory effects of coffee. *Clin Pharmacol Ther* 40:431–437
- Smits P, Schouten J, Thien T (1989) Cardiovascular effects of two xanthines and the relation to adenosine antagonism. *Clin Pharmacol Ther* 45:593–599
- Smits P, Thien T, Van't Laar A (1985a) The cardiovascular effects of regular and decaffeinated coffee. *Br J Clin Pharmacol* 19:852–854
- Smits P, Thien T, Laar A (1985b) Circulatory effects of coffee in relation to the pharmacokinetics of caffeine. *Am J Cardiol* 56:958–963
- Sofi F, Conti AA, Gori AM et al (2007) Coffee consumption and risk of coronary heart disease: a meta-analysis. *Nutr Metab Cardiovasc Dis* 17:209–223
- Song Y, Shryock JC, Knot HJ, Belardinelli L (2001) Selective attenuation by adenosine of arrhythmogenic action of isoproterenol on ventricular myocytes. *Am J Physiol Heart Circ Physiol* 280:H2789–H2795

- Stolzenberg-Solomon RZ, Miller ER III, Maguire MG, Selhub J, Appel LJ (1999) Association of dietary protein intake and coffee consumption with serum homocysteine concentrations in an older population. *Am J Clin Nutr* 69:467–475
- Strandhagen E, Thelle DS (2003) Filtered coffee raises serum cholesterol: results from a controlled study. *Eur J Clin Nutr* 57:1164–1168
- Strandhagen E, Zetterberg H, Aires N et al (2004) The methylenetetrahydrofolate reductase C677T polymorphism is a major determinant of coffee-induced increase of plasma homocysteine: a randomized placebo controlled study. *Int J Mol Med* 13:811–815
- Superko HR, Bortz W Jr, Williams PT, Albers JJ, Wood PD (1991) Caffeinated and decaffeinated coffee effects on plasma lipoprotein cholesterol, apolipoproteins, and lipase activity: a controlled, randomized trial. *Am J Clin Nutr* 54:599–605
- Superko HR, Myll J, DiRicco C, Williams PT, Bortz WM, Wood PD (1994) Effects of cessation of caffeinated-coffee consumption on ambulatory and resting blood pressure in men. *Am J Cardiol* 73:780–784
- Sutton-Tyrrell K, Najjar SS, Boudreau RM et al (2005) Elevated aortic pulse wave velocity, a marker of arterial stiffness, predicts cardiovascular events in well-functioning older adults. *Circulation* 111:3384–3390
- Taddei S, Virdis A, Mattei P, Favilla S, Salvetti A (1991) Adenosine causes angiotensin II release in human forearm arterioles. *J Hypertens Suppl* 9:S232–S233
- Tawfik HE, Schnermann J, Oldenburg PJ, Mustafa SJ (2005) Role of A<sub>1</sub> adenosine receptors in regulation of vascular tone. *Am J Physiol Heart Circ Physiol* 288:H1411–H1416
- Thelle DS, Heyden S, Fodor JG (1987) Coffee and cholesterol in epidemiological and experimental studies. *Atherosclerosis* 67:97–103
- Timmers HJ, Rongen GA, Karemaker JM, Wieling WW, Marres HA, Lenders JW (2004) The role of carotid chemoreceptors in the sympathetic activation by adenosine in humans. *Clin Sci (Lond)* 106:75–82
- Tsuzuki J, Newburgh RW (1975) Inhibition of 5'-nucleotidase in rat brain by methylxanthines. *J Neurochem* 25:895–896
- Uiterwaal CS, Verschuren WM, Bueno-de-Mesquita HB et al (2007) Coffee intake and incidence of hypertension. *Am J Clin Nutr* 85:718–723
- Umemura T, Ueda K, Nishioka K et al (2006) Effects of acute administration of caffeine on vascular function. *Am J Cardiol* 98:1538–1541
- Urgert R, Katan MB (1997) The cholesterol-raising factor from coffee beans. *Annu Rev Nutr* 17:305–324
- Urquhart RA, Broadley KJ (1992a) The effects of P1- and muscarinic-receptor agonists upon cAMP-dependent and independent inotropic responses of guinea-pig cardiac preparations. *Gen Pharmacol* 23:619–626
- Urquhart RA, Broadley KJ (1992b) The indirect negative inotropic effects of the P1-receptor agonist, L-phenylisopropyladenosine, in guinea-pig isolated cardiac preparations: comparison with cromakalim. *Can J Physiol Pharmacol* 70:910–915
- van Dam RM, Feskens EJ (2002) Coffee consumption and risk of type 2 diabetes mellitus. *Lancet* 360:1477–1478
- van Dam RM, Hu FB (2005) Coffee consumption and risk of type 2 diabetes: a systematic review. *JAMA* 294:97–104
- van Dam RM, Willett WC, Manson JE, Hu FB (2006) Coffee, caffeine, and risk of type 2 diabetes: a prospective cohort study in younger and middle-aged U.S. women. *Diab Care* 29:398–403
- van Woudenberg GJ, Vliegenthart R, van Rooij FJA et al (2008) Coffee consumption and coronary calcification: the Rotterdam coronary calcification study. *Arterioscler Thromb Vasc Biol* 28:1018–1023
- van Dusseldorp M, Smits P, Lenders JW, Thien T, Katan MB (1991) Boiled coffee and blood pressure. A 14-week controlled trial. *Hypertension* 18:607–613
- Varani K, Portaluppi F, Gessi S et al (2000) Dose and time effects of caffeine intake on human platelet adenosine A<sub>2A</sub> receptors: functional and biochemical aspects. *Circulation* 102:285–289

- Varani K, Portaluppi F, Merighi S, Ongini E, Belardinelli L, Borea PA (1999) Caffeine alters A<sub>2A</sub> adenosine receptors and their function in human platelets. *Circulation* 99:2499–2502
- Verhoef P, Pasman WJ, Van VT, Urgert R, Katan MB (2002) Contribution of caffeine to the homocysteine-raising effect of coffee: a randomized controlled trial in humans. *Am J Clin Nutr* 76:1244–1248
- Vlachopoulos C, Hirata K, O'Rourke MF (2003) Effect of caffeine on aortic elastic properties and wave reflection. *J Hypertens* 21:563–570
- Weggemans RM, Zock PL, Ordovas JM, Pedro-Botet J, Katan MB (2001a) Apoprotein E genotype and the response of serum cholesterol to dietary fat, cholesterol and cafestol. *Atherosclerosis* 154:547–555
- Weggemans RM, Zock PL, Ordovas JM, Ramos-Galluzzi J, Katan MB (2001b) Genetic polymorphisms and lipid response to dietary changes in humans. *Eur J Clin Invest* 31:950–957
- Wierema TK, Houben AJ, Kroon AA et al (2005) Mechanisms of adenosine-induced renal vasodilatation in hypertensive patients. *J Hypertens* 23:1731–1736
- Winkelmayr WC, Stampfer MJ, Willett WC, Curhan GC (2005) Habitual caffeine intake and the risk of hypertension in women. *JAMA* 294:2330–2335
- Woo OF, Pond SM, Benowitz NL, Olson KR (1984) Benefit of hemoperfusion in acute theophylline intoxication. *J Toxicol Clin Toxicol* 22:411–424
- Yang JN, Bjorklund O, Lindstrom-Tornqvist K et al (2009) Mice heterozygous for both A<sub>1</sub> and A<sub>2A</sub> adenosine receptor genes show similarities to mice given long-term caffeine. *J Appl Physiol* 106:631–639
- Ye Y, Abu Said G, Lin Y et al (2008) Caffeinated coffee blunts the myocardial protective effects of statins against ischemia-reperfusion injury in the rat. *Cardiovasc Drugs Ther* 22:275–282
- Yellon DM, Downey JM (2003) Preconditioning the myocardium: from cellular physiology to clinical cardiology. *Physiol Rev* 83:1113–1151
- Zhang W, Lopez-Garcia E, Li TY, Hu FB, van Dam RM (2009) Coffee consumption and risk of cardiovascular diseases and all-cause mortality among men with type 2 diabetes. *Diab Care* 32:1043–1045

# Methylxanthines in Asthma

Stephen L. Tilley

## Contents

1	Introduction .....	440
2	Historical Background .....	440
3	Actions of Methylxanthines in the Lung .....	442
4	Molecular Mechanisms of Action .....	444
4.1	ASM and Bronchodilation .....	444
4.2	Molecular Mechanisms in Immune Cells .....	447
5	Clinical Use of Methylxanthines for the Treatment of Asthma .....	449
5.1	Acute Severe and Near-Fatal Asthma .....	449
5.2	Chronic Asthma .....	450
6	Methylxanthines and Respiration .....	450
7	Conclusions .....	451
	References .....	451

**Abstract** Methylxanthines represent a unique class of drugs for the treatment of asthma. The methylxanthine theophylline has demonstrated efficacy in attenuating the three cardinal features of asthma – reversible airflow obstruction, airway hyper-responsiveness, and airway inflammation. At doses achieving relatively high serum levels in which toxic side effects are sometimes observed, direct bronchodilatory effects of theophylline are recognized. At lower serum concentrations, theophylline is a weak bronchodilator but retains its capacity as an immunomodulator, anti-inflammatory, and bronchoprotective drug. Intense investigation into the molecular mechanisms of action of theophylline has identified several different points of action. Phosphodiesterase inhibition and adenosine receptor antagonism have both been implicated in promoting airway smooth muscle relaxation and bronchodilation.

---

S.L. Tilley

Department of Medicine, Division of Pulmonary and Critical Care Medicine, and Center for Environmental Medicine, Asthma, and Lung Biology, University of North Carolina, Chapel Hill, NC 27599, USA

e-mail: stephen\_tilley@med.unc.edu



Similar mechanisms of action may explain the inhibitory effects of theophylline on immune cells. At lower concentrations that fail to inhibit phosphodiesterase, effects on histone deacetylase activity are believed to contribute to the immunomodulatory actions of theophylline. Since anti-inflammatory and immunomodulatory effects of methylxanthines are realized at lower serum concentrations than are required for bronchodilation, theophylline's predominant role in asthma treatment is as a controller medication for chronic, persistent disease.

**Keywords** Adenosine receptors · Airway smooth muscle · Asthma · Mast cell · Methylxanthines · Phosphodiesterase · Theophylline

## Abbreviations

ASM	Airway smooth muscle
cAMP	Cyclic AMP
cGMP	Cyclic GMP
HDAC	Histone deacetylase
ICS	Inhaled corticosteroids
PDE	Phosphodiesterase
PPAR $\gamma$	Peroxisome-proliferator-activated receptor $\gamma$
RyR	Ryanodine receptor

## 1 Introduction

The methylxanthine theophylline is one of the most widely prescribed medications for the treatment of asthma worldwide. Guidelines for the treatment of asthma, however, recommend that theophylline be used as a second- and third-line agent owing to its potential for toxicity, and the availability of effective alternatives, namely, inhaled corticosteroids (ICS) and  $\beta$ -agonists. For patients who do not achieve optimal asthma control on ICS and  $\beta$ -agonists, or those without access to these medications, theophylline represents an important component of our therapeutic armamentarium.

## 2 Historical Background

As early as the eighteenth century, physicians recognized that strong coffee was effective at improving symptoms in asthmatics. In 1860, medical textbooks recommended coffee for the treatment of dyspnea in bronchial asthma (Salter 1860). In the

early 1900s, the bronchodilating effects of 1,3,7-trimethylxanthine (caffeine) and 3,7-dimethylxanthine (theobromine) were demonstrated experimentally in bovine bronchial smooth muscle and guinea-pig lung (Trendelenburg 1912; Baehr and Pick 1913; Pal 1912). Shortly thereafter, pharmacologists at Johns Hopkins University set out to study the antispasmodic actions of a number of popular remedies for asthma, including methylxanthines, using bronchial smooth muscle strips from pigs. Macht and Ting (1921) published their findings in 1921, noting that 1,3-dimethylxanthine (theophylline) was a much more effective bronchodilator than caffeine. The first clinical description of theophylline use in asthmatics was reported by Hirsch (1922) from Germany in 1922, when he described four patients who responded well to the rectal administration of “Spasmopurin,” a mixture of 66.7% theophylline and 33.3% theobromine. Hirsch also tested his theophylline/theobromine combination on bovine bronchial smooth muscle strips, noting smooth muscle relaxation, and thus concluded that dimethylxanthines act by producing relaxation of bronchial smooth muscle (Schultze-Werninghaus and Meier-Sydow 1982). Despite his recommendation in his publication that theophylline be considered for clinical use both acutely and prophylactically in asthma, methylxanthines did not receive further attention for this indication until 1936, when numerous antidotal reports emerged touting the efficacy of theophylline in patients with asthma. In 1937, two concurrent but independent clinical trials reported, as did Hirsch in 1922, that methylxanthines were efficacious in asthma. Interestingly, both investigations were initially focused on the use of aminophylline, the more soluble ethylenediamine salt of theophylline, as a diuretic for the relief of dyspnea in heart failure, only incidentally noting its efficacy in asthmatics. Herrmann et al. (1937) reported the “prompt, complete, persistent relief” of extreme dyspnea in 14 of 16 patients with status asthmaticus, while Greene et al. (1937) reported the relief of asthma in 11 allergic asthmatics treated with aminophylline, and showed improvement in pulmonary function following its administration. After the “rediscovery” of theophylline for bronchial asthma by Herrmann et al. and Greene et al. in the 1930s, numerous additional reports emerged on the subject, and the Food and Drug Administration approved the use of theophylline for asthma in the USA in 1940.

Among the available drugs for the treatment of asthma at the time (methylxanthines, anticholinergics, adrenergic agonists), the methylxanthines were effective bronchodilators that acted quickly and had a reasonable duration of action. As concerns about the safety of systemic use of adrenergic agonists rose, methylxanthines became the cornerstone of asthma therapy worldwide. Clinical trials clearly demonstrated that theophylline could reduce the symptoms of chronic asthma, improve lung function, improve exercise tolerance, reduce the need for rescue medication, and facilitate the withdrawal of oral corticosteroids (reviewed in McFadden 1985; Weinberger and Hendeles 1996)). Slow-release formulations of theophylline were developed, allowing the dosing interval to be increased from 6 to 12 h, and eventually to 24 h, thus improving patient compliance. The golden age of methylxanthines had been reached. However, owing to the development of more effective therapies, and rising concerns about potential toxicities of methylxanthines, the use of theophylline in industrialized nations began to fall.

Declines in theophylline use began with the development of selective  $\beta_2$ -adrenergic agonists, and studies demonstrating the inferiority of methylxanthines to these new agents in acute asthma exacerbations. In the 1980s, several studies from emergency departments reported that aerosolized albuterol, a selective  $\beta_2$ -agonist, consistently outperformed aminophylline and theophylline (Rossing et al. 1980; Fanta et al. 1986; Siegel et al. 1985), resulting in dramatic reductions in the use of methylxanthines for acute exacerbations of asthma.

With an increased appreciation that airway inflammation plays a critical role in the pathogenesis of asthma, potent ICS have been developed and have largely replaced theophylline as the mainstay of treatment for chronic persistent asthma in developed countries. In addition, continued investigation into the pathogenesis of asthma has resulted in the development of therapies targeting the leukotriene pathway (e.g., monteleukast, zileuton) and IgE binding to mast cells (omaluzimab). As a result, the use of theophylline as a controller therapy for chronic asthma has declined in parts of the world with access to these anti-inflammatory and immunomodulatory drugs.

### 3 Actions of Methylxanthines in the Lung

The primary effect of methylxanthines in the human asthmatic lung was for many years assumed to be their capacity to relax airway smooth muscle (ASM). More recent observations, however, suggest that additional mechanisms may be responsible for the therapeutic effects of this class of drugs. These mechanisms include inhibition of immune cell activation and proliferation, effects on mucociliary transport, and reduction in proinflammatory gene expression via the induction of histone deacetylase (HDAC) activity.

The bronchodilating effects of methylxanthines on ASM from animals was first reported in 1912, and shortly thereafter the efficacy of theophylline to relax ASM in bronchial strips from pigs was recognized (Trendelenburg 1912; Baehr and Pick 1913; Pal 1912). In the early 1980s, independent laboratories in France and Sweden demonstrated that theophylline relaxed ASM in both large and small airways from ex vivo human lung samples obtained from patients undergoing lung surgery (Guillot et al. 1984; Finney et al. 1985), suggesting a mechanism for the beneficial clinical effects of theophylline observed in asthmatics. Interestingly, these studies showed that approximately 25% of the specimens failed to relax with  $\beta$ -agonists, but demonstrated universal relaxation with theophylline. Theophylline is a relatively weak bronchodilator, with an  $EC_{50}$  for relaxation of human ASM in vitro ranging from 14 to 67  $\mu\text{g}/\text{mL}$  (Guillot et al. 1984; Cortijo et al. 1993). Pharmacodynamic studies in humans have demonstrated dose-dependent bronchodilation with serum theophylline concentrations between 5 and 20  $\mu\text{g}/\text{mL}$ , with maximal effects requiring the maintenance of serum levels at the upper end of this range (Mitenko and Ogilvie 1973a, b). Observations that lower serum concentrations of theophylline were effective at preventing exercise-induced bronchoconstriction and

airway inflammation suggested that ASM-independent effects may be responsible for its efficacy. These observations were particularly important since the serum levels required for direct bronchodilation are difficult to achieve owing to dose-limiting toxicities of nausea, vomiting, and tachycardia.

Theophylline can inhibit bronchoconstriction induced by several stimuli, including exercise, methacholine, histamine, and antigens (Magnussen et al. 1987, 1988; McWilliams et al. 1984; Pauwels et al. 1985). For many of these stimuli, the bronchoprotective effects of theophylline are realized at low serum levels (4–10  $\mu\text{g/mL}$ ), and do not correlate with the degree of acute bronchodilation. These observations suggest that the bronchodilation and bronchoprotection afforded by theophylline may occur by independent mechanisms.

Mast cells are central to the pathogenesis of asthma, and bronchoconstriction resulting from a variety of stimuli occurs indirectly as a result of mast cell activation. The clinical observations of bronchoprotection by theophylline to challenge by numerous provoking stimuli suggest that theophylline may be acting on mast cells to produce its beneficial clinical effects. Indeed, studies with both rodent and human mast cells have shown that theophylline can dose-dependently inhibit mediator release by cells activated by antigens and other stimuli (Sydbom and Fredholm 1982; Pearce et al. 1982; Louis and Radermecker 1990; Weston et al. 1997).

Theophylline has demonstrated inhibitory effects on many other cell types important to asthma pathogenesis, including eosinophils, neutrophils, lymphocytes, and macrophages, as well as in animal models of asthma. Theophylline can inhibit eosinophil chemotaxis to a number of stimuli (Numao et al. 1991), reduce leukotriene  $\text{C}_4$  release (Tenor et al. 1996), and reduce superoxide production by eosinophils (Yasui et al. 2000a). Theophylline induces apoptosis of activated eosinophils from asthmatics (Takeuchi et al. 1999). Similar to observations in eosinophils, theophylline can promote apoptosis and reduce chemotaxis and superoxide production by neutrophils (Yasui et al. 2000b). T-lymphocyte proliferation and activation can be suppressed by theophylline at concentrations that are used therapeutically in patients with asthma (Scordamaglia et al. 1988; Singer et al. 1992; Rosenthal et al. 1992). Theophylline can also suppress mitogen-induced secretion of proinflammatory cytokines from human peripheral blood lymphocytes (Scordamaglia et al. 1988; Prieur and Granger 1975). Finally, theophylline displays a number of inhibitory effects on macrophage function, including chemotaxis and migration into tissues and production of reactive oxygen species, arachidonic acid products, and proinflammatory cytokines (Stephens and Snyderman 1982; Godfrey et al. 1987; Calhoun et al. 1991; Bailly et al. 1990; Prabhakar et al. 1993). Very low concentrations of theophylline (1.8  $\mu\text{g/mL}$ ) can suppress endotoxin-induced  $\text{TNF-}\alpha$  production by monocytes *in vivo* (Spatafora et al. 1994).

Methylxanthines have been shown to reduce allergic inflammation in many species, including rat, guinea pig, and rabbit (Pauwels 1987; Manzini et al. 1993; Ali et al. 1992). While the inhibitory actions of theophylline on each type of immune cell *in vitro* and in animal models strongly suggested that methylxanthines might reduce inflammation in asthmatics, the doses used in most of these investigations exceeded clinically achievable doses in humans.

A number of clinical studies have established that theophylline has clinically relevant anti-inflammatory properties in asthmatics in doses that produce little to no toxicity. In a double-blind, placebo-controlled study of chronic treatment of mild allergic asthmatics for 6 weeks using a theophylline dose that produced plasma levels of 6.6  $\mu\text{g/mL}$ , biopsies following antigen challenge showed significantly reduced numbers of EG2+ eosinophils and CD4+ lymphocytes in the bronchial wall (Sullivan et al. 1994). In a similarly designed study, theophylline treatment resulted in reduced IL-4 expression and a decrease in the number of epithelial CD8+ cells in bronchial biopsies (Finnerty et al. 1996). Improved asthma control was observed in the theophylline-treated group. In patients with nocturnal asthma, low-dose theophylline has been shown to inhibit the early morning influx of neutrophils and eosinophils into the lung (Kraft et al. 1996). Finally, low-dose theophylline can reduce the number of CD4+ and CD8+ T cells in bronchoalveolar lavage fluid after allergen challenge (Jaffar et al. 1996). These reductions in lymphocyte numbers were observed in patients already using high-dose ICS, suggesting that the molecular mechanisms responsible for the anti-inflammatory effects of theophylline are different from those of corticosteroids.

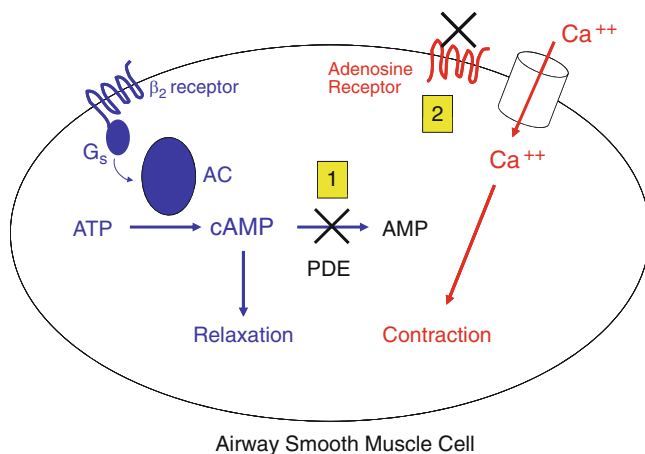
## 4 Molecular Mechanisms of Action

Several mechanisms have been proposed for the observed bronchodilatory and immunomodulatory effects of theophylline in the asthmatic airway. Phosphodiesterase (PDE) inhibition and adenosine receptor antagonism are the most established. Effects on endogenous catecholamine release, calcium ion flux, modulation of HDAC activity, and induction of peroxisome-proliferator-activated receptor  $\gamma$  (PPAR $\gamma$ ) expression have also been reported.

### 4.1 ASM and Bronchodilation

#### 4.1.1 PDE Inhibition

Elevation of intracellular cyclic AMP (cAMP) concentration, and to a lesser extent cyclic GMP (cGMP) concentration, is a major mechanism promoting ASM relaxation and resultant bronchodilation. Theophylline is a nonselective inhibitor of PDE, the enzymes which break down cyclic nucleotides. As a result, theophylline treatment can increase intracellular cAMP and cGMP concentrations in ASM cells and by this mechanism induce bronchodilation (Fig. 1). Dose-dependent ASM relaxation and concomitant cAMP elevation has been demonstrated in ex vivo airways from both animals and humans (Rabe et al. 1995; Fredholm et al. 1979). Theophylline is a weak PDE inhibitor, and the degree of PDE inhibition is low at therapeutically achievable serum levels. For example, total PDE activity in



**Fig. 1** Effects of methylxanthines on airway smooth muscle function. (1) Phosphodiesterase inhibition by methylxanthines inhibits cyclic AMP (cAMP) degradation and promotes airway smooth muscle (ASM) relaxation by raising intracellular cAMP levels. (2) Adenosine receptor antagonism. Activation of adenosine receptors increases intracellular calcium levels. In some species, adenosine activates adenosine receptors on ASM; in other species, the effect is indirect via activation of adenosine receptors on mast cells and neurons, resulting in paracrine signaling to ASM by additional mediators, promoting ASM contraction (leukotrienes, histamine). By antagonizing adenosine receptors, methylxanthines block calcium-activated ASM contraction

human lung extracts is inhibited by only 5–10% at therapeutic concentrations of theophylline (Polson et al. 1978). However, some PDE isoenzymes appear to be more sensitive, with selected isoforms demonstrating 50% inhibition by theophylline concentrations in the high-therapeutic range (18  $\mu\text{g}/\text{mL}$ ) (Bergstrand and Lundquist 1978). These data are consistent with clinical observations of increased bronchodilation if plasma theophylline levels are pushed to greater than 15  $\mu\text{g}/\text{mL}$ . While the effects of theophylline on airway tone at lower plasma levels may be due in part to some degree of PDE inhibition, it is believed that its actions on other cell types may indirectly influence airway caliber.

#### 4.1.2 Adenosine Receptor Antagonism

In addition to their actions as PDE inhibitors, methylxanthines also act as adenosine receptor antagonists (Fredholm et al. 1979; Fredholm and Persson 1982). Since exogenous adenosine produces bronchoconstriction in asthmatics, it has been postulated that theophylline's bronchodilatory actions may result from antagonism of adenosine receptors (Cushley et al. 1983a, b). The specific adenosine receptors involved in adenosine-induced bronchoconstriction have been extensively investigated. In rabbits, activation of  $A_1$  receptors elicits ASM contraction *ex vivo* and bronchoconstriction *in vivo* (Ali et al. 1992, 1994a, b; Abebe and Mustafa 1998). In mice, adenosine-induced bronchoconstriction occurs indirectly through activation

of A<sub>1</sub> receptors on neurons and A<sub>3</sub> receptors on mast cells (Hua et al. 2007; Tilley et al. 2003). ASM contraction occurs as a result of acute increases in intracellular calcium levels. Thus, methylxanthines can inhibit these actions in ASM directly through antagonism of adenosine receptors on ASM (Fig. 1), and indirectly by decreasing the levels of paracrine mediators released by adjacent or embedded mast cells through inhibition of mast cell activation by adenosine, as described in more detail below.

The pathways mediating adenosine-induced bronchoconstriction in human asthmatics have not been fully elucidated. The mechanism appears to be indirect through mast cell activation, since the response can be largely attenuated by mast-cell-stabilizing drugs and antihistamines (Phillips et al. 1987, 1989a, b). In contrast to rodents, *in vitro* studies with the malignant human mast cell line HMC-1 have implicated the A<sub>2B</sub> receptor in adenosine-induced mast cell activation (Feoktistov and Biaggioni 1995; Feoktistov et al. 2001). Clinical studies have shown enprofylline to have superior efficacy to theophylline in the treatment of asthma (Persson et al. 1986). While initially felt to be devoid of antagonist activity at adenosine receptors, enprofylline is now believed to be a reasonably selective A<sub>2B</sub> receptor antagonist (Fredholm and Persson 1982; Feoktistov and Biaggioni 1995; Auchampach et al. 1997). Collectively, these data have suggested the A<sub>2B</sub> adenosine receptor as an attractive therapeutic target in asthma (Feoktistov et al. 1998).

### 4.1.3 Endogenous Catecholamine Release

Animal studies have demonstrated that methylxanthines provoke the release of epinephrine from the adrenal gland (Peach 1972; Poisner 1973; Berkowitz and Spector 1971). In normal humans 3 h following the administration of aminophylline (5 mg/kg), epinephrine levels increased twofold (Higbee et al. 1982). Similar effects of aminophylline were found in asthmatic children in a study which examined the relationship between pulmonary function and plasma catecholamine levels (Ishizaki et al. 1988). During a constant 72-h infusion of aminophylline, peak expiratory flow progressively increased during the first 48 h. In contrast, epinephrine levels rose rapidly but returned to normal levels within 24 h. These results suggest that methylxanthine-induced epinephrine release may contribute to the immediate bronchodilation observed by raising the cAMP concentration in ASM, via stimulation of  $\beta_2$  adrenergic receptors but that the sustained and continued bronchodilatory effects of methylxanthines occur by an alternative mechanism.

### 4.1.4 Ryanodine Receptors and Calcium

A fourth mechanism by which methylxanthines may influence airway caliber is through their effects on intracellular calcium in ASM cells. Intracellular calcium regulates both ASM contraction and relaxation. While acute, step increases in intracellular calcium levels elicit ASM contraction via phosphorylation of myosin

light-chain kinase, slower increases in intracellular calcium levels can dephosphorylate myosin light-chain kinase and induce relaxation. In 1977, it was suggested that calcium ion uptake into storage sites in smooth muscle cells could be influenced by theophylline (Kolbeck et al. 1979). Caffeine is now a well-recognized ryanodine receptor (RyR) agonist inducing calcium release from internal stores (Herrmann-Frank et al. 1999; Dettbarn et al. 1994; Cheek et al. 1993). Caffeine, aminophylline, and theophylline have all been demonstrated to potentiate luminal calcium activation of RyRs, reduce the threshold for spontaneous calcium release, and increase basal activity of RyRs in cardiac myocytes (Kong et al. 2008). While it has been postulated that these effects of methylxanthines on calcium homeostasis may be responsible for their arrhythmogenic potential, this same mechanism, if present in ASM, might explain the beneficial effects of these drugs on bronchomotor tone.

## 4.2 *Molecular Mechanisms in Immune Cells*

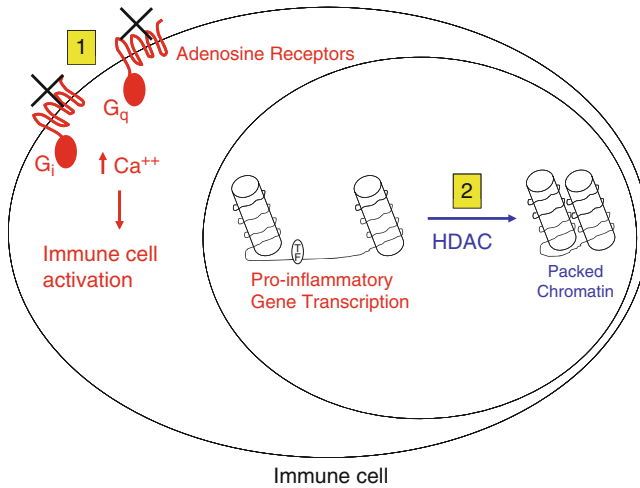
While PDE inhibition and cAMP level elevation results in inhibition of immune cell proliferation and activation, a number of lines of evidence suggest that the immunomodulatory effects of theophylline occur predominantly by alternative mechanisms. Unlike ASM, where relaxation *in vitro* is not observed following exposure to low concentrations of theophylline, inhibitory effects on many immune cells have been demonstrated at very low theophylline levels. Several mechanisms have been proposed for the immunomodulatory properties of low-dose theophylline.

### 4.2.1 Adenosine Receptor Antagonism

Adenosine can elicit both pro- and anti-inflammatory effects in immune cells as a result of signaling through G-protein-coupled receptors with different intracellular signaling pathways. A<sub>1</sub> and A<sub>3</sub> receptors couple to G<sub>i</sub>, and their activation results in rises in intracellular calcium levels, which typically activates immune cells. As depicted in Fig. 2, antagonism of these adenosine receptors will inhibit immune cell activation. In contrast, A<sub>2A</sub> and A<sub>2B</sub> receptors couple to G<sub>s</sub>, and their activation results in rises in intracellular cAMP levels, resulting in inhibition of immune cell function. In the HMC-1 mast cell line, A<sub>2B</sub> receptors couple to G<sub>s</sub> as well as G<sub>q</sub> (Ryzhov et al. 2006). The complexity is further increased by the differential expression of each adenosine receptor by specific immune cells, changes in receptor expression during inflammation, differing ligand affinities of each receptor, and changing adenosine concentrations depending upon the intensity and duration of the asthmatic attack. In asthma, attention has been largely directed to the proinflammatory effects of adenosine.

In the asthmatic lung, adenosine acutely degranulates airway mast cells. Adenosine receptor antagonism by theophylline can block adenosine-induced mast cell





**Fig. 2** Anti-inflammatory and immunomodulatory actions of methylxanthines in immune cells. (1) Methylxanthines antagonize cell-surface adenosine receptors, blocking the proinflammatory actions of adenosine on immune cells. (2) Methylxanthines increase histone deacetylase activity, promoting chromatin packing, preventing proinflammatory gene transcription

activation; thus, inhibition of mast cell function by blocking adenosine receptor–G<sub>q</sub>–calcium signaling is one plausible mechanism of action of theophylline (Fig. 2). However, many immune cells, including mast cells, express G<sub>s</sub>-coupled A<sub>2A</sub> and A<sub>2B</sub> adenosine receptors, and their activation results in the accumulation of intracellular cAMP, a potent inhibitor of immune cell activation and migration. Antagonism of these G<sub>s</sub>-coupled receptors by theophylline would be expected to activate rather than inhibit the function of immune cells, including T-cells and eosinophils, suggesting that additional mechanisms may be responsible for the chronic, immunosuppressive effects of methylxanthines.

#### 4.2.2 Modulation of HDAC Activity and Induction of PPAR $\gamma$

The acetylation of core histones by coactivator proteins results in DNA “unwinding,” facilitating RNA polymerase binding and transcription of proinflammatory cytokines. Corepressor proteins that have HDAC activity suppress inflammatory gene transcription by promoting the repacking of chromatin (Fig. 2). In bronchial biopsies and alveolar macrophages from asthmatics, HDAC activity is reduced relative to that in normal subjects (Ito et al. 2002a; Cosio et al. 2004). Theophylline has been shown to directly increase HDAC enzymatic activity in airway epithelia and macrophages (Ito et al. 2002b). Neither PDE inhibition nor adenosine receptor antagonism mimicked the effects of theophylline on HDAC activity, supporting the conclusion that direct interactions of theophylline with HDAC proteins is a novel mechanism of action. While the exact means by which theophylline activates

HDAC is not certain, data from this study suggest that it may occur by allosteric action and/or phosphorylation. Since corticosteroids induce HDAC gene transcription, resulting in increased HDAC protein levels, theophylline and steroids may act synergistically to suppress proinflammatory mediator production by both resident structural cells and infiltrating immune cells in the asthmatic lung. Since low doses of theophylline can influence HDAC activity independent of PDE inhibition and adenosine receptor antagonism, clinically beneficial effects may be realized without the major recognized side effects of theophylline (nausea, vomiting, seizures, cardiac arrhythmias) which are believed to result from antagonism of these PDE enzymes and adenosine receptors. The human studies described below support the concept that these biochemical observations regarding HDAC activity may be clinically relevant.

PPAR $\gamma$  is a nuclear receptor regulating immune cell function. Synthetic agonists for PPAR $\gamma$  have been shown to attenuate factor-induced eosinophil survival and chemotaxis. In one study using human peripheral blood eosinophils, theophylline markedly enhanced both messenger RNA and protein levels of PPAR $\gamma$  in eosinophils (Usami et al. 2006). It is likely that several of the mechanisms described above act together to mediate the anti-inflammatory actions of methylxanthines.

## 5 Clinical Use of Methylxanthines for the Treatment of Asthma

### 5.1 *Acute Severe and Near-Fatal Asthma*

Methylxanthines are not generally recommended for patients presenting to the emergency department with acute exacerbations of asthma. Inhaled  $\beta$ -agonists in combination with early systemic steroids and anticholinergics are highly effective, and studies looking at the addition of intravenously administered aminophylline to this regimen have yielded mixed results (Fanta et al. 1986; Siegel et al. 1985; Wrenn et al. 1991; Ream et al. 2001; D'Avila et al. 2008). In a meta-analysis of 13 clinical trials comparing nebulized  $\beta$ -agonists with or without intravenous administration of aminophylline, no additional benefit from adding aminophylline was observed (Littenberg 1988). In addition to a potential lack of added efficacy, theophylline has a very narrow therapeutic/toxic index, and several studies have reported increased side effects when it is added to  $\beta$ -agonists therapy (Fanta et al. 1986; Siegel et al. 1985). In a study from England, out of 43 asthma deaths analyzed, there was a significantly greater percentage of toxic theophylline concentrations (21%) in patients who died compared with matched controls (7%) (Eason and Markowe 1989). Taken together, these studies suggest methylxanthines should not be used routinely for acute exacerbations of asthma. They may have a role, however, in patients presenting with status asthmaticus who are refractory to standard care or those with impending respiratory failure (Levy et al. 1998; Self et al. 2002; Aubier et al. 1981).

## 5.2 *Chronic Asthma*

Owing to the superior efficacy of ICS and long-acting  $\beta$ -agonists, methylxanthines are no longer recommended as first-line therapy for the treatment of chronic persistent asthma. However, numerous clinical trials with theophylline have demonstrated efficacy, and asthma guidelines continue to endorse the use of this class of drugs as an add-on therapy in noncontrolled asthmatics, and as alternatives to ICS and long-acting  $\beta$ -agonists (e.g., if access to these agents is restricted owing to financial or social barriers) (Bateman et al. 2008). Several trials have shown the efficacy of low-dose theophylline as an add-on therapy to ICS in chronic persistent asthma. Evans et al. (1997) reported that adding theophylline to low-dose ICS was as effective as doubling the ICS dose. Consistent with this study, subsequent trials have reported similar improvements in lung function and asthma control in subjects treated with low-dose ICS and theophylline compared with those taking high-dose ICS alone (Ukena et al. 1997; Lim et al. 2000; Spears et al. 2009).

## 6 **Methylxanthines and Respiration**

Methylxanthines are respiratory stimulants. They have been used clinically to treat infants with apnea of prematurity as well as adults with central sleep apnea and periodic breathing–Cheyne–Stokes respiration (Kelly and Shannon 1981; Espinoza et al. 1987; Dowdell et al. 1990). Apnea of prematurity occurs in 85% of infants born earlier than 34 weeks of gestation (Barrington and Finer 1991). Aminophylline, theophylline, and caffeine reduce the frequency of apnea and the need for mechanical ventilation, and are the mainstay of treatment for this common disorder (Henderson-Smart and Steer 2001).

The effect of theophylline on central apnea, periodic breathing, and obstructive sleep apnea has been examined in several clinical trials. A blinded, placebo-controlled study showed that theophylline administered intravenously overnight improved central, but not obstructive, apneas (Espinoza et al. 1987). In a small study of men with heart failure and periodic breathing with central apneas, orally administered theophylline decreased central apneas from a mean value of 26 per hour to six per hour (Javaheri et al. 1996). In contrast, studies in patients with obstructive sleep apnea have failed to show a significant improvement in apnea by methylxanthines (Guilleminault and Hayes 1983; Mulloy and McNicholas 1992).

The mechanism by which methylxanthines stimulate respiration has been extensively investigated, and several potential mechanisms have been proposed. These include augmentation of hypoxic and hypercapnic ventilatory responses, inhibition of the ventilatory depressant effect of adenosine, increasing metabolic rate, and improving respiratory muscle performance (Javaheri and Guerra 1990; Lakshminarayan et al. 1978; Murciano et al. 1987, 1984; Fredholm 1984; Eldridge et al. 1983). Relevant to sleep-disordered breathing associated with heart failure,

theophylline exerts positive inotropic actions on the heart which may indirectly improve periodic breathing by decreasing circulation time.

## 7 Conclusions

Since their discovery, methylxanthines have cycled in and out of favor for the treatment of asthma in developed nations. Worldwide they continue to represent one of the most commonly used therapies for bronchial asthma. Recognition that methylxanthines have immunomodulatory effects at low serum concentrations, and that they may act synergistically with corticosteroids, suggests that they can be exploited as immunomodulators rather than bronchodilators – without systemic toxicity. Intense investigation into the cellular and molecular mechanisms by which caffeine and theophylline modulate airway inflammation in asthma has helped identify a number of novel biological pathways that may serve as future drug targets for asthma and other inflammatory diseases. Until such molecular scalpels are developed and tested, the continued use of theophylline will be important for relieving dyspnea and improving the quality of life for patients afflicted with this most common immune-mediated pulmonary disease.

## References

- Abebe W, Mustafa SJ (1998) A1 adenosine receptor-mediated Ins(1,4,5)P<sub>3</sub> generation in allergic rabbit airway smooth muscle. *Am J Physiol* 275:L990
- Ali S, Mustafa SJ, Metzger WJ (1992) Adenosine-induced bronchoconstriction in an allergic rabbit model: antagonism by theophylline aerosol. *Agents Actions* 37:165
- Ali S, Mustafa SJ, Metzger WJ (1994a) Adenosine receptor-mediated bronchoconstriction and bronchial hyperresponsiveness in allergic rabbit model. *Am J Physiol* 266:L271
- Ali S, Mustafa SJ, Metzger WJ (1994b) Adenosine-induced bronchoconstriction and contraction of airway smooth muscle from allergic rabbits with late-phase airway obstruction: evidence for an inducible adenosine A1 receptor. *J Pharmacol Exp Ther* 268:1328
- Aubier M, De Troyer A, Sampson M, Macklem PT, Roussos C (1981) Aminophylline improves diaphragmatic contractility. *N Engl J Med* 305:249
- Auchampach JA, Jin X, Wan TC, Caughey GH, Linden J (1997) Canine mast cell adenosine receptors: cloning and expression of the A<sub>3</sub> receptor and evidence that degranulation is mediated by the A<sub>2B</sub> receptor. *Mol Pharmacol* 52:846
- Baehr G, Pick EP (1913) *Pharmakologische Studien an der Bronchialmuskulatur der uberlebenden Meerschweinchenlunge*. *Arch Exp Pathol Pharmacol* 74:40
- Bailly S, Ferrua B, Fay M, Gougerot-Pocidalo MA (1990) Differential regulation of IL 6, IL 1 A, IL 1 beta and TNF alpha production in LPS-stimulated human monocytes: role of cyclic AMP. *Cytokine* 2:205
- Barrington K, Finer N (1991) The natural history of the appearance of apnea of prematurity. *Pediatr Res* 29:372
- Bateman ED, Hurd SS, Barnes PJ, Bousquet J, Drazen JM, FitzGerald M, Gibson P, Ohta K, O'Byrne P, Pedersen SE, Pizzichini E, Sullivan SD, Wenzel SE, Zar HJ (2008) Global strategy for asthma management and prevention: GINA executive summary. *Eur Respir J* 31:143

- Bergstrand H, Lundquist B (1978) Partial purification and characterization of cyclic nucleotide phosphodiesterases from human bronchial tissue. *Mol Cell Biochem* 21:9
- Berkowitz BA, Spector S (1971) Effect of caffeine and theophylline on peripheral catecholamines. *Eur J Pharmacol* 13:193
- Calhoun WJ, Stevens CA, Lambert SB (1991) Modulation of superoxide production of alveolar macrophages and peripheral blood mononuclear cells by beta-agonists and theophylline. *J Lab Clin Med* 117:514
- Cheek TR, Moreton RB, Berridge MJ, Stauderman KA, Murawsky MM, Bootman MD (1993) Quantal  $\text{Ca}^{2+}$  release from caffeine-sensitive stores in adrenal chromaffin cells. *J Biol Chem* 268:27076
- Cortijo J, Bou J, Beleta J, Cardelus I, Llenas J, Morcillo E, Gristwood RW (1993) Investigation into the role of phosphodiesterase IV in bronchorelaxation, including studies with human bronchus. *Br J Pharmacol* 108:562
- Cosio BG, Mann B, Ito K, Jazrawi E, Barnes PJ, Chung KF, Adcock IM (2004) Histone acetylase and deacetylase activity in alveolar macrophages and blood monocytes in asthma. *Am J Respir Crit Care Med* 170:141
- Cushley MJ, Tattersfield AE, Holgate ST (1983a) Inhaled adenosine and guanosine on airway resistance in normal and asthmatic subjects. *Br J Clin Pharmacol* 15:161
- Cushley MJ, Tattersfield AE, Holgate ST (1983b) Adenosine antagonism as an alternative mechanism of action of methylxanthines in asthma. *Agents Actions Suppl* 13:109
- D'Avila RS, Piva JP, Marostica PJ, Amantea SL (2008) Early administration of two intravenous bolus of aminophylline added to the standard treatment of children with acute asthma. *Respir Med* 102:156
- Dettbarn C, Gyorke S, Palade P (1994) Many agonists induce "quantal"  $\text{Ca}^{2+}$  release or adaptive behavior in muscle ryanodine receptors. *Mol Pharmacol* 46:502
- Dowdell WT, Javaheri S, McGinnis W (1990) Cheyne-Stokes respiration presenting as sleep apnea syndrome. Clinical and polysomnographic features. *Am Rev Respir Dis* 141:871
- Eason J, Markowe HL (1989) Aminophylline toxicity—how many hospital asthma deaths does it cause? *Respir Med* 83:219
- Eldridge FL, Millhorn DE, Waldrop TG, Kiley JP (1983) Mechanism of respiratory effects of methylxanthines. *Respir Physiol* 53:239
- Espinoza H, Antic R, Thornton AT, McEvoy RD (1987) The effects of aminophylline on sleep and sleep-disordered breathing in patients with obstructive sleep apnea syndrome. *Am Rev Respir Dis* 136:80
- Evans DJ, Taylor DA, Zetterstrom O, Chung KF, O'Connor BJ, Barnes PJ (1997) A comparison of low-dose inhaled budesonide plus theophylline and high-dose inhaled budesonide for moderate asthma. *N Engl J Med* 337:1412
- Fanta CH, Rossing TH, McFadden ER Jr (1986) Treatment of acute asthma. Is combination therapy with sympathomimetics and methylxanthines indicated? *Am J Med* 80:5
- Feoktistov I, Biaggioni I (1995) Adenosine A2b receptors evoke interleukin-8 secretion in human mast cells. An enprofylline-sensitive mechanism with implications for asthma. *J Clin Invest* 96:1979
- Feoktistov I, Polosa R, Holgate ST, Biaggioni I (1998) Adenosine A2B receptors: a novel therapeutic target in asthma? *Trends Pharmacol Sci* 19:148
- Feoktistov I, Garland EM, Goldstein AE, Zeng D, Belardinelli L, Wells JN, Biaggioni I (2001) Inhibition of human mast cell activation with the novel selective adenosine A(2B) receptor antagonist 3-isobutyl-8-pyrrolidinioxanthine (IPDX)(2). *Biochem Pharmacol* 62:1163
- Finnerty JP, Lee C, Wilson S, Madden J, Djukanovic R, Holgate ST (1996) Effects of theophylline on inflammatory cells and cytokines in asthmatic subjects: a placebo-controlled parallel group study. *Eur Respir J* 9:1672
- Finney MJ, Karlsson JA, Persson CG (1985) Effects of bronchoconstrictors and bronchodilators on a novel human small airway preparation. *Br J Pharmacol* 85:29

- Fredholm BB (1984) Effects of methylxanthines on skeletal muscle and on respiration. *Prog Clin Biol Res* 158:365
- Fredholm BB, Persson CG (1982) Xanthine derivatives as adenosine receptor antagonists. *Eur J Pharmacol* 81:673
- Fredholm BB, Brodin K, Strandberg K (1979) On the mechanism of relaxation of tracheal muscle by theophylline and other cyclic nucleotide phosphodiesterase inhibitors. *Acta Pharmacol Toxicol (Copenh)* 45:336
- Godfrey RW, Manzi RM, Gennaro DE, Hoffstein ST (1987) Phospholipid and arachidonic acid metabolism in zymosan-stimulated human monocytes: modulation by cAMP. *J Cell Physiol* 131:384
- Greene JA, Paul WD, Faller AE (1937) The action of theophylline with ethylenediamine on intrathecal and venous pressures in cardiac failure and on bronchial obstruction in cardiac failure and in bronchial asthma. *J Am Med Assoc* 109:1712
- Guilleminault C, Hayes B (1983) Naloxone, theophylline, bromocriptine, and obstructive sleep apnea. Negative results. *Bull Eur Physiopathol Respir* 19:632
- Guillot C, Fornaris M, Badier M, Orehek J (1984) Spontaneous and provoked resistance to isoproterenol in isolated human bronchi. *J Allergy Clin Immunol* 74:713
- Henderson-Smart DJ, Steer P (2001) Methylxanthine treatment for apnea in preterm infants. *Cochrane Database Syst Rev* CD000140
- Herrmann G, Aynesworth MB, Martin J (1937) Successful treatment of persistent extreme dyspnea status asthmaticus: use of theophylline ethylene diamine (aminophylline, USP) intravenously. *J Lab Clin Med* 23:135
- Herrmann-Frank A, Luttgau HC, Stephenson DG (1999) Caffeine and excitation-contraction coupling in skeletal muscle: a stimulating story. *J Muscle Res Cell Motil* 20:223
- Higbee MD, Kumar M, Galant SP (1982) Stimulation of endogenous catecholamine release by theophylline: a proposed additional mechanism of action for theophylline effects. *J Allergy Clin Immunol* 70:377
- Hirsch S (1922) Klinischer und experimenteller Beitrag zur krampflösenden Wirkung der Purinderivate. *Klin Wochschr* 1:615
- Hua X, Erikson CJ, Chason KD, Rosebrock CN, Deshpande DA, Penn RB, Tilley SL (2007) Involvement of A1 adenosine receptors and neural pathways in adenosine-induced bronchoconstriction in mice. *Am J Physiol Lung Cell Mol Physiol* 293:L25
- Ishizaki T, Minegishi A, Morishita M, Odajima Y, Kanagawa S, Nagai T, Yamaguchi M (1988) Plasma catecholamine concentrations during a 72-hour aminophylline infusion in children with acute asthma. *J Allergy Clin Immunol* 82:146
- Ito K, Caramori G, Lim S, Oates T, Chung KF, Barnes PJ, Adcock IM (2002a) Expression and activity of histone deacetylases in human asthmatic airways. *Am J Respir Crit Care Med* 166:392
- Ito K, Lim S, Caramori G, Cosio B, Chung KF, Adcock IM, Barnes PJ (2002b) A molecular mechanism of action of theophylline: induction of histone deacetylase activity to decrease inflammatory gene expression. *Proc Natl Acad Sci USA* 99:8921
- Jaffar ZH, Sullivan P, Page C, Costello J (1996) Low-dose theophylline modulates T-lymphocyte activation in allergen-challenged asthmatics. *Eur Respir J* 9:456
- Javaheri S, Guerra L (1990) Lung function, hypoxic and hypercapnic ventilatory responses, and respiratory muscle strength in normal subjects taking oral theophylline. *Thorax* 45:743
- Javaheri S, Parker TJ, Wexler L, Liming JD, Lindower P, Roselle GA (1996) Effect of theophylline on sleep-disordered breathing in heart failure. *N Engl J Med* 335:562
- Kelly DH, Shannon DC (1981) Treatment of apnea and excessive periodic breathing in the full-term infant. *Pediatrics* 68:183
- Kolbeck RC, Speir WA Jr, Carrier GO, Bransome ED Jr (1979) Apparent irrelevance of cyclic nucleotides to the relaxation of tracheal smooth muscle induced by theophylline. *Lung* 156:173
- Kong H, Jones PP, Koop A, Zhang L, Duff HJ, Chen SR (2008) Caffeine induces  $Ca^{2+}$  release by reducing the threshold for luminal  $Ca^{2+}$  activation of the ryanodine receptor. *Biochem J* 414:441

- Kraft M, Torvik JA, Trudeau JB, Wenzel SE, Martin RJ (1996) Theophylline: potential anti-inflammatory effects in nocturnal asthma. *J Allergy Clin Immunol* 97:1242
- Lakshminarayan S, Sahn SA, Weil JV (1978) Effect of aminophylline on ventilatory responses in normal man. *Am Rev Respir Dis* 117:33
- Levy BD, Kitch B, Fanta CH (1998) Medical and ventilatory management of status asthmaticus. *Intensive Care Med* 24:105
- Lim S, Jatakanon A, Gordon D, Macdonald C, Chung KF, Barnes PJ (2000) Comparison of high dose inhaled steroids, low dose inhaled steroids plus low dose theophylline, and low dose inhaled steroids alone in chronic asthma in general practice. *Thorax* 55:837
- Littenberg B (1988) Aminophylline treatment in severe, acute asthma. A meta-analysis. *JAMA* 259:1678
- Louis RE, Radermecker MF (1990) Substance P-induced histamine release from human basophils, skin and lung fragments: effect of nedocromil sodium and theophylline. *Int Arch Allergy Appl Immunol* 92:329
- Macht DI, Ting G-C (1921) A study of antispasmodic drugs on the bronchus. *J Pharmacol Exp Ther* 18:373
- Magnussen H, Reuss G, Jorres R (1987) Theophylline has a dose-related effect on the airway response to inhaled histamine and methacholine in asthmatics. *Am Rev Respir Dis* 136:1163
- Magnussen H, Reuss G, Jorres R (1988) Methylxanthines inhibit exercise-induced bronchoconstriction at low serum theophylline concentration and in a dose-dependent fashion. *J Allergy Clin Immunol* 81:531
- Manzini S, Perretti F, Abelli L, Evangelista S, Seeds EA, Page CP (1993) Isbufylline, a new xanthine derivative, inhibits airway hyperresponsiveness and airway inflammation in guinea pigs. *Eur J Pharmacol* 249:251
- McFadden ER Jr (1985) Methylxanthine therapy and reversible airway obstruction. *Am J Med* 79:1
- McWilliams BC, Menendez R, Kelly HW, Howick J (1984) Effects of theophylline on inhaled methacholine and histamine in asthmatic children. *Am Rev Respir Dis* 130:193
- Mitenko PA, Ogilvie RI (1973a) Rational intravenous doses of theophylline. *N Engl J Med* 289:600
- Mitenko PA, Ogilvie RI (1973b) Pharmacokinetics of intravenous theophylline. *Clin Pharmacol Ther* 14:509
- Mulloy E, McNicholas WT (1992) Theophylline in obstructive sleep apnea. A double-blind evaluation. *Chest* 101:753
- Murciano D, Aubier M, Lecocguic Y, Pariente R (1984) Effects of theophylline on diaphragmatic strength and fatigue in patients with chronic obstructive pulmonary disease. *N Engl J Med* 311:349
- Murciano D, Aubier M, Viires N, Mal H, Pariente R (1987) Effects of theophylline and enprofylline on diaphragmatic contractility. *J Appl Physiol* 63:51
- Numao T, Fukuda T, Akutsu I, Makino S (1991) Effects of anti-asthmatic drugs on human eosinophil chemotaxis. *Nihon Kyobu Shikkan Gakkai Zasshi* 29:65
- Pal J (1912) Über toxische Reaktionen der Koronararterien und Bronchien. *Dtsch Med Wochenschr* 38:5
- Pauwels R (1987) The effects of theophylline on airway inflammation. *Chest* 92:32S
- Pauwels R, Van Renterghem D, Van der Straeten M, Johannesson N, Persson CG (1985) The effect of theophylline and enprofylline on allergen-induced bronchoconstriction. *J Allergy Clin Immunol* 76:583
- Peach MJ (1972) Stimulation of release of adrenal catecholamine by adenosine 3':5'-cyclic monophosphate and theophylline in the absence of extracellular  $Ca^{2+}$ . *Proc Natl Acad Sci USA* 69:834
- Pearce FL, Befus AD, Gaudie J, Bienenstock J (1982) Mucosal mast cells. II. Effects of anti-allergic compounds on histamine secretion by isolated intestinal mast cells. *J Immunol* 128:2481

- Persson CG, Andersson KE, Kjellin G (1986) Effects of enprofylline and theophylline may show the role of adenosine. *Life Sci* 38:1057
- Phillips GD, Rafferty P, Beasley R, Holgate ST (1987) Effect of oral terfenadine on the bronchoconstrictor response to inhaled histamine and adenosine 5'-monophosphate in non-atopic asthma. *Thorax* 42:939
- Phillips GD, Polosa R, Holgate ST (1989a) The effect of histamine-H1 receptor antagonism with terfenadine on concentration-related AMP-induced bronchoconstriction in asthma. *Clin Exp Allergy* 19:405
- Phillips GD, Scott VL, Richards R, Holgate ST (1989b) Effect of nedocromil sodium and sodium cromoglycate against bronchoconstriction induced by inhaled adenosine 5'-monophosphate. *Eur Respir J* 2:210
- Poinsner AM (1973) Direct stimulant effect of aminophylline on catecholamine release from the adrenal medulla. *Biochem Pharmacol* 22:469
- Polson JB, Krzanowski JJ, Fitzpatrick DF, Szentivanyi A (1978) Studies on the inhibition of phosphodiesterase-catalyzed cyclic AMP and cyclic GMP breakdown and relaxation of canine tracheal smooth muscle. *Biochem Pharmacol* 27:254
- Prabhakar U, Lipshutz D, Truneh A (1993) Inhibition of CD44, CD45 and LFA-3 mediated cytokine release from human monocytes by SK&F 86002 and pentoxifylline. *Int J Immunopharmacol* 15:205
- Prieur AM, Granger GA (1975) The effect of agents which modulate levels of the cyclic nucleotides on human lymphotoxin secretion and activity in vitro. *Transplantation* 20:331
- Rabe KF, Magnussen H, Dent G (1995) Theophylline and selective PDE inhibitors as bronchodilators and smooth muscle relaxants. *Eur Respir J* 8:637
- Ream RS, Loftis LL, Albers GM, Becker BA, Lynch RE, Mink RB (2001) Efficacy of IV theophylline in children with severe status asthmaticus. *Chest* 119:1480
- Rosenthal LA, Taub DD, Moors MA, Blank KJ (1992) Methylxanthine-induced inhibition of the antigen- and superantigen-specific activation of T and B lymphocytes. *Immunopharmacology* 24:203
- Rossing TH, Fanta CH, Goldstein DH, Snapper JR, McFadden ER Jr (1980) Emergency therapy of asthma: comparison of the acute effects of parenteral and inhaled sympathomimetics and infused aminophylline. *Am Rev Respir Dis* 122:365
- Ryzhov S, Goldstein AE, Biaggioni I, Feoktistov I (2006) Cross-talk between G(s)- and G(q)-coupled pathways in regulation of interleukin-4 by A(2B) adenosine receptors in human mast cells. *Mol Pharmacol* 70:727
- Salter H (1860) *Asthma. Its pathology and treatment.* London
- Schultz-Werninghaus G, Meier-Sydow J (1982) The clinical and pharmacological history of theophylline: first report on the bronchospasmolytic action in man by S. R. Hirsch in Frankfurt (Main) 1922. *Clin Allergy* 12:211
- Scordamaglia A, Ciprandi G, Ruffoni S, Caria M, Paolieri F, Venuti D, Canonica GW (1988) Theophylline and the immune response: in vitro and in vivo effects. *Clin Immunol Immunopathol* 48:238
- Self TH, Redmond AM, Nguyen WT (2002) Reassessment of theophylline use for severe asthma exacerbation: is it justified in critically ill hospitalized patients? *J Asthma* 39:677
- Siegel D, Sheppard D, Gelb A, Weinberg PF (1985) Aminophylline increases the toxicity but not the efficacy of an inhaled beta-adrenergic agonist in the treatment of acute exacerbations of asthma. *Am Rev Respir Dis* 132:283
- Singer JW, Bianco JA, Takahashi G, Simrell C, Petersen J, Andrews DF 3rd (1992) Effect of methylxanthine derivatives on T cell activation. *Bone Marrow Transplant* 10:19
- Spatafora M, Chiappara G, Merendino AM, D'Amico D, Bellia V, Bonsignore G (1994) Theophylline suppresses the release of tumour necrosis factor-alpha by blood monocytes and alveolar macrophages. *Eur Respir J* 7:223
- Spears M, Donnelly I, Jolly L, Brannigan M, Ito K, McSharry C, Lafferty J, Chaudhuri R, Braganza G, Adcock IM, Barnes PJ, Wood S, Thomson NC (2009) Effect of low-dose



- theophylline plus beclometasone on lung function in smokers with asthma: a pilot study. *Eur Respir J* 33:1010
- Stephens CG, Snyderman R (1982) Cyclic nucleotides regulate the morphologic alterations required for chemotaxis in monocytes. *J Immunol* 128:1192
- Sullivan P, Bekir S, Jaffar Z, Page C, Jeffery P, Costello J (1994) Anti-inflammatory effects of low-dose oral theophylline in atopic asthma. *Lancet* 343:1006
- Sydbom A, Fredholm BB (1982) On the mechanism by which theophylline inhibits histamine release from rat mast cells. *Acta Physiol Scand* 114:243
- Takeuchi M, Hayakawa A, Takagi K, Hiramatsu K, Shimizu Y, Matsumoto S, Hiramatsu T, Ito Y, Kume H, Suzuki R, Yamaki K (1999) Theophylline induces apoptosis of the IL-3 activated eosinophils of patients with bronchial asthma. *Apoptosis* 4:461
- Tenor H, Hatzelmann A, Church MK, Schudt C, Shute JK (1996) Effects of theophylline and rolipram on leukotriene C4 (LTC4) synthesis and chemotaxis of human eosinophils from normal and atopic subjects. *Br J Pharmacol* 118:1727
- Tilley SL, Tsai M, Williams CM, Wang ZS, Erikson CJ, Galli SJ, Koller BH (2003) Identification of A3 receptor- and mast cell-dependent and -independent components of adenosine-mediated airway responsiveness in mice. *J Immunol* 171:331
- Trendelenburg P (1912) Physiologische und pharmakologische Untersuchungen an der isolierten Bronchialmuskulatur. *Arch Exp Pathol Pharmacol* 69:79
- Ukena D, Harnest U, Sakalauskas R, Magyar P, Vetter N, Steffen H, Leichtl S, Rathgeb F, Keller A, Steinijans VW (1997) Comparison of addition of theophylline to inhaled steroid with doubling of the dose of inhaled steroid in asthma. *Eur Respir J* 10:2754
- Usami A, Ueki S, Ito W, Kobayashi Y, Chiba T, Mahemuti G, Oyamada H, Kamada Y, Fujita M, Kato H, Saito N, Kayaba H, Chihara J (2006) Theophylline and dexamethasone induce peroxisome proliferator-activated receptor-gamma expression in human eosinophils. *Pharmacology* 77:33
- Weinberger M, Hendeles L (1996) Theophylline in asthma. *N Engl J Med* 334:1380
- Weston MC, Anderson N, Peachell PT (1997) Effects of phosphodiesterase inhibitors on human lung mast cell and basophil function. *Br J Pharmacol* 121:287
- Wrenn K, Slovis CM, Murphy F, Greenberg RS (1991) Aminophylline therapy for acute bronchospastic disease in the emergency room. *Ann Intern Med* 115:241
- Yasui K, Agematsu K, Shinozaki K, Hokibara S, Nagumo H, Yamada S, Kobayashi N, Komiyama A (2000a) Effects of theophylline on human eosinophil functions: comparative study with neutrophil functions. *J Leukoc Biol* 68:194
- Yasui K, Agematsu K, Shinozaki K, Hokibara S, Nagumo H, Nakazawa T, Komiyama A (2000b) Theophylline induces neutrophil apoptosis through adenosine A2A receptor antagonism. *J Leukoc Biol* 67:529

# Methylxanthines and Inflammatory Cells

György Haskó and Bruce Cronstein

## Contents

1	Introduction .....	458
2	Adenosine Receptor Expression in the Immune System .....	458
3	Adenosine Receptor Signaling in the Immune System .....	459
4	Caffeine and Theophylline Are Phosphodiesterase Inhibitors at High Concentrations ...	460
5	Theophylline as a Histone Deacetylase Activator .....	460
6	Effect of Methylxanthines on Inflammatory Cells .....	461
6.1	Caffeine .....	461
6.2	Theophylline .....	461
7	Enprofylline is Anti-inflammatory by Blocking A <sub>2B</sub> Receptors .....	463
8	Effects of Methylxanthines in Inflammatory States .....	463
8.1	Caffeine Exacerbates Acute Inflammatory Liver Injury by Blocking the Physiological Anti-inflammatory Effect of Endogenous Adenosine .....	463
8.2	Caffeine and Theophylline Reverse the Anti-inflammatory Effects of Methotrexate .....	464
9	Conclusion .....	464
	References .....	465

**Abstract** Both caffeine and theophylline have a variety of roles in regulating inflammatory responses. At pharmacologically relevant concentrations most of the effects of these commonly used methylxanthines are attributable to adenosine receptor blockade and histone deacetylase activation. In addition, at higher concentrations methylxanthines can suppress inflammation by inhibiting phosphodiesterases, thereby elevating intracellular cyclic adenosine monophosphate levels. In summary, methylxanthines regulate inflammation by multiple mechanisms.

---

G. Haskó

Department of Surgery, University of Medicine and Dentistry of New Jersey-New Jersey Medical School, 185 South Orange Avenue, Newark, NJ 07103, USA

B. Cronstein (✉)

Department of Medicine, New York University School of Medicine, 550 First Avenue, New York, NY 10016, USA

**Keywords** Asthma · Arthritis · Chronic obstructive pulmonary disease · Cytokine · Macrophage · Methothrexate · Monocyte · Neutrophil · Tumor necrosis factor · Protein kinase A

## 1 Introduction

Caffeine is a nonselective adenosine receptor antagonist, and it is believed that at concentrations achieved during normal human consumption, caffeine exerts its biological effects through antagonism of adenosine receptors (Fredholm et al. 1999). Similar to caffeine, theophylline is a nonselective albeit more potent antagonist of adenosine receptors (Fredholm et al. 2001). Adenosine is an extracellular purine nucleoside signaling molecule, which regulates cell and tissue function both in health and in disease (Hasko et al. 2008). Adenosine is generated following the degradation of its precursor, ATP, a process which can occur both extra- and intracellularly. ATP, a mostly intracellular molecule, is liberated from the cell following stressful and injurious events, and is metabolized to adenosine via a cascade of ectonucleotidases, including CD39 (nucleoside triphosphate diphosphorylase) and CD73 (5'-ectonucleotidase) (Yegutkin 2008). Adenosine that accumulates intracellularly following ATP metabolism is extruded from the cell via nucleoside transporters (Volonte and D'Ambrosi 2009). Cells of the immune/inflammatory system, including neutrophils, mast cells, endothelial cells, lymphocytes, and platelets, have been appreciated as the most prodigious sources of extracellular adenosine (Eltzschig et al. 2008; Hasko et al. 2008). In addition to serving as a source for adenosine release, immune cells are also among the most widely studied cell types targeted by the regulatory influences of adenosine (Deaglio et al. 2007; Erdmann et al. 2005; Feoktistov and Biaggioni 1995; Fozard et al. 1996; Hasko et al. 2007, 2008; Holgate 2005; Sitkovsky 2009).

## 2 Adenosine Receptor Expression in the Immune System

Adenosine produces its biological effects by binding to and activating one or more of four membrane-spanning adenosine receptors, designated  $A_1$ ,  $A_{2A}$ ,  $A_{2B}$ , and  $A_3$ . All four adenosine receptors contain seven transmembrane domains and couple to intracellular GTP binding proteins (G proteins). Adenosine elicits activation of  $A_1$ ,  $A_{2A}$ , and  $A_3$  receptors with  $EC_{50}$  values that range from 0.01 to 1  $\mu\text{M}$ , and  $A_{2B}$  receptor activation occurs at adenosine levels that exceed 10  $\mu\text{M}$  ( $EC_{50}$  24  $\mu\text{M}$ ) (Fredholm et al. 2001). Because physiological adenosine concentrations are less than 1  $\mu\text{M}$ , physiological levels of adenosine can activate only  $A_1$ ,  $A_{2A}$ , and  $A_3$  receptors, and  $A_{2B}$  receptor activation requires pathophysiological conditions (Fredholm 2007). In addition to adenosine concentrations at the cell surface, receptor density and the functionality of the intracellular signaling pathways coupled to adenosine receptors are also key factors in dictating the nature and

magnitude of the effect of adenosine on the cell. For example,  $A_{2A}$  receptor activation inhibits production of the T helper (Th)-1-inducing cytokine interleukin (IL)-12 more potently by human monocytes that are pretreated with the proinflammatory cytokine IL-1 or tumor necrosis factor (TNF)- $\alpha$ , mediators that also increase  $A_{2A}$  receptor expression in these cells (Khoa et al. 2001). In addition, the effect of adenosine can also be affected by the polarized localization of adenosine receptors:  $A_3$  adenosine receptors accrue at the leading edge of migrating neutrophils and are instrumental in directing the movement of cells in response to chemotactic mediators (Chen et al. 2006). Finally, it is important to keep in mind that results regarding adenosine receptor function in one species cannot be readily extrapolated to another one, because sequence differences in cloned adenosine receptors have been shown to be associated with differential pharmacological responses to selective agonists and antagonists. In this regard,  $A_3$  receptors were not even discovered until they were cloned (Zhou et al. 1992), because the  $A_3$  receptor, especially in rodents, is insensitive to caffeine and theophylline, antagonists which had been pivotal in identifying adenosine-receptor-mediated effects.

### 3 Adenosine Receptor Signaling in the Immune System

Adenosine receptors in general dictate cell function through coupling to G proteins, but some G-protein-independent actions have also been reported (Fredholm et al. 2007). Adenosine receptors were initially classified as  $A_1$  [cyclic AMP (cAMP)-decreasing] or  $A_2$  (cAMP-increasing) receptors (van Calker et al. 1979). Subsequently the cAMP-increasing  $A_2$  receptors were divided into two groups: high-affinity  $A_{2A}$  receptors and low-affinity  $A_{2B}$  receptors (Bruns et al. 1986). More recent studies have revealed that in addition to  $A_1$  receptors,  $A_3$  receptors also decrease intracellular cAMP concentrations (Jin et al. 1997). In addition to signaling via the adenylyl cyclase–cAMP system, adenosine receptors can signal through a variety of other pathways.  $A_{2A}$  receptors, as other Gs-protein-coupled receptors, signal chiefly via the adenylate cyclase–cAMP–protein kinase A (PKA) canonical pathway, but they can also activate exchange factor directly activated by cAMP (Epac) (Fredholm et al. 2007). Signaling downstream from PKA occurs through phosphorylation of the transcription factor CREB on serine residue 133, leading to direct CREB-mediated transcriptional activation (Nemeth et al. 2003). Activated CREB can also modulate gene expression indirectly by competing with nuclear factor (NF)- $\kappa$ B or other transcription factors for an important cofactor, CBP (Fredholm et al. 2007). In other cell types, adenosine  $A_{2A}$  receptors stimulate collagen production via mitogen-activated protein (MAP) kinases (Che et al. 2007) and inhibit neutrophil superoxide production through activation of protein phosphatases (Revan et al. 1996). Furthermore, recent results implicated CEBP $\beta$  in the stimulatory effect of  $A_{2A}$  receptor agonists on IL-10 production by *Escherichia coli*-challenged macrophages (Csoka et al. 2007).  $A_{2B}$  receptor stimulation can induce both adenylyl cyclase activation via Gs and phospholipase C activation via

Gq (Feoktistov and Biaggioni 1997). Interaction between these two pathways is important for upregulation of IL-4 production by mast cells upon A<sub>2B</sub> receptor activation (Ryzhov et al. 2006). Specifically, Gq-mediated activation of phospholipase C $\beta$  causes calcium mobilization and an increase in NFATc1-dependent IL-4 transcription, whose response is further facilitated by Gs-mediated NFATc1 protein accumulation. Traditionally, A<sub>3</sub> receptor activation is linked to Gi-mediated inhibition of adenylyl cyclase and Gq-mediated stimulation of phospholipase C (Gessi et al. 2008) and A<sub>3</sub> receptors can activate phospholipase D, RhoA, WNT, MAP kinase and phosphatidylinositol 3-kinase pathways in governing cell function. For example, A<sub>3</sub>-receptor-mediated augmentation of histamine released in sensitized murine mast cells was blocked by inactivating Gi proteins with pertussis toxin and by using pharmacological phosphatidylinositol 3-kinase inhibitors (Zhong et al. 2003). Caffeine and theophylline block A<sub>1</sub>, A<sub>2A</sub>, and A<sub>2B</sub> receptors at pharmacologically relevant concentrations but it appears that these methylxanthines are not potent A<sub>3</sub> receptor antagonists (Fredholm et al. 1999). Although there is controversy regarding what constitutes relevant caffeine concentrations in the immune system (Horrigan et al. 2006), the inhibitory effect of adenosine on formyl-Met-Leu-Phe (fMLP)-stimulated respiratory burst in neutrophil leukocytes was reversed by 30  $\mu$ M caffeine (Fredholm et al. 1996), a concentration that occurs in plasma following consumption of caffeine-containing beverages (Fredholm et al. 1999). In the same study, theophylline proved to be an antagonist of adenosine receptors and it was more potent than caffeine. In addition to its effect on neutrophil respiratory burst, caffeine inhibited the adenosine-receptor mediated accumulation of cAMP in rat thymocytes with a  $K_i$  value of approximately 20  $\mu$ M (Fredholm and Sandberg 1983). Theophylline was slightly more potent and its  $K_i$  value was approximately 10  $\mu$ M.

#### **4 Caffeine and Theophylline Are Phosphodiesterase Inhibitors at High Concentrations**

Higher concentrations of both caffeine and theophylline inhibit cAMP phosphodiesterase (PDE) (Beavo et al. 1971). Inhibition of PDEs by caffeine or theophylline generally requires concentrations of 100–1,000  $\mu$ M, which typically exceeds the concentrations observed in blood following normal oral dosing and can be associated with toxicity (Sawynok and Yaksh 1993). Nevertheless, it is plausible that some of the biological effects of these methylxanthines are due to PDE inhibition.

#### **5 Theophylline as a Histone Deacetylase Activator**

Expression of inflammatory gene expression is governed by a balance between histone acetylation and deacetylation (Barnes 2006). Inflammatory stimuli activate transcription factors such as NF- $\kappa$ B, which leads to histone acetylation and

increased inflammatory gene expression. This process is reversed by recruitment of histone deacetylases to the promoter of inflammatory genes. Recent studies have shown that theophylline at clinically relevant concentrations is an activator of histone deacetylases, which explains its anti-inflammatory effects especially in combination with glucocorticoids (Cosio et al. 2004).

## 6 Effect of Methylxanthines on Inflammatory Cells

### 6.1 Caffeine

On the basis of the fact that caffeine plasma concentrations rarely exceed 50–60  $\mu\text{M}$  during normal human consumption (Fredholm et al. 1999), we will first consider studies that showed caffeine being efficacious at altering inflammatory cell function at these low concentrations. Sullivan et al. (1995) demonstrated that caffeine increased chemiluminescence and myeloperoxidase release by lipopolysaccharide (LPS)-primed, fMLP-stimulated mixed leukocyte cultures, and increases were already evident at 10  $\mu\text{M}$  caffeine. Consistent with the fact that caffeine antagonizes adenosine receptors at this concentration, addition of the adenosine-degrading enzyme adenosine deaminase abolished the effect of caffeine, indicating that caffeine acted by altering an endogenous adenosine response. In addition, the possibility of PDE inhibition as a mechanism for the enhancing effect of caffeine on leukocyte activation was excluded by a lack of effect of caffeine on leukocyte cAMP levels. In a more recent study, 50  $\mu\text{M}$  caffeine decreased TNF- $\alpha$  production by LPS-activated cord blood (neonatal) monocytes by 20% (Chavez-Valdez et al. 2009). In the same study, however, caffeine failed to affect TNF- $\alpha$  production by adult monocytes.

Caffeine at concentrations of 100  $\mu\text{M}$  or higher appears to downregulate inflammatory cell function. Horrigan et al. (2004) found that caffeine suppressed TNF- $\alpha$  release by LPS-stimulated human whole blood at 100  $\mu\text{M}$  by approximately 40% in a cAMP-PKA dependent fashion. In addition, the production of IL-1 $\beta$ , IL-12, and IL-10 was not affected by 100  $\mu\text{M}$  caffeine (Horrigan et al. 2004). In another study, caffeine was shown to decrease colony formation in cultures of murine bone-marrow-derived macrophages, and again, 100  $\mu\text{M}$  was the lowest effective concentration (Inouye and Wharton 1986).

### 6.2 Theophylline

Theophylline has been used as a bronchodilator in the therapy of asthma and chronic obstructive pulmonary disease (COPD) for several decades. Its therapeutic concentration is between 55 and 110  $\mu\text{M}$  and it causes unacceptable side effects above 110  $\mu\text{M}$  (Ito et al. 2002). It has long been suggested that theophylline may

exert some of its beneficial effects in asthma and COPD by downregulating inflammation. One widely held view is that the beneficial effect of theophylline is secondary to its ability to block  $A_{2B}$  receptors (Haskó et al. 2009). This is based, in part, on recent *in vivo* evidence (Mustafa et al. 2007) documenting that selective antagonists of adenosine  $A_{2B}$  receptors inhibit airway inflammation and airway reactivity induced by allergen or AMP in a murine asthma model. The contribution of  $A_{2B}$  receptor blockade to the anti-inflammatory effects of theophylline is also underlined by the observation that at therapeutic concentrations, theophylline is a relatively potent  $A_{2B}$  receptor antagonist. Although  $A_{2B}$  receptors on mast cells seem to be major players in triggering the lung inflammatory response in asthma and COPD (Polosa and Holgate 2006), proinflammatory effects of  $A_{2B}$  receptor stimulation have also been observed with human bronchial smooth-muscle cells (Zhong et al. 2004), human bronchial epithelial cells (Zhong et al. 2006), and human lung fibroblasts (Zhong et al. 2005), which produce increased levels of IL-6 (Zhong et al. 2004, 2005) and IL-19 (Zhong et al. 2006) following  $A_{2B}$  receptor activation.

In addition to  $A_{2B}$  receptors, theophylline blocks  $A_1$  and  $A_{2A}$  receptors at therapeutically relevant concentrations and thus it can reverse the effects of endogenously released adenosine, resulting in both pro- and anti-inflammatory effects depending on which adenosine receptors are expressed. For example, 10  $\mu\text{M}$  theophylline enhanced TNF- $\alpha$  production by human monocytes stimulated with advanced glycation end products, an effect that was postulated to be secondary to blockade of anti-inflammatory adenosine ( $A_{2A}$  or  $A_3$ ) receptors on monocytes (Meiners et al. 2004). Similar to TNF- $\alpha$  production by monocytes, theophylline at therapeutically relevant concentrations (maximal efficacy at 50  $\mu\text{M}$ ) augmented superoxide production by fMLP-activated neutrophils (Yasui et al. 2000b). Because 8-sulfophenyltheophylline, a nonselective adenosine receptor antagonist, reproduced the stimulatory effect of theophylline on superoxide production, it was concluded that theophylline acts by antagonizing adenosine receptors. Finally, 50  $\mu\text{M}$  theophylline was shown to accelerate human granulocyte apoptosis (Yasui et al. 1997, 2000a), an observation consistent with an adenosine-receptor-mediated effect, because adenosine can delay the apoptosis of granulocytes (Walker et al. 1997) and other adenosine receptor antagonists can mimic the effect of theophylline (Yasui et al. 2000a).

In addition to blockade of adenosine receptors, recent studies have revealed a further mechanism that can explain the anti-inflammatory effects of theophylline in asthma. This new evidence is based on the idea that inflammatory gene expression is regulated by a balance between histone acetylation and deacetylation (Barnes 2006). It appears that theophylline is able to tip this balance in favor of histone deacetylation, resulting in decreased inflammatory gene expression (Ito et al. 2002).

Lastly, theophylline has anti-inflammatory effects in various *in vitro* cellular systems when used at therapeutically irrelevant, high concentrations. Theophylline inhibited TNF- $\alpha$  production by LPS-stimulated human mononuclear cells at 200–1,000  $\mu\text{M}$  (Endres et al. 1991); this effect was mediated by inhibition of PDE. In addition, theophylline decreased arachidonate (Hichami et al. 1995),

thromboxane B<sub>2</sub>, (Baker and Fuller 1992), and lysosomal enzyme (Hichami et al. 1995) release by stimulated mononuclear cells at suprathreshold concentrations.

## 7 Enprofylline is Anti-inflammatory by Blocking A<sub>2B</sub> Receptors

On the basis of the efficacy of theophylline as an antiasthma drug (Holgate 2005), efforts to develop a similar xanthine-based compound with a better safety profile led to the development of enprofylline (3-propyl xanthine). It was subsequently demonstrated that similar to theophylline, enprofylline can weakly block A<sub>2B</sub> receptors on mast cells, which might explain its efficacy as an antiasthma agent. The advantage of enprofylline over theophylline is that enprofylline is a selective A<sub>2B</sub> antagonist, whereas theophylline blocks other adenosine receptors as well (Fozard and Hannon 1999).

## 8 Effects of Methylxanthines in Inflammatory States

### 8.1 *Caffeine Exacerbates Acute Inflammatory Liver Injury by Blocking the Physiological Anti-inflammatory Effect of Endogenous Adenosine*

Ohta and Sitkovsky (2001) demonstrated that endogenous adenosine by engaging A<sub>2A</sub> receptors has a nonredundant role in the prevention of inflammatory liver damage induced by concanavalin A. They showed that mice deficient in A<sub>2A</sub> receptors exhibited increased and protracted production of proinflammatory cytokines, including TNF- $\alpha$  and interferon- $\gamma$ , which was paralleled by augmented biochemical and histological signs of liver injury. These results raised the possibility that caffeine acting as an A<sub>2A</sub> receptor antagonist might interfere with the endogenous protective mechanism rendered by adenosine–A<sub>2A</sub> receptor interaction. To address this possibility, Ohta et al. (2007) injected mice with 20 mg/kg caffeine, which corresponds with caffeine amounts ingested during normal human consumption (Fredholm et al. 1999), before inducing liver injury using concanavalin A. The results showed that caffeine exacerbated liver injury as determined by biochemical and histological analysis, and the increased liver injury coincided with increased production of harmful proinflammatory cytokines.

To investigate the role of A<sub>2A</sub>-receptor-mediated signaling in the exacerbation of liver injury following caffeine administration, caffeine (10 and 20 mg/kg) was injected into A<sub>2A</sub> receptor knockout and wild-type mice and liver injury was induced using concanavalin A (Ohta et al. 2007). While caffeine produced an exacerbated liver injury in wild-type mice, knockout mice failed to respond to caffeine with increased acute liver injury, confirming that caffeine enhances liver injury through blocking the action of endogenous adenosine. Importantly, when



caffeine was administered at the supratherapeutic dose of 100 mg/kg, it no longer enhanced inflammation; it actually suppressed it. This anti-inflammatory effect is consistent with caffeine being a PDE inhibitor at high concentrations, because other PDE inhibitors have similar anti-inflammatory effects in general acute inflammation (Hasko et al. 1998; Nemeth et al. 1997).

## ***8.2 Caffeine and Theophylline Reverse the Anti-inflammatory Effects of Methotrexate***

Low-dose orally administered methotrexate is currently the gold standard therapy for the treatment of rheumatoid arthritis. Recent studies have demonstrated that most of the anti-inflammatory effects of methotrexate are mediated by enhanced adenosine release and activation of anti-inflammatory adenosine receptors (Cronstein 2005). Indeed, most of the genetic markers associated with response or lack of response to methotrexate are in enzymes inhibited by methotrexate polyglutamates that are involved in adenosine generation (Dervieux 2009; Dervieux et al. 2004, 2005, 2009; Weisman et al. 2006; Wessels et al. 2006a, b).

Because it is difficult to sample adenosine levels at inflamed sites in patients with rheumatoid arthritis and it is, therefore, difficult to validate the hypothesis that adenosine mediates the anti-inflammatory effects of methotrexate therapy, experiments were carried out with the adjuvant arthritis model of rheumatoid arthritis using adenosine receptor antagonists. In these studies, both theophylline and caffeine reversed the anti-inflammatory effects of methotrexate in rats with adjuvant arthritis (Montesinos et al. 2000) without affecting the course of the arthritis itself. This finding suggested that such commonly used adenosine receptor antagonists as caffeine might reverse the therapeutic effects of methotrexate therapy in patients as well. Indeed, a small prospective study by Neshet et al. (2003) demonstrated that patients started on methotrexate but told to abstain from caffeine responded significantly better than those who did not. Similar observations were made by Silke et al. (2001) of methotrexate in patients as well. In contrast, a larger retrospective study (Benito-Garcia et al. 2006) did not confirm the effect of caffeine consumption on the response to methotrexate; however, since all of the patients in this study had been taking methotrexate for longer than 2 years, it is likely that any patients who did not respond at all had already stopped methotrexate therapy by the time that the survey was done.

## **9 Conclusion**

Both caffeine and theophylline play a variety of roles in regulating inflammatory responses. Although many of the effects of these commonly used methylxanthines are attributable to adenosine receptor blockade, these methylxanthines regulate inflammation by multiple mechanisms.

## References

- Baker AJ, Fuller RW (1992) Effect of cyclic adenosine monophosphate, 5'-(N-ethylcarboxyamido)-adenosine and methylxanthines on the release of thromboxane and lysosomal enzymes from human alveolar macrophages and peripheral blood monocytes in vitro. *Eur J Pharmacol* 211:157–161
- Barnes PJ (2006) Theophylline for COPD. *Thorax* 61:742–744
- Beavo JA, Rogers NL, Crofford OB, Baird CE, Hardman JG, Sutherland EW, Newman EV (1971) Effects of phosphodiesterase inhibitors on cyclic AMP levels and on lipolysis. *Ann N Y Acad Sci* 185:129–136
- Benito-Garcia E, Heller JE, Chibnik LB, Maher NE, Matthews HM, Bilics JA, Weinblatt ME, Shadick NA (2006) Dietary caffeine intake does not affect methotrexate efficacy in patients with rheumatoid arthritis. *J Rheumatol* 33:1275–1281
- Bruns RF, Lu GH, Pugsley TA (1986) Characterization of the A2 adenosine receptor labeled by [3H]NECA in rat striatal membranes. *Mol Pharmacol* 29:331–346
- Chavez-Valdez R, Wills-Karp M, Ahlawat R, Cristofalo EA, Nathan A, Gauda EB (2009) Caffeine modulates TNF-alpha production by cord blood monocytes: the role of adenosine receptors. *Pediatr Res* 65:203–208
- Che J, Chan ES, Cronstein BN (2007) Adenosine A2A receptor occupancy stimulates collagen expression by hepatic stellate cells via pathways involving protein kinase A, Src, and extracellular signal-regulated kinases 1/2 signaling cascade or p38 mitogen-activated protein kinase signaling pathway. *Mol Pharmacol* 72:1626–1636
- Chen Y, Corriden R, Inoue Y, Yip L, Hashiguchi N, Zinkernagel A, Nizet V, Insel PA, Junger WG (2006) ATP release guides neutrophil chemotaxis via P2Y2 and A3 receptors. *Science* 314:1792–1795
- Cosio BG, Tsaprouni L, Ito K, Jazrawi E, Adcock IM, Barnes PJ (2004) Theophylline restores histone deacetylase activity and steroid responses in COPD macrophages. *J Exp Med* 200:689–695
- Cronstein BN (2005) Low-dose methotrexate: a mainstay in the treatment of rheumatoid arthritis. *Pharmacol Rev* 57:163–172
- Csoka B, Nemeth ZH, Virag L, Gergely P, Leibovich SJ, Pacher P, Sun CX, Blackburn MR, Vizi ES, Deitch EA, Hasko G (2007) A2A adenosine receptors and C/EBPbeta are crucially required for IL-10 production by macrophages exposed to *Escherichia coli*. *Blood* 110:2685–2695
- Deaglio S, Dwyer KM, Gao W, Friedman D, Usheva A, Erat A, Chen JF, Enjoji K, Linden J, Oukka M, Kuchroo VK, Strom TB, Robson SC (2007) Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression. *J Exp Med* 204:1257–1265
- Dervieux T (2009) Methotrexate pharmacogenomics in rheumatoid arthritis: introducing false-positive report probability. *Rheumatology (Oxford)* 48:597–598
- Dervieux T, Furst D, Lein DO, Capps R, Smith K, Walsh M, Kremer J (2004) Polyglutamation of methotrexate with common polymorphisms in reduced folate carrier, aminoimidazole carboxamide ribonucleotide transformylase, and thymidylate synthase are associated with methotrexate effects in rheumatoid arthritis. *Arthritis Rheum* 50:2766–2774
- Dervieux T, Furst D, Lein DO, Capps R, Smith K, Caldwell J, Kremer J (2005) Pharmacogenetic and metabolite measurements are associated with clinical status in patients with rheumatoid arthritis treated with methotrexate: results of a multicentred cross sectional observational study. *Ann Rheum Dis* 64:1180–1185
- Dervieux T, Wessels JA, van der Straaten T, Penrod N, Moore JH, Guchelaar HJ, Kremer JM (2009) Gene-gene interactions in folate and adenosine biosynthesis pathways affect methotrexate efficacy and tolerability in rheumatoid arthritis. *Pharmacogenet Genomics*
- Eltzschig HK, Macmanus CF, Colgan SP (2008) Neutrophils as sources of extracellular nucleotides: functional consequences at the vascular interface. *Trends Cardiovasc Med* 18:103–107

- Endres S, Fulle HJ, Sinha B, Stoll D, Dinarello CA, Gerzer R, Weber PC (1991) Cyclic nucleotides differentially regulate the synthesis of tumour necrosis factor- $\alpha$  and interleukin-1  $\beta$  by human mononuclear cells. *Immunology* 72:56–60
- Erdmann AA, Gao ZG, Jung U, Foley J, Borenstein T, Jacobson KA, Fowler DH (2005) Activation of Th1 and Tc1 cell adenosine A2A receptors directly inhibits IL-2 secretion in vitro and IL-2-driven expansion in vivo. *Blood* 105:4707–4714
- Feoktistov I, Biaggioni I (1995) Adenosine A2b receptors evoke interleukin-8 secretion in human mast cells. An enprofylline-sensitive mechanism with implications for asthma. *J Clin Invest* 96:1979–1986
- Feoktistov I, Biaggioni I (1997) Adenosine A2B receptors. *Pharmacol Rev* 49:381–402
- Fozard JR, Hannon JP (1999) Adenosine receptor ligands: potential as therapeutic agents in asthma and COPD. *Pulm Pharmacol Ther* 12:111–114
- Fozard JR, Pfannkuche HJ, Schuurman HJ (1996) Mast cell degranulation following adenosine A3 receptor activation in rats. *Eur J Pharmacol* 298:293–297
- Fredholm BB (2007) Adenosine, an endogenous distress signal, modulates tissue damage and repair. *Cell Death Differ* 14:1315–1323
- Fredholm BB, Sandberg G (1983) Inhibition by xanthine derivatives of adenosine receptor-stimulated cyclic adenosine 3',5'-monophosphate accumulation in rat and guinea-pig thymocytes. *Br J Pharmacol* 80:639–644
- Fredholm BB, Zhang Y, van der Ploeg I (1996) Adenosine A2A receptors mediate the inhibitory effect of adenosine on formyl-Met-Leu-Phe-stimulated respiratory burst in neutrophil leucocytes. *Naunyn Schmiedebergs Arch Pharmacol* 354:262–267
- Fredholm BB, Battig K, Holmen J, Nehlig A, Zvartau EE (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fredholm BB, Irenius E, Kull B, Schulte G (2001) Comparison of the potency of adenosine as an agonist at human adenosine receptors expressed in Chinese hamster ovary cells. *Biochem Pharmacol* 61:443–448
- Fredholm BB, Chern Y, Franco R, Sitkovsky M (2007) Aspects of the general biology of adenosine A2A signaling. *Prog Neurobiol* 83:263–276
- Gessi S, Merighi S, Varani K, Leung E, Mac Lennan S, Borea PA (2008) The A3 adenosine receptor: an enigmatic player in cell biology. *Pharmacol Ther* 117:123–140
- Hasko G, Szabo C, Nemeth ZH, Salzman AL, Vizi ES (1998) Suppression of IL-12 production by phosphodiesterase inhibition in murine endotoxemia is IL-10 independent. *Eur J Immunol* 28:468–472
- Hasko G, Pacher P, Deitch EA, Vizi ES (2007) Shaping of monocyte and macrophage function by adenosine receptors. *Pharmacol Ther* 113:264–275
- Hasko G, Linden J, Cronstein B, Pacher P (2008) Adenosine receptors: therapeutic aspects for inflammatory and immune diseases. *Nat Rev Drug Discov* 7:759–770
- Hasko G, Csoka B, Nemeth ZH, Vizi ES, Pacher P (2009) A(2B) adenosine receptors in immunity and inflammation. *Trends Immunol* 30:263–270
- Hichami A, Boichot E, Germain N, Legrand A, Moodley I, Lagente V (1995) Involvement of cyclic AMP in the effects of phosphodiesterase IV inhibitors on arachidonate release from mononuclear cells. *Eur J Pharmacol* 291:91–97
- Holgate ST (2005) The Quintiles Prize Lecture 2004. The identification of the adenosine A2B receptor as a novel therapeutic target in asthma. *Br J Pharmacol* 145:1009–1015
- Horrigan LA, Kelly JP, Connor TJ (2004) Caffeine suppresses TNF- $\alpha$  production via activation of the cyclic AMP/protein kinase A pathway. *Int Immunopharmacol* 4:1409–1417
- Horrigan LA, Kelly JP, Connor TJ (2006) Immunomodulatory effects of caffeine: friend or foe? *Pharmacol Ther* 111:877–892
- Inouye LK, Wharton W (1986) The relationship between intracellular cyclic AMP concentrations and the in vitro growth of macrophages. *J Leukoc Biol* 39:657–670

- Ito K, Lim S, Caramori G, Cosio B, Chung KF, Adcock IM, Barnes PJ (2002) A molecular mechanism of action of theophylline: induction of histone deacetylase activity to decrease inflammatory gene expression. *Proc Natl Acad Sci USA* 99:8921–8926
- Jin X, Shepherd RK, Duling BR, Linden J (1997) Inosine binds to A3 adenosine receptors and stimulates mast cell degranulation. *J Clin Invest* 100:2849–2857
- Khoa ND, Montesinos MC, Reiss AB, Delano D, Awadallah N, Cronstein BN (2001) Inflammatory cytokines regulate function and expression of adenosine A(2A) receptors in human monocytic THP-1 cells. *J Immunol* 167:4026–4032
- Meiners I, Hauschildt S, Nieber K, Munch G (2004) Pentoxifylline and propentofylline are inhibitors of TNF-alpha release in monocytes activated by advanced glycation endproducts. *J Neural Transm* 111:441–447
- Montesinos C, Yap JS, Desai A, Posadas I, McCrary CT, Cronstein BN (2000) Reversal of the antiinflammatory effects of methotrexate by the nonselective adenosine receptor antagonists theophylline and caffeine. Evidence that the antiinflammatory effects of methotrexate are mediated via multiple adenosine receptors in rat adjuvant arthritis. *Arthritis Rheum* 43:656–663
- Mustafa SJ, Nadeem A, Fan M, Zhong H, Belardinelli L, Zeng D (2007) Effect of a specific and selective A(2B) adenosine receptor antagonist on adenosine agonist AMP and allergen-induced airway responsiveness and cellular influx in a mouse model of asthma. *J Pharmacol Exp Ther* 320:1246–1251
- Nemeth ZH, Hasko G, Szabo C, Vizi ES (1997) Amrinone and theophylline differentially regulate cytokine and nitric oxide production in endotoxemic mice. *Shock* 7:371–375
- Nemeth ZH, Leibovich SJ, Deitch EA, Sperlagh B, Virag L, Vizi ES, Szabo C, Hasko G (2003) Adenosine stimulates CREB activation in macrophages via a p38 MAPK-mediated mechanism. *Biochem Biophys Res Commun* 312:883–888
- Nesher G, Mates M, Zevin S (2003) Effect of caffeine consumption on efficacy of methotrexate in rheumatoid arthritis. *Arthritis Rheum* 48:571–572
- Ohta A, Sitkovsky M (2001) Role of G-protein-coupled adenosine receptors in downregulation of inflammation and protection from tissue damage. *Nature* 414:916–920
- Ohta A, Lukashov D, Jackson EK, Fredholm BB, Sitkovsky M (2007) 1, 3, 7-trimethylxanthine (caffeine) may exacerbate acute inflammatory liver injury by weakening the physiological immunosuppressive mechanism. *J Immunol* 179:7431–7438
- Polosa R, Holgate ST (2006) Adenosine receptors as promising therapeutic targets for drug development in chronic airway inflammation. *Curr Drug Targets* 7:699–706
- Revan S, Montesinos MC, Naime D, Landau S, Cronstein BN (1996) Adenosine A2 receptor occupancy regulates stimulated neutrophil function via activation of a serine/threonine protein phosphatase. *J Biol Chem* 271:17114–17118
- Ryzhov S, Goldstein AE, Biaggioni I, Feoktistov I (2006) Cross-talk between G(s)- and G(q)-coupled pathways in regulation of interleukin-4 by A(2B) adenosine receptors in human mast cells. *Mol Pharmacol* 70:727–735
- Sawynok J, Yaksh TL (1993) Caffeine as an analgesic adjuvant: a review of pharmacology and mechanisms of action. *Pharmacol Rev* 45:43–85
- Silke C, Murphy MS, Buckley T, Busted S, Molloy MG, Phelan M (2001) The effects of caffeine ingestion on the efficacy of methotrexate. *Rheumatology (Oxford)* 40(Suppl 1):34
- Sitkovsky MV (2009) T regulatory cells: hypoxia-adenosinergic suppression and re-direction of the immune response. *Trends Immunol* 30(3):102–108
- Sullivan GW, Luong LS, Carper HT, Barnes RC, Mandell GL (1995) Methylxanthines with adenosine alter TNF alpha-primed PMN activation. *Immunopharmacology* 31:19–29
- van Calker D, Muller M, Hamprecht B (1979) Adenosine regulates via two different types of receptors, the accumulation of cyclic AMP in cultured brain cells. *J Neurochem* 33:999–1005
- Volonte C, D'Ambrosi N (2009) Membrane compartments and purinergic signalling: the purinome, a complex interplay among ligands, degrading enzymes, receptors and transporters. *FEBS J* 276:318–329

- Walker BA, Rocchini C, Boone RH, Ip S, Jacobson MA (1997) Adenosine A2a receptor activation delays apoptosis in human neutrophils. *J Immunol* 158:2926–2931
- Weisman MH, Furst DE, Park GS, Kremer JM, Smith KM, Wallace DJ, Caldwell JR, Dervieux T (2006) Risk genotypes in folate-dependent enzymes and their association with methotrexate-related side effects in rheumatoid arthritis. *Arthritis Rheum* 54:607–612
- Wessels JA, de Vries-Bouwstra JK, Heijmans BT, Slagboom PE, Goekoop-Ruiterman YP, Allaart CF, Kerstens PJ, van Zeben D, Breedveld FC, Dijkman BA, Huizinga TW, Guchelaar HJ (2006a) Efficacy and toxicity of methotrexate in early rheumatoid arthritis are associated with single-nucleotide polymorphisms in genes coding for folate pathway enzymes. *Arthritis Rheum* 54:1087–1095
- Wessels JA, Kooloos WM, Jonge RD, De Vries-Bouwstra JK, Allaart CF, Linsen A, Collee G, Sonnaville PD, Lindemans J, Huizinga TW, Guchelaar HJ (2006b) Relationship between genetic variants in the adenosine pathway and outcome of methotrexate treatment in patients with recent-onset rheumatoid arthritis. *Arthritis Rheum* 54:2830–2839
- Yasui K, Hu B, Nakazawa T, Agematsu K, Komiyama A (1997) Theophylline accelerates human granulocyte apoptosis not via phosphodiesterase inhibition. *J Clin Invest* 100:1677–1684
- Yasui K, Agematsu K, Shinozaki K, Hokibara S, Nagumo H, Nakazawa T, Komiyama A (2000a) Theophylline induces neutrophil apoptosis through adenosine A2A receptor antagonism. *J Leukoc Biol* 67:529–535
- Yasui K, Agematsu K, Shinozaki K, Hokibara S, Nagumo H, Yamada S, Kobayashi N, Komiyama A (2000b) Effects of theophylline on human eosinophil functions: comparative study with neutrophil functions. *J Leukoc Biol* 68:194–200
- Yegutkin GG (2008) Nucleotide- and nucleoside-converting ectoenzymes: important modulators of purinergic signalling cascade. *Biochim Biophys Acta* 1783(5):673–694
- Zhong H, Shlykov SG, Molina JG, Sanborn BM, Jacobson MA, Tilley SL, Blackburn MR (2003) Activation of murine lung mast cells by the adenosine A3 receptor. *J Immunol* 171:338–345
- Zhong H, Belardinelli L, Maa T, Feoktistov I, Biaggioni I, Zeng D (2004) A(2B) adenosine receptors increase cytokine release by bronchial smooth muscle cells. *Am J Respir Cell Mol Biol* 30:118–125
- Zhong H, Belardinelli L, Maa T, Zeng D (2005) Synergy between A2B adenosine receptors and hypoxia in activating human lung fibroblasts. *Am J Respir Cell Mol Biol* 32:2–8
- Zhong H, Wu Y, Belardinelli L, Zeng D (2006) A2B adenosine receptors induce IL-19 from bronchial epithelial cells, resulting in TNF-alpha increase. *Am J Respir Cell Mol Biol* 35:587–592
- Zhou QY, Li C, Olah ME, Johnson RA, Stiles GL, Civelli O (1992) Molecular cloning and characterization of an adenosine receptor: the A3 adenosine receptor. *Proc Natl Acad Sci USA* 89:7432–7436

# Methylxanthines, Inflammation, and Cancer: Fundamental Mechanisms

Akio Ohta and Michail Sitkovsky

## Contents

1	Caffeine and Liver Inflammation .....	471
1.1	Possible Prevention of Liver Diseases by Habitual Coffee Consumption .....	471
1.2	Caffeine May Promote the Induction of Acute Hepatitis .....	472
2	Caffeine and Tumor Immunology .....	476
3	Control of Inflammation by Methylxanthines .....	477
	References .....	477

**Abstract** Methylxanthines are an integral part of everyday food and drink consumption even though the majority of humans do not identify them by their chemical name. The breakthrough in understanding the action(s) of methylxanthines was in large part due to the understanding that methylxanthines can function as antagonists of adenosine receptors. This represented an example of scientific search and was instructive in view of both new therapeutic options and alarming realizations. It was the subsequent demonstration of the *in vivo* critical role of A2A adenosine receptors in controlling excessive collateral inflammatory damage that attracted the attention

---

A. Ohta (✉)

Department of Pharmaceutical Sciences, New England Inflammation and Tissue Protection Institute, Northeastern University, 134 Mugar Building, 360 Huntington Avenue, Boston, MA 02115, USA

e-mail: a.ohta@neu.edu

M. Sitkovsky

Department of Pharmaceutical Sciences, New England Inflammation and Tissue Protection Institute, Northeastern University, 134 Mugar Building, 360 Huntington Avenue, Boston, MA 02115, USA

Cancer Vaccine Center, Dana-Farber Cancer Institute, Harvard Institutes of Medicine, 77 Avenue Pasteur, Room 418, Boston, MA 02115, USA

of immunologists to the A2A-adenosine-receptor-antagonizing methylxanthines. We summarize here data showing that caffeine is capable of preventing the inhibition of antitumor T cells in a hypoxic tumor microenvironment. On the other hand, caffeine may exacerbate liver damage by weakening the tissue-protecting A2A adenosine receptor signaling during episodes of acute liver inflammation. However, methylxanthines may also prevent the excessive hepatic connective tissue deposition that is associated with the progression of chronic hepatitis to cirrhosis, which is one of the common causes of mortality.

**Keywords** Adenosine · Adenosine receptor · Autoimmunity · Caffeine · Hepatitis · Hypoxia · Tumor

## Abbreviations

A2AR	A2A adenosine receptors
ALT	Alanine aminotransferase
AST	Aspartate aminotransferase
cAMP	Cyclic AMP
Con A	Concanavalin A
GGT	$\gamma$ -Glutamyltransferase
LPS	Lipopolysaccharide

In this chapter we will provide an overview of both pro- and anti-inflammatory properties of caffeine. This is potentially important since caffeine is now consumed without appreciation that it may affect the course of ongoing inflammatory response. Caffeine has long been considered to be anti-inflammatory because of its activity as a phosphodiesterase inhibitor (Horrigan et al. 2006). To inhibit phosphodiesterase, however, caffeine must be present at a high concentration, which may not be achieved during normal consumption in humans (May et al. 1982; Smith et al. 1982; Tiffin et al. 1995). It was subsequently understood that the effects of caffeine at doses that reflect the routine levels of consumption are mediated by the ability to antagonize the adenosine receptors (Fredholm et al. 1999). Recent studies investigated adenosine receptor antagonism by clinically relevant dose of caffeine and found that caffeine rather promotes inflammation (Ohta et al. 2007), which is opposite to the effects as a phosphodiesterase inhibitor. In this chapter, we discuss the effects of this representative methylxanthine on inflammation, focusing on pathogenesis of inflammatory liver diseases. We also describe the possible use of caffeine as a proinflammatory anticancer drug to be combined with tumor immunotherapy.

## 1 Caffeine and Liver Inflammation

### 1.1 *Possible Prevention of Liver Diseases by Habitual Coffee Consumption*

Inflammation of the liver results from a variety of causes. The most prevalent cause of hepatic inflammatory diseases is hepatitis virus infection (Lai et al. 2003; Poynard et al. 2003). There are also many patients suffering from alcohol-associated hepatitis, autoimmune hepatitis, and drug-induced liver injury (Lee 2004; Czaja 2005). After the acute phase, different degrees of severity of inflammation may persist as chronic hepatitis in a considerable number of viral, autoimmune, and drug-induced hepatitis patients. The repeated liver tissue remodeling caused by frequent inflammation often leads to development of fibrosis and cirrhosis (Nakamoto et al. 1998; Freeman et al. 2003; Fattovich et al. 2004). Furthermore, repeated liver inflammation may result in the development of liver cancer (Nakamoto et al. 1998; Fattovich et al. 2004).

Inflammatory cells play a major role both in liver damage and in tissue repair (Chisari 1997; Rehmann 2000; Ohta et al. 2000). The immune response of viral antigen-specific T cells plays a pivotal role in the destruction of infected hepatocytes. Recent studies have suggested that even in cases of chemical hepatotoxicity, immune cells, including T cells, participate in expanding liver damage after initial chemical insult (Luster et al. 2001; Simeonova et al. 2001). Furthermore, T cells are suggested to regulate development of cirrhosis by affecting fibrogenesis after hepatic damage (Shi et al. 1997; Safadi et al. 2004).

During the last two decades, epidemiology studies consistently showed an inverse relation between habitual coffee consumption and the risk of liver diseases. Coffee consumption has been shown to inversely correlate with the risk of cirrhosis (Klatsky and Armstrong 1992; Klatsky et al. 1993; Corrao et al. 2001; Tverdal and Skurtveit 2003; Ruhl and Everhart 2005a). In 1992, it was reported that the risk of alcoholic cirrhosis decreased to 20% by consumption of four or more cups of coffee per day (Klatsky and Armstrong 1992). The same dataset also showed a lower risk of cirrhosis death in coffee drinkers (Klatsky et al. 1993). Subsequent studies from Italy showed a decreased risk by coffee drinking in both alcoholic and nonalcoholic cirrhosis (Corrao et al. 2001). The relative risk was correlated with the amount of coffee, e.g., 47% in those who drink one cup per day, 23% for two cups, 16% for four or more cups compared with noncoffee drinkers. A mortality follow-up study from Norway confirmed the existence of an inverse relation between coffee and cirrhosis (Tverdal and Skurtveit 2003). In a recent study from the USA, subjects who consumed more than two cups of coffee had a lower incidence of chronic liver diseases (relative risk 0.43) compared with those who consumed less than one cup per day (Ruhl and Everhart 2005a). Such a protective effect of coffee was clear in the high-risk group for liver diseases, i.e., heavy alcohol intake, overweight, diabetes, or high iron load.



The serum levels of  $\gamma$ -glutamyltransferase (GGT), a relevant marker of cirrhotic risk, were found to be lower in coffee drinkers. GGT levels decreased in association with coffee consumption in the general population (Casiglia et al. 1993), more evidently among heavy alcohol drinkers and heavy smokers (Tanaka et al. 1998). There is also an inverse relation between coffee and serum transaminase levels, aspartate aminotransferase (AST) and alanine aminotransferase (ALT). In the general population, the relative risk for elevated AST and ALT levels (more than 40 U/L) decreased in coffee drinkers, correlating with the daily coffee consumption (Honjo et al. 2001). A study from the USA showed that lower ALT levels (less than 43 U/L) were associated with increasing coffee consumption. The relative risk was 0.56 in persons who drank more than two cups compared with noncoffee drinkers (Ruhl and Everhart 2005b). The observed protective effect of coffee was independent of the type of risk for liver injury. These studies indicate that coffee consumption is associated with decreased hepatocyte damage in individuals at high risk of liver injury.

For caffeine-containing beverages other than coffee, a few studies have shown controversial results. The epidemiological relation between caffeine consumption and liver diseases, however, is not as clear as discussed above for coffee consumption. Initially, tea and other caffeine-containing beverages were reported to have no correlation with serum GGT levels and cirrhosis (Klatsky et al. 1993; Corrao et al. 2001). However, recent studies have shown an inverse relation between tea consumption and serum ALT levels and the incidence of chronic liver diseases in the high-risk population (Ruhl and Everhart 2005a, b). Indeed, tea consumption was as effective as coffee consumption in the latter studies.

## ***1.2 Caffeine May Promote the Induction of Acute Hepatitis***

Biological effects of caffeine have been studied for a long time (Fredholm et al. 1999; Foukas et al. 2002). Importantly, caffeine can modulate intracellular levels of cyclic AMP (cAMP), which is a regulator of immune cell functions. The increase of the level of cAMP inhibits activation of inflammatory effector cells such as T cells and granulocytes. The induction of cAMP strongly inhibits oxidative burst in neutrophils and suppresses inflammation (Nielson et al. 1990). In T cells, cAMP downregulates various cellular functions, including proliferation, cytokine production, and cytotoxicity mediated by Fas ligand and granule exocytosis (Sitkovsky et al. 1988; Sugiyama et al. 1997). Caffeine is capable of increasing cAMP levels through inhibition of cAMP phosphodiesterase (Fredholm et al. 1999). Caffeine can exert an anti-inflammatory effect, which is a property shared with many other cAMP phosphodiesterase inhibitors. When rats were injected with D-galactosamine and/or lipopolysaccharide (LPS), coffee and other caffeine-containing beverages decreased liver damage, but decaffeinated coffee was not hepatoprotective (He et al. 2001; Akashi et al. 2009). An anti-inflammatory effect of caffeine was directly implicated in the suppression of liver damage by a high dose of caffeine.

Anti-inflammatory action by phosphodiesterase inhibition is well established; however, the clinical relevance of this pathway might be limited since the inhibition of phosphodiesterase requires a higher concentration of caffeine than the concentration achieved by normal human caffeine consumption (Fredholm et al. 1999). A caffeine concentration as high as 100  $\mu\text{M}$  was needed to inhibit production of LPS-induced cytokines in a cAMP/protein kinase A dependent manner (Horrigan et al. 2004). However, an acute dose of 2–2.5 mg/kg caffeine raised the blood concentration of caffeine only to 20–25  $\mu\text{M}$  in humans; this dose corresponds to two cups of coffee (May et al. 1982; Smith et al. 1982; Tiffin et al. 1995).

It is accepted now that it is the antagonism of adenosine receptors that is mostly operational at clinically relevant concentrations (Fredholm et al. 1999; Foukas et al. 2002). The antagonism of adenosine receptors requires approximately 20 times lower concentration of caffeine than the inhibition of phosphodiesterase (Fredholm et al. 1999). Currently, the behavioral activation by caffeine is largely accounted for by the interference with tonic activation of adenosine receptor signaling, especially A<sub>2</sub>A adenosine receptors (A<sub>2</sub>AR).

There are four different cell-surface receptors for extracellular adenosine: A<sub>1</sub>, A<sub>2</sub>A, A<sub>2</sub>B, and A<sub>3</sub> receptors (Fredholm et al. 2001). The high-affinity A<sub>1</sub> receptor and the low-affinity A<sub>3</sub> receptor are coupled to G<sub>i</sub> protein. The cAMP-elevating G<sub>s</sub>-protein-coupled A<sub>2</sub> receptors are subdivided into high-affinity A<sub>2</sub>AR and low-affinity A<sub>2</sub>B adenosine receptors. Immune cells express A<sub>2</sub>AR at high levels, and adenosine binding to A<sub>2</sub>AR inhibits immune activation through cAMP–protein kinase A signaling pathway. Pharmacological stimulation by an A<sub>2</sub>AR agonist results in the inhibition of proliferation (Huang et al. 1997), production of cytokines (Lappas et al. 2005), cytotoxicity of T cells (Koshiba et al. 1997), and activation of monocytes (Link et al. 2000) and granulocytes (Cronstein et al. 1990). The anti-inflammatory action of A<sub>2</sub>AR agonists has also been shown *in vivo*, including in ischemia/reperfusion injury (Lappas et al. 2006), airway inflammation (Fozard et al. 2002), inflammatory bowel disease (Odashima et al. 2005), and acute hepatitis (Ohta and Sitkovsky 2001).

Adenosine is an abundant compound in cells, and an important source of extracellular adenosine accumulation is tissue hypoxia (Sitkovsky et al. 2004; Sitkovsky and Ohta 2005). Hypoxia is associated with the increase of ATP catabolism into adenosine by the functions of apyrase (CD39) and 5'-ectonucleotidase (CD73) (Eltzschig et al. 2004). ATP released into the extracellular space undergoes phosphohydrolysis by CD39, which converts ATP/ADP to AMP in tandem with CD73, which then converts AMP to adenosine. Interestingly, hypoxia induces these two cell-surface enzymes, CD39 and CD73, promoting an increase of extracellular adenosine accumulation (Eltzschig et al. 2004; Kobie et al. 2006; Deaglio et al. 2007). Hypoxia also inhibits conversion of adenosine into AMP by adenosine kinase activity (Morote-Garcia et al. 2008). Furthermore, uptake of extracellular adenosine to the intracellular compartment is inhibited under the hypoxic condition. Therefore, the overall outcome of tissue hypoxia will be an increase of extracellular adenosine levels and a subsequent increase of adenosine signaling (Sitkovsky et al. 2004; Sitkovsky and Ohta 2005).

The physiological relevance of endogenous adenosine in the downregulation of inflammation is proven to be tissue-protective by preventing the collateral damage in a negative-feedback manner. The excessive damage to endothelial cells and the microcirculation leads to the interruption of the normal blood and oxygen supply, resulting in tissue hypoxia (Sitkovsky et al. 2004; Sitkovsky and Ohta 2005). Tissue hypoxia caused by inflammatory damage to the blood circulation can induce an increase of local adenosine levels (Driver et al. 1993; Martin et al. 2000). In the inflamed tissue, ATP release from damaged/dead cells may also contribute to the elevation of extracellular adenosine levels (Trautmann 2009). The increase in adenosine–A2AR signaling subsequent to inflammatory tissue damage then downregulates immune reactions in local inflamed tissue. This hypothesis was proven in a study demonstrating exacerbated inflammation in A2AR-deficient mice (Ohta and Sitkovsky 2001). Induction of acute hepatitis in A2AR-deficient mice caused much more devastating liver damage than in wild-type mice. Administration of an A2AR antagonist to wild-type mice also showed the exacerbation of hepatitis (Ohta and Sitkovsky 2001). These results indicate that extracellular increase of the level of adenosine is of great importance in the control of inflammatory responses. Adenosine produced from damaged tissue is the negative-feedback signal to immune cells necessary to prevent continuing excessive inflammatory damage to the vital organ that can lead to critical tissue dysfunction. Augmented inflammation in A2AR-deficient mice was not limited to hepatitis. It was also observed in arthritis, ischemia-reperfusion damage, sepsis, and lung inflammation (Montesinos et al. 2003; Day et al. 2004; Thiel et al. 2005; Németh et al. 2006). There are many other endogenous anti-inflammatory molecules that play an important role in the resolution of inflammation (Lawrence et al. 2002); however, it is interesting that no other active anti-inflammatory pathway is able to compensate for the absence of A2AR (Ohta and Sitkovsky 2001).

The demonstration of adenosine–A2AR interaction as a critical pathway in physiological downregulation of acute inflammation and protection of tissues raised the possibility that A2AR antagonists may be able to enhance inflammation. Caffeine is a nonselective antagonist of various adenosine receptor subtypes and it can antagonize A2AR (Fredholm et al. 1999). This, in turn, suggested the possibility that caffeine might exacerbate tissue damage if consumed during an acute inflammation episode (Ohta et al. 2007).

To clarify this clinically important issue, we examined the effects of caffeine on inflammation using a concanavalin A (Con A)-induced murine hepatitis model. Intravenous injection of Con A into mice induces T-cell-dependent necroinflammatory liver damage, which resembles human viral/autoimmune hepatitis (Tiegs et al. 1992). Con A-induced liver injury has been studied extensively, and the involvement of T cells, natural killer T cells (Toyabe et al. 1997; Kaneko et al. 2000), and Kupffer cells (Schumann et al. 2000) as well as the production of cytokines have been shown to be indispensable to the induction of the liver damage. As expected, Con A-induced liver damage was strongly exacerbated by caffeine at a dose of 10–20 mg/kg body weight (Ohta et al. 2007). The liver damage in caffeine-coinjected mice accompanied upregulation of proinflammatory cytokines, including interferon- $\gamma$ . Caffeine was

confirmed to exacerbate liver damage through antagonism of A2AR because it failed to increase liver damage in A2AR-deficient mice (Ohta et al. 2007).

In these assays caffeine was tested using a clinically relevant dose, which is equivalent to two or three cups of coffee consumption in humans. Injection of 20 mg/kg caffeine into mice raised the serum caffeine concentration to 40  $\mu$ M after 30 min (Ohta et al. 2007). This concentration of caffeine is sufficient to antagonize A2AR but not to inhibit cAMP phosphodiesterase (Fredholm et al. 1999). When caffeine was given at a high dose (100 mg/kg), which is not often achieved, the mice were strongly protected from Con A-induced liver damage accompanying downregulation of proinflammatory cytokines and a significant increase of anti-inflammatory cytokine IL-10 (Ohta et al. 2007).

This study showed that consumption of a normal amount of caffeine by humans may result in blockade of A2AR signaling in immune cells and may lead to exacerbation of acute inflammation. Because of multiple effects of caffeine on biological functions (Fredholm et al. 1999; Foukas et al. 2002), the effect of caffeine on inflammation is biphasic, dependent on the doses. Indeed, caffeine has opposite (pro- vs anti-inflammatory) effects on inflammation, with the dose determining the overall effect. In contrast to the inhibition of cAMP phosphodiesterase at high doses, a low dose of caffeine regulates cAMP levels by a different mechanism, i.e., inhibition of adenosine-mediated cAMP induction by antagonism of A2AR. The exacerbation of inflammation by a low dose of caffeine may be more clinically relevant since humans habitually consume lower, “proinflammatory” doses of caffeine (May et al. 1982; Smith et al. 1982; Tiffin et al. 1995).

The proinflammatory role of caffeine may not be easily revealed in epidemiology studies, which consistently show a beneficial effect of coffee consumption for liver diseases. One of the reasons could be that besides caffeine, coffee contains many other components, such as polyphenolic compounds (chlorogenic acid, caffeic acid, protocatechuic acid), lignan (secoisolariciresinol), diterpenes (cafestol, kahweol), and trigonelline (George et al. 2008). It is possible that some of these compounds may be responsible for the protection against liver diseases.

While the studies described above defined adenosine as the endogenous negative regulator of acute inflammation, it should be noted that adenosine also plays an important role in the resolution of inflammation. It has been shown that A2AR signaling promotes remodeling of damaged tissue by promotion of angiogenesis through upregulation of vascular endothelial growth factor (Montesinos et al. 2004) and downregulation of antiangiogenic thrombospondin-1 (Desai et al. 2005). Moreover, adenosine can upregulate collagen production and promote fibrogenesis (Nakav et al. 2009). Recent studies showed that A2AR-deficient mice are less susceptible to hepatic fibrosis induction (Chan et al. 2006). Importantly, A2AR antagonists, including caffeine, also prevented promotion of hepatic fibrosis (Nakav et al. 2009; Chan et al. 2006; Fernández et al. 2008). Taken together, these findings show that adenosine not only inhibits induction of inflammation but also promotes tissue remodeling through angiogenesis and fibrogenesis. While it is still to be carefully investigated, it is possible to speculate that caffeine may prevent cirrhosis development by the inhibition of the A2AR-mediated fibrosis-promoting pathway.

## 2 Caffeine and Tumor Immunology

Proinflammatory effects and subsequent exacerbation of tissue damage by A2AR antagonists are detrimental if they take place in vital organs, but the same effects may be highly desirable if they enhance the antipathogen response and inflammatory damage in cancerous tissues. Here we will provide an overview of the path to improve the immunotherapy of cancer by taking advantage of the proinflammatory effects of caffeine.

Difficulties in the efficient induction of inflammatory damage in tumors have been a big obstacle in immunotherapy of cancer (Gajewski et al. 2006; Mellor and Munn 2008) even though there have been very impressive improvements in the induction of antitumor immunity, e.g., cancer vaccines, genetically engineered antitumor T cells, and immune modulatory molecules (Gattinoni et al. 2006; Dougan and Dranoff 2009). These failures are explained by the tumor's escape due to the tumor's fortification with many different immunosuppressive mechanisms that can critically impair the efficacy of the induced antitumor effectors (Drake et al. 2005; Bai et al. 2008). The presence of regulatory T cells and myeloid suppressor cells may play a significant role in the intratumoral immunosuppressive mechanism. Anti-inflammatory cytokines such as IL-10 and TGF- $\beta$  are secreted from tumors (and from the suppressor cells at least in part). Expression of programmed death ligand-1 can induce apoptosis of T cells. Glucose deprivation and L-arginine deficiency by the expression of indoleamine-2,3-dioxygenase causes nutrient deficits in antitumor effector cells (Gattinoni et al. 2006; Dougan and Dranoff 2009).

Another important immunosuppressive mechanism that caffeine could be interfering with is hypoxia and subsequent increase of the level of extracellular adenosine in the tumor microenvironment (Ohta et al. 2006). Many solid tumors are characterized by an insufficient oxygen supply and transient or chronic hypoxia (Harris 2002; Vaupel and Mayer 2007; Sitkovsky et al. 2008). Tumor hypoxia may contribute to the propagation of oncogenic signals in the tumor microenvironment as was shown in the switch to the angiogenic phenotype (Laderoute et al. 2000), and tumor hypoxia is associated with poor prognosis (Evans et al. 2000; Giatromanolaki et al. 2001). Reflecting local hypoxia in tumors, adenosine levels are high in the tumor microenvironment (Ohta et al. 2006). The tumor-protecting role of adenosine was demonstrated by a dramatic improvement of tumor rejection in A2AR-deficient mice (Ohta et al. 2006). In this study, A2AR-deficient mice, but none of the wild-type mice, could survive tumor challenge, suggesting that hypoxic tumors protect themselves from incoming antitumor immune cells by producing extracellular adenosine, which then inhibits immune cells through A2AR.

In this experiment, treatment with A2AR antagonists improved T cell-dependent tumor eradication in wild-type mice (Ohta et al. 2006). One such antagonist was caffeine, which enabled T-cell-mediated elimination of tumor nodules by interfering with adenosine–A2AR interactions. This immunopotentiating activity of caffeine is consistent with the exacerbation of inflammatory tissue damage in acute hepatitis described above. Furthermore, caffeine treatment decreased the number of

microvessels in the tumor and increased the number of apoptotic tumor cells (Ohta et al. 2006). This suggests that caffeine not only served as an immunoenhancer but also improved antitumor responses through inhibition of adenosine-mediated angiogenesis.

### 3 Control of Inflammation by Methylxanthines

In summary, the use of methylxanthines is a promising approach to promote immune responses against pathogens and against tumors. These compounds interrupt the immunosuppressive adenosine–A2AR signaling and may enhance the antipathogen immunity and pathogen destruction by immune cells. It is expected that caffeine could be effective as an adjuvant in vaccination against many other pathogens and with cancer vaccines. However, although a strong antiviral immune response is beneficial to the elimination of pathogens, suppression of the tissue-protecting adenosine–A2AR pathway may cause the uncontrolled activation of immune cells and severe inflammation. For example, exacerbation of acute viral hepatitis can result in fulminant hepatitis, the serious and high-mortality complication. Consumption of A2AR-antagonizing methylxanthines may require caution in patients with ongoing acute inflammation. While viral fulminant hepatitis causes acute liver failure, the main cause of acute liver failure is the drug-induced hepatitis, most often because of acetaminophen overdose (Caraceni and Van Thiel 1995; Lee 2003). Since activated inflammatory cells are often found in drug-induced hepatitis (Luster et al. 2001; Simeonova et al. 2001), methylxanthines may also enhance the tissue damage during drug-induced hepatitis. In contrast, during chronic hepatitis, A2AR antagonists are expected to prevent excessive hepatic connective tissue deposition resulting from fibrogenesis. Methylxanthines may prevent excessive hepatic connective tissue deposition that is associated with the progression of hepatitis to cirrhosis, which is one of the common causes of mortality. Since methylxanthines can affect many of the events during acute/chronic hepatitis, it is important that studies are conducted to determine whether the use of A2AR-antagonizing methylxanthines should be informed by the individual's inflammation status.

### References

- Akashi I, Kagami K, Hirano T, Oka K (2009) Protective effects of coffee-derived compounds on lipopolysaccharide/D-galactosamine induced acute liver injury in rats. *J Pharm Pharmacol* 61:473–478
- Bai A, Higham E, Eisen HN, Wittrup KD, Chen J (2008) Rapid tolerization of virus-activated tumor-specific CD8+ T cells in prostate tumors of TRAMP mice. *Proc Natl Acad Sci USA* 105:13003–13008
- Caraceni P, Van Thiel DH (1995) Acute liver failure. *Lancet* 345:163–169

- Casiglia E, Spolaore P, Ginocchio G, Ambrosio GB (1993) Unexpected effects of coffee consumption on liver enzymes. *Eur J Epidemiol* 9:293–297
- Chan ES, Montesinos MC, Fernandez P, Desai A, Delanoc DL, Yee H, Reiss AB, Pillinger MH, Chen JF, Schwarzschild MA, Friedman SL, Cronstein BN (2006) Adenosine A<sub>2A</sub> receptors play a role in the pathogenesis of hepatic cirrhosis. *Br J Pharmacol* 148:1144–1155
- Chisari FV (1997) Cytotoxic T cells and viral hepatitis. *J Clin Invest* 99:1472–1477
- Corrao G, Zambon A, Bagnardi V, D'Amicis A, Klatsky A, Group CSIDECIR (2001) Coffee, caffeine, and the risk of liver cirrhosis. *Ann Epidemiol* 11:458–465
- Cronstein BN, Daguma L, Nichols D, Hutchison AJ, Williams M (1990) The adenosine/neutrophil paradox resolved: human neutrophils possess both A<sub>1</sub> and A<sub>2</sub> receptors that promote chemotaxis and inhibit O<sub>2</sub> generation, respectively. *J Clin Invest* 85:1150–1157
- Czaja AJ (2005) Current concepts in autoimmune hepatitis. *Ann Hepatol* 4:6–24
- Day YJ, Marshall MA, Huang L, McDuffie MJ, Okusa MD, Linden J (2004) Protection from ischemic liver injury by activation of A<sub>2A</sub> adenosine receptors during reperfusion: inhibition of chemokine induction. *Am J Physiol Gastrointest Liver Physiol* 286:G285–G293
- Deaglio S, Dwyer KM, Gao W, Friedman D, Usheva A, Erat A, Chen JF, Enjoji K, Linden J, Oukka M, Kuchroo VK, Strom TB, Robson SC (2007) Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression. *J Exp Med* 204:1257–1265
- Desai A, Victor-Vega C, Gadangi S, Montesinos MC, Chu CC, Cronstein BN (2005) Adenosine A<sub>2A</sub> receptor stimulation increases angiogenesis by down-regulating production of the anti-angiogenic matrix protein thrombospondin 1. *Mol Pharmacol* 67:1406–1413
- Dougan M, Dranoff G (2009) Immune therapy for cancer. *Annu Rev Immunol* 27:83–117
- Drake CG, Doody AD, Mihalyo MA, Huang CT, Kelleher E, Ravi S, Hipkiss EL, Flies DB, Kennedy EP, Long M, McGary PW, Coryell L, Nelson WG, Pardoll DM, Adler AJ (2005) Androgen ablation mitigates tolerance to a prostate/prostate cancer-restricted antigen. *Cancer Cell* 7:239–249
- Driver AG, Kukoly CA, Ali S, Mustafa SJ (1993) Adenosine in bronchoalveolar lavage fluid in asthma. *Am Rev Respir Dis* 148:91–97
- Eltzschig HK, Thompson LF, Karhausen J, Cotta RJ, Ibla JC, Robson SC, Colgan SP (2004) Endogenous adenosine produced during hypoxia attenuates neutrophil accumulation: coordination by extracellular nucleotide metabolism. *Blood* 104:3986–3992
- Evans SM, Hahn S, Pook DR, Jenkins WT, Chalian AA, Zhang P, Stevens C, Weber R, Weinstein G, Benjamin I, Mirza N, Morgan M, Rubin S, McKenna WG, Lord EM, Koch CJ (2000) Detection of hypoxia in human squamous cell carcinoma by EF5 binding. *Cancer Res* 60:2018–2024
- Fattovich G, Stroffolini T, Zagni I, Donato F (2004) Hepatocellular carcinoma in cirrhosis: incidence and risk factors. *Gastroenterology* 127:S35–S50
- Fernández P, Trzaska S, Wilder T, Chiriboga L, Blackburn MR, Cronstein BN, Chan ES (2008) Pharmacological blockade of A<sub>2A</sub> receptors prevents dermal fibrosis in a model of elevated tissue adenosine. *Am J Pathol* 172:1675–1682
- Foukas LC, Daniele N, Ktori C, Anderson KE, Jensen J, Shepherd PR (2002) Direct effects of caffeine and theophylline on p110 delta and other phosphoinositide 3-kinases. Differential effects on lipid kinase and protein kinase activities. *J Biol Chem* 277:37124–37130
- Fozard JR, Ellis KM, Villela Dantas MF, Tigani B, Mazzoni L (2002) Effects of CGS 21680, a selective adenosine A<sub>2A</sub> receptor agonist, on allergic airways inflammation in the rat. *Eur J Pharmacol* 438:183–188
- Fredholm BB, Bättig K, Holmén J, Nehlig A, Zvartau EE (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fredholm BB, IJzerman AP, Jacobson KA, Klotz KN, Linden J (2001) International Union of Pharmacology. XXV. Nomenclature and classification of adenosine receptors. *Pharmacol Rev* 53:527–552

- Freeman AJ, Law MG, Kaldor JM, Dore GJ (2003) Predicting progression to cirrhosis in chronic hepatitis C virus infection. *J Viral Hepat* 10:285–293
- Gajewski TF, Meng Y, Blank C, Brown I, Kacha A, Kline J, Harlin H (2006) Immune resistance orchestrated by the tumor microenvironment. *Immunol Rev* 213:131–145
- Gattinoni L, Powell DJ Jr, Rosenberg SA, Restifo NP (2006) Adoptive immunotherapy for cancer: building on success. *Nat Rev Immunol* 6:383–393
- George SE, Ramalakshmi K, Mohan Rao LJ (2008) A perception on health benefits of coffee. *Crit Rev Food Sci Nutr* 48:464–486
- Giatromanolaki A, Koukourakis MI, Sivridis E, Pastorek J, Wykoff CC, Gatter KC, Harris AL (2001) Expression of hypoxia-inducible carbonic anhydrase-9 relates to angiogenic pathways and independently to poor outcome in non-small cell lung cancer. *Cancer Res* 61:7992–7998
- Harris AL (2002) Hypoxia – a key regulatory factor in tumour growth. *Nat Rev Cancer* 2:38–47
- He P, Noda Y, Sugiyama K (2001) Suppressive effect of coffee on lipopolysaccharide-induced hepatitis in D-galactosamine-sensitized rats. *Biosci Biotechnol Biochem* 65:1924–1927
- Honjo S, Kono S, Coleman MP, Shintchi K, Sakurai Y, Todoroki I, Umeda T, Wakabayashi K, Imanishi K, Nishikawa H, Ogawa S, Katsurada M, Nakagawa K, Yoshizawa N (2001) Coffee consumption and serum aminotransferases in middle-aged Japanese men. *J Clin Epidemiol* 54:823–829
- Horrigan LA, Kelly JP, Connor TJ (2004) Caffeine suppresses TNF-alpha production via activation of the cyclic AMP/protein kinase A pathway. *Int Immunopharmacol* 4:1409–1417
- Horrigan LA, Kelly JP, Connor TJ (2006) Immunomodulatory effects of caffeine: friend or foe? *Pharmacol Ther* 111:877–892
- Huang S, Apasov S, Koshiba M, Sitkovsky M (1997) Role of A2a extracellular adenosine receptor-mediated signaling in adenosine-mediated inhibition of T-cell activation and expansion. *Blood* 90:1600–1610
- Kaneko Y, Harada M, Kawano T, Yamashita M, Shibata Y, Gejyo F, Nakayama T, Taniguchi M (2000) Augmentation of Valpha14 NKT cell-mediated cytotoxicity by interleukin 4 in an autocrine mechanism resulting in the development of concanavalin A-induced hepatitis. *J Exp Med* 191:105–114
- Klatsky AL, Armstrong MA (1992) Alcohol, smoking, coffee, and cirrhosis. *Am J Epidemiol* 136:1248–1257
- Klatsky AL, Armstrong MA, Friedman GD (1993) Coffee, tea, and mortality. *Ann Epidemiol* 3:375–381
- Kobie JJ, Shah PR, Yang L, Rebhahn JA, Fowell DJ, Mosmann TR (2006) T regulatory and primed uncommitted CD4 T cells express CD73, which suppresses effector CD4 T cells by converting 5'-adenosine monophosphate to adenosine. *J Immunol* 177:6780–6786
- Koshiba M, Kojima H, Huang S, Apasov S, Sitkovsky MV (1997) Memory of extracellular adenosine/A2a purinergic receptor-mediated signaling in murine T cells. *J Biol Chem* 272:25881–25889
- Laderoute KR, Alarcon RM, Brody MD, Calaoagan JM, Chen EY, Knapp AM, Yun Z, Denko NC, Giaccia AJ (2000) Opposing effects of hypoxia on expression of the angiogenic inhibitor thrombospondin 1 and the angiogenic inducer vascular endothelial growth factor. *Clin Cancer Res* 6:2941–2950
- Lai CL, Ratziu V, Yuen MF, Poynard T (2003) Viral hepatitis B. *Lancet* 362:2089–2094
- Lappas CM, Rieger JM, Linden J (2005) A2A adenosine receptor induction inhibits IFN-gamma production in murine CD4+ T cells. *J Immunol* 174:1073–1080
- Lappas CM, Day YJ, Marshall MA, Engelhard VH, Linden J (2006) Adenosine A2A receptor activation reduces hepatic ischemia reperfusion injury by inhibiting CD1d-dependent NKT cell activation. *J Exp Med* 203:2639–2648
- Lawrence T, Willoughby DA, Gilroy DW (2002) Anti-inflammatory lipid mediators and insights into the resolution of inflammation. *Nat Rev Immunol* 2:787–795
- Lee WM (2003) Acute liver failure in the United States. *Semin Liver Dis* 23:217–226
- Lee WM (2004) Acetaminophen and the U.S. Acute Liver Failure Study Group: lowering the risks of hepatic failure. *Hepatology* 40:6–9



- Link AA, Kino T, Worth JA, McGuire JL, Crane ML, Chrousos GP, Wilder RL, Elenkov IJ (2000) Ligand-activation of the adenosine A2a receptors inhibits IL-12 production by human monocytes. *J Immunol* 164:436–442
- Luster MI, Simeonova PP, Gallucci RM, Bruccoleri A, Blazka ME, Yuceosoy B (2001) Role of inflammation in chemical-induced hepatotoxicity. *Toxicol Lett* 120:317–321
- Martin C, Leone M, Viviani X, Ayem ML, Guieu R (2000) High adenosine plasma concentration as a prognostic index for outcome in patients with septic shock. *Crit Care Med* 28: 3198–3202
- May DC, Jarboe CH, VanBakel AB, Williams WM (1982) Effects of cimetidine on caffeine disposition in smokers and nonsmokers. *Clin Pharmacol Ther* 31:656–661
- Mellor AL, Munn DH (2008) Creating immune privilege: active local suppression that benefits friends, but protects foes. *Nat Rev Immunol* 8:74–80
- Montesinos MC, Desai A, Delano D, Chen JF, Fink JS, Jacobson MA, Cronstein BN (2003) Adenosine A2A or A3 receptors are required for inhibition of inflammation by methotrexate and its analog MX-68. *Arthritis Rheum* 48:240–247
- Montesinos MC, Shaw JP, Yee H, Shamamian P, Cronstein BN (2004) Adenosine A(2A) receptor activation promotes wound neovascularization by stimulating angiogenesis and vasculogenesis. *Am J Pathol* 164:1887–1892
- Morote-Garcia JC, Rosenberger P, Kuhlicke J, Eltzschig HK (2008) HIF-1-dependent repression of adenosine kinase attenuates hypoxia-induced vascular leak. *Blood* 111:5571–5580
- Nakamoto Y, Guidotti LG, Kuhlen CV, Fowler P, Chisari FV (1998) Immune pathogenesis of hepatocellular carcinoma. *J Exp Med* 188:341–350
- Nakav S, Kachko L, Vorobiov M, Rogachev B, Chaimovitz C, Zlotnik M, Douvdevani A (2009) Blocking adenosine A2A receptor reduces peritoneal fibrosis in two independent experimental models. *Nephrol Dial Transplant* 24:2392–2399
- Németh ZH, Csóka B, Wilmanski J, Xu D, Lu Q, Ledent C, Deitch EA, Pacher P, Spolarics Z, Haskó G (2006) Adenosine A2A receptor inactivation increases survival in polymicrobial sepsis. *J Immunol* 176:5616–5626
- Nielson CP, Vestal RE, Sturm RJ, Heaslip R (1990) Effects of selective phosphodiesterase inhibitors on the polymorphonuclear leukocyte respiratory burst. *J Allergy Clin Immunol* 86: 801–808
- Odashima M, Bamias G, Rivera-Nieves J, Linden J, Nast CC, Moskaluk CA, Marini M, Sugawara K, Kozaiwa K, Otaka M, Watanabe S, Cominelli F (2005) Activation of A2A adenosine receptor attenuates intestinal inflammation in animal models of inflammatory bowel disease. *Gastroenterology* 129:26–33
- Ohta A, Sitkovsky M (2001) Role of G-protein-coupled adenosine receptors in downregulation of inflammation and protection from tissue damage. *Nature* 414:916–920
- Ohta A, Sekimoto M, Sato M, Koda T, Nishimura S, Iwakura Y, Sekikawa K, Nishimura T (2000) Indispensable role for TNF- $\alpha$  and IFN- $\gamma$  at the effector phase of liver injury mediated by Th1 cells specific to hepatitis B virus surface antigen. *J Immunol* 165:956–961
- Ohta A, Gorelik E, Prasad SJ, Ronchese F, Lukashev D, Wong MK, Huang X, Caldwell S, Liu K, Smith P, Chen JF, Jackson EK, Apasov S, Abrams S, Sitkovsky M (2006) A2A adenosine receptor protects tumors from antitumor T cells. *Proc Natl Acad Sci USA* 103:13132–13137
- Ohta A, Lukashev D, Jackson EK, Fredholm BB, Sitkovsky M (2007) 1,3,7-Trimethylxanthine (caffeine) may exacerbate acute inflammatory liver injury by weakening the physiological immunosuppressive mechanism. *J Immunol* 179:7431–7438
- Poynard T, Yuen MF, Ratziu V, Lai CL (2003) Viral hepatitis C. *Lancet* 362:2095–2100
- Rehermann B (2000) Intrahepatic T cells in hepatitis B: viral control versus liver cell injury. *J Exp Med* 191:1263–1268
- Ruhl CE, Everhart JE (2005a) Coffee and tea consumption are associated with a lower incidence of chronic liver disease in the United States. *Gastroenterology* 129:1928–1936
- Ruhl CE, Everhart JE (2005b) Coffee and caffeine consumption reduce the risk of elevated serum alanine aminotransferase activity in the United States. *Gastroenterology* 128:24–32

- Safadi R, Ohta M, Alvarez CE, Fiel MI, Bansal M, Mehal WZ, Friedman SL (2004) Immune stimulation of hepatic fibrogenesis by CD8 cells and attenuation by transgenic interleukin-10 from hepatocytes. *Gastroenterology* 127:870–882
- Schumann J, Wolf D, Pahl A, Brune K, Papadopoulos T, van Rooijen N, Tiegs G (2000) Importance of Kupffer cells for T-cell-dependent liver injury in mice. *Am J Pathol* 157:1671–1683
- Shi Z, Wakil AE, Rockey DC (1997) Strain-specific differences in mouse hepatic wound healing are mediated by divergent T helper cytokine responses. *Proc Natl Acad Sci USA* 94:10663–10668
- Simeonova PP, Gallucci RM, Hulderman T, Wilson R, Kommineni C, Rao M, Luster MI (2001) The role of tumor necrosis factor- $\alpha$  in liver toxicity, inflammation, and fibrosis induced by carbon tetrachloride. *Toxicol Appl Pharmacol* 177:112–120
- Sitkovsky MV, Ohta A (2005) The ‘danger’ sensors that STOP the immune response: the A2 adenosine receptors? *Trends Immunol* 26:299–304
- Sitkovsky MV, Trenn G, Takayama H (1988) Cyclic AMP-dependent protein kinase as a part of the possible down-regulating pathway in the antigen receptor-regulated cytotoxic T lymphocyte conjugate formation and granule exocytosis. *Ann N Y Acad Sci* 532:350–358
- Sitkovsky MV, Lukashev D, Apasov S, Kojima H, Koshiba M, Caldwell C, Ohta A, Thiel M (2004) Physiological control of immune response and inflammatory tissue damage by hypoxia-inducible factors and adenosine A2A receptors. *Annu Rev Immunol* 22:657–682
- Sitkovsky MV, Kjaergaard J, Lukashev D, Ohta A (2008) Hypoxia-adenosinergic immunosuppression: tumor protection by T regulatory cells and cancerous tissue hypoxia. *Clin Cancer Res* 14:5947–5952
- Smith JM, Pearson S, Marks V (1982) Plasma caffeine concentration in outpatients. *Lancet* 2:985–986
- Sugiyama H, Chen P, Hunter M, Sitkovsky M (1997) Perturbation of the expression of the catalytic subunit Ca of cyclic AMP-dependent protein kinase inhibits TCR-triggered secretion of IL-2 by T-helper cells. *J Immunol* 158:171
- Tanaka K, Tokunaga S, Kono S, Tokudome S, Akamatsu T, Moriyama T, Zakouji H (1998) Coffee consumption and decreased serum gamma-glutamyltransferase and aminotransferase activities among male alcohol drinkers. *Int J Epidemiol* 27:438–443
- Thiel M, Chouker A, Ohta A, Jackson E, Caldwell C, Smith P, Lukashev D, Bittmann I, Sitkovsky MV (2005) Oxygenation inhibits the physiological tissue-protecting mechanism and thereby exacerbates acute inflammatory lung injury. *PLoS Biol* 3:e174
- Tiegs G, Hentschel J, Wendel A (1992) A T cell-dependent experimental liver injury in mice inducible by concanavalin A. *J Clin Invest* 90:196–203
- Tiffin P, Ashton H, Marsh R, Kamali F (1995) Pharmacokinetic and pharmacodynamic responses to caffeine in poor and normal sleepers. *Psychopharmacology* 121:494–502
- Toyabe S, Seki S, Iiai T, Takeda K, Shirai K, Watanabe H, Hiraide H, Uchiyama M, Abo T (1997) Requirement of IL-4 and liver NK1+ T cells for concanavalin A-induced hepatic injury in mice. *J Immunol* 159:1537–1542
- Trautmann A (2009) Extracellular ATP in the immune system: more than just a “danger signal”. *Sci Signal* 2:pe6
- Tverdal A, Skurtveit S (2003) Coffee intake and mortality from liver cirrhosis. *Ann Epidemiol* 13:419–423
- Vaupel P, Mayer A (2007) Hypoxia in cancer: significance and impact on clinical outcome. *Cancer Metastasis Rev* 26:225–239

# Methylxanthines and Drug Dependence: A Focus on Interactions with Substances of Abuse

Micaela Morelli and Nicola Simola

## Contents

1	Methylxanthine Dependence .....	484
2	Methylxanthine Tolerance .....	486
3	Methylxanthine Withdrawal .....	487
4	Methylxanthine Abuse Potential .....	488
5	Interactions Between Methylxanthines and Other Psychoactive Drugs .....	490
5.1	Amphetamines .....	491
5.2	Cocaine .....	493
5.3	Cannabis Derivates .....	495
5.4	Ethanol .....	495
5.5	Nicotine .....	497
5.6	Methylphenidate .....	498
5.7	Antagonists of the Glutamate NMDA Receptors .....	498
5.8	Opiates .....	499
5.9	Anxiolytic–Hypnotics .....	500
5.10	Miscellaneous Psychoactive Drugs .....	500
6	General Considerations on Methylxanthines and Drug Dependence Phenomena .....	501
	References .....	501

**Abstract** This chapter examines the psychostimulant actions of methylxanthines, with a focus on the consequences of their excessive use. Consumption of methylxanthines is pervasive and their use is often associated with that of substances known to produce dependence and to have abuse potential. Therefore, the consequences

---

M. Morelli (✉)

Department of Toxicology, University of Cagliari, Via Ospedale, 72, 09124 Cagliari, Italy  
Centre of Excellence for Neurobiology of Dependence, University of Cagliari, 09124 Cagliari, Italy

CNR Institute of Neuroscience, Cagliari, Italy

e-mail: morelli@unica.it

N. Simola

Department of Toxicology, University of Cagliari, Via Ospedale, 72, 09124 Cagliari, Italy

of this combined use are taken into consideration in order to evaluate whether, and to what extent, methylxanthines could influence dependence on or abuse of other centrally active substances, leading to either amplification or attenuation of their effects. Since the methylxanthine that mostly influences mental processes and readily induces psychostimulation is caffeine, this review mainly focuses on caffeine as a prototype of methylxanthine-produced dependence, examining, at the same time, the risks related to caffeine use.

**Keywords** Adenosine receptors · Amphetamine · Caffeine · Cocaine · Nicotine

## Abbreviations

CPP	Conditioned place preference
D <sub>2</sub> High	High-affinity D <sub>2</sub>
DARPP-32	Dopamine- and cyclic-AMP-regulated 32-kDa phosphoprotein
DSM IV	<i>Diagnostic and Statistical Manual of Mental Disorders</i> , fourth edition
IEGs	Immediate early genes
NGFI-A	Nerve growth factor I-A
NMDA	<i>N</i> -Methyl-D-aspartate

## 1 Methylxanthine Dependence

Methylxanthines, which include caffeine, theophylline, paraxanthine, and theobromine, are present in several dietary products, including coffee, tea, soft and “energy drinks”, maté, cakes, candies, and chocolate. Moreover, theophylline is used as an antiasthmatic, and caffeine is added to cold remedies and analgesic medications (Brice and Smith 2002). Owing to the large diffusion of these products, it can be assumed that the world population consumes methylxanthines every day in any of these forms. It must be noted that the majority of caffeine, the most popular of the methylxanthines, is degraded by the hepatic microsomal enzymatic system to paraxanthine and partially to theobromine and theophylline (Svenningsson et al. 1999).

Psychoactive substances are often associated with dependence phenomena, the definition of which identifies the presence of a pattern of behaviors focused on repetitive and compulsive seeking and taking of drugs. Compulsive and repetitive drug use may result in tolerance to the effect of the drug and withdrawal symptoms when use is reduced or stopped. The *Diagnostic and Statistical Manual of Mental Disorders*, fourth edition (DSM IV), by the American Psychiatric Association (1994) delineates seven criteria to define dependence (Table 1).

**Table 1** Characterization of methylxanthines according to the criteria of drug dependence proposed by the *Diagnostic and Statistical Manual of Mental Disorders*, fourth edition (American Psychiatric Association 1994)

Criterion of drug dependence	Fulfillment by methylxanthines
Induction of tolerance	Caffeine partially fulfills this criterion, as it induces tolerance to some (e.g., cardiovascular effects), but not all, of its effects. Tolerance to caffeine usually disappears after a short period the use of the substance has been discontinued
Induction of withdrawal symptoms	Caffeine partially fulfills this criterion, as it can induce withdrawal symptoms such as fatigue, headache, irritability, and depressed mood. Although these symptoms can be severe in certain individuals, caffeine withdrawal syndrome is usually not harmful and resolves in a few days after use of the substance has been discontinued
Taking the substance in larger amounts or over a period longer than intended	Methylxanthines can partially fulfill this criterion. However, it has to be considered that methylxanthines are usually consumed in the form of coffee, tea, soft drinks, and chocolate. Besides their methylxanthine content, the organoleptic properties of such dietary sources (e.g., flavor and sugar content), can greatly influence their prolonged intake
Persistent desire and unsuccessful efforts to cut down, or control, use	Methylxanthines, in general, do not fulfill this criterion. Similarly to what was observed for the previous criterion, excessive consumption of methylxanthine-containing dietary sources may be due to factors other than methylxanthines (see Sect. 4)
Spending a great deal of time in activities necessary to obtain the substance or recover from the effects of the substance	Methylxanthines do not fulfill this criterion
Giving up or reducing important social, occupational, or recreational activities because of substance use	Methylxanthines do not fulfill this criterion. On the contrary, conviviality may drive the consumption of methylxanthines
Continued use despite knowledge of persistent or recurrent physical or psychological problems exacerbated by the substance	Methylxanthines, in general, do not fulfill this criterion. On the other hand, the use of caffeine is often described as self-regulating, owing to the onset of unwanted aversive effects at high doses of the substance

Although possible dependence on caffeine has been considered by several research groups (Gilliland and Bullock 1984; Griffiths et al. 1990; Nehlig 1999), methylxanthines do not fulfill the criteria for drugs producing dependence, whereas caffeine partially fulfills these criteria (Table 1). The two symptoms of dependence

quoted by the DSM IV that methylxanthines may be associated with more closely are tolerance and withdrawal. Tolerance to a drug and withdrawal do not define characteristics of dependence, although they typically accompany dependence on certain drugs.

Since caffeine use is associated with very limited physical and psychological consequences and with weak reinforcing properties and withdrawal symptoms, the definition of caffeine as an “atypical drug of dependence” given by Daly and Fredholm (1998) is still widely accepted. Methylxanthine consumption is often associated with a group of factors varying from social conviviality and relaxation to stimulation, as an aid to regulate sleep and to increase attention and concentration. All these factors may have an important role when considering the different aspects influencing dependence.

In addition to caffeine dependence, the DSM IV describes caffeine use as associated with several distinct psychiatric syndromes: caffeine intoxication, caffeine-induced sleep disorder, and caffeine-induced anxiety disorder. Moreover, caffeine produces biphasic effects, with low doses eliciting increased attention, concentration, a feeling of well-being, increased energy, desire to socialize, and motivation for work, and high doses resulting in anxiety, tension, restlessness, and sleeplessness. One of the interesting effects of caffeine ingestion is that it tends to be self-regulating. Unlike other psychoactive substances such as heroin, ethanol, and psychostimulants such as cocaine and amphetamine, the use of which tends to increase without bound, the amount of caffeine that people consume is limited by the onset of unwanted side effects.

## 2 Methylxanthine Tolerance

Tolerance is a pharmacological phenomenon where the dose of a drug needs to be continually increased in order to achieve and maintain the same effect. Tolerance has been clearly described for some of the effects of caffeine.

In humans, tolerance to the behavioral effects of caffeine is of low magnitude and incomplete (Fredholm et al. 1999; Watson et al. 2002). It develops to the central effects of caffeine such as increase in tension, anxiety, and jitteriness, as well as to peripheral effects such as modifications in blood pressure, heart rate, and diuresis (Fredholm et al. 1999; Nehlig 2004). In line with the widespread use of caffeine for its psychostimulant effects, minimal tolerance develops to caffeine-induced attention and wakefulness.

In experimental animals, tolerance develops to the motor-stimulant effects of caffeine and theophylline and to the cerebral electrical activation, seizures, and disruption of operant behavior for food reward produced by caffeine (Finn and Holtzman 1988; Lau and Falk 1995) but not to the increase in brain 2-deoxyglucose uptake (Nehlig 2004). Of particular interest is the finding that long-term caffeine administration decreases the susceptibility of mice to *N*-methyl-D-aspartate (NMDA)-induced seizures (Georgiev et al. 1993).

Tolerance does not appear to be related to modifications in the total level of active metabolites of methylxanthine in the brain. Instead, disruption of dopaminergic functions by downregulated levels of adenosine A<sub>2A</sub> receptors, which largely interact with dopamine receptors, and increased expression of adenosine A<sub>1</sub> receptors in specific brain areas were found in caffeine-tolerant rats (Svenningsson et al. 1999).

### 3 Methylxanthine Withdrawal

The general definition of withdrawal takes into consideration the presence of symptoms that occur upon the abrupt discontinuation or a decrease in dosage of a drug. In addition, in order to experience the symptoms of withdrawal, dependence on the drug must be present.

In humans, the caffeine withdrawal syndrome, characterized by headache, fatigue, drowsiness, irritability, depressed mood, and anxiety, starts after 12–24 h of abstinence, peaks 20–48 h later, and does not appear to be related to the quantity of caffeine ingested (Fredholm et al. 1999; Griffiths et al. 1990; Nehlig 2004). Interestingly, there is a relationship between headache and cerebral blood flow (Couturier et al. 1997). Symptoms of caffeine withdrawal are not necessarily manifested all at the same time and their intensity may vary considerably between different individuals. Caffeine withdrawal syndrome is usually not harmful, it is self-limiting, and no reliable modifications in social behavior have been observed (Comer et al. 1997).

In accordance with the vast and increasing number of adolescents who drink caffeinated beverages, caffeine dependence and withdrawal symptoms are increasingly being reported in teenagers (Bernstein et al. 2002). Withdrawal symptoms were also apparent in newborns of mothers who were heavy drinkers of coffee or maté, which contain both caffeine and theobromine (McGowan et al. 1988; Martín et al. 2007).

In the adult North American population, coffee accounts for three quarters of daily methylxanthine consumption, whereas in the teenage population the majority of caffeine (two thirds) is consumed from soft drinks (Bernstein et al. 2002).

Theophylline withdrawal has been reported in some adult asthmatic patients in whom exacerbation of asthma associated with a significant decrease in peripheral blood monocytes, activated CD4<sup>+</sup> T lymphocytes, and CD8<sup>+</sup> T cells was observed without the presence of psychotropic symptoms (Kidney et al. 1995).

In experimental animals, withdrawal induces a decrease in locomotor activity (Kaplan et al. 1993; Nikodijević et al. 1993) and disrupts operant behavior (Mumford et al. 1988). In rodents, the magnitude and duration of withdrawal appear to be a function of the amount of caffeine assumed.

## 4 Methylxanthine Abuse Potential

Reinforcement has a major role in the abuse potential of drugs. This term refers to the efficacy of a substance in establishing and maintaining a behavior on which the delivery of the substance is dependent.

In humans, methylxanthines – particularly caffeine, on which the majority of studies are focused – have mild reinforcing properties, which maintain the self-administration of beverages containing these substances. Reinforcement occurs in both moderate and heavy consumers, although at different rates. Moreover, a clear U-shaped response exists, with high doses associated with aversive effects (Fredholm et al. 1999; Nehlig 2004). The presence of reinforcing properties is the principal but not the only determinant of caffeine ingestion, since in some studies its intake has been shown to be driven by the need to avoid withdrawal symptoms (Schuh and Griffiths 1997). Interestingly, however, humans discriminate caffeine and theophylline from a placebo or amphetamine without needing to be in a withdrawal state (Oliveto et al. 1993; Griffiths and Mumford 1995).

Although not yet clearly demonstrated, it has been suggested that factors unrelated to methylxanthine content – such as the smell and flavor of coffee, tea, and chocolate, together with the social environment that accompanies their consumption – may have an important role in their abuse potential (Benton 2004; Nehlig 2004).

In experimental animals, one of the first parameters examined by studies aimed at evaluating the abuse potential of methylxanthines was discrimination. The results of these studies showed that caffeine produced a slight increase in responding, which was approximately twice as high as with theophylline, while amphetamine produced a much greater increase (Modrow et al. 1981; Carney et al. 1985). Moreover, in rats able to discriminate caffeine from saline, theobromine did not evoke caffeine-like responses (Carney et al. 1985). Caffeine and theophylline were also compared with theobromine. The rank order of potency was caffeine > theophylline > theobromine (Carney 1982). In follow-up studies focused on caffeine and dopamine receptor agonist drugs, it was reported that low doses of caffeine produced cues resembling a weak dopaminergic stimulus (Harland et al. 1989). A similar type of generalization was found in nonhuman primates (Holtzman 1996). These studies demonstrated the presence of differences and similarities between methylxanthines and psychostimulants such as amphetamine and cocaine, and are in line with studies in humans that show low reinforcing properties of methylxanthines compared with other psychostimulants (Heishman and Henningfield 1992). More recent studies that focused on the abuse potential of methylxanthines have shown that while both caffeine and theophylline at low doses induce conditioned place preference (CPP) in rodents (Zarrindast and Moghadamnia 1997; Bedingfield et al. 1998), caffeine maintains self-administration behavior in all or a subset of animals depending on the protocol of administration, whereas theophylline is not self-administered



(Griffiths and Mumford 1995; Sahraei et al. 1999). An irregular pattern of caffeine self-administration was also observed in nonhuman primates, confirming that, in contrast to other known psychostimulants, caffeine acts as a reinforcer in limited conditions (Griffiths and Mumford 1995). In addition, it is important to emphasize that self-administration studies utilize intravenous administration, a way that magnifies the reinforcing effects of a drug, whereas caffeine is consumed orally. In line with this, intravenous administration of caffeine increased the amount of self-administered cocaine, whereas drinking caffeine reduced it (Kuzmin et al. 2000).

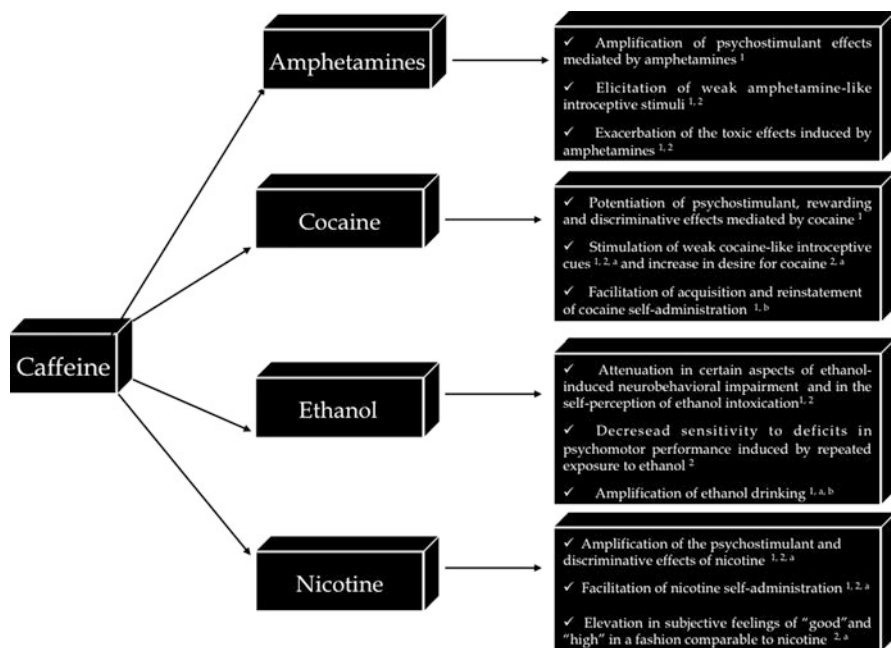
Regarding the role of methylxanthines in “chocolate addicts,” different issues should be considered. Chocolate is often consumed to relieve distress; however, the amount of cocoa consumed, although chronic, remains moderate. Chocolate addicts rarely display other addictive behaviors (Dallard et al. 2001) and do not seem to suffer from eating disorders, but may represent a population vulnerable to depression or anxiety. Methylxanthines contained in chocolate – caffeine and primarily theobromine – may contribute to the popularity of chocolate. However, their content is far too low to reach levels high enough to produce stimulation. Other attributes are probably much more important in determining chocolate’s special appeal and in explaining related self-reports of chocolate cravings and “chocoloholism” (Smit et al. 2004). Moreover, the reinforcing properties of chocolate, which are similar to those of palatable food (Maccioni et al. 2008), have never been demonstrated to depend on theobromine, which does not appear to have abuse potential.

Several studies have investigated the biochemical basis of methylxanthine abuse potential and, similar to behavioral studies, concluded that methylxanthines, and particularly caffeine, differ profoundly from other psychostimulants recognized to have abuse potential. All these studies suggest that functional interactions between adenosine and dopamine receptors, whose activation is critically involved in promoting psychostimulation, are crucial to caffeine-elicited abuse effects (Ferré et al. 1997). Interactions between adenosine and neurotransmitters other than dopamine, such as glutamate, serotonin, and acetylcholine, may be also important to caffeine-induced central effects. In particular, adenosine–glutamate interactions have been suggested to participate in caffeine-elicited psychostimulation (Ferré 2008).

Particularly important to this subject are the studies examining the response in dopamine release and cerebral glucose utilization in the shell of the nucleus accumbens, an area deeply involved in addictive properties of drugs (Di Chiara 2002). Caffeine – differently from drugs having overt abuse potential such as cocaine, amphetamine, and nicotine – increases dopamine release or glucose utilization in the nucleus accumbens shell only at high doses, while it increases dopamine release in the medial prefrontal cortex at doses corresponding to those assumed to result from its recreational consumption (Nehlig and Boyet 2000; Quarta et al. 2004; De Luca et al. 2007).

## 5 Interactions Between Methylxanthines and Other Psychoactive Drugs

Several investigations in experimental animals have demonstrated the ability of methylxanthines, and in particular of caffeine, to modulate the psychopharmacological effects of many psychoactive substances, including drugs of abuse (Tuazon et al. 1992; Shoaib et al 1999; Kunin et al. 2000; Gasior et al. 2002; Green and Schenk 2002) (Fig. 1). Similar effects of methylxanthines have also been observed in humans, and epidemiology studies have shown that caffeine consumption is often a correlate in drug dependence (Istvan and Matarazzo 1984; Kozlowski et al. 1993). On the basis of these findings, concerns have emerged about the possibility that intake of methylxanthines may facilitate either the development of dependence on other substances or relapse in former drug addicts. Such concerns have grown ever since the introduction to the market of the so-called “energy drinks”, which contain caffeine at quite high concentrations and are being increasingly consumed, often in combination with substances with abuse potential. This habit could favor the pharmacological interactions between caffeine and addictive substances,



**Fig. 1** General overview of the main pharmacological interactions between the methylxanthine caffeine and substances bearing abuse potential. *1* an effect observed in experimental animals, *2* an effect observed in humans, *a* an effect for which contradictory reports are present in the literature, *b* an effect observed in experimental animals and the occurrence of which is hypothesized to take place also in humans

potentially influencing the effects of the latter. Furthermore, unintentional consumption might also play a role in the interactions between methylxanthines and substances of abuse. Several chemical determinations have in fact demonstrated that methylxanthines are present at significant levels, as either contaminants or additives, in several illicit drugs (Fucci and De Giovanni 1998; Cheng et al. 2006).

Similarities and differences in the effects and interactions between methylxanthines and other psychoactive substances have both been described. Moreover, it has been ascertained that these often involve effects other than those related to dependence phenomena, potentially leading to harmful, unwanted consequences.

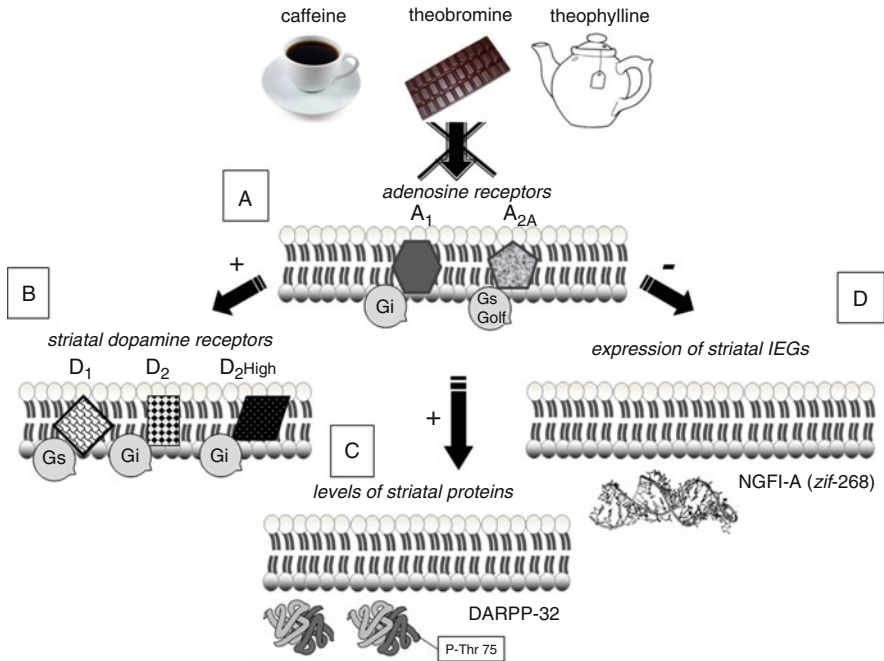
This part of the chapter summarizes current knowledge on the interactions between methylxanthines, in particular caffeine, and other psychoactive substances, including addictive, recreational, and prescribed drugs, describing the most representative ones. The implications of these interactions are discussed, with particular emphasis on drug dependence.

## 5.1 *Amphetamines*

Several studies have addressed the issue that methylxanthines and amphetamines may influence each other's actions.

Drug-discrimination studies performed in experimental animals have demonstrated that caffeine can, to some extent, engender interoceptive stimuli resembling those triggered by amphetamines (Holtzman 1986; Jain and Holtzman 2005). Such a similarity with amphetamines can also be envisioned for other methylxanthines, as some substances in this class are known to share the discriminative properties of caffeine (Carney et al. 1985). Methylxanthines can also potentiate the effects of amphetamines. However, while such a facilitatory influence does not seem to involve enhancement of the rewarding properties of amphetamines (Tuazon et al. 1992), it has nevertheless been observed for several effects, such as stimulation of motor activity, food-maintained operant behavior, and expression of immediate early genes (Jaszyna et al. 1998; Tronci et al. 2006). It must be emphasized here that the facilitation of amphetamine-mediated effects by methylxanthines observed in experimental animals can persist even in the absence of concomitant exposure to caffeine (Cauli et al. 2003; Tronci et al. 2006). Moreover, it is also worth recalling that evidence in experimental animals indicates that the amphetamine-like stimulus effect of caffeine does not undergo tolerance (Jain and Holtzman 2005). Taken together, the previous findings are particularly interesting when considering the possible role that consumption of methylxanthines may have in amphetamine abuse (Fig. 1).

Methylxanthines exert their central effects chiefly by antagonizing the  $A_1$  and  $A_{2A}$  receptors (Fredholm et al. 1999). It is well known that  $A_{2A}$  receptors and dopamine  $D_2$  receptors interact in opposite functional ways (Ferré et al. 1997). In line with this, investigations in rodents have clearly shown, on the one hand, that many of the behavioral and neurochemical effects of caffeine have a dopaminergic component, as dopamine receptor blockade can suppress them (Garrett and Griffiths 1997). On the



**Fig. 2** General overview of the molecular mechanisms involved in the interactions between methylxanthines and psychoactive substances having abuse potential. Methylxanthines act as competitive antagonists of both A<sub>1</sub> and A<sub>2A</sub> receptors (A). Such antagonism of adenosine receptors can potentiate striatal dopamine transmission by indirectly facilitating the signal mediated by D<sub>1</sub> and D<sub>2</sub> receptors and by elevating the relative proportion of high-affinity D<sub>2</sub> receptors (D<sub>2High</sub>) (B). Additional mechanisms of this pharmacological interactions related to methylxanthine-mediated antagonism of adenosine receptors may involve the elevation of the levels of dopamine- and cyclic-AMP-regulated 32-kDa phosphoprotein (DARPP-32), and of its Thr-75 phosphorylated isoform (C) and the depressed expression of the immediate early gene (IEG) NGFI-A (or *zif-268*) (D) in the corpus striatum. Effects on dopamine receptors, striatal proteins, and IEGs have chiefly been demonstrated for caffeine, but can also be hypothesized for other methylxanthines, on the basis of the antagonism of adenosine receptors these substances may exert

other hand, it has been determined that exposure to caffeine can engender facilitation of dopamine transmission in the corpus striatum of rodents. This effect has been shown to involve proteins such as dopamine- and cyclic-AMP-regulated 32-kDa phosphoprotein (Lindskog et al. 2002; Vaugeois 2002; Hsu et al. 2009a), immediate early genes such as nerve growth factor I-A also known as *zif-268* (Tronci et al. 2006), and high-affinity D<sub>2</sub> (D<sub>2High</sub>) receptors (Simola et al. 2008) (Fig. 2). Therefore, modulation of dopamine transmission could be an important mechanism underlying the interactions between amphetamines and methylxanthines, and it may also be involved in the influence methylxanthines exert on the effects of other psychostimulants (see Sect. 5.2).

Future studies should also focus on another major issue in methylxanthine–amphetamine interactions, namely, the ability of methylxanthines to enhance the

toxic effects of amphetamines. To date, this phenomenon has been characterized with regard to the interactions between caffeine and 3,4-methylenedioxymethamphetamine (also known as MDMA and ecstasy) and methamphetamine. In rodents, combined administration of caffeine powerfully exacerbates the toxic effects of these amphetamines (Delle Donne and Sonsalla 1994; McNamara et al. 2006).

Studies performed in humans have also evaluated interactions and similarities between methylxanthines and amphetamines. In this regard, drug-discrimination studies with caffeine are particularly interesting as they confirm what has been observed in experimental animals by showing that caffeine is capable of triggering amphetamine-like interoceptive stimuli, though with weaker intensity (Heishman and Henningfield 1992) (Fig. 1). Since interoceptive cues are an important component in drug abuse (Stolerman 1993), such an effect could potentially favor either the development or the relapse of an amphetamine dependence in individuals jointly consuming amphetamines and methylxanthines. Further investigation is necessary and should also address the potential additive toxicological interactions involving methylxanthines and amphetamines, as the occurrence of such a phenomenon has been reported in humans as well (Lambrecht et al. 1993).

## 5.2 Cocaine

Individuals addicted to cocaine often consume it in combination with various psychoactive substances, either licit or illicit, in what is defined as “polydrug use” (Leri et al. 2003). On the basis of the popularity of methylxanthines and of the similarity in the human consumption of amphetamines and cocaine, several studies have examined the interactions between methylxanthines and cocaine, and explored whether consumption of methylxanthines may influence cocaine abuse.

Investigations in experimental animals demonstrated that methylxanthines may exert effects that are to some degree similar to those triggered by cocaine and that methylxanthines can potentiate cocaine-mediated effects, although such a potentiation has not always been observed. Drug-discrimination experiments have demonstrated that rats trained to discriminate cocaine generalize to caffeine, although in an incomplete fashion, and that caffeine can amplify cocaine-mediated discriminative effects (Harland et al. 1989). A facilitatory influence of methylxanthines has also been described on other effects mediated by cocaine in rodents, such as motor stimulation and eliciting CPP (Schenk et al. 1990; Bedingfield et al. 1998). This latter finding appears interesting, as it suggests that methylxanthines could enhance cocaine-mediated reward. Most notably, evidence in rodents shows that methylxanthines dramatically influence cocaine self-administration. Thus, a faster acquisition of cocaine self-administration has been observed in rats exposed to chronic caffeine (Horger et al. 1991). Such a facilitatory influence of caffeine has also been observed in rhesus monkeys trained to self-administer smoked cocaine (Comer and Carroll 1996). Furthermore, both caffeine and theophylline

have been shown to efficiently reinstate cocaine self-administration in experienced rats in which this behavior was extinguished (Green and Schenk 2002) (Fig. 1). Although no consensus can be reached on the basis of the results of studies in experimental animals on the ability of methylxanthines to enhance cocaine-mediated reinforcement (Hogger et al. 1991; Kuzmin et al. 2000), the previous findings are of interest. In fact, the facilitation and reinstatement of cocaine self-administration by methylxanthines provides straightforward evidence that consumption of methylxanthines might be causally linked to an increase in cocaine intake.

Akin to what is observed for amphetamines, studies in experimental animals have indicated that methylxanthines might amplify cocaine-mediated effects by facilitating dopamine transmission (Green and Schenk 2002). Hence, the elevation in the level of D<sub>2</sub>High receptors in the striatum of rats exposed to caffeine for a long time could be an important mechanism (Simola et al. 2008), since an increase in the level of D<sub>2</sub>High receptors has also been observed in rats trained to self-administer cocaine (Briand et al. 2008) (Fig. 2). In this regard, it has, however, to be mentioned that the existence of two interchangeable affinity states for the dopamine D<sub>2</sub> receptors (D<sub>2</sub>High and low-affinity D<sub>2</sub> receptors, having high and low affinity for dopamine, respectively) has been clearly demonstrated only by *in vitro* experiments (George et al. 1985). On the other hand, whether such a phenomenon exists also *in vivo* is still controversial (McCormick et al. 2008), although evidence supporting this hypothesis has recently been obtained (Seeman 2009).

In line with the findings described above, investigations in humans have revealed the existence of similarities in the effects and reciprocal interactions between methylxanthines and cocaine. Cocaine addicts may consume more caffeine than the general population, suggesting a causal link could underlie the combined intake of these substances (Budney et al. 1993). Moreover, caffeine has been shown to elicit subjective effects resembling those of cocaine, although in a weaker fashion, and to increase cocaine “wanting” in cocaine abusers (Rush et al. 1995; Oliveto et al. 1998). These effects of caffeine, however, could not always be replicated (Liguori et al. 1997) (Fig. 1). It should be noted, however, that human studies addressing methylxanthine–cocaine interactions have often been performed under strikingly discrepant experimental conditions with respect to the procedures used and the participants recruited (e.g., former abusers, abusers, naïve individuals). Such methodological differences may have greatly influenced the outcomes observed in different investigations. Therefore, although conclusive evidence has not been obtained demonstrating that consumption of methylxanthines can promote cocaine abuse in humans, data demonstrating that methylxanthines can amplify certain effects of cocaine deserve full consideration. It is worth mentioning that development and relapse of cocaine dependence are powerfully modulated by environmental and pharmacological cues, including the consumption of psychoactive substances exerting subjective effects resembling those of cocaine (Spealman et al. 1999). Therefore, the ability of methylxanthines to engender cocaine-like interoceptive stimuli may have particular relevance to cocaine abuse.

### 5.3 *Cannabis Derivates*

Although cannabis is one of the most popular drugs of abuse, little is known on the pharmacological interactions between methylxanthines and cannabis derivatives, such as marijuana and hashish, and on the implications such interactions might have on drug dependence.

Studies in humans have shown that teenagers and college students often consume caffeine together with marijuana (Bernstein et al. 2002; Miller 2008). However, whether caffeine might be a gateway drug for marijuana dependence has not been proven.

Studies in experimental animals support this view, with evidence indicating that a functional interaction exists between  $A_{2A}$  and cannabinoid  $CB_1$  receptors. Such an interaction takes place at both the neurochemical level (as a dependence of  $CB_1$ -mediated transmission on  $A_{2A}$  receptors has been reported in cotransfected cell lines) and the behavioral level (as antagonism of  $A_{2A}$  receptors counteracts the motor-depressant effects mediated by  $CB_1$  receptor stimulation in experimental animals) (Carriba et al. 2007). Recent investigations support a role for adenosine transmission, and thus methylxanthines, in cannabis abuse. Studies in mice have shown that  $A_{2A}$  receptors are involved in rewarding effects and physical dependence induced by  $\Delta^9$ -tetrahydrocannabinol (Soria et al. 2004). Moreover, long-term exposure to caffeine has been shown to facilitate  $CB_1$ -mediated transmission in the rat corpus striatum (Rossi et al. 2009). On the basis of these findings, it is reasonable to hypothesize that consumption of caffeine, and/or other methylxanthines, may also have some relevance to cannabis use in humans.

### 5.4 *Ethanol*

Epidemiology studies have shown that a positive correlation may exist between the consumption of caffeine and that of ethanol (Istvan and Matarazzo 1984; Kozlowski et al. 1993). Moreover, anecdotal reports describing caffeine as a “hangover helper” have long provided an empirical justification for the combined consumption of these substances. A wealth of studies in both experimental animals and humans have been performed to elucidate the features of methylxanthine–ethanol interactions. This issue has been attracting even greater interest ever since the introduction to the market of the highly caffeinated “energy drinks.” In fact, such beverages are increasingly being consumed in combination with ethanol, and evidence exists that this habit is often associated with health-threatening consequences (Marczinski and Fillmore 2006; O’Brien et al. 2008).

In experimental animals, both caffeine and theophylline ameliorate several behavioral parameters indicative of ethanol intoxication (Dar et al. 1987; Connole et al. 2004). Results showing a worsening of ethanol-induced impairment have, however, also been reported, depending on the specific methylxanthine and the



effect of ethanol considered (Kuribara and Tadokoro 1992). Therefore, such studies do not definitively clarify the precise effects of methylxanthines on ethanol intoxication, and suggest that when an amelioration of intoxication by methylxanthines exists, it is often narrowed to certain aspects of the phenomenon. In addition, experiments in rats have shown that caffeine can promote ethanol drinking (Kunin et al. 2000) (Fig. 1). This finding would provide a direct link between caffeine intake and ethanol consumption, although it must be acknowledged that such an effect of caffeine has not always been observed (Potthoff et al. 1983).

It is still not clear what molecular mechanism could underlie the stimulation of ethanol drinking by caffeine. The mechanisms at the basis of the counteraction caffeine may exert on effects related to ethanol intoxication, however, appear to be more defined. In particular, antagonism of  $A_1$  and  $A_{2A}$  receptors appears crucial to this action of caffeine, though the specific receptor involved varies according to the effect of ethanol considered. Hence,  $A_1$  receptors seem to mediate caffeine-elicited reversal of deficits in motor coordination induced by ethanol (Barwick and Dar 1998; Connole et al. 2004), and it is reasonable to think this may be because of the high enrichment of these receptors in areas governing motor coordination such as the cortex and cerebellum (Ribeiro et al. 2002). However, blockade of  $A_{2A}$  rather than  $A_1$  receptors seems to be involved in caffeine-mediated counteraction of hypnosis induced by ethanol (El Yacoubi et al. 2003).

Studies in humans examining methylxanthine–ethanol interactions have mostly focused on the influence caffeine exerts on ethanol intoxication, and have yielded mixed results (Liguori and Robinson 2001; Drake et al. 2003). Nevertheless, it is worth mentioning that these studies converge on the point that caffeine consumed in association with ethanol, rather than improving ethanol-induced impairments, would reduce the self-perception of ethanol intoxication (Fig. 1). This has been suggested to be a major risk associated with joint ethanol–caffeine consumption, since an altered perception of one’s psychophysical integrity could lead to the performance of hazardous activities. Notably, this view has received support from a recent investigation showing an increase in the performance of risky behaviors (e.g., driving under ethanol intoxication) by college students who reported combined drinking of caffeinated “energy drinks” and ethanol (O’Brien et al. 2008).

As previously mentioned, epidemiology studies suggest the existence of a positive correlation between the consumption of caffeine and the consumption of ethanol. Interestingly, caffeine can promote ethanol drinking in rats (Kunin et al. 2000), whereas human data show that caffeine enhances tolerance to ethanol (Fillmore 2003), the onset of which is critical to ethanol dependence (DSM IV). Hence, it can be hypothesized that a caffeine-mediated increase in ethanol tolerance could promote the escalation of ethanol consumption. On the other hand, the studies in ethanol consumers demonstrating that caffeine reduces the feelings of ethanol intoxication seem to suggest that joint ethanol–caffeine drinking might reflect a sort of self-medication, in which caffeine serves to counteract the adverse effects - of ethanol (Grattan-Miscio and Vogel-Sprott 2005). To date, little is known on the interactions between ethanol and methylxanthines other than caffeine. Investigation of this issue appears warranted, on the basis of studies in experimental



animals showing that theophylline may interact with ethanol in a different way from caffeine (Dar et al. 1987).

## 5.5 *Nicotine*

A positive relationship between the consumption of nicotine in the form of tobacco smoking and caffeine in the form of drinking coffee has long been described by anecdotal reports and more recently by epidemiology studies (Istvan and Matarazzo 1984; Swanson et al. 1994). Several investigations have been performed to elucidate the features of this habit; however, very scarce information is available on the interactions between nicotine and methylxanthines other than caffeine.

Studies in experimental animals have demonstrated that caffeine can amplify many behavioral effects of nicotine (Shoaib et al. 1999; Gasior et al. 2002; Celik et al. 2006). The results obtained from drug-discrimination and self-administration experiments are particularly interesting and relevant to nicotine abuse. Although it does not engender nicotine-like subjective cues, caffeine has been shown to amplify the discriminative effects of nicotine when given in joint administration (Gasior et al. 2002). Moreover, a faster acquisition of nicotine self-administration has been reported in rats chronically exposed to caffeine, suggesting that caffeine may potentiate the reinforcing effects of nicotine (Shoaib et al. 1999) (Fig. 1). Notably, the regimen of caffeine exposure used in these studies was found not to significantly affect the pharmacokinetics of nicotine (Gasior et al. 2002). This, therefore, suggests that caffeine facilitates the effects of nicotine through pharmacodynamic mechanisms, and lends support for caffeine consumption being a risky habit for nicotine abuse (Gasior et al. 2002).

The precise molecular mechanisms through which caffeine amplifies nicotine-mediated effects are not yet completely understood. Caffeine is known to affect the dopaminergic component of the nicotine discriminative stimulus in the rat (Gasior et al. 1999), and so it is reasonable to hypothesize the involvement of the dopaminergic system in caffeine–nicotine interactions. Although caffeine and nicotine preferentially target adenosine and nicotinic receptors, respectively, one should bear in mind that both drugs have a very complex pharmacological profile (Fredholm et al. 1999; Barik and Wonnacott 2009). Therefore, different neurotransmitters could be involved in caffeine–nicotine interactions.

Studies in humans have extended preclinical findings by ascertaining both similarities and additive interactions between caffeine and nicotine. It has been reported that caffeine and nicotine elevate subjective ratings of “good effect”, liking, and “high” in a comparable fashion (Garrett and Griffiths 2001). Moreover, caffeine has been found to potentiate the stimulant and reinforcing effects of nicotine, and it has been observed that some subjective effects of caffeine, such as feelings of comfort, may be more marked in smokers (Perkins et al. 2001; Jones and Griffiths 2003). However, differences between the subjective effects of caffeine and nicotine have also been reported (Garrett and Griffiths 2001). Further,

potentiation of nicotine-reinforcing effects by caffeine has not always been observed (Perkins et al. 2005; Blank et al. 2007) (Fig. 1). It is worth noting that the use of different methodological approaches may have affected the outcomes of these studies. For example, the smoking history of the individuals investigated is known to critically influence caffeine–nicotine interactions, as long-term nicotine use may elicit tolerance to some of its effects (Sobel et al. 2004). Similarly, other factors, such as caffeine history, route of administration of either substance, drug use, and the specific effects evaluated, are known to influence the features of caffeine–nicotine interactions (Garrett and Griffiths 2001; Jones and Griffiths 2003). Mechanisms other than pharmacological potentiation of nicotine effects have also been proposed to underlie joint consumption of caffeine and nicotine. On the basis of the evidence that smoking hastens caffeine metabolism in humans, it has been suggested that high caffeine intake by smokers could stem from a self-adjustment of the caffeine dose owing to its increased metabolism (Benowitz et al. 1989). Avoiding nicotine abstinence has also been taken into question, as caffeine counteracts some symptoms of nicotine withdrawal (Cohen et al. 1994; Sobel et al. 2004). Finally, other studies hypothesize that joint consumption of caffeine and nicotine might arise from personality traits, rather than pharmacological interactions between the substances (Gurpegui et al. 2007).

## 5.6 *Methylphenidate*

Pharmacological similarities and interactions between methylxanthines and methylphenidate have been described.

Studies in experimental animals have shown, on the one hand, that caffeine amplifies the motor-stimulant effects of methylphenidate through a dopaminergic mechanism (Boeck et al. 2009) and, on the other hand, that caffeine and methylphenidate share discriminative properties (Holtzman 1986).

In human studies, the ability of caffeine to engender methylphenidate-like introceptive cues has been replicated (Oliveto et al. 1993). Together with preclinical data, this finding suggests that consumption of caffeine might influence and potentially amplify the effects of methylphenidate in humans. This consideration is particularly relevant as methylphenidate is being increasingly used without medical prescription as a performance enhancer, chiefly by college students (Dupont et al. 2008), and individuals who improperly use methylphenidate are at higher risk for abusing other drugs and performing risky behavior (McCabe et al. 2005).

## 5.7 *Antagonists of the Glutamate NMDA Receptors*

Substances acting as antagonists of glutamate NMDA receptors, such as phencyclidine and its derivative ketamine, have long been used as recreational drugs

(Wolff and Winstock 2006); however, systematic studies evaluating their effects in humans are lacking.

Behavioral studies in experimental animals have observed a modulation of NMDA-mediated transmission by methylxanthines (de Oliveira et al. 2005). In line with this, caffeine has been reported to potentiate the psychostimulant effects of ketamine and to exert additive effects with phencyclidine in rodents (Powell and Holtzman 1998; Hsu et al. 2009b). Interestingly, in the rat, caffeine has been found to potentiate the rewarding effects elicited by NMDA receptor blockade, suggesting that caffeine consumption might impact the addictive properties of antagonists at such receptors (Bespalov et al. 2006), although to date this has not been proven conclusively. Nevertheless, and irrespective of drug abuse, the combined consumption of methylxanthines and NMDA receptor antagonists, in particular ketamine, appears inadvisable. In fact, studies in experimental animals have demonstrated that caffeine can boost the toxic and lethal effects of ketamine (Hsu et al. 2009b), suggesting that joint consumption of these substances can be potentially dangerous.

## 5.8 Opiates

It is not clear whether there is a correlation between intake of methylxanthines and opiate abuse, and hardly any evidence is available on this issue.

Studies performed in experimental animals have demonstrated a potentiation of morphine-stimulated locomotion by acute caffeine in mice; however, an inversion of this effect has been observed over time (Kuribara 1995; Weisberg and Kaplan 1999). Mixed findings have also been reported with regard to the way methylxanthines influence the rewarding/reinforcing properties of opiates. Theophylline has been reported to attenuate morphine-induced CPP in rats, suggesting that this methylxanthine could blunt the rewarding properties of morphine (Sahraei et al. 2006). Theophylline has also been found to affect morphine self-administration in rats, suggesting an influence on the reinforcing properties of opiates. However, such an effect appears very complex, as theophylline has been reported to either facilitate or depress morphine self-administration (Sahraei et al. 1999). Interestingly, a depressant influence on morphine self-administration has also been observed in rats treated with caffeine (Sudakov et al. 2003).

Both  $A_1$  and  $A_{2A}$  receptors seem to participate in the interactions between methylxanthines and opiates. The specific subtype involved, however, depends on the particular effect of the opiates considered, and this might explain the mixed outcomes from studies in experimental animals. In fact, while the  $A_1$  receptors are mostly involved in opiate-stimulated locomotion, the  $A_{2A}$  receptors have a major role in opiate reinforcing properties (Sahraei et al. 1999; Weisberg and Kaplan 1999). Moreover, a role for dopamine transmission in the modulation of opiate effects by methylxanthines has been suggested (Kuribara 1995).

The data in experimental animals summarized above do not account for an overt facilitatory influence of methylxanthines on opiate-induced effects, and accordingly

do not provide support for the consumption of methylxanthines being a major risk for opiate abuse in humans. Nevertheless, it is worth mentioning that, in experimental animals, blockade of adenosine transmission can precipitate the symptoms of opiate withdrawal (Khalili et al. 2001; Stella et al. 2003).

## 5.9 *Anxiolytic–Hypnotics*

Anxiolytic–hypnotics, including benzodiazepines and barbiturates, are often used without medical prescription or for a time longer than is required for them to exert their therapeutic effects (Licata and Rowlett 2008). Several investigations have examined the pharmacological interactions between methylxanthines and these compounds.

Investigations in experimental animals have shown that methylxanthines and anxiolytic–hypnotics interact at the pharmacokinetic level (Lau and Wang 1996). Conversely, negligible pharmacodynamic interactions seem to take place between these substances (Lau et al. 1997).

The findings of studies in humans are in line with what has been reported in experimental animals, showing the existence of pharmacokinetic interactions between methylxanthines and benzodiazepines and the occurrence of mixed pharmacodynamic effects following the joint consumption of these substances (Roache and Griffiths 1987; Cysneiros et al. 2007).

It has been reported that the improper use of benzodiazepines may be associated with heavy caffeine consumption (Lekka et al. 1997), but conflicting data on this issue also exist (Cooper et al. 2004). Notably, even when such an association was observed, it was not possible to ascertain whether caffeine consumption triggered the use of benzodiazepines or whether caffeine was consumed as self-medication to counteract some unwanted effect of benzodiazepines (Lekka et al. 1997). Similarly, hardly any data exist that suggest a causal correlation between consumption of methylxanthines and abuse of barbiturates. Therefore, on the basis of current evidence, consumption of methylxanthines does not appear to be a major risk factor for the improper use of anxiolytic–hypnotic drugs.

## 5.10 *Miscellaneous Psychoactive Drugs*

Studies in experimental animals have demonstrated that, in addition to the substances described already, methylxanthines can interact with diverse psychoactive drugs, such as antipsychotics, antidepressants, phenylethylamines, and volatile solvents (Young et al. 1998; Chan and Chen 2003; Enríquez-Castillo et al. 2008; Varty et al. 2008). The implications of such interactions in terms of drug dependence, however, appear very limited.

## 6 General Considerations on Methylxanthines and Drug Dependence Phenomena

The data summarized in this chapter indicate that consumption of methylxanthines, such as caffeine, may promote the onset of dependence – although the features of such dependence appear less marked than those typical of dependence on other psychoactive substances such as psychostimulants, ethanol, nicotine, and opiates. Dependence on caffeine is generally compatible with social and productive life, although, as suggested by studies in experimental animals, it might carry a risk of favoring the establishment of a dependence on other substances. In fact, as described already, several pieces of evidence demonstrate that methylxanthines can amplify the effects of diverse addictive substances, potentially leading to an increase in their liability for abuse. Such a potential influence of methylxanthines on dependence phenomena appears particularly relevant to cocaine and nicotine abuse. In addition to this, consumption of methylxanthines has been shown to be associated with the intake of addictive substances even without overtly influencing their abuse liability. This phenomenon appears evident with ethanol, the consumption of which is likely promoted by the methylxanthine caffeine by means of a mechanism involving a reduced self-perception of ethanol intoxication. Interactions between methylxanthines and other psychoactive substances appear more complex, and their relevance to drug dependence is often undetermined. In this regard, a potential facilitatory influence of methylxanthines on the addictive effects of drugs such as amphetamines, methylphenidate, and cannabis derivatives is postulated, but not conclusively proven. An issue of particular relevance when examining the role played by methylxanthines in dependence phenomena is that adenosine receptors, to which methylxanthines bind with high affinity, can interact with several neurochemical pathways involved in the effects of addictive psychoactive substances. Of paramount interest in this regard is the finding that methylxanthines, and in particular caffeine, may interact with the dopaminergic system, which plays a crucial role in drug-dependence phenomena (Fig. 2).

In conclusion, data in this chapter indicate that caution should be exercised in the combined consumption of methylxanthines and other psychoactive substances. In fact, although consumption of methylxanthines does not seem to be particularly harmful per se, it could nevertheless result in modification of the effects of other psychoactive substances, and this might have implications in terms of both drug dependence and drug-induced toxicity.

## References

- American Psychiatric Association (1994) Diagnostic and statistical manual of mental disorders, 4th edn. American Psychiatric Association, Washington
- Barik J, Wonnacott S (2009) Molecular and cellular mechanisms of action of nicotine in the CNS. *Handb Exp Pharmacol* 192:173–207

- Barwick VS, Dar MS (1998) Adenosinergic modulation of ethanol-induced motor incoordination in the rat motor cortex. *Prog Neuropsychopharmacol Biol Psychiatry* 22:587–607
- Bedingfield JB, King DA, Holloway FA (1998) Cocaine and caffeine: conditioned place preference, locomotor activity, and additivity. *Pharmacol Biochem Behav* 61:291–296
- Benowitz NL, Hall SM, Modin G (1989) Persistent increase in caffeine concentrations in people who stop smoking. *BMJ* 298:1075–1076
- Benton D (2004) The biology and psychology of chocolate craving. In: Nehlig A (ed) *Coffee, tea chocolate and the brain*. CRC, Boca Raton
- Bernstein GA, Carroll ME, Thuras PD et al (2002) Caffeine dependence in teenagers. *Drug Alcohol Depend* 66:1–6
- Bespalov A, Dravolina O, Belozertseva I et al (2006) Lowered brain stimulation reward thresholds in rats treated with a combination of caffeine and N-methyl-D-aspartate but not alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionate or metabotropic glutamate receptor-5 receptor antagonists. *Behav Pharmacol* 17:295–302
- Blank MD, Kleykamp BA, Jennings JM et al (2007) Caffeine's influence on nicotine's effects in nonsmokers. *Am J Health Behav* 31:473–483
- Boeck CR, Marques VB, Valvassori SS et al (2009) Early long-term exposure with caffeine induces cross-sensitization to methylphenidate with involvement of DARPP-32 in adulthood of rats. *Neurochem Int* 55:318–322
- Briand LA, Flagel SB, Seeman P et al (2008) Cocaine self-administration produces a persistent increase in dopamine D2 High receptors. *Eur Neuropsychopharmacol* 18:551–556
- Brice CF, Smith AP (2002) Factors associated with caffeine consumption. *Int J Food Sci Nutr* 53:55–64
- Budney AJ, Higgins ST, Hughes JR et al (1993) Nicotine and caffeine use in cocaine-dependent individuals. *J Subst Abuse* 5:117–130
- Carney JM (1982) Effects of caffeine, theophylline and theobromine on scheduled controlled responding in rats. *Br J Pharmacol* 75:451–454
- Carney JM, Holloway FA, Modrow HE (1985) Discriminative stimulus properties of methylxanthines and their metabolites in rats. *Life Sci* 36:913–920
- Carriba P, Ortiz O, Patkar K et al (2007) Striatal adenosine A2A and cannabinoid CB1 receptors form functional heteromeric complexes that mediate the motor effects of cannabinoids. *Neuropsychopharmacology* 32:2249–2259
- Cauli O, Pinna A, Valentini V et al (2003) Subchronic caffeine exposure induces sensitization to caffeine and cross-sensitization to amphetamine ipsilateral turning behavior independent from dopamine release. *Neuropsychopharmacology* 28:1752–1759
- Celik E, Uzbay IT, Karakas S (2006) Caffeine and amphetamine produce cross-sensitization to nicotine-induced locomotor activity in mice. *Prog Neuropsychopharmacol Biol Psychiatry* 30:50–55
- Chan MH, Chen HH (2003) Toluene exposure increases aminophylline-induced seizure susceptibility in mice. *Toxicol Appl Pharmacol* 193:303–308
- Cheng JY, Chan MF, Chan TW et al (2006) Impurity profiling of ecstasy tablets seized in Hong Kong by gas chromatography-mass spectrometry. *Forensic Sci Int* 162:87–94
- Cohen C, Pickworth WB, Bunker EB et al (1994) Caffeine antagonizes EEG effects of tobacco withdrawal. *Pharmacol Biochem Behav* 47:919–936
- Comer SD, Carroll ME (1996) Oral caffeine pretreatment produced modest increases in smoked cocaine self-administration in rhesus monkeys. *Psychopharmacology* 126:281–285
- Comer SD, Haney M, Foltin RW et al (1997) Effects of caffeine withdrawal on humans living in a residential laboratory. *Exp Clin Psychopharmacol* 5:399–403
- Connole L, Harkin A, Maginn M (2004) Adenosine A1 receptor blockade mimics caffeine's attenuation of ethanol-induced motor incoordination. *Basic Clin Pharmacol Toxicol* 95:299–304
- Cooper M, Safran M, Eberhardt M (2004) Caffeine consumption among adults on benzodiazepine therapy: United States 1988–1994. *Psychol Rep* 95:183–191

- Couturier EG, Laman DM, van Duijn MA et al (1997) Influence of caffeine and caffeine withdrawal on headache and cerebral blood flow velocities. *Cephalalgia* 17:188–190
- Cysneiros RM, Farkas D, Harmatz JS et al (2007) Pharmacokinetic and pharmacodynamic interactions between zolpidem and caffeine. *Clin Pharmacol Ther* 82:54–62
- Dallard I, Cathebras P, Sauron C et al (2001) Is cocoa a psychotropic drug? Psychopathologic study of a population of subjects self-identified as chocolate addicts. *Encephale* 27:181–186
- Daly JW, Fredholm BB (1998) Caffeine—an atypical drug of dependence. *Drug Alcohol Depend* 51:199–206
- Dar MS, Jones M, Close G et al (1987) Behavioral interactions of ethanol and methylxanthines. *Psychopharmacology* 91:1–4
- Delle Donne KT, Sonsalla PK (1994) Protection against methamphetamine-induced neurotoxicity to neostriatal dopaminergic neurons by adenosine receptor activation. *J Pharmacol Exp Ther* 271:1320–1326
- De Luca MA, Bassareo V, Bauer A et al (2007) Caffeine and accumbens shell dopamine. *J Neurochem* 103:157–163
- de Oliveira RV, Dall'Igna OP, Tort AB et al (2005) Effect of subchronic caffeine treatment on MK-801-induced changes in locomotion, cognition and ataxia in mice. *Behav Pharmacol* 16:79–84
- Di Chiara G (2002) Nucleus accumbens shell and core dopamine: differential role in behavior and addiction. *Behav Brain Res* 137:75–114
- Drake CL, Roehrs T, Turner L et al (2003) Caffeine reversal of ethanol effects on the multiple sleep latency test, memory, and psychomotor performance. *Neuropsychopharmacology* 28:371–378
- Dupont RL, Coleman JJ, Bucher RH et al (2008) Characteristics and motives of college students who engage in nonmedical use of methylphenidate. *Am J Addict* 17:167–171
- El Yacoubi M, Ledent C, Parmentier M et al (2003) Caffeine reduces hypnotic effects of alcohol through adenosine A2A receptor blockade. *Neuropharmacology* 45:977–985
- Enríquez-Castillo A, Alamilla J, Barral J et al (2008) Differential effects of caffeine on the antidepressant-like effect of amitriptyline in female rat subpopulations with low and high immobility in the forced swimming test. *Physiol Behav* 94:501–509
- Ferré S (2008) An update on the mechanisms of the psychostimulant effects of caffeine. *J Neurochem* 105:1067–1079
- Ferré S, Fredholm BB, Morelli M et al (1997) Adenosine-dopamine receptor-receptor interactions as an integrative mechanism in the basal ganglia. *Trends Neurosci* 20:482–487
- Fillmore MT (2003) Alcohol tolerance in humans is enhanced by prior caffeine antagonism of alcohol-induced impairment. *Exp Clin Psychopharmacol* 11:9–17
- Finn IB, Holtzman SG (1988) Tolerance and cross-tolerance to theophylline-induced stimulation of locomotor activity in rats. *Life Sci* 42:2475–2482
- Fredholm BB, Bättig K, Holmén J et al (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51:83–133
- Fucci N, De Giovanni N (1998) Adulterants encountered in the illicit cocaine market. *Forensic Sci Int* 95:247–252
- Garrett BE, Griffiths RR (1997) The role of dopamine in the behavioral effects of caffeine in animals and humans. *Pharmacol Biochem Behav* 57:533–541
- Garrett BE, Griffiths RR (2001) Intravenous nicotine and caffeine: subjective and physiological effects in cocaine abusers. *J Pharmacol Exp Ther* 296:486–494
- Gasior M, Jaszyna M, Munzar P et al (2002) Caffeine potentiates the discriminative-stimulus effects of nicotine in rats. *Psychopharmacology* 162:385–395
- Gasior M, Shoaib M, Yasar S et al (1999) Acquisition of nicotine discrimination and discriminative stimulus effects of nicotine in rats chronically exposed to caffeine. *J Pharmacol Exp Ther* 288:1053–1073
- George SR, Watanabe M, Di Paolo T et al (1985) The functional state of the dopamine receptor in the anterior pituitary is in the high affinity form. *Endocrinology* 117:690–697

- Georgiev V, Johansson B, Fredholm BB (1993) Long-term caffeine treatment leads to a decreased susceptibility to NMDA-induced clonic seizures in mice without changes in adenosine A1 receptor number. *Brain Res* 612:271–277
- Gilliland K, Bullock W (1984) Caffeine: a potential drug of abuse. *Adv Alcohol Subst Abuse* 3:53–73
- Grattan-Miscio KE, Vogel-Sprott M (2005) Alcohol, intentional control, and inappropriate behavior: regulation by caffeine or an incentive. *Exp Clin Psychopharmacol* 13:48–55
- Green TA, Schenk S (2002) Dopaminergic mechanism for caffeine-produced cocaine seeking in rats. *Neuropsychopharmacology* 26:422–430
- Griffiths RR, Evans SM, Heishman SJ et al (1990) Low-dose caffeine physical dependence in humans. *J Pharmacol Exp Ther* 255:1123–1132
- Griffiths RR, Mumford GK (1995) Caffeine: a drug of abuse? In: Bloom FE, Kupfer DJ (eds) *Psychopharmacology: the fourth generation of progress*. Raven, New York
- Gurpegui M, Jurado D, Luna JD et al (2007) Personality traits associated with caffeine intake and smoking. *Prog Neuropsychopharmacol Biol Psychiatry* 31:997–1005
- Harland RD, Gauvin DV, Michaelis RC et al (1989) Behavioral interaction between cocaine and caffeine: a drug discrimination analysis in rats. *Pharmacol Biochem Behav* 32:1017–1023
- Heishman SJ, Henningfield JE (1992) Stimulus functions of caffeine in humans: relation to dependence potential. *Neurosci Biobehav Rev* 16:273–287
- Holtzman SG (1986) Discriminative stimulus properties of caffeine in the rat: noradrenergic mediation. *J Pharmacol Exp Ther* 239:706–714
- Holtzman SG (1996) Discriminative effects of CGS 15943, a competitive adenosine receptor antagonist, in monkeys: comparison to methylxanthines. *J Pharmacol Exp Ther* 277:739–746
- Horger BA, Wellman PJ, Morien A et al (1991) Caffeine exposure sensitizes rats to the reinforcing effects of cocaine. *Neuroreport* 2:53–56
- Hsu CW, Chen CY, Wang CS et al (2009a) Caffeine and a selective adenosine A2A receptor antagonist induce reward and sensitization behavior associated with increased phospho-Thr75-DARPP-32 in mice. *Psychopharmacology* 204:313–325
- Hsu HR, Mei YY, Wu CY et al (2009b) Behavioural and toxic interaction profile of ketamine in combination with caffeine. *Basic Clin Pharmacol Toxicol* 104:379–383
- Istvan J, Matarazzo JD (1984) Tobacco, alcohol, and caffeine use: a review of their interrelationships. *Psychol Bull* 95:301–326
- Jain R, Holtzman SG (2005) Caffeine induces differential cross tolerance to the amphetamine-like discriminative stimulus effects of dopaminergic agonists. *Brain Res Bull* 65:415–421
- Jaszyna M, Gasior M, Shoab M et al (1998) Behavioral effects of nicotine, amphetamine and cocaine under a fixed-interval schedule of food reinforcement in rats chronically exposed to caffeine. *Psychopharmacology* 140:257–271
- Jones HE, Griffiths RR (2003) Oral caffeine maintenance potentiates the reinforcing and stimulant subjective effects of intravenous nicotine in cigarette smokers. *Psychopharmacology* 165:280–290
- Kaplan GB, Greenblatt DJ, Kent MA et al (1993) Caffeine treatment and withdrawal in mice: relationships between dosage, concentrations, locomotor activity and A1 adenosine receptor binding. *J Pharmacol Exp Ther* 266:1563–1572
- Khalili M, Semnani S, Fathollahi Y (2001) Caffeine increases paraventricular neuronal firing rate and induces withdrawal signs in morphine-dependent rats. *Eur J Pharmacol* 412:239–245
- Kidney J, Dominguez M, Taylor PM et al (1995) Immunomodulation by theophylline in asthma. Demonstration by withdrawal of therapy. *Am J Respir Crit Care Med* 151:1907–1914
- Kozlowski LT, Henningfield JE, Keenan RM et al (1993) Patterns of alcohol, cigarette, and caffeine and other drug use in two drug abusing populations. *J Subst Abuse Treat* 10:171–179
- Kunin D, Gaskin S, Rogan F et al (2000) Caffeine promotes ethanol drinking in rats. Examination using a limited-access free choice paradigm. *Alcohol* 21:271–277



- Kuribara H (1995) Caffeine enhances acute stimulant effect of morphine but inhibits morphine sensitization when assessed by ambulation of mice. *Prog Neuropsychopharmacol Biol Psychiatry* 19:313–321
- Kuribara H, Tadokoro S (1992) Caffeine does not effectively ameliorate, but rather may worsen the ethanol intoxication when assessed by discrete avoidance in mice. *Jpn J Pharmacol* 59:393–398
- Kuzmin A, Johansson B, Semenova S et al (2000) Differences in the effect of chronic and acute caffeine on self-administration of cocaine in mice. *Eur J Neurosci* 12:3026–3032
- Lambrecht GL, Malbrain ML, Chew SL et al (1993) Intranasal caffeine and amphetamine causing stroke. *Acta Neurol Belg* 93:146–149
- Lau CE, Falk JL (1995) Dose-dependent surmountability of locomotor activity in caffeine tolerance. *Pharmacol Biochem Behav* 52:139–143
- Lau CE, Wang J (1996) Alprazolam, caffeine and their interaction: relating DRL performance to pharmacokinetics. *Psychopharmacology* 126:115–124
- Lau CE, Wang Y, Falk JL (1997) Differential reinforcement of low rate performance, pharmacokinetics and pharmacokinetic-pharmacodynamic modeling: independent interaction of alprazolam and caffeine. *J Pharmacol Exp Ther* 281:1013–1029
- Lekka NP, Paschalis C, Beratis S (1997) Nicotine, caffeine and alcohol use in high- and low-dose benzodiazepine users. *Drug Alcohol Depend* 45:207–212
- Leri F, Bruneau J, Stewart J (2003) Understanding polydrug use: review of heroin and cocaine co-use. *Addiction* 98:7–22
- Licata SC, Rowlett JK (2008) Abuse and dependence liability of benzodiazepine-type drugs: GABA(A) receptor modulation and beyond. *Pharmacol Biochem Behav* 90:74–89
- Liguori A, Hughes JR, Goldberg K et al (1997) Subjective effects of oral caffeine in formerly cocaine-dependent humans. *Drug Alcohol Depend* 49:17–24
- Liguori A, Robinson JH (2001) Caffeine antagonism of alcohol-induced driving impairment. *Drug Alcohol Depend* 63:123–129
- Lindskog M, Svenningsson P, Pozzi L et al (2002) Involvement of DARPP-32 phosphorylation in the stimulant action of caffeine. *Nature* 418:774–778
- Maccioni P, Pes D, Carai MA et al (2008) Suppression by the cannabinoid CB1 receptor antagonist, rimonabant, of the reinforcing and motivational properties of a chocolate-flavoured beverage in rats. *Behav Pharmacol* 19:197–209
- Marczinski CA, Fillmore MT (2006) Clubgoers and their trendy cocktails: implications of mixing caffeine into alcohol on information processing and subjective reports of intoxication. *Exp Clin Psychopharmacol* 14:450–458
- Martín I, López-Vílchez MA, Mur A et al (2007) Neonatal withdrawal syndrome after chronic maternal drinking of mate. *Ther Drug Monit* 29:127–129
- McCabe SE, Knight JR, Teter CJ (2005) Non-medical use of prescription stimulants among US college students: prevalence and correlates from a national survey. *Addiction* 100:96–106
- McCormick PN, Kapur S, Seeman P et al (2008) Dopamine D2 receptor radiotracers [(11C)(+)-PHNO and (3H)raclopride are indistinguishably inhibited by D2 agonists and antagonists ex vivo. *Nucl Med Biol* 35:11–17
- McGowan JD, Altman RE, Kanto WP (1988) Neonatal withdrawal symptoms after chronic maternal ingestion of caffeine. *South Med J* 81:1092–1094
- McNamara R, Kerans A, O'Neill B et al (2006) Caffeine promotes hyperthermia and serotonergic loss following co-administration of the substituted amphetamines, MDMA (“Ecstasy”) and MDA (“Love”). *Neuropharmacology* 50:69–80
- Miller KE (2008) Energy drinks, race, and problem behaviors among college students. *J Adolesc Health* 43:490–497
- Modrow HE, Holloway FA, Carney JM (1981) Caffeine discrimination in the rat. *Pharmacol Biochem Behav* 14:683–688
- Mumford GK, Neill DB, Holtzman SG (1988) Caffeine elevates reinforcement threshold for electrical brain stimulation: tolerance and withdrawal changes. *Brain Res* 459:163–167

- Nehlig A (1999) Are we dependent upon coffee and caffeine? A review on human and animal data. *Neurosci Biobehav Rev* 23:563–576
- Nehlig A (2004) Dependence upon coffee and caffeine: an update. In: Nehlig A (ed) *Coffee, tea, chocolate and the brain*. CRC, Boca Raton
- Nehlig A, Boyet S (2000) Dose-response study of caffeine effects on cerebral functional activity with a specific focus on dependence. *Brain Res* 858:71–77
- Nikodijević O, Jacobson KA, Daly JW (1993) Locomotor activity in mice during chronic treatment with caffeine and withdrawal. *Pharmacol Biochem Behav* 44:199–216
- O'Brien MC, McCoy TP, Rhodes SD et al (2008) Caffeinated cocktails: energy drink consumption, high-risk drinking, and alcohol-related consequences among college students. *Acad Emerg Med* 15:453–460
- Oliveto AH, Bickel WK, Hughes JR et al (1993) Pharmacological specificity of the caffeine discriminative stimulus in humans: effects of theophylline, methylphenidate and bupropion. *Behav Pharmacol* 4:237–246
- Oliveto AH, McCance-Katz E, Singha A et al (1998) Effects of d-amphetamine and caffeine in humans under a cocaine discrimination procedure. *Behav Pharmacol* 9:207–217
- Perkins KA, Fonte C, Ashcom J et al (2001) Subjective responses to nicotine in smokers may be associated with responses to caffeine and to alcohol. *Exp Clin Psychopharmacol* 9:91–100
- Perkins KA, Fonte C, Stolinski A et al (2005) The influence of caffeine on nicotine's discriminative stimulus, subjective, and reinforcing effects. *Exp Clin Psychopharmacol* 13:275–281
- Potthoff AD, Ellison G, Nelson L (1983) Ethanol intake increases during continuous administration of amphetamine and nicotine, but not several other drugs. *Pharmacol Biochem Behav* 18:489–493
- Powell KR, Holtzman SG (1998) Lack of NMDA receptor involvement in caffeine-induced locomotor stimulation and tolerance in rats. *Pharmacol Biochem Behav* 59:433–438
- Quarta D, Borycz J, Solinas M et al (2004) Adenosine receptor-mediated modulation of dopamine release in the nucleus accumbens depends on glutamate neurotransmission and N-methyl-D-aspartate receptor stimulation. *J Neurochem* 91:873–880
- Ribeiro JA, Sebastião AM, de Mendonça A (2002) Adenosine receptors in the nervous system: pathophysiological implications. *Prog Neurobiol* 68:377–392
- Roache JD, Griffiths RR (1987) Interactions of diazepam and caffeine: behavioral and subjective dose effects in humans. *Pharmacol Biochem Behav* 26:801–812
- Rossi S, De Chiara V, Musella A et al (2009) Caffeine drinking potentiates cannabinoid transmission in the striatum: interaction with stress effects. *Neuropharmacology* 56:590–597
- Rush CR, Sullivan JT, Griffiths RR (1995) Intravenous caffeine in stimulant drug abusers: subjective reports and physiological effects. *J Pharmacol Exp Ther* 273:351–358
- Sahraei H, Barzegari AA, Shams J et al (2006) Theophylline inhibits tolerance and sensitization induced by morphine: a conditioned place preference paradigm study in female mice. *Behav Pharmacol* 17:621–628
- Sahraei H, Motamedi F, Khoshbaten A et al (1999) Adenosine A(2) receptors inhibit morphine self-administration in rats. *Eur J Pharmacol* 383:107–113
- Schenk S, Horger B, Snow S (1990) Caffeine preexposure sensitizes rats to the motor activating effects of cocaine. *Behav Pharmacol* 1:447–451
- Schuh KJ, Griffiths RR (1997) Caffeine reinforcement: the role of withdrawal. *Psychopharmacology* 130:320–326
- Seeman P (2009) Dopamine D<sub>2</sub>High receptors measured ex vivo are elevated in amphetamine-sensitized animals. *Synapse* 63:186–192
- Shoaib M, Swanner LS, Yasar S et al (1999) Chronic caffeine exposure potentiates nicotine self-administration in rats. *Psychopharmacology* 142:327–333
- Simola N, Morelli M, Seeman P (2008) Increase of dopamine D<sub>2</sub>(High) receptors in the striatum of rats sensitized to caffeine motor effects. *Synapse* 62:394–397
- Smit HJ, Gaffan EA, Rogers PJ (2004) Methylxanthines are the psycho-pharmacologically active constituents of chocolate. *Psychopharmacology* 176:412–419

- Sobel BF, Sigmon SC, Griffiths RR (2004) Transdermal nicotine maintenance attenuates the subjective and reinforcing effects of intravenous nicotine, but not cocaine or caffeine, in cigarette-smoking stimulant abusers. *Neuropsychopharmacology* 29:991–1003
- Soria G, Castañé A, Berrendero F et al (2004) Adenosine A2A receptors are involved in physical dependence and place conditioning induced by THC. *Eur J Neurosci* 20:2203–2213
- Spealman RD, Barrett-Larimore RL, Rowlett JK et al (1999) Pharmacological and environmental determinants of relapse to cocaine-seeking behavior. *Pharmacol Biochem Behav* 64:327–336
- Stella L, De Novellis V, Vitelli MR et al (2003) Interactive role of adenosine and dopamine in the opiate withdrawal syndrome. *Naunyn Schmiedebergs Arch Pharmacol* 368:113–118
- Stolerman IP (1993) Components of drug dependence: reinforcement, discrimination and adaptation. *Biochem Soc Symp* 59:1–12
- Sudakov SK, Rusakova IV, Medvedeva OF (2003) Effect of chronic caffeine consumption on changes in locomotor activity of WAG/G and Fischer-344 rats induced by nicotine, ethanol, and morphine. *Bull Exp Biol Med* 136:563–565
- Svenningsson P, Nomikos GG, Fredholm BB (1999) The stimulatory action and the development of tolerance to caffeine is associated with alterations in gene expression in specific brain regions. *J Neurosci* 19:4011–4022
- Swanson JA, Lee JW, Hopp JW (1994) Caffeine and nicotine: a review of their joint use and possible interactive effects in tobacco withdrawal. *Addict Behav* 19:229–256
- Tronci E, Simola N, Carta AR et al (2006) Potentiation of amphetamine-mediated responses in caffeine-sensitized rats involves modifications in A2A receptors and zif-268 mRNAs in striatal neurons. *J Neurochem* 98:1078–1089
- Tuazon DB, Suzuki T, Misawa M et al (1992) Methylxanthines (caffeine and theophylline) blocked methamphetamine-induced conditioned place preference in mice but enhanced that induced by cocaine. *Ann N Y Acad Sci* 654:531–533
- Varty GB, Hodgson RA, Pond AJ et al (2008) The effects of adenosine A2A receptor antagonists on haloperidol-induced movement disorders in primates. *Psychopharmacology* 200:393–401
- Vaugeois JM (2002) Signal transduction: positive feedback from coffee. *Nature* 418:734–736
- Watson J, Deary I, Kerr D (2002) Central and peripheral effects of sustained caffeine use: tolerance is incomplete. *Br J Clin Pharmacol* 54:400–406
- Weisberg SP, Kaplan GB (1999) Adenosine receptor antagonists inhibit the development of morphine sensitization in the C57BL/6 mouse. *Neurosci Lett* 264:89–92
- Wolff K, Winstock AR (2006) Ketamine: from medicine to misuse. *CNS Drugs* 20:199–218
- Young R, Gabryszuk M, Glennon RA (1998) (-)Ephedrine and caffeine mutually potentiate one another's amphetamine-like stimulus effects. *Pharmacol Biochem Behav* 61:169–173
- Zarrindast MR, Moghadamnia AA (1997) Adenosine receptor agents and conditioned place preference. *Gen Pharmacol* 29:285–289

# Methylxanthines and Human Health: Epidemiological and Experimental Evidence

Marie-Soleil Beaudoin and Terry E. Graham

## Contents

1	Introduction .....	510
2	Caffeine, Coffee, and Carbohydrate Homeostasis .....	513
2.1	Acute Ingestion of Caffeine and Carbohydrate Homeostasis .....	513
2.2	Acute Ingestion of Coffee and Carbohydrate Homeostasis .....	518
2.3	Chronic Ingestion of Caffeine and Coffee and Carbohydrate Management .....	518
3	Target Tissues and Actions of Caffeine .....	522
3.1	Skeletal Muscle .....	523
3.2	Adrenal Medulla and the Sympathetic Nervous System .....	525
3.3	Adipose Tissue .....	527
3.4	Liver .....	527
3.5	Gastrointestinal System .....	529
3.6	Pancreatic $\beta$ Cells .....	531
4	Caffeine, Coffee, and Cardiovascular Disease: Epidemiology and Mechanisms .....	531
5	Coffee, Caffeine, and Other Health Effects .....	537
6	Summary .....	540
	References .....	541

**Abstract** When considering methylxanthines and human health, it must be recognized that in many countries most caffeine is consumed as coffee. This is further confounded by the fact that coffee contains many bioactive substances in addition to caffeine; it is rich in phenols (quinides, chlorogenic acid, and lactones) and also has diterpenes (fatty acid esters), potassium, niacin, magnesium, and the vitamin B<sub>3</sub> precursor trigonelline. There is a paradox as consumption of either caffeine or caffeinated coffee results in a marked insulin resistance and yet habitual coffee consumption has repeatedly been reported to markedly reduce the risk for type 2 diabetes. There is strong evidence that caffeine reduces insulin sensitivity in

---

M.-S. Beaudoin and T.E. Graham (✉)

Department of Human Health and Nutritional Sciences, University of Guelph, 50 Stone Road East,  
Guelph, ON, Canada N1G 2W1  
e-mail: terrygra@uoguelph.ca

skeletal muscle and this may be due to a combination of direct antagonism of A<sub>1</sub> receptors and indirectly  $\beta$ -adrenergic stimulation as a result of increased sympathetic activity. Caffeine may also induce reduced hepatic glucose output. With the exception of bone mineral, there is little evidence that caffeine impacts negatively on other health issues. Coffee does not increase the risk of cardiovascular diseases or cancers and there is some evidence suggesting a positive relationship for the former and for some cancers, particularly hepatic cancer.

**Keywords** Caffeine · Coffee · Insulin resistance · Type 2 diabetes · Adenosine · Cardiovascular disease

## Abbreviations

AUC	Area under the curve
cAMP	Cyclic AMP
CGA	Chlorogenic acid
CNS	Central nervous system
CVD	Cardiovascular disease
CYP1A2	Cytochrome P450 1A2
FFA	Free fatty acid
GIP	Glucose-dependent insulinotropic polypeptide
GLP-1	Glucagon-like peptide-1
ISI	Insulin sensitivity index
OGTT	Oral glucose tolerance test
SNS	Sympathetic nervous system
T2D	Type 2 diabetes

## 1 Introduction

The topic of this chapter is methylxanthines and human health. However, caffeine is the dominant methylxanthine consumed by humans and, furthermore, humans rarely consume caffeine by itself. The vast majority of caffeine consumption is as coffee. Therefore, in order to address human health practically, we have chosen to review both caffeine and coffee. It is very important to note that coffee is a complex food that is composed of thousands of compounds and that caffeine is only one of at least a hundred that are biologically active (Ranheim and Halvorsen 2005). In addition, much of the laboratory-based research with humans and animal models is with pure caffeine and yet the majority of epidemiology research is based on coffee consumption. One should not transpose the effects of caffeine to those of coffee without strong supporting data. A further challenge in establishing the effects of caffeine on human health is to consider the physiological effects of

caffeine and not the pharmacological actions. Humans normally experience systemic plasma concentrations of up to 30–60  $\mu\text{mol/L}$ , while many investigations use levels that are orders of magnitude greater.

The focus of the chapter is on human health and thus we will restrict the review to the ingestion of caffeine or other methylxanthines and coffee rather than other forms of administration. There appears to be an endless supply of novel commercial products that contain caffeine. These are designed for ingestion, skin application, or even to be aspirated. For example, Reissig et al. (2009) reviewed caffeinated energy drinks; they reported that, in the USA, bottles/cans of these items provided from 50 mg to over 500 mg of caffeine and their popularity is reflected by the estimation that sales in 2006 were US \$5.4 billion. Nevertheless, coffee remains by far the most common dietary source of caffeine in North America, Europe, and Brazil (Health Canada 2007; Frary et al. 2005; Camargo et al. 1999). In addition, coffee is the main source of caffeine for athletes, used as a way to improve performance (Tunicliffe et al. 2008). On the other hand, although tea generally contains less caffeine than coffee, it constitutes a major source of caffeine in heavy tea-consuming countries such as Japan (Yamada et al. 2009) and the UK (Lundsberg 1998). Similarly, mate is the main contributor to caffeine intake in Argentina (Olmos et al. 2009). Despite the heterogeneity of caffeine dietary sources, health issues have mostly been investigated in relation to pure caffeine or coffee. Consequently, while the current chapter will focus on coffee, caffeine, and health, it will also acknowledge the potential health effects of other caffeine sources, such as tea, when the information is available.

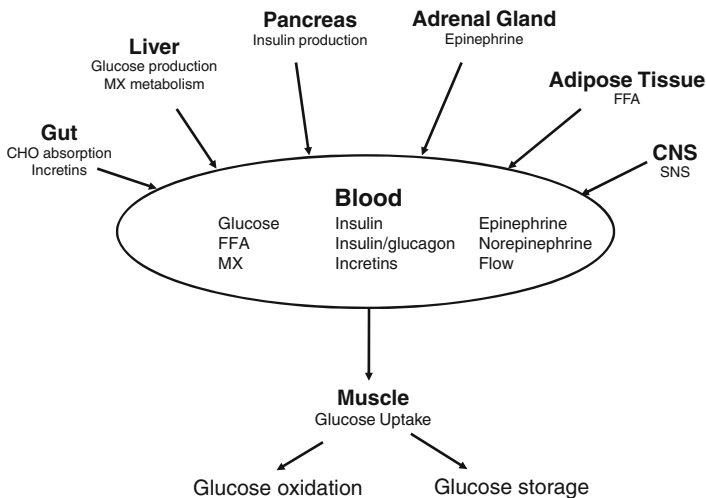
There have been a number of broad reviews on either caffeine or methylxanthine-containing foods and human health. Nawrot et al. (2003) concluded that moderate caffeine consumption (approximately 400 mg/day) was not associated with any adverse effects, including cardiovascular health and cancers. Others (van Dam 2008; Ranheim and Halvorsen 2005) have pointed out that consumption of boiled/unfiltered coffee increases the levels of blood lipids and is a risk for cardiovascular disease (CVD). However, filtered coffee reduces the risk of CVD and type 2 diabetes (T2D). It is commonly proposed that this may be due to the antioxidant, heterocyclic compounds in coffee. Similarly, Engler and Engler (2006) concluded that cocoa and chocolate have beneficial cardiovascular effects owing to their high flavonol content.

One often assumes that ill-health issues of obesity and the associated diseases such as T2D (and the complications of neuropathies, blindness, renal failure, amputations) and CVD are predominantly problems for the developed countries. While obesity is obviously a major problem in countries such as Canada and the USA, it is now a significant issue in countries such as Brazil, Guatemala, Nigeria, India, and China (Misra and Khurana 2008). Given this, it is not surprising that T2D is a global health problem. For example, almost 80% of diabetes deaths occur in low- and middle-income countries (WHO 2010). The World Health Organization estimates that over 4% of the world population will have T2D in two decades. Thus, if caffeine and coffee are nutritional factors associated with this disease, this is important in global health.

For both ethical and economic reasons, it is also important to note that not only is the frequency of occurrence of T2D increasing dramatically, but also that the average age of diagnosis is becoming younger. It is well documented that key risk factors are inactivity and obesity. Lifestyle intervention (physical activity and a healthy diet) is one of the best treatments and is also very important in preventing or delaying T2D. In this chapter the main focus will be on the relationship between caffeine and coffee and carbohydrate homeostasis and the primary health issue will be T2D.

The regulation of insulin and carbohydrate homeostasis is complex, involving many tissues and processes before one superimposes caffeine, a biologically active compound that can affect almost every tissue of the body. It also is a very important question as caffeine is a stable part of the diet of most adults globally. To add to the complexity, the most common source of caffeine is coffee, which contains many bioactive substances in addition to caffeine.

In order to evaluate the potential health impact of methylxanthines, it is fundamental that their impact on each tissue is understood. Methylxanthines are known to directly affect many tissues and this is a major complication for establishing the primary actions of caffeine in the human body. Once any one tissue has responded to caffeine, this response can result in a secondary action that is indirectly associated with the ingestion of caffeine. Figure 1 summarizes this with reference to the potential for caffeine to interfere with the actions of insulin on skeletal muscle. For example, caffeine stimulates the central nervous system (CNS) and the resultant changes in activity in the sympathetic nervous system (SNS) output could result in



**Fig. 1** Summary of the putative primary and secondary effects of caffeine on factors affecting carbohydrate homeostasis in some of the biological systems in the human. Caffeine affects several tissues simultaneously, which complicates the understanding of the physiological effects of caffeine. *CHO* carbohydrate, *CNS* central nervous system, *FFA* free fatty acids, *MX* methylxanthine, *SNS* sympathetic nervous system

further actions from a number of tissues. As will be reviewed briefly herein and has been discussed at length in other chapters, the main action of caffeine in its physiological range appears to be that of an adenosine receptor antagonist. Thus, any tissue that expresses such receptors could respond to caffeine. However, there are several subtypes of adenosine receptors and they are found in most tissues. This results in a vast array of possible primary and secondary responses. Owing to this complexity and multiple interactions, many scientists restrict their investigations to merely one tissue or cell line and only to caffeine. While this reductionist approach provides many scientific advantages, it dramatically restricts one's ability to understand the physiological effects of caffeine on the human, let alone the effects of coffee ingestion.

## 2 Caffeine, Coffee, and Carbohydrate Homeostasis

### 2.1 *Acute Ingestion of Caffeine and Carbohydrate Homeostasis*

Caffeine and coffee have been extensively studied in regard to their effects on insulin resistance and T2D. As mentioned, caffeine and coffee constitute two distinct metabolic challenges and appear to have different impacts on human health. In the case of insulin resistance, extensive literature supports that caffeine acutely decreases insulin sensitivity. In contrast, a similar body of literature documents that chronic consumption of coffee decreases the risk of T2D.

Acute administration of alkaloid caffeine impairs glucose homeostasis in healthy (Graham et al. 2001; Dekker et al. 2007; Battram et al. 2006; Keijzers et al. 2002; Norager et al. 2006), obese (Petrie et al. 2004; Lee et al. 2005), and diabetic (Robinson et al. 2004; Lee et al. 2005; Lane et al. 2004, 2008) subjects. In responsive populations, consumption of caffeine 1 h before an oral glucose tolerance test (OGTT) has been consistently shown to increase insulin area under the curve (AUC) by 25–42% (Graham et al. 2001; Robinson et al. 2004; Dekker et al. 2007; Thong and Graham 2002). Despite this exaggerated insulin response, most (Pizziol et al. 1998; Graham et al. 2000; Battram et al. 2006; Robinson et al. 2004; Dekker et al. 2007), but not all (Petrie et al. 2004; Thong and Graham 2002), studies reported exaggerated blood glucose response and elevated glucose AUC. An insulin sensitivity index (ISI) was developed to estimate insulin sensitivity based on the insulin and glucose AUC (Matsuda and DeFronzo 1999). Caffeine has consistently reduced the ISI by 14–25% compared with a placebo (Table 1) (Battram et al. 2006; Petrie et al. 2004; Robinson et al. 2004; Thong and Graham 2002).

In addition, the effects of caffeine on insulin sensitivity have been examined with the euglycemic–hyperinsulinemic clamp technique. In agreement with the studies employing the OGTT method, ingesting 5 mg caffeine per kilogram of body weight prior to the insulin clamp reduced the glucose infusion rate (i.e., created an insulin resistance) by 13–37% compared with a placebo (Fig. 2, Table 2)

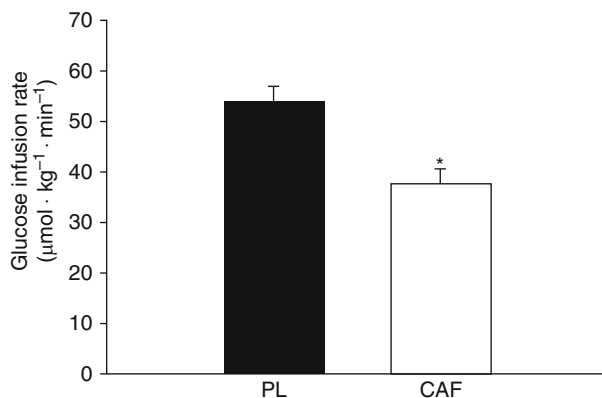


**Table 1** Summary of contemporary studies that examined the effects of caffeine on glucose and insulin metabolism during an oral carbohydrate challenge in humans

Reference	Subjects	Treatments	Study design	Results
Pizziol et al. (1998)	30 (12 males, 18 females) healthy subjects	CAF (200 mg) or DECAF (50 mL)	DECAF or DECAF + CAF 5 min prior to OGTT (4 h)	DECAF + CAF increased glucose AUC. No significant difference for insulin AUC
Graham et al. (2001)	18 young, fit males	CAF (5 mg/kg BW) or PL	Ingestion of capsule 1 h prior to OGTT (2 h).	CAF increased C-peptide (37%), insulin (60%), and glucose (24%) AUC
Thong and Graham (2002)	7 active males	PL, CAF (5 mg/kg BW), PRO (80 mg), CAF + PRO	Ingestion 90 min prior to OGTT (2 h)	CAF increased insulin (42%) and C-peptide (37%) AUC. No change in glucose AUC. CAF + PRO did not differ from PL
Lane et al. (2004)	14 (11 males, 3 females) T2D, CAF users	CAF (375 mg) or PL	250 mg CAF 1 h prior to 125 mg CAF + boost (75 g CHO). Blood samples drawn from 0 to 180 min	CAF increased glucose AUC (21%) and insulin AUC (48%)
Petrie et al. (2004)	9 obese, healthy males	CAF (5 mg/kg BW) or PL	Ingestion of capsule 1 h prior to OGTT (2 h) before and after 12-week weight-loss program	CAF increased insulin AUC before and after weight loss. No change in glucose AUC
Robinson et al. (2004)	12 males with T2D	CAF (5 mg/kg BW) or PL	Ingestion of capsule 1 h prior to OGTT (3 h)	CAF increased glucose AUC (16%) and insulin AUC (25%). CAF decreased insulin sensitivity by 14%
Norager et al. (2006)	30 (15 males, 15 females) elderly (> 70-year-old) subjects	CAF (6 mg/kg BW) or PL	Cycling at 65% of maximum O <sub>2</sub> uptake 1 h after capsule ingestion	CAF increased insulin resistance (HOMA-IR) by 23% at the end of exercise
Batram et al. (2007b)	14 tetraplegic patients	CAF (4 mg/kg BW) or PL	Ingestion of capsule 1 h prior to 2 h-OGTT	No effects of CAF on glucose, insulin, proinsulin or C-peptide

Dekker et al. (2007)	12 males, noncaffeine users	CAF (5 mg/kg BW) or PL	Habituation. 1 PL/A and 3 CAF trials separated by 7 days + consumption of 5 mg/kg CAF every day	CAF increased glucose AUC at day 0 and insulin AUC at day 0 and day 14
Lane et al. (2008)	10 (5 males, 5 females) T2D, CAF users	CAF (500 mg) or PL	250 mg CAF with breakfast (Boost Plus – 90 g CHO) + 250 mg CAF with lunch. Continuous blood glucose monitoring for 72 h	CAF increased average daytime glucose and postprandial glucose responses
Robinson et al. (2009)	27 pregnant women (19 without GDM; 8 with GDM)	CAF (3 mg/kg prepregnancy BW) or PL	Ingestion of capsule 1 h prior to an OGTT (2 h)	CAF increased glucose AUC and C-peptide AUC in GDM women. CAF decreased insulin sensitivity by 18% in GDM women

*AUC* area under the curve, *BW* body weight, *CAF* caffeine, *CHO* carbohydrate, *DECAF* decaffeinated coffee, *GDM* gestational diabetes mellitus, *HOMA-IR* homeostasis model assessment of insulin resistance, *OGTT* oral glucose tolerance test, *PL* placebo, *PRO* propranolol, *T2D* type 2 diabetes



**Fig. 2** Caffeine-induced insulin resistance shown during a euglycemic–hyperinsulinemic clamp. Glucose infusion rates for a placebo (PL) and caffeine (CAF) are shown for seven healthy males. The *asterisk* indicates that the ingestion of caffeine lowered the glucose disposal compared with the placebo. (Reproduced from Thong et al. 2002 with permission)

(Keijzers et al. 2002; Greer et al. 2001; Thong et al. 2002; Battram et al. 2005; Lee et al. 2005; Battram et al. 2007a). As discussed below, this likely results from caffeine antagonism of adenosine receptors in skeletal muscle.

It must be noted that studies using euglycemic–hyperinsulinemic clamps investigate different aspects of metabolism than those using OGTTs. The former is the gold standard to examine peripheral tissues responses to insulin. On the other hand, an OGTT provides valuable data on postprandial metabolism and takes into account other tissues, such as the gut, pancreatic  $\beta$  cells, and the liver. Given that caffeine may affect gut incretins, liver glucose output,  $\beta$ -cell function, as well as adipose tissue and muscle, the information from both of these techniques is essential to obtain a better understanding of the effect of caffeine on carbohydrate metabolism. The consistent finding using either approach that caffeine resulted in a 15–30% reduction in insulin's effectiveness is impressive. It may suggest that the major reason for this immediate effect is the same in both situations: impaired glucose disposal in skeletal muscle.

While all of these investigations examined caffeine and focused on hyperglycemia and applications for T2D, it is noteworthy that de Galan et al. (2002) infused the dimethylxanthine theophylline, used type 1 diabetics, and infused glucose to maintain hypoglycemia rather than euglycemia. They found that this methylxanthine also caused an insulin resistance, but noted that for a person with type 1 diabetes who is hypoglycemic, this would be beneficial. A practical extension of this finding could be to suggest that in a hypoglycemic state, ingesting a beverage such as a cola would be particularly helpful. Not only would this provide a large amount of easily absorbed carbohydrate, but presumably the caffeine would also cause insulin resistance, thus further increasing the low amount of blood glucose.

In summary, the available evidence strongly supports that acute ingestion of caffeine impairs insulin sensitivity and glucose disposal in lean, obese, and diabetic

**Table 2** Summary of contemporary studies that examined the effects of caffeine on glucose disposal during a euglycemic-hyperinsulinemic clamp in humans

Reference	Subjects	Treatments	Study design	Results
Greer et al. (2001)	9 sedentary, lean male, non-CAF users	CAF (5 mg/kg BW) or PL	Ingestion of capsule immediately followed by 180-min clamp	CAF decreased GIR by 24% and decreased CHO storage by 35%
Keijzers et al. (2002)	12 healthy subjects (6 females, 6 males)	CAF infusion (3 mg/kg BW) or saline	120-min clamp started 30 min after the start of the CAF infusion	CAF decreased insulin sensitivity by 15%
Thong et al. (2002)	7 recreationally active males	CAF (5 mg/kg BW) or PL	1-leg extension (1 h), rest (2 h), CAF or PL, rest (1 h), 100-min clamp	CAF decreased GIR (30%), decreased leg glucose uptake (51% exercised leg vs. 55% rested leg)
Battram et al. (2005)	12 healthy, young males; CAF users	CAF (5 mg/kg BW), PL and PL + high or low epinephrine	Ingestion of CAF or infusion of epinephrine 30 min prior to 120-min clamp	CAF and high EPI decreased GIR by 34 and 13%, respectively. EPI levels similar between CAF and low EPI
Lee et al. (2005)	Lean ( $n = 8$ ), obese ( $n = 7$ ), and T2D ( $n = 8$ ) males	CAF (5 mg/kg BW) or PL	Ingestion of capsule 30 min prior to 180-min clamp before and after a 13-week aerobic exercise program	CAF decreased insulin sensitivity by 23–37% in all three groups before and after the exercise program
Battram et al. (2007a)	8 healthy males	PL + saline; CAF (5 mg/kg BW) + saline; PL + EPI; CAF + EPI	Ingestion of capsule 30 min prior to 120-min clamp	CAF (26%), EPI (24%), and CAF + EPI (42%) decreased insulin-corrected GIR vs. PL. Trend for CAF + EPI to be different from EPI or CAF ( $p < 0.08$ )

*BW* body weight, *CAF* caffeine, *CHO* carbohydrate, *EPI* epinephrine, *GIR* glucose infusion rate, *PL* placebo, *T2D* type 2 diabetes

subjects. While data from studies investigating caffeine with either an OGTT or a euglycemic–hyperinsulinemic clamp are very convincing, such studies use a reductionist approach, investigating caffeine in isolation. As noted previously, caffeine is most often consumed in the form of beverages or foods. In North America, 60–75% of caffeine intake comes from coffee (Health Canada 2007). Therefore, investigating the effects of caffeinated coffee on insulin sensitivity and glucose uptake is highly relevant.

## ***2.2 Acute Ingestion of Coffee and Carbohydrate Homeostasis***

Similar to caffeine, acute consumption of coffee (note that the term “coffee” will refer to caffeinated coffee) is detrimental to glucose tolerance in healthy subjects. Coffee has been shown to acutely increase glucose AUC compared with decaffeinated coffee in healthy men during an OGTT (Battram et al. 2006; Greenberg et al. 2009; Johnston et al. 2003). In addition, Moisey et al. (2008) showed that, in comparison with decaffeinated coffee, regular coffee increased glucose AUC, insulin AUC, and decreased the ISI when consumed prior to both high and low glycemic index cereal meals (Table 3). Furthermore, this effect was also observed following a second meal (Moisey et al. 2010).

In these studies, coffee significantly impaired glucose disposal, when compared with decaffeinated coffee. One can then ask whether this effect is due to deleterious effects of coffee and/or to possible beneficial effects of decaffeinated coffee. There is limited information in this regard and the results are not consistent. While Battram et al. (2006) found that the glucose AUC during an OGTT was significantly lower following consumption of decaffeinated coffee compared with a placebo, Kacker (2003) did not report differences in insulin or glucose AUCs between decaffeinated coffee and water treatments during an OGTT. Recently, van Dijk et al. (2009) also failed to show an effect of decaffeinated coffee ingested prior to an OGTT and in contrast to all of these investigations, Greenberg et al. (2009) found that decaffeinated coffee resulted in impaired glucose metabolism compared with a placebo. While there is limited information and no consensus regarding decaffeinated coffee, taken collectively, a large number of investigations have revealed that acute ingestion of regular coffee is detrimental to glucose tolerance.

## ***2.3 Chronic Ingestion of Caffeine and Coffee and Carbohydrate Management***

Investigation of the effect of coffee consumption on carbohydrate homeostasis highlights a discrepancy between acute and chronic exposure to coffee. On one hand, acute consumption of coffee is detrimental to glucose tolerance. On the other hand, regular consumption of coffee is associated with a reduced risk of T2D.

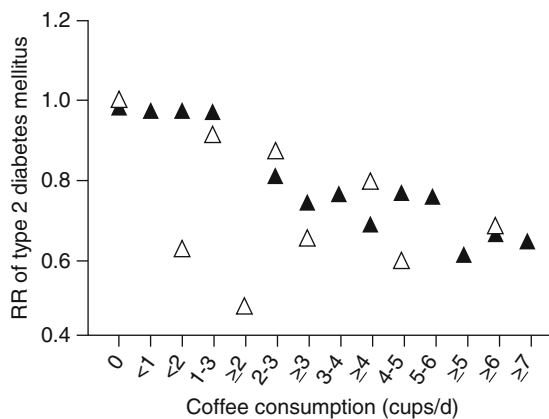
**Table 3** Summary of contemporary studies investigating the acute effects of coffee (caffeinated and decaffeinated) on glucose and insulin metabolisms in humans

Reference	Subjects	Treatments	Study design	Results
Johnston et al. (2003)	9 (4 males, 5 females) lean subjects	COF, DECAF, or water	400-mL drink ingested simultaneously with 25 g glucose. Blood samples were taken for 3 h	Between 0 and 30 min, COF increased glucose and insulin AUC vs. DECAF. COF also increased glucose AUC vs. PL
Batram et al. (2006)	11 healthy, young males	DECAF, COF, caffeine, PL	Ingestion of drink 1 h prior to OGTT	Caffeine increased glucose and insulin AUC. DECAF decreased glucose AUC vs. PL
Moisey et al. (2008)	10 healthy males	COF (5 mg/kg BW) or DECAF	Ingestion of drink 1 h prior to high GI or low GI cereal	COF increased glucose AUC, insulin AUC (NS for high GI), and C-peptide AUC for both low and high GI cereals
van Dijk et al. (2009)	15 overweight males	DECAF (12 g), trigonelline (500 mg), CGA (1 g), or PL	Ingestion 30 min prior to 2 h OGTT	CGA and trigonelline reduced glucose and insulin concentrations 15 min into OGTT
Moisey et al. (2010)	10 healthy males	COF (5 mg/kg BW), DECAF, or water	Ingestion of treatment + high glycemic cereal 3 h prior to a 3-h OGTT	No change in AUC In the OGTT, COF increased insulin AUC by 49 and 57% compared with DECAF and water, respectively
Greenberg et al. (2009)	11 healthy young men	COF (6 mg/kg BW), DECAF, caffeine, or water	Ingestion of treatment 1 h prior to OGTT	COF increased ISI compared with DECAF and water during the OGTT In the OGTT, DECAF had lower glucose AUC than caffeine The PL had lower insulin AUC than DECAF, COF, and caffeine

AUC area under the curve, CGA chlorogenic acid, COF caffeinated coffee, DECAF decaffeinated coffee, GI glycemic index, ISI insulin sensitivity index, NS nonsignificant, OGTT oral glucose tolerance test, PL placebo

While habitual intake of caffeine can lead to habituation and dampened responses in parameters such as increased blood pressure in some individuals (Debrah et al. 1995; Lovallo et al. 2004), Dekker et al. (2007) found that caffeine continued to result in an insulin resistance in caffeine-naïve subjects after 14 days of caffeine ingestion. In addition, in the many studies summarized above, most subjects had been regular consumers of coffee for years or decades. While there was a only brief period of withdrawal (1–2 days) prior to testing, they responded to caffeine or coffee consumption with a marked insulin resistance. Thus, any habituation to caffeine appears to be minimal and/or rapidly reversible.

In a meta-analysis of consumption of coffee and T2D, the relative risks for heavy (six or more cups per day) and moderate (four to six cups daily) coffee drinkers were 0.65 and 0.72, respectively, compared with the lowest coffee consumers (zero to two cups daily) (van Dam and Hu 2005). These findings have been replicated across sexes, geographical locations, and obesity levels (reviewed in van Dam 2008; van Dam and Hu 2005; van Dam et al. 2006; Pereira et al. 2006; Salazar-Martinez et al. 2004; Agardh et al. 2004; Tuomilehto et al. 2004), and this relationship is also documented in patients with impaired glucose tolerance at the baseline (Smith et al. 2006). Coffee appears to have dose-dependent protective effects on T2D (Fig. 3). A recent meta-analysis suggested that for every additional cup of coffee consumed daily, the risk of T2D is reduced by 7% (Huxley et al. 2009). An inverse, linear relationship has also been reported between T2D and the consumption of decaffeinated coffee and tea, although fewer studies investigated this association (Huxley et al. 2009). In fact, drinking daily three or four cups of regular coffee, decaffeinated coffee, or tea decreased the risk of T2D by 25, 33, and



**Fig. 3** Summary of the relative risk of type 2 diabetes with habitual coffee consumption. This figure summarizes the data from various epidemiology studies that were summarized by Huxley et al. (2009). The *closed symbols* represent the consumption of caffeinated coffee and the *open symbols* represent the consumption of decaffeinated coffee. Each additional daily cup of coffee consumed reduced the relative risk by 7% and consuming three or four cups of coffee a day resulted in a 25% lower risk compared with not drinking coffee. Similar results were found for decaffeinated coffee, but the number of studies was limited

20%, respectively (Huxley et al. 2009). Interestingly, a small prospective study found that moderate consumption of regular coffee prior to pregnancy, but not decaffeinated coffee, decreased the risk of gestational diabetes by 50% compared with the risk for nonconsumers (Adeney et al. 2007). It is noteworthy that Robinson et al. (2009) found that acute ingestion of caffeine did not cause insulin resistance in pregnant women who did not have gestational diabetes and yet those with this illness responded to caffeine with an insulin resistance (Table 1). In summary, regular coffee drinking is protective against diabetes (T2D and gestational diabetes) in a dose-dependent manner and across ethnic groups and a wide range of drinking habits.

The beneficial effects of coffee consumption are attributed to coffee compounds other than caffeine. Caffeine constitutes only approximately 2% of coffee, and some of the bioactive compounds in coffee are still unknown (Tunicliffe and Shearer 2008). The concentrations of these will vary with the type of bean, the growing conditions, the roasting techniques, and the method of brewing and filtering the coffee. Tse (1991, 1992) isolated an unidentified cholinomimetic compound from coffee and infusion into rodents resulted in a decrease in blood pressure and heart rate. Coffee also contains diterpenes (fatty acid esters), which elevate the level of blood cholesterol, and minerals such as potassium, niacin, and magnesium. Furthermore, coffee is a major dietary source of the vitamin B<sub>3</sub> precursor trigonelline, and is also rich in phenols [quinides, chlorogenic acid (CGA), and lactones]. These compounds are formed during the roasting of the beans and appear to have a variety of biological actions, including being antioxidants.

Oxidative stress occurs in the body when free radicals overcome the protective effects of antioxidants. The resulting cellular damage is associated with several health problems, including diabetes, CVD, and cancer. Fruits and vegetables are typical sources of antioxidants in the diet. However, the volume and frequency of coffee consumption in Western countries make coffee the major source of dietary antioxidants (Tunicliffe and Shearer 2008). The extent of their efficacy in humans *in vivo* is still debated.

Two classes of compounds have been specifically suggested to be beneficial to insulin sensitivity: CGAs and quinides. In diabetic rats, plant extracts containing mostly 5-caffeoylquinic acid (5-CQA), a common CGA in coffee, lowered plasma glucose levels (Andrade-Cetto and Wiedenfeld 2001). In humans, 1 g CGA and 500 mg trigonelline independently improved glucose and insulin concentrations early during an OGTT. However, this effect was not seen with decaffeinated coffee providing 264 mg CGA and 72 mg trigonelline (van Dijk et al. 2009). The possibility remains that prolonged or high-concentration exposure to these compounds may be necessary before there is a health benefit.

Shearer et al. (2007) provided direct evidence that short-term consumption of decaffeinated coffee had a beneficial result. They fed rats a high-fat diet for 28 days along with decaffeinated coffee, decaffeinated coffee with caffeine added, or water as the available fluid. The decaffeinated coffee treatment created an insulin-sensitive state as an increased glucose infusion rate was required during an insulin clamp compared with decaffeinated coffee with caffeine and water treatments.



The concentration of CGAs in coffee reaches 2.5 mmol/L but very little CGA is detected in the plasma of coffee drinkers because it is hydrolyzed rapidly (Tunicliffe and Shearer 2008). This suggests that advantageous effects of CGA could occur in the gastrointestinal system, before CGA is absorbed and degraded, or could be mediated indirectly, through its main product, caffeic acid (McCarthy 2005).

Quinides, another potentially beneficial class of compounds, are formed from CGAs during the roasting process of coffee beans and are thus specific to coffee. A euglycemic–hyperinsulemic clamp on rats simultaneously with the infusion of decaffeinated coffee extract, synthetic quinide, or saline showed that the synthetic quinide treatment increased the glucose infusion rate (Shearer et al. 2003). While these data strongly support the potential positive impact of decaffeinated coffee and particularly that of CGAs and/or quinides on glucose homeostasis, further research is necessary to confirm their efficacy in humans and their mechanistic bases. Epidemiological evidence documents that drinking coffee can reduce considerably the risk of T2D. This effect is attributed to coffee compounds other than caffeine. However, most products in coffee are still unknown; thus, it is difficult to attribute the positive effects of coffee to one particular class of compounds. It is important to point out that few of the epidemiology studies controlled for the type of coffee consumed. The methods of coffee preparation (e.g., espresso, boiled, filtered, decaffeination) influence the composition of coffee (Ranheim and Halvorsen 2005) and it is unknown how those varied types of coffee modulate metabolic responses.

### 3 Target Tissues and Actions of Caffeine

Adenosine receptors are ubiquitous, occurring throughout the nervous system, and in the vascular endothelium, heart, liver, adipose tissues, and muscle (Reppert and Weaver 1991; Dixon et al. 1996; Fredholm et al. 1999). Thus, the actions that result from caffeine are dependent on which type of adenosine receptors it blocks and in which tissue the receptors are located. It is unlikely that any one tissue is “dominating” the response, but rather it likely results from a combination of the actions of caffeine on various tissues. With regard to insulin resistance, it appears that the major tissue affected is skeletal muscle. Pertinent to this discussion, the effects of caffeine on muscle could be, in part, secondary to the initial effects on other tissues (Fig. 1). To gain a better understanding for how caffeine can mediate its multiple effects and influence one’s health, it is important to examine the actions of caffeine on various tissues. Two to three cups/mugs of coffee can result in plasma caffeine levels of 20–40  $\mu\text{mol/L}$  (as will ingestion of approximately 5 mg/kg of caffeine) and this is accompanied with lower levels of the dimethylxanthines. Of the latter, paraxanthine is the most abundant, usually reaching 5–8  $\mu\text{mol/L}$  (McLean and Graham 2002). Caffeine has a half-life of approximately 4–6 h (McLean and Graham 2002) and, at biological concentrations (5–50  $\mu\text{mol/L}$ ), it is an antagonist to adenosine receptors. This is addressed in detail in other chapters of

this book. The receptors are associated with intracellular pathways that influence cyclic AMP (cAMP) production, phospholipase C, and mitogen-activated protein kinases (Schulte and Fredholm 2003). The receptors have several isoforms ( $A_1$ ,  $A_{2a}$ ,  $A_{2b}$ , and  $A_3$ ) and caffeine is believed to antagonize all except the  $A_3$  form (Daly and Fredholm 2004). Thus, the action of caffeine on a given tissue depends on its complement of receptor isoforms. The  $A_1$  and  $A_2$  adenosine receptors have opposite actions, with the former associated with  $G_i$  protein input onto adenylate cyclase, decreasing the level of intracellular cAMP, while the latter is associated with  $G_s$  protein and increases in the level of cAMP. The  $A_1$  receptor is also associated with  $G_{\beta\gamma}$  subunits of the heterotrimeric G protein that can have actions separate from those of the  $G_\alpha$  subunit that mediates the inhibition of adenylate cyclase. This  $G_{\beta\gamma}$  subunit affects calcium release, potassium channels, and voltage-sensitive calcium channels. With such diverse potential effects and a vast number of tissues expressing adenosine receptors, even the primary responses in a person who consumes caffeine are complex and when the secondary events are added to this it becomes an enormous challenge. This section will attempt to address these interactions as they apply to lipid and carbohydrate metabolism in muscle. The distribution of the subtypes of adenosine receptors will not be addressed as this has been addressed elsewhere in the text. It will only be noted in tissues that apply directly to the main topic of lipid and carbohydrate management.

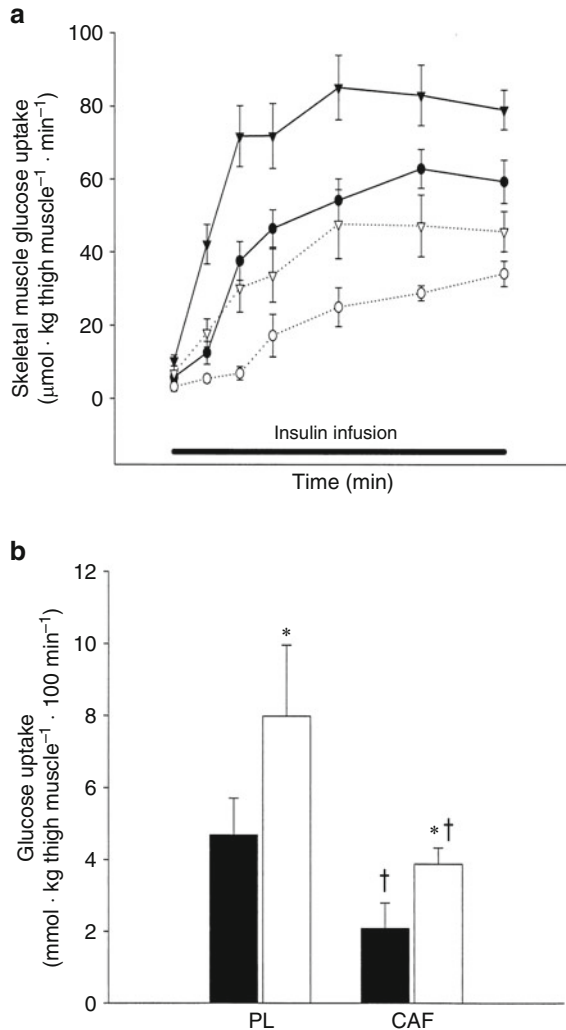
### 3.1 *Skeletal Muscle*

Skeletal muscle is the dominant tissue for glucose disposal as it is insulin-sensitive and represents 35–40% of the body mass. Whole-body insulin resistance is commonly associated with limitations in this tissue. Thong et al. (2002) found that caffeine ingestion resulted in a 30% decrease in whole-body glucose disposal (Fig. 2) and this was associated with a 50–55% decrease in leg glucose uptake. This was found for the leg after exercise (when glucose uptake is normally enhanced) as well as when there was no exercise (Fig. 4). Thus, one of caffeine's dominant actions is to impair blood glucose disposal, particularly in skeletal muscle. The mechanisms by which this occurs have been difficult to establish.

Various studies (Vergauwen et al. 1994, 1997; Challis et al. 1984; Han et al. 1998) have demonstrated that functionally muscle has  $A_1$  receptors and that caffeine's effects on rodent skeletal muscle are via the antagonism of  $A_1$  receptors and subsequent elevation of the level of cAMP. However, until recently, only  $A_2$  receptors had been demonstrated (Lyngne and Hellsten 2000). Thong et al. (2007) have now shown that indeed  $A_1$  receptors do physically exist in the plasma membrane of rat soleus muscle and, furthermore,  $A_1$  antagonism in vitro resulted in a 30% decrease in glucose uptake. They have also demonstrated that human muscle experiences an increase in the level of cAMP (Thong et al. 2002). In addition, Cheng et al. (2000) demonstrated that streptozotocin-induced (i.e., insulin-free) diabetic rats experienced a decrease in blood glucose level when treated

**Fig. 4** Caffeine-induced insulin resistance in the leg of human subjects.

**a** Summary of the net leg glucose uptake responses during a hyperinsulinemic–euglycemic clamp. The *open symbols* are for the subjects' leg that was not exercised and the *filled symbols* are for the previously exercised leg. The *circles* represent the caffeine-ingestion condition and the *triangles* represent the placebo. **b** Area under the curve data of **a**, with the *filled bars* representing the rested leg and the exercised leg is indicated by the *open bars*. The *asterisk* indicates a significant difference for exercise versus rest and the *cross* reflects a difference in caffeine versus the control. (Reproduced from Thong et al. 2002 with permission)



with dipyridamole to increase endogenous adenosine, an  $A_1$  receptor agonist, while antagonism of  $A_1$  receptors blocked the response. Furthermore, they found that the former treatment enhanced glucose uptake and glycogen synthesis, while  $A_1$  receptor antagonism blocked the uptake of glucose.

There may also be an additional effect of caffeine on muscle carbohydrate metabolism. Several studies (Greer et al. 2001; Thong et al. 2002; Lee et al. 2005) noted that when there was a decreased glucose disposal due to caffeine, the whole-body metabolic rate and respiratory exchange ratio were not altered. Thus, the amount of carbohydrate oxidized was not affected and the amount of carbohydrate stored must account for the depressed glucose disposal. Lee et al.

(2005) estimated this to be a 50–65% depression in carbohydrate storage. Thong et al. (2002) were able to make these measures and estimates both for the whole body (Fig. 1) and the leg (Fig. 4). Their data demonstrated that this preferential impairment of storage rather than oxidation occurred in the leg, i.e., muscle.

Caffeine could have direct intracellular effects, as it is also known to cross cell membranes. Caffeine ingestion resulted in an increase in the level of cAMP in both exercised and rested legs and this was accompanied with a decrease in the level of glycogen synthase (17% fractional velocity and 35% I form) (Thong et al. 2002). Rush and Spriet (2001) reported that physiological concentrations of caffeine could directly inhibit glycogen phosphorylase. The findings are preliminary but they do present the possibility that caffeine could have direct actions on metabolic enzymes associated with carbohydrate storage. Thus, while less glucose enters this tissue during caffeine exposure, of that which does enter, it may be that a smaller fraction than normal is directed into glycogen storage.

We are only aware of one investigation that directly examined coffee and skeletal muscle glucose uptake. Shearer et al. (2007) fed rats a high-fat diet for 4 weeks to induce insulin resistance and they also consumed either water, decaffeinated coffee, or decaffeinated coffee plus caffeine during this time. Those consuming decaffeinated coffee had an increased insulin sensitivity of approximately 100%, but when the decaffeinated coffee had caffeine added, this positive effect was not present. They also found that decaffeinated coffee specifically resulted in increased glucose uptake in a variety of skeletal muscles, while this was not seen when caffeine was consumed and, furthermore, this caffeine treatment also depressed glucose uptake by the heart. Thus, while some components of coffee may increase insulin sensitivity in skeletal muscle, caffeine can result in insulin resistance and the latter could be due to antagonizing A<sub>1</sub> receptors. However, there are a variety of other possible actions of caffeine that could also impact on muscle.

### ***3.2 Adrenal Medulla and the Sympathetic Nervous System***

A number of areas of the CNS are affected by caffeine, but of particular interest for this topic is the SNS. As discussed previously, the SNS is stimulated when the CNS is exposed to caffeine. Epinephrine originates from efferent stimulation of the adrenal medulla, but circulating norepinephrine is the result of “spillover” or “washout” from the SNS activity of specific tissues. There are a number of reports that caffeine increases circulating levels of epinephrine, but changes in norepinephrine concentration have been inconsistent (Robertson et al. 1981; Arciero et al. 1995; Van Soeren et al. 1993; Graham and Spriet 1995). We (Graham et al. 2000) have reported an increase in norepinephrine “spillover” from the leg of humans during exercise. These endocrine responses may in turn precipitate some of the metabolic responses associated with caffeine ingestion. Increased SNS activity can directly result in decreased glucose clearance and can also mobilize free fatty acid (FFA) and the latter could result in insulin resistance.

Epinephrine constitutes a potential mechanistic explanation as elevations can result in a decrease in glucose uptake in muscle (Battram et al. 2007a; Howlett et al. 1999). Comparison of people with tetraplegia (Van Soeren et al. 1996; Mohr et al. 1998; Battram et al. 2007b) and able-bodied individuals provides insight into one of the actions of caffeine. Battram et al. (2007b) found that caffeine consumption followed by an OGTT in people with tetraplegia did not result in any indication of insulin resistance. This may be due to changes associated with the profound muscle atrophy that they experience or may be due to the SNS. Able-bodied individuals respond to caffeine ingestion with an increase of about 100% in the level of circulating epinephrine. However, patients with tetraplegic injuries have a denervated adrenal medulla and very little circulating epinephrine and when they ingest caffeine, there is no increase in the level of epinephrine (Van soeren et al. 1996; Battram et al. 2007b). This strongly suggests that the caffeine–epinephrine response is not a direct action on the medulla but rather is secondary to stimulation of the CNS. This finding also suggests that caffeine-induced insulin resistance could be a product of increased levels of catecholamines. The importance of epinephrine as a mediator of caffeine effects was also supported when simultaneous ingestion of caffeine and propranolol, a  $\beta$ -adrenergic receptor blocker, was shown to counteract the deleterious effects of caffeine alone on glucose disposal (Thong and Graham 2002). Thus, it appears that epinephrine is essential to mediate caffeine effects on glucose tolerance.

Mechanistically, epinephrine activation of  $\beta$ -adrenergic receptors decreases whole-body glucose metabolism by 40%, probably through inhibition of glucose transporter type 4 activity (Bonen et al. 1992; Han and Bonen 1998). This effect is of a magnitude similar to what is reported with caffeine alone (Tables 1, 2). However, epinephrine levels ranging from 2 to 4 nmol/L are necessary to generate this magnitude of a decrease in glucose disposal, while ingestion of 5 mg of caffeine per kilogram of body weight only increases the plasma epinephrine concentration to 0.3–0.6 nmol/L (Graham et al. 1998; Thong and Graham 2002). Owing to this difference in plasma concentrations, it is unlikely that epinephrine alone mediates the effects of caffeine on glucose disposal. In addition, Battram et al. (2005, 2007a) reported that the glucose infusion rate during a euglycemic–hyperinsulinemic clamp was reduced by 26 and 24% with caffeine and epinephrine infusions, respectively. This similar impairment was achieved even though the epinephrine concentration in the caffeine trial was 50% less than that in the epinephrine trial. Most importantly, they found that simultaneous ingestion of caffeine and infusion of epinephrine resulted in a far greater impact on glucose disposal (42%), despite lower epinephrine levels in this treatment compared with that of epinephrine infusion alone. In addition, as mentioned previously, Thong et al. (2007) showed that  $A_1$  antagonism resulted in a 30% decrease in glucose uptake in skeletal muscle *in vitro* and there was no epinephrine present in the medium. Taken together, these data suggest that epinephrine is likely an important factor, but not sufficient, to entirely explain the caffeine effect. There is also clear evidence that antagonism of  $A_1$  receptors in muscle is very important.

### 3.3 *Adipose Tissue*

Intravenous infusion of a triglyceride emulsion plus heparin for at least 3 h decreases glucose disposal during a euglycemic–hyperinsulinemic clamp (Boden et al. 2001; Belfort and Mandarino 2005; Dresner et al. 1999; Kruszynska et al. 2002; Itani et al. 2002; Homko et al. 2003). It is well known (Battram et al. 2006; Petrie et al. 2004; Thong et al. 2002; Dekker et al. 2007) that caffeine results in an elevation of FFA and thus this in turn could also induce insulin resistance. Catecholamines can directly increase the level of plasma FFA and it is not clear whether or not the FFA mobilization is due to a direct action of caffeine on adipocytes or is due to the increase in SNS activity described earlier. Adipocytes express adenosine  $A_1$  receptors and these are tonically active, inhibiting adenylate cyclase and reducing both the level of cAMP and lipolysis (Liang et al. 2002). Adipocytes also have adrenergic  $\beta_2$  receptors and thus they could mediate the lipolysis. Investigations with different subject populations have assisted in establishing critical aspects of the hierarchy of metabolic events associated with caffeine. When people with tetraplegia ingest caffeine, they experience no increase in the levels of epinephrine as noted earlier and yet there is a large increase in circulating FFA concentration that is comparable with that observed in able-bodied subjects (Van soeren et al. 1996; Battram et al. 2007b). This clearly demonstrates that caffeine directly enhances lipolysis independent of the SNS. Similarly, Johansson et al. (2007) studied rodent adipocytes with the  $A_1$  receptor knocked out and concluded that this receptor was indeed responsible for the antilipolytic actions of caffeine.

Regardless of the mechanism of FFA mobilization, there are at least two reasons to believe that the rise in FFA concentration is not critical for the skeletal muscle insulin resistance. First, Greer et al. (2001) performed hyperinsulinemic–euglycemic clamps immediately after subjects had ingested caffeine. They observed a marked decrease in glucose disposal (Table 2) and yet the circulating FFA levels decreased rapidly and markedly from the onset of the hyperinsulinemia. In addition, as noted above, Battram et al. (2007b) found that caffeine followed by an OGTT did not result in insulin resistance in people with tetraplegia and yet there was a normal increase in FFA concentration. Thus, it appears that the mechanisms that influence skeletal muscle insulin sensitivity are (1) caffeine-induced stimulation of the SNS and increased circulation of epinephrine and (2) direct caffeine antagonism of  $A_1$  receptors on the muscle tissue.

### 3.4 *Liver*

With regard to the topic of caffeine and carbohydrate homeostasis, most of the research focus has been on skeletal muscle, but the liver could be involved both in the regulation of the circulating levels of caffeine and also in the regulation of blood glucose concentrations. In addition, this tissue experiences concentrations of caffeine and glucose that far exceed those of other tissues. Once the caffeine or

glucose is absorbed, it enters the portal circulation and is delivered to the liver via the portal artery. The liver can take up the glucose to store it as glycogen and can also liberate glucose into the systemic circulation. In addition, the hepatic P450 system metabolizes the caffeine to dimethylxanthines. These dimethylxanthines plus caffeine that were not catabolized in the first pass then enter the vena cava and their concentrations are diluted as they circulate systemically. Similarly, glucose that is not cleared, as well as any that is released from the liver owing to either glycogenolysis or gluconeogenesis, will enter the systemic circulation. Thus, the liver is exposed to a far higher concentration of caffeine than any other tissue of the body and also is fundamental to the dynamic regulation of blood glucose.

Rarely has the liver been investigated *in vivo* as its portal circulation and complex metabolism present methodological challenges. In dogs, adenosine infusion has been shown to increase hepatic glucose output and to inhibit the suppressive effects of insulin (McLane et al. 1990). Pencek et al. (2004) infused caffeine and glucose into the portal vein of conscious dogs during a hyperinsulinemic–hyperglycemic clamp. When the hepatic portal artery, portal vein, and a systemic (hepatic) artery were simultaneously sampled, the respective concentrations were 101, 83, and 35  $\mu\text{mol/L}$ , illustrating the differences in caffeine levels throughout the circulation. The hepatocytes experienced threefold greater caffeine concentration compared with other tissues. In this study no differences were found in the levels of arterial glucagon, cortisol, and norepinephrine, but caffeine did induce a very modest increase in the level of epinephrine. Nevertheless, caffeine resulted in an approximately 100% increased net hepatic glucose uptake. Despite this, whole-body glucose level did not improve, implying that there was a decreased peripheral glucose uptake. Thus, the hepatic response to caffeine did not contribute to the exaggerated blood glucose response associated with caffeine and carbohydrate ingestion, but rather could minimize it.

The few studies of either adenosine or caffeine on hepatic carbohydrate metabolism support that the  $A_1$  receptor enhances glucose release and caffeine reduces hepatic glucose output, but the metabolic factors involved have not been established. Buxton et al. (1987) reported that adenosine stimulated glucose release from isolated rat liver; they also found that adenosine stimulated glycogen phosphorylase activity in isolated hepatocytes. However, when Pencek et al. (2004) infused caffeine and glucose into the portal vein of conscious dogs, they found that the ratio of hepatic glycogen synthase to phosphorylase activity did not change. However, the level of hepatic glucose 6-phosphatase increased and lactate production increased by 40%. This may account for the observation (Graham et al. 2000) that caffeine consumption by human subjects resulted in elevated arterial lactate levels both at rest and during exercise despite no change in leg lactate production. We are not aware of any investigations of liver carbohydrate metabolism and coffee; however, both CGA (Arion et al. 1997) and a derivative (Herling et al. 1999) have been found to inhibit hepatic glucose 6-phosphatase, a key reaction in the production of glucose by the liver. Furthermore, Herling et al. (1999) also demonstrated a dose-dependent reduction in blood glucose level in rats exposed to a synthetic derivative of CGA. As mentioned previously, infusing rats with a

synthetic quinide during a euglycemic–hyperinsulinemic clamp increased the glucose infusion rate by approximately 40% (Shearer et al. 2003). There was no change in the glucose uptake of a variety of skeletal muscles or the heart and they proposed that this positive impact on whole-body insulin sensitivity was due to a reduced net hepatic glucose output. This presents the possibility that caffeine and other components of coffee could all reduce hepatic glucose release.

### 3.5 *Gastrointestinal System*

While skeletal muscle, the pancreas, and the liver have been recognized as essential tissues in modulating insulin responses, intestinal-derived factors are now believed to play an important role in carbohydrate homeostasis. Two peptides, glucagon-like peptide-1 (GLP-1) and glucose-dependent insulinotropic peptide (GIP), can enhance glucose-induced insulin secretion of  $\beta$  cells. These incretin hormones are secreted from the gastrointestinal tract and their resulting actions are mediated through G-protein-coupled receptors in a variety of tissues, most importantly on the pancreas. In humans, incretin actions on pancreatic  $\beta$  cells are thought to account for approximately 50–70% of the total postprandial insulin secretion (Baggio and Drucker 2007).

Both dietary carbohydrates and lipids stimulate GLP-1 and GIP secretion probably through direct interaction between nutrients and cells. Potential mechanisms of incretins' insulinotropic effects include activation of insulin gene transcription, enhanced messenger RNA stability, and improved insulin bioavailability (Baggio and Drucker 2007). In addition to well-documented effects on the pancreas, extra-pancreatic incretin actions have also been described. For instance, Sandhu et al. (1999) described that infusion of GLP-1 during a hyperinsulinemic clamp in depancreatized dogs increased the glucose infusion rate and whole-body glucose utilization. This insulin-sensitivity effect was absent during a clamp with lower (fasting) levels of insulin. The main tissue involved in glucose disposal is skeletal muscle, and they found evidence of GLP-1 receptor messenger RNA in skeletal muscle. This contrasts with other studies that did not find the presence of GLP-1 receptors in muscle in humans and rats (Bullock et al. 1996; Wei and Mojsov 1996). This could suggest a species-specific expression of GLP-1 receptors and actions; therefore, additional studies are necessary before this finding can be extended to humans.

These hormones are currently the focus of a great deal of pharmacological investigation regarding T2D. Intravenous injection of GLP-1 in fasted, T2D patients increased the level of C-peptide concurrently with the level of insulin, suggesting that at least some of the effects of incretin mimetics occur at the pancreas level, potentiating insulin secretion after a meal (Meier et al. 2004). In addition, with long-term treatment, GLP-1 mimetics and dipeptidyl peptidase-4 inhibitors can reduce the level of glycosylated hemoglobin (HbA<sub>1c</sub>) by 0.8–1.75% (Drucker and Nauck 2006).



To the best of our knowledge, the impact of caffeine on the secretion of incretins has not been investigated. However, Johnston et al. (2003) examined the effects of ingesting coffee or water concurrent with 25 g dextrose. They showed that coffee consumption resulted in a blunted GIP response and there was a trend for decaffeinated coffee to induce a lower response than that of regular coffee. However, these findings are controversial as recently Greenberg et al. (2009) failed to find any effect of either decaffeinated coffee or regular coffee on GIP responses for the first 2 h of an OGTT challenge.

Johnston et al. (2003) suggested that antioxidants present in coffee, especially CGA, could act to delay glucose absorption, hence explaining the blunted GIP secretion with coffee. It is not known if caffeine itself could have an impact on carbohydrate homeostasis prior to its absorption from the gastrointestinal system. The effects reported for caffeine on the gastrointestinal system have been inconsistent regarding responses such as gastric secretion, gastroesophageal reflux, and jejunal secretions. Van Nieuwenhoven et al. (2000) conducted a comprehensive investigation of gastrointestinal function and caffeine. However, it was a complicated study as the caffeine was ingested with a carbohydrate–electrolyte solution and after a rest period, 90 min of steady-state exercise was performed. They found that the caffeine ingestion did not affect gastric emptying, orocecal transit time, gastroesophageal reflux, gastric pH, or gastrointestinal transit time. While intestinal permeability was unaffected, intestinal glucose uptake was greater following the caffeine–carbohydrate–electrolyte ingestion as compared with carbohydrate–electrolyte consumption alone. The authors speculated that caffeine stimulated the sodium–glucose-linked transporter that is associated with glucose uptake by the jejunum. The authors also pointed out that the lack of detrimental effects of caffeine could be due to the low dose (120 mg). Yeo et al. (2005) reported that coingesting caffeine and a glucose solution during prolonged exercise resulted in the greatest increase in carbohydrate oxidation and this was due to increased exogenous carbohydrate oxidation. However, this is in contrast to previous reports in which it was stated that caffeine did not influence carbohydrate absorption or oxidation during exercise (Jacobson et al. 2001; Sasaki et al. 1987).

Incretin effects on carbohydrate homeostasis likely include both pancreatic and extrapancreatic responses. It is unknown if caffeine or coffee affects the incretin response. This is an area of science that is in its infancy regarding the incretin impact both on the pancreatic  $\beta$  cells and on other tissues, especially skeletal muscle. Given the emphasis that is placed on altered incretin responses in the T2D state, the putative roles of coffee and caffeine need to be explored in detail. While caffeine may have effects on gastrointestinal glucose absorption, the understanding is far from complete at this time. The possibility that aspects of coffee may blunt some of the incretin response needs to be investigated further. It is not clear how this would be associated with the positive effect of chronic consumption of coffee on the risk for T2D. It would also seem highly unlikely that increased absorption would result in the increased glucose AUC and insulin resistance, but it could be a contributing factor.

### 3.6 Pancreatic $\beta$ Cells

With regard to pancreatic  $\beta$  cells, caffeine could also have a direct effect on insulin secretion. The  $\beta$  cells have  $A_1$  receptors that inhibit insulin secretion (Hilliare-Buys et al. 1989; Töpfer et al. 2008) and exposure to caffeine can increase insulin release (Töpfer et al. 2008). In studies in which caffeine and OGTT challenges are given to humans, normally the caffeine is ingested 1 h prior to the OGTT. The reports summarized in Table 1 did not observe an increase in the level of either insulin or C-peptide prior to the OGTT. Furthermore, if the caffeine did promote insulin secretion, this should lower the blood glucose level, but this was not observed either prior to or during the OGTT. Finally, during insulin clamp studies the insulin secretion would not be a factor. Thus, it appears that a caffeine-induced insulin secretion is not a factor in carbohydrate management following a single ingestion of caffeine. In contrast, Wu et al. (2005) found that habitual consumption of coffee was inversely associated with fasting concentrations of C-peptide. This result was found for both decaffeinated and caffeinated coffee, but not for tea, suggesting that bioactive factors in coffee, other than caffeine, accounted for this change. It is noteworthy that the C-peptide reduction was greater for obese (27%) or overweight (20%) women compared with normal-weight women (11%).

Figure 1 summarizes the known effects of caffeine and caffeinated coffee that may acutely affect carbohydrate homeostasis. The major site of the insulin resistance is clearly skeletal muscle and it appears that the metabolic alteration is initially the inhibition of uptake of glucose and subsequently its storage, while carbohydrate oxidation is unaffected. An increase in SNS activity or at least the presence of a normal concentration of epinephrine appears to be important, but role these play has not been established. In addition, there may be an enhanced gastrointestinal absorption of glucose and there appears to be a decreased net hepatic glucose output. So little is known regarding the habitual effects of the noncaffeine components of coffee on these tissues and mechanisms to allow us to speculate regarding how coffee consumption decreases the risk for T2D. Van Dam et al. (2006) proposed that since the relative risk for T2D is similar in obese and nonobese coffee drinkers, this must mean that the absolute reduction in risk is greater for the obese coffee drinkers and they may particularly benefit from coffee!

## 4 Caffeine, Coffee, and Cardiovascular Disease: Epidemiology and Mechanisms

The association of caffeine and CVD is reviewed in depth in Riksen et al. (2010). Once caffeine enters the systemic circulation, every tissue of the body is exposed to this substance. It is well known that caffeine and coffee ingestion result in a modest increase in vascular resistance and thus an increase in blood pressure, particularly the diastolic pressure (Jee et al. 1999; Smits et al. 1983, 1991; Quinlan et al. 2000). Smits et al. (1991) also showed that a caffeine challenge as little as 1 mg/kg (plasma

concentration of 3.1  $\mu\text{mol/L}$ ) was enough to increase the pressure and higher levels of caffeine did not result in a larger response. Furthermore, caffeine almost completely attenuated the hemodynamic responses to the adenosine transport antagonist dipyridamole, suggesting that indeed caffeine is acting as an adenosine receptor antagonist. Similarly, Quinlan et al. (2000) confirmed that plasma caffeine concentrations of 5–10  $\mu\text{mol/L}$  caused an increase in blood pressure, but the response was quite modest. Graham et al. (2000) reported that caffeine ingestion resulted in a 5 mm Hg increase in mean blood pressure and a 21% increase in leg vascular resistance but no difference in leg blood flow or oxygen uptake. While these effects could also be the result of increased levels of epinephrine (see the discussion earlier), Smits et al. (1990, 1991) demonstrated that the hemodynamic effects of caffeine were independent of a catecholamine effect.

Martin et al. (2006a) reported that approximately half of their healthy subjects demonstrated a blunted response to a forearm vascular conductance test and an adenosine infusion. The exercise hyperemia was the same in both groups but when nitric oxide production was inhibited, this affected only the “responders.” They concluded that those with a blunted response to adenosine also had less contribution from nitric oxide. When the methylxanthine derivative aminophylline was employed, the response to adenosine was markedly blunted, but only in the adenosine “responders” (Martin et al. 2006b). This suggests that there could be caffeine “responders” and “nonresponders”. However, this does not appear to be due to a CYP1A2\*1F polymorphism as the differences were abolished when the subjects were infused with dipyridamole (Martin et al. 2007). The scientists proposed that the nonresponders could have a greater activity in the adenosine transporter and thus less endogenous adenosine available for the receptors.

Caffeinated coffee consumption has also been extensively investigated in relation to CVD risk. Early epidemiology studies reported increased risk of CVD in regular coffee drinkers (reviewed in Higdon and Frei 2006; Ranheim and Halvorsen 2005; van Dam 2008). This was based on the knowledge that acute consumption of coffee leads to a small increase in blood pressure and may be associated with hypertension in the long term (Jee et al. 1999). Despite these early suggestions, recent evidence does not support a role of coffee consumption in CVD prevalence. It is possible that the early studies did not adequately control for unhealthy behaviors associated with coffee drinking (e.g., cigarette smoking, physical inactivity, and alcohol consumption), therefore mistakenly associating increased CVD risk with coffee consumption.

In recent prospective studies (summarized in Table 4), high coffee consumption (four or more cups daily) did not increase the risk of heart-failure hospitalization/mortality (Ahmed et al. 2009), coronary heart disease (Kleemola et al. 2000), or cardiovascular mortality (Mineharu et al. 2009). Furthermore, coffee consumption was not associated with elevated CVD risk in postmyocardial infarction patients (Silletta et al. 2007; Mukamal et al. 2009). Similarly, in women, no significant relationships were found between the risks of stroke (Lopez-Garcia et al. 2009), myocardial infarction (Rosner et al. 2006), cardiovascular mortality (Mineharu et al. 2009), and moderate-to-heavy coffee consumption (four or five cups daily).

**Table 4** Summary of contemporary studies examining the effects of long-term moderate and heavy coffee consumption on cardiovascular disease risk and mortality

Reference	Study design	Subjects	Dependent variable	Results	Conclusions
Jee et al. (1999)	Meta-analysis	11 trials; 522 subjects; mean duration: 56 days; mean coffee consumption: 5 cups/day	Systolic and diastolic BP	Systolic BP increased by 2.4 mm Hg Diastolic BP increased by 1.2 mm Hg	Consumption of coffee is associated with increased BP
Kleemola et al. (2000)	Cohort study; 10-year FU	20,179 Finnish men and women; age 30–59	Fatal and nonfatal CHD; total mortality	Adjusted RR for CHD events and mortality did not differ between low (1–3 cups/day) and high ( $\geq 7$ cups/day) coffee consumers, in both men and women	Coffee consumption does not increase the risk of CHD or death
Hammar et al. (2003)	Case-control study	Swedish men and women; age 45–70	First nonfatal MI	Increased incidence of MI when consuming boiled coffee vs. filtered coffee: Men RR: 1.41 Women RR: 1.63	Consumption of boiled coffee increased incidence of first nonfatal MI
Bidel et al. (2006)	Cohort study; 20.8-year FU	3,837 Finnish patients with T2D, age: 25–74	CVD, CHD, and stroke mortality	HR for total mortality: 0–2 cups/day: 1.00 3–4 cups/day: 0.77 5–6 cups/day: 0.68 $\geq 7$ cups/day: 0.70 Same trend for CVD and CHD mortality	Coffee drinking in T2D patients is associated with reduced total, CVD, and CHD mortality
Rosner et al. (2006)	Cohort study; 5-year FU	32,650 Swedish women; age 40–74	MI events	RR for MI compared with 0–4 cups coffee/week: 5–7 cups/week: 0.84 2–3 cups/day: 0.65 4–5 cups/day: 0.64 $\geq 6$ cups/day: 0.65	Coffee consumption does not increase MI risk. Drinking $\geq 5$ cups/week decreased MI risk nonsignificantly
Silletta et al. (2007)	Cohort study; 42-month FU	11,231 Italian patients with recent MI	Fatal and nonfatal CVD events	RR for CVD event compared with noncoffee consumers: $< 2$ cups/day: 1.02	No association between moderate coffee consumption and CVD events in post-MI patients

*(continued)*

Table 4 (continued)

Reference	Study design	Subjects	Dependent variable	Results	Conclusions
Ahmed et al. (2009)	Cohort study; 9-year FU	37,315 healthy men at the baseline	Heart failure events or mortality	2-4 cups/day: 0.91 >4 cups/day: 0.88 RR compared with $\leq 1$ cup coffee/day: 2 cups/day: 0.87 3 cups/day: 0.89 4 cups/day: 0.89 $\geq 5$ cups/day: 0.89	High coffee consumption does not increase risk of heart failure
Lopez-Garcia et al. (2009)	Cohort study; 24-year FU	83,076 healthy women at the baseline	Stroke events	RR compared with $\leq 1$ cup coffee/month: 1 cup/month to 4 cups/week: 0.98 5-7 cups/week: 0.88 2-3 cups/day: 0.81 $\geq 4$ cups/day: 0.80	Long-term coffee consumption may modestly reduce risk of stroke in women
Mukamal et al. (2009)	Cohort study; 7-9-year FU	1,369 Swedish patients with first acute MI	Mortality after first acute MI	HR compared with $< 1$ cup coffee/day: 1 to $< 3$ cups/day: 0.68 3 to $< 5$ cups/day: 0.56 5 to $< 7$ cups/day: 0.52 $\geq 7$ cups/day: 0.58	Coffee consumption at the time of hospitalization was inversely related to mortality after the first MI
Zhang et al. (2009)	Cohort study; 24-year FU	7,170 women with T2D, but free of CVD or cancer at the baseline	CVD and total mortality	Compared with noncoffee consumers, RR for CVD: $\geq 4$ cups CC/day: 0.76 $\geq 2$ cups DECAF/day: 0.96 Same trend for total mortality	CC and DECAF consumption was not associated with increased CVD or all-cause mortality

In all studies, relative risks and hazard risks were adjusted for possible confounders. None of the studies supported that long-term coffee consumption (except for consumption of boiled coffee) increase cardiovascular disease events. Some studies (Bidel et al. 2006; Rosner et al. 2006; Lopez-Garcia et al. 2009; Mukamal et al. 2009) even suggested protective effects of coffee. Except in the Zhang et al. (2009) paper, no distinctions were made between caffeinated and decaffeinated coffee. *BP* blood pressure, *CC* caffeinated coffee, *CHD* coronary heart disease, *CVD* cardiovascular disease, *DECAF* decaffeinated coffee, *FU* follow-up, *HR* hazard ratio, *MI* myocardial infarction, *RR* relative risk, *T2D* type 2 diabetes mellitus

Some authors have even suggested that coffee could act protectively against CVD (Lopez-Garcia et al. 2008, 2009; Rosner et al. 2006), but more research is needed to confirm this hypothesis. In summary, it does not appear that coffee is detrimental to CVD in men and women, and there is a possibility that coffee may protect against CVD (Lopez-Garcia et al. 2008).

T2D increases the risk of CVD by up to threefold (Stamler et al. 1993), and thus nutritional interventions that alter this risk are very important. The impact of coffee consumption on CVD prevalence in patients with T2D has been investigated. Zhang et al. (2009) reported that neither caffeinated nor decaffeinated coffee increased the risk for total or CVD mortality in diabetic women consuming four or more cups of coffee per day (relative risk for coffee 0.80 and for decaffeinated coffee 0.76). Moreover, neither caffeinated nor decaffeinated coffee increased blood lipid levels, while only decaffeinated coffee (two or more cups daily) reduced glycosylated hemoglobin levels. Another prospective study (mean follow-up 20.8 years) documented that coffee consumption in Finnish diabetic men significantly reduced the relative risk of total mortality, CVD, and coronary heart disease (respective relative risks for five or six cups of coffee per day of 0.68, 0.70, and 0.70) (Bidel et al. 2006).

The possible negative relationship between coffee consumption and CVD risk would likely be mediated through antioxidants present in coffee. As discussed previously, CGAs and quinides are the main candidates with antioxidative properties in coffee. Reduction of inflammation and endothelial dysfunction due to antioxidants in coffee may partly explain the benefits of coffee. Nevertheless, the association between long-term coffee consumption and CVD is still equivocal. Several confounders may explain the discrepancy in the data presented. First, the difference between the effects of caffeinated coffee and decaffeinated coffee has not been investigated. For instance, these beverages have differential effects on hypertension, a mechanism that could be involved in a potential coffee-induced increase in CVD risk. Indeed, acute ingestion of alkaloid caffeine (200–250 mg) can increase systolic and diastolic blood pressure by approximately 2.4 and 1.2 mm Hg, respectively (Jee et al. 1999). The ingestion of caffeinated coffee induces a similar, but lessened, effect, while decaffeinated coffee does not increase blood pressure (Ricksen et al. 2009). Therefore, the hypertensive effects of coffee are attributed to caffeine. Furthermore, only partial habituation to the pressor action of coffee occurs, so even regular coffee drinkers experience the acute hypertensive effects of caffeine. On the basis of this knowledge, the lack of control for caffeine content in coffee may have introduced error and variation in previous investigations. Given the increased risk of CVD in T2D patients and the potential for regular coffee ingestion to reduce the risk for T2D, it becomes even more important to resolve the association between coffee consumption and CVD.

Recent studies have also documented that genetic polymorphisms may modify the association between coffee consumption and CVD risks. However, it should be noted that the data are particular to caffeine rather than to coffee in a general sense. For instance, intake of coffee was only associated with myocardial infarction risk in a subpopulation carrying the CYP1A2\*1F allele (Cornelis et al. 2006). CYP1A2 is

the gene encoding for cytochrome P450 1A2, a key liver enzyme that is responsible for metabolizing caffeine. Individuals carrying the CYP1A2\*1F allele are slow metabolizers of caffeine, compared with individuals who are homozygous for the more prevalent CYP1A2\*1A allele. Hypothetically, slower metabolism of caffeine may increase plasma caffeine concentration and exacerbate a possible detrimental effect on myocardial function. However, this study should be considered as preliminary, as its conclusions have been questioned (Ingelman-Sundberg et al. 2006). Nevertheless, it is likely that a number of polymorphisms play roles in the responses to caffeine and other coffee components, and it will require an extensive body of work in order to resolve these components. For instance, the same CYP1A2\*1F polymorphism could exacerbate the hypertensive effects of coffee, whereas this effect was nonsignificant in the homozygous CYP1A2\*1A subsample (Palatini et al. 2009). Moreover, individuals homozygous for the AA genotype in the CYP1A2 gene did not benefit from the protective effect of coffee on breast cancer (Kotsopoulos et al. 2007) and were more sensitive to its deleterious effects on ovarian cancer (Goodman et al. 2003). The importance of genotype in the strength of the association between caffeine, coffee, and health outcomes is a very new field of research. This present knowledge is very preliminary and undoubtedly represents a very small fraction of the gene–environment interactions that pertain to caffeine and/or coffee. Even within the one situation of the impact of caffeine on insulin sensitivity, there are many factors where a transporter, a receptor, or a second messenger may be affected by a single nucleotide polymorphism. Therefore, the inevitable genetic variability in a population may explain some of the variation and discrepancy in the epidemiology studies presented here.

The method of brewing is also known to modulate the composition of coffee. Cafestol and kahweol are two compounds present in coffee that are known to increase total and low-density lipoprotein cholesterol, two important risk factors for CVD (Higdon and Frei 2006). However, these two molecules are trapped in filter paper during coffee preparation, in such way that filter coffee contains up to 80–100-fold less cafestol and kahweol compared with boiled coffee (Jee et al. 2001). Consequently, different types of coffee brewing techniques may have influenced the data presented and this is a clear illustration of an effect of coffee that is independent of caffeine. Total cholesterol, low-density lipoprotein cholesterol, and apolipoprotein B levels increased in subjects randomized to consumed boiled coffee for 79 days compared with participants who did not drink coffee or who consumed filtered coffee (Van Dusseldorp et al. 1991). In accordance with its effects on serum lipids, the consumption of boiled coffee was associated with increased incidence of myocardial infarction compared with the consumption of filtered coffee in both Swedish males and females (Hammar et al. 2003). In addition, the association between the elevation in serum lipids and coffee was stronger in hyperlipidemic patients (Jee et al. 2001). Taken together, these data suggest that the risk of dyslipidemia following coffee consumption may be exacerbated by drinking boiled coffee and in at-risk populations.

Coffee can also increase the blood content of homocysteine, an amino acid that is associated with cardiovascular risk (Antoniade et al. 2009). Coffee had a



dose-dependent positive relationship with homocysteine content in several clinical trials (Nygard et al. 1997; reviewed in Higdon and Frei 2006) and this effect has been attributed, at least in part, to caffeine and CGA (reviewed in Ranheim and Halvorsen 2005). It is unknown how brewing techniques and/or genetic polymorphisms may influence the coffee–homocysteine relationship.

In summary, the health effects of coffee on CVD are still equivocal, although recent studies have pointed toward a neutral and/or a positive effect of long-term coffee consumption for most individuals. On one side, there is evidence that at least one type of coffee (boiled) can be a risk factor for CVD and that certain individuals could be more sensitive to the effects of caffeine. In contrast, there is a possible protective effect of coffee on CVD due to the presence of antioxidants, namely, CGAs and quinides, in coffee.

## 5 Coffee, Caffeine, and Other Health Effects

The health effects of coffee and caffeine have been extensively studied regarding CVD and T2D. Given the ubiquitous nature of the adenosine receptors and the large number of bioactive compounds in coffee and its effects on numerous tissues, coffee and caffeine are likely to modulate other health outcomes. Here, we will review the available information about the impact of caffeine or coffee consumption on weight control, different forms of cancer, osteoporosis, and cognitive diseases.

There are many weight-loss-oriented commercial products that contain caffeine. Caffeine ingestion can stimulate epinephrine secretion and can mobilize FFAs. This would appear to lend support to the concept that caffeine would also increase metabolism and hence promote weight loss. Fundamental to this topic, any weight-loss supplement must either decrease energy intake (i.e., decrease appetite) or increase energy expenditure. To accomplish the latter, there must either be an increased demand for ATP or a decrease in the efficiency of the production of ATP (i.e., the oxygen cost of producing ATP must become greater perhaps owing to uncoupling proteins in mitochondria). Greenberg et al. (2005) reported that the negative association of coffee consumption and reduced risk for T2D was also associated with weight loss and that this could account for some of the benefit of coffee consumption. However, a direct association with coffee or caffeine and weight control has proven to be elusive. While an increase in energy output of merely a few kilocalories per day can result in a large difference in body weight over years, it is extremely difficult to measure accurately in a short-term experiment. The necessary long-term, longitudinal investigations have not been conducted.

There are a number of accounts that acute caffeine or coffee ingestion can increase the resting or diet-induced thermogenesis by approximately 10% (Acheson et al. 1980; Dulloo et al. 1989; Arciero et al. 1995; Bracco et al. 1995; Astrup et al. 1990; Jung et al. 1981). This would be a significant contribution to the energy balance if it was maintained over decades of caffeine/coffee ingestion. It is interesting to note



that comparisons of lean and obese or postobese subjects consistently report that the effect is less in obese and postobese individuals (Acheson et al. 1980; Bracco et al. 1995; Jung et al. 1981). Acheson et al. (2004) found that a large (10 mg/kg) dose of caffeine increased the metabolic rate by 13%, doubled the turnover of lipids, and modestly increased FFA oxidation. However, Bracco et al. (1995) noted that 24 h after caffeine ingestion, the response was blunted. Lopez-Garcia et al. (2006) conducted an epidemiological, prospective study of body weight and caffeine intake of almost 65,000 people for 12 years. While caffeine intake was related to weight loss, the effect was very modest, less than 0.5 kg during the 12 years. There is very little evidence that coffee or caffeine can promote significant changes in body weight.

Recently, Thom (2007) evaluated a new product (Coffee Slender) that contains a green coffee bean extract that is rich in CGAs. The preliminary assessment demonstrated that a single ingestion of the product resulted in a modest decreased blood glucose response to a carbohydrate challenge. More notably, ingestion of the product for 12 weeks resulted in a 5.4-kg weight loss. It was speculated that the CGAs inhibited that absorption of glucose, but no data were provided to support this hypothesis and the findings should be considered preliminary.

The strongest beneficial association between coffee and cancer is perhaps that for liver cancer. Several cohort, case-control, and meta-analysis investigations documented a decreased risk of hepatocellular carcinoma with coffee consumption (Tanaka et al. 2007; Bravi et al. 2007; Montella et al. 2007; Shimazu et al. 2005; Hu et al. 2008; Inoue et al. 2005, 2009; Kurozawa et al. 2005; Gallus et al. 2002a). This negative, dose-dependent association is evident even in low consumers (one cup of coffee daily), but was not reported with tea or decaffeinated coffee in one study (Montella et al. 2007). The underlying mechanisms for this strong association are still unknown, but the beneficial impacts of coffee on liver function have also been documented for liver enzyme function (Ruhl and Everhart 2005a; Honjo et al. 2001) and chronic liver disease (Ruhl and Everhart 2005b; Gallus et al. 2002b; Freedman et al. 2009), suggesting that coffee acts on the whole spectrum of liver function. As with CVD and T2D, one cannot dismiss the effects of antioxidants as a plausible mechanism. Moreover, owing to the lack of differentiation between caffeinated and decaffeinated coffee (except in one study) in the previously reviewed studies, it is not possible to say whether the protective effects are related to the caffeine content of coffee.

There have been some suggestions that coffee consumption may decrease the risk of other diseases, such as colorectal, breast, lung, and bladder cancers. The research did not find a strong association between long-term coffee consumption and any of those types of cancer (La Vecchia and Tavani 2007). The relative risk for colorectal cancer was between 0.91 and 1.0 in high-coffee versus low-coffee consumers (not significant) (Je et al. 2009; Naganuma et al. 2007; Larsson et al. 2006; Michels et al. 2005). Similarly, most large-scales studies (Ishitani et al. 2008; Ganmaa et al. 2008), but not all (Baker and Beehler 2006), did not report significant relationships between coffee and breast cancer in the general public. However, coffee may be protective in specific populations, such as postmenopausal women (Ganmaa et al. 2008) or women who carry high-risk mutations BRCA1 and BRCA2

(Nkondjock et al. 2006). Similar equivocal results have been reported for lung (Kabagambe and Wellons 2009) and bladder (Zeegers et al. 2001; Villanueva et al. 2009) cancers, but the strong association between coffee drinking and smoking, a major risk factor for cancer, complicates data interpretation. After adjustments for smoking, the associations between coffee consumption and lung and bladder cancers were much less pronounced (Villanueva et al. 2009; Pelluchi and La Vecchia 2009; Kabagambe and Wellons 2009). In summary, there are sparse data to support a protective effect of coffee on cancer risks (except for liver cancer), but the current data clearly show that coffee is not an independent risk factor for cancer.

Coffee has also been suggested to be protective against some cognitive diseases (for a detailed discussion see Sawynok 2010). The strongest evidence is related to Parkinson's disease. Consumption of caffeinated coffee (Hu et al. 2007; Tan et al. 2003; Ascherio et al. 2001, 2003; Hernan et al. 2002), tea (Hu et al. 2007; Tan et al. 2003; Ascherio et al. 2001), but not decaffeinated coffee (Ascherio et al. 2001), was linked to a dose-dependent reduction in the incidence of Parkinson's disease. The hazard ratios for the consumption of five or more cups of coffee or three or more cups of tea daily were 0.40 and 0.41, respectively (Hu et al. 2007). In addition, the association between caffeine and Parkinson's disease may be modified by the use of hormone replacement therapy in postmenopausal women. In these women, caffeine intake increased Parkinson's disease risk, whereas in women who did not use hormone replacement therapy, caffeine was protective (Ascherio et al. 2003). Similar caffeine-induced beneficial effects have been suggested for Alzheimer's disease, but the evidence in humans is very sparse. Coffee consumption was inversely associated with Alzheimer's disease in preliminary investigations (Barranco Quintana et al. 2007; Maia and de Mendonca 2002), but more research is needed to confirm this link. De Felice et al. (2009) investigated the role of insulin in Alzheimer's disease with highly differentiated, cultured, hippocampal nerve cells. It is interesting to note that insulin completely prevented the development of synapse abnormalities that are characteristic of Alzheimer's disease and furthermore, this protection was potentiated by rosiglitazone, an insulin-sensitizing drug for T2D treatment. (This has led to the lay press referring to Alzheimer's disease as type 3 diabetes.) While it is quite speculative, given that habitual heavy intake of coffee leads to a decreased risk of T2D, it could be that a positive association between coffee intake and Alzheimer's disease could be associated with increased insulin sensitivity.

Despite strong epidemiological evidence highlighting the benefits of coffee in many health aspects, coffee is nonetheless associated with some negative health outcomes. Most investigations (Harris and Dawson-Hugues 1994; Rapuri et al. 2001; Hernandez-Avila et al. 1991; Kiel et al. 1990; Hallström et al. 2006), but not all (Lloyd et al. 2000), reported that high caffeine consumption (250–450 mg) is associated with increased risk of low bone mineral density, osteoporosis, and osteoporotic fractures in middle-aged women. This situation may be exacerbated in women with low calcium intake (Harris and Dawson-Hugues 1994; Hallström et al. 2006), in lean (BMI < 25.1) subjects (Korpelainen et al. 2003), and in

women who carry a polymorphism in the vitamin D receptor gene (Rapuri et al. 2001).

Despite its wide acceptance and consumption, caffeine remains a drug, and, as such, may be associated with adverse effects related to abuse, dependence, and with withdrawal. In some individuals, caffeine at doses that can be found in several cups of coffee can be associated with harmful side effects, such as headache, tachycardia, tremor, insomnia, nausea, and diarrhea (Higdon and Frei 2006). In addition, caffeine withdrawal has been clearly characterized in patients who abstain from doses as low as 100 mg caffeine daily. The onset of caffeine withdrawal occurs 12–24 h after abstinence and can last up to 9 days. The most common symptoms include headache, fatigue, decreased alertness, drowsiness, depressed mood, irritability, and difficulty concentrating (Juliano and Griffiths 2004).

Consumption of caffeine and caffeinated coffee has been associated with several health outcomes. It appears that coffee may be protective not only against T2D and CVD, but also against liver cancer, Parkinson's disease, and Alzheimer's disease. On the other hand, coffee consumption is associated with adverse effects for osteoporosis and caffeine-withdrawal symptoms. Finally, coffee's relationship with several other conditions, such as bladder, colorectal, breast, and lung cancers, is still equivocal and deserves further attention. With the exception of T2D, seldom has the difference between coffee and caffeine been addressed. In any case, it is well recognized that caffeine and coffee are both bioactive substances that have extended effects on several tissues, organs, systems, and ultimately on human health.

## 6 Summary

The topic of methylxanthines and human health is founded by coffee, the major food source of caffeine, having many bioactive compounds, some of which have antioxidant functions. There is a paradox as consumption of either caffeine or caffeinated coffee results in a marked insulin resistance and yet habitual coffee consumption repeatedly been reported to reduce the risk for T2D. There is strong evidence that caffeine reduces insulin sensitivity in skeletal muscle and this may be due to a combination of direct antagonism of A<sub>1</sub> receptors and  $\beta$ -adrenergic stimulation due to increased sympathetic activity. Caffeine may also induce reduced hepatic glucose output. With the exception of bone mineral, there is little evidence that caffeine impacts negatively on other health issues. Coffee does not increase the risk of CVDs or cancers and there is some evidence suggesting a positive relationship for the former and for some cancers, particularly hepatic cancer. There is limited evidence that caffeine or coffee is effective in promoting weight loss, but there is some evidence that coffee may have a positive effect on neurodegenerative diseases.

**Acknowledgements** The work by the authors was supported by the Natural Science and Engineering Research Council of Canada (NSERC). M.-S.B. received an Ontario Graduate Scholarship.

## References

- Acheson KJ, Zahorska-Markiewicz B et al (1980) Caffeine and coffee: their influence on metabolic rate and substrate utilization in normal weight and obese individuals. *Am J Clin Nutr* 33(5):989–997
- Acheson KJ, Gremaud G et al (2004) Metabolic effects of caffeine in humans: lipid oxidation or futile cycling? *Am J Clin Nutr* 79(1):40–46
- Adeney KL, Williams MA et al (2007) Coffee consumption and the risk of gestational diabetes mellitus. *Acta Obstet Gynecol Scand* 86(2):161–166, Abstract
- Agardh EE, Carlsson S et al (2004) Coffee consumption, type 2 diabetes and impaired glucose tolerance in Swedish men and women. *J Intern Med* 255(6):645–652
- Ahmed HN, Levitan E et al (2009) Coffee consumption and risk of heart failure in men: an analysis from the cohort of Swedish men. *Am Heart J* 158:667–672
- Andrade-Cetto A, Wiedenfeld H (2001) Hypoglycemic effect of *Cecropia obtusifolia* on streptozotocin diabetic rats. *J Ethnopharmacol* 78(2–3):145–149
- Antoniade C, Antonopoulos AS et al (2009) Homocysteine and coronary atherosclerosis: from folate fortification to the recent clinical trials. *Eur Heart J* 30(1):6–15
- Arciero PJ, Gardner AW et al (1995) Effects of caffeine ingestion on NE kinetics, fat oxidation, and energy expenditure in younger and older men. *Am J Physiol* 268(6Pt1):E1192–E1198
- Arion WJ, Canfield WK et al (1997) Chlorogenic acid and hydroxynitrobenzaldehyde: new inhibitors of hepatic glucose 6-phosphatase. *Arch Biochem Biophys* 339(2):315–322
- Ascherio A, Zhang SM et al (2001) Prospective study of caffeine consumption and risk of Parkinson's disease in men and women. *Ann Neurol* 50(1):56–63
- Ascherio A, Chen H et al (2003) Caffeine, postmenopausal estrogen, and risk of Parkinson's disease. *Neurology* 60(5):790–795
- Astrup A, Toubro S et al (1990) Caffeine: a double-blind, placebo-controlled study of its thermogenic, metabolic, and cardiovascular effects in healthy volunteers. *Am J Clin Nutr* 51(5):759–767
- Baggio LL, Drucker DJ (2007) Biology of incretins: GLP-1 and GIP. *Gastroenterology* 132(6):2131–2157
- Baker JA, Beehler GP (2006) Consumption of coffee, but not black tea, is associated with decreased risk of premenopausal breast cancer. *J Nutr* 136(1):166–171
- Barranco Quintana JL, Allam MF et al (2007) Alzheimer's disease and coffee: a quantitative review. *Neurol Res* 29(1):91–95
- Batram DS, Graham TE et al (2005) The effect of caffeine on glucose kinetics in humans – influence of adrenaline. *J Physiol* 569(1):347–355
- Batram DS, Arthur R et al (2006) The glucose intolerance induced by caffeinated coffee ingestion is less pronounced than that due to alkaloid caffeine in men. *J Nutr* 136(5):1276–1280
- Batram DS, Graham TE et al (2007a) Caffeine's impairment of insulin-mediated glucose disposal cannot be solely attributed to adrenaline in humans. *J Physiol* 583(3):1069–1077
- Batram DS, Bugaresti J et al (2007b) Acute caffeine ingestion does not impair glucose tolerance in persons with tetraplegia. *J Appl Physiol* 102(1):374–381
- Belfort R, Mandarino L (2005) Dose-response effect of elevated plasma free fatty acid on insulin signaling. *Diabetes* 54(6):1640–1648
- Bidel S, Hu G et al (2006) Coffee consumption and risk of total and cardiovascular mortality among patients with type 2 diabetes. *Diabetologia* 49:2618–2626
- Boden G, Lebed B et al (2001) Effects of acute changes of plasma free fatty acids on intramyocellular fat content and insulin resistance in healthy subjects. *Diabetes* 50(7):1612–1617
- Bonen A, Megeney LA et al (1992) Epinephrine administration stimulates GLUT4 translocation but reduces glucose transport in muscle. *Biochem Biophys Res Commun* 187(2):685–691
- Bracco D, Ferrara JM et al (1995) Effects of caffeine on energy metabolism, heart rate, and methylxanthine metabolism in lean and obese women. *Am J Physiol* 269(4 Pt 1):E671–E678
- Bravi F, Bosetti C et al (2007) Coffee drinking and hepatocellular carcinoma risk: a meta-analysis. *Hepatology* 46(2):430–435

- Bullock BP, Heller RS et al (1996) Tissue distribution of messenger ribonucleic acid encoding the rat glucagon-like peptide-1 receptor. *Endocrinology* 137(7):2968–2978
- Buxton DB, Fisher RA et al (1987) Stimulation of glycogenolysis and vasoconstriction by adenosine analogs in the perfused rat liver. *Biochem J* 248:35–41
- Camargo MC, Toledo MC et al (1999) Caffeine daily intake from dietary sources in Brazil. *Food Addit Contam* 16(2):79–87
- Challis RAJ, Budohoski L et al (1984) Effects of an adenosine-receptor antagonist on insulin-resistance in soleus muscle from obese Zucker rats. *Biochem J* 221:915–917
- Cheng JT, Chi TC et al (2000) Activation of adenosine A<sub>1</sub> receptors by drugs to lower plasma glucose in streptozotocin-induced diabetic rats. *Auton Neurosci* 83:127–133
- Cornelis MC, El-Sohemy A et al (2006) Coffee, CYP1A2 genotype, and risk of myocardial infarction. *JAMA* 295(10):1135–1141
- Daly JW, Fredholm BB (2004) Mechanisms of action of caffeine on the nervous system. In: Nehlig A (ed) *Coffee, tea, chocolate and the brain*, 1st edn. CRC Press, Boca Raton, FL
- De Felice FG, Verira MNN et al (2009) Protection of synapses against Alzheimer's-linked toxins: insulin signaling prevents the pathogenic binding of A beta oligomers. *Proc Natl Acad Sci USA* 106:1971–1976
- de Galan BE, Tack CE et al (2002) Theophylline improves hypoglycemia unawareness in type 1 diabetes. *Diabetes* 51:790–796
- Debrah K, Haigh R et al (1995) Effects of acute and chronic caffeine use on the cerebrovascular, cardiovascular and hormonal responses to orthostasis in healthy volunteers. *Clin Sci (Lond)* 89(5):475–480
- Dekker MJ, Gusba JE et al (2007) Glucose homeostasis remains altered by acute caffeine ingestion following 2 weeks of daily caffeine consumption in previously non-caffeine-consuming males. *Br J Nutr* 98:556–562
- Dixon AK, Gubitza AK et al (1996) Tissue distribution of adenosine receptor mRNAs in the rat. *Br J Pharmacol* 118(6):1461–1468
- Dresner A, Laurent D et al (1999) Effects of free fatty acids on glucose transport and IRS-1-associated phosphatidylinositol 3-kinase activity. *J Clin Invest* 103(2):253–259
- Drucker DJ, Nauck MA (2006) The incretin system: glucagon-like peptide-1 receptor agonists and dipeptidyl peptidase-4 inhibitors in type 2 diabetes. *Lancet* 368(9548):1696–1705
- Dulloo AG, Geissler CA et al (1989) Normal caffeine consumption: influence on thermogenesis and daily energy expenditure in lean and postobese human volunteers. *Am J Clin Nutr* 49(1):44–50
- Engler MB, Engler MM (2006) The emerging role of flavonoid-rich cocoa and chocolate in cardiovascular health and disease. *Nutr Rev* 64(3):109–118
- Frary CD, Johnson RK, Wang MQ (2005) Food sources and intakes of caffeine in the diets of persons in the United States. *J Am Diet Assoc* 105(1):110–113
- Fredholm BB, Bättig K et al (1999) Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol Rev* 51(1):83–133
- Freedman ND, Everhart JE et al (2009) Coffee intake is associated with lower rates of liver disease progression in chronic hepatitis C. *Hepatology* 50(5):1360–1369
- Gallus S, Bertuzzi M et al (2002a) Does coffee protect against hepatocellular carcinoma? *Br J Cancer* 87:956–959
- Gallus S, Tavani A et al (2002b) Does coffee protect against liver cirrhosis? *Ann Epidemiol* 12(3):202–205
- Ganmaa D, Willett WC et al (2008) Coffee, tea, caffeine and risk of breast cancer: a 22-year follow-up. *Int J Cancer* 122:2071–2076
- Goodman MT, Tung KH et al (2003) Association of caffeine intake and CYP1A2 genotype with ovarian cancer. *Nutr Cancer* 46(1):23–29
- Graham TE, Spriet LL (1995) Metabolic, catecholamine, and exercise performance responses to various doses of caffeine. *J Appl Physiol* 78(3):867–874
- Graham TE, Hibbert E et al (1998) Metabolic and exercise endurance effects of coffee and caffeine ingestion. *J Appl Physiol* 85(3):883–889

- Graham TE, Helge JW et al (2000) Caffeine ingestion does not alter carbohydrate of fat metabolism in human skeletal muscle during exercise. *J Physiol* 529(3):837–847
- Graham TE, Sathasivam P et al (2001) Caffeine ingestion elevates plasma insulin response in humans during an oral glucose tolerance test. *Can J Physiol Pharmacol* 79:559–565
- Greenberg JA, Axen KV et al (2005) Coffee, tea and diabetes: the role of weight loss and caffeine. *Int J Obes (Lond)* 29(9):1121–1129
- Greenberg JA, Owen DR et al (2009) Decaffeinated coffee and glucose metabolism in young men. *Diabetes Care* 33(2):278–280
- Greer F, Hudson R et al (2001) Caffeine ingestion decreases glucose disposal during a hyperinsulinemic-euglycemic clamp in sedentary humans. *Diabetes* 50(10):2349–2354
- Hallström H, Wolk A et al (2006) Coffee, tea and caffeine consumption in relation to osteoporotic fracture risk in a cohort of Swedish women. *Osteoporos Int* 17:1055–1064
- Hammar N, Andersson T et al (2003) Association of boiled and filtered coffee with incidence of first nonfatal myocardial infarction: the SHEEP and the VHEEP study. *J Intern Med* 253:653–659
- Han XX, Bonen A (1998) Epinephrine translocates GLUT-4 but inhibits insulin-stimulated glucose transport in rat muscle. *Am J Physiol* 274(4Pt1):E700–E707
- Han DH, Hansen PA et al (1998) Removal of adenosine decreases the responsiveness of muscle glucose transport to insulin and contractions. *Diabetes* 47(11):1671–1675
- Harris SS, Dawson-Hughes B (1994) Caffeine and bone loss in healthy postmenopausal women. *Am J Clin Nutr* 60:573–578
- Health Canada (2007) Caffeine. [http://www.hc-sc.gc.ca/fn-an/securit/facts-faits/caffeine\\_e.html](http://www.hc-sc.gc.ca/fn-an/securit/facts-faits/caffeine_e.html). Cited 8 Feb 2010.
- Herling AW, Burger H et al (1999) Alterations of carbohydrate and lipid intermediary metabolism during inhibition of glucose-6-phosphatase in rats. *Eur J Pharmacol* 368(1):75–82
- Hernan MA, Takkouche B et al (2002) A meta-analysis of coffee drinking, cigarette smoking, and the risk of Parkinson's disease. *Ann Neurol* 52(3):276–284
- Hernandez-Avila M, Colditz GA et al (1991) Caffeine, moderate alcohol intake, and risk of fractures of the hip and forearm in middle-aged women. *Am J Clin Nutr* 54:157–163
- Higdon JV, Frei B (2006) Coffee and health: a review of recent human research. *Crit Rev Food Sci Nutr* 46:101–123
- Hilliare-Buys D, Gross R et al (1989) Effect of pertussis toxin on A<sub>1</sub> receptor-mediated inhibition of insulin secretion. *Br J Pharmacol* 96:3–4
- Homko CJ, Cheung P et al (2003) Effects of free fatty acids on glucose uptake and utilization in healthy women. *Diabetes* 52(2):487–491
- Honjo S, Kono S et al (2001) Coffee consumption and serum aminotransferases in middle-aged Japanese men. *J Clin Epidemiol* 54(8):823–829
- Howlett K, Galbo H et al (1999) Effect of adrenaline on glucose kinetics during exercise in adrenalectomised humans. *J Phys* 519(3):911–921
- Hu G, Bidel S et al (2007) Coffee and tea consumption and the risk of Parkinson's disease. *Mov Disord* 22(15):2242–2248
- Hu G, Tuomilehto J et al (2008) Joint effects of coffee consumption and serum gamma-glutamyl-transferase on the risk of liver cancer. *Hepatology* 48(1):129–136
- Huxley R, Lee CMY et al (2009) Coffee, decaffeinated coffee, and tea consumption in relation to incident type 2 diabetes mellitus. *Arch Intern Med* 169(22):2053–2063
- Ingelman-Sundberg M, Sim SC et al (2006) Coffee, myocardial infarction, and CYP nomenclature. *JAMA* 296(7):764–765
- Inoue M, Yoshimi I et al (2005) Influence of coffee drinking on subsequent risk of hepatocellular carcinoma: a prospective study in Japan. *J Natl Cancer Inst* 97:293–300
- Inoue M, Kurahashi N et al (2009) Effect of coffee and green tea consumption on the risk of liver cancer: cohort analysis by hepatitis virus infection status. *Cancer Epidemiol Biomark Prev* 18(6):1746–1753
- Ishitani K, Lin J et al (2008) Caffeine consumption and risk of breast cancer in a large prospective cohort of women. *Arch Intern Med* 168(18):2022–2031

- Itani SI, Ruderman NB et al (2002) Lipid-induced insulin resistance in human muscle is associated with changes in diacylglycerol, protein kinase C, and IkappaB-alpha. *Diabetes* 51(17): 2005–2011
- Jacobson TL, Febbraio MA et al (2001) Effect of caffeine co-ingested with carbohydrate or fat on metabolism and performance in endurance-trained men. *Exp Physiol* 86(1):137–144
- Je Y, Liu W et al (2009) Coffee consumption and risk of colorectal cancer: a systematic review and meta-analysis of prospective cohort studies. *Int J Cancer* 124:1662–1668
- Jee SH, He J et al (1999) The effect of chronic coffee drinking on blood pressure: a meta-analysis of controlled clinical trials. *Hypertension* 33:647–652
- Jee SH, He J et al (2001) Coffee consumption and serum lipids: a meta-analysis of randomized controlled clinical trials. *Am J Epidemiol* 153:353–362
- Johansson SM, Yang JN et al (2007) Eliminating the antilipolytic adenosine A1 receptor does not lead to compensatory changes in the antilipolytic actions of PGE2 and nicotinic acid. *Acta Physiol (Oxf)* 190(1):87–96
- Johnston KL, Clifford MN et al (2003) Coffee acutely modifies gastrointestinal hormone secretion and glucose tolerance in humans: glycemic effects of chlorogenic acid and caffeine. *Am J Clin Nutr* 78(4):728–733
- Juliano LM, Griffiths RR (2004) A critical review of caffeine withdrawal: empirical validation of symptoms and signs, incidence, severity, and associated features. *Psychopharmacology* 176(1):1–29
- Jung RT, Shetty PS et al (1981) Caffeine: its effect on catecholamines and metabolism in lean and obese humans. *Clin Sci (Lond)* 60(5):527–535
- Kabagambe EK, Wellons MF (2009) Benefits and risk of caffeine and caffeinated beverages. In Eamranond P (ed) *UpToDate*, version 17.3
- Kacker S (2003) Ingestion of caffeinated coffee impairs blood glucose homeostasis in response to either high or low glycemic index cereals in non-obese males. Master thesis, University of Guelph
- Keijzers GB, DeGalan BE et al (2002) Caffeine can decrease insulin sensitivity in humans. *Diabetes Care* 25:364–369
- Kiel DP, Felson DT et al (1990) Caffeine and the risk of hip fracture: the Framingham study. *Am J Epidemiol* 132:675–684
- Kleemola P, Jousilahti P et al (2000) Coffee consumption and the risk of coronary heart disease and death. *Arch Intern Med* 160:3393–3400
- Korpelainen R, Korpelainen J et al (2003) Lifestyle factors are associated with osteoporosis in lean women but not in normal and overweight women: a population-based cohort study of 1222 women. *Osteoporos Int* 14(1):34–43
- Kotsopoulos J, Ghadiria P et al (2007) The CYP1A2 genotype modifies the association between coffee consumption and breast cancer risk among BRCA1 mutation carriers. *Cancer Epidemiol Biomark Prev* 16(5):912–916
- Kruszynska YT, Worrall DS et al (2002) Fatty acid-induced insulin resistance: decreases muscle PI3K activation but unchanged Akt phosphorylation. *J Clin Endocrinol Metab* 87(1):226–234
- Kurozawa Y, Ogimoto I et al (2005) Coffee and risk of death from hepatocellular carcinoma in a large cohort study in Japan. *Br J Cancer* 93:607–610
- La Vecchia C, Tavani A (2007) Coffee and cancer risk: an update. *Eur J Cancer Prev* 16:385–389
- Lane JD, Barkauskas CE et al (2004) Caffeine impairs glucose metabolism in type 2 diabetes. *Diabetes Care* 27(8):2047–2048
- Lane JD, Feinglos MN et al (2008) Caffeine increases ambulatory glucose and postprandial responses in coffee drinkers with type 2 diabetes. *Diabetes Care* 31(2):221–222
- Larsson SC, Bergkvist L et al (2006) Coffee consumption and incidence of colorectal cancer in two prospective cohort studies of Swedish women and men. *Am J Epidemiol* 163:638–644
- Lee S, Hudson R et al (2005) Caffeine ingestion is associated with reductions in glucose uptake independent of obesity and type 2 diabetes before and after exercise training. *Diabetes Care* 28(3):566–572

- Liang HX, Belardinelli L et al (2002) Tonic activity of the rat adipocyte A1-adenosine receptor. *Br J Pharmacol* 135(6):1457–1466
- Lloyd T, Johnson-Rollings N et al (2000) Bone status among postmenopausal women with different habitual caffeine intakes: a longitudinal investigation. *J Am Coll Nutr* 19(2): 256–261
- Lopez-Garcia E, van Dam RM et al (2006) Changes in caffeine intake and long-term weight change in men and women. *Am J Clin Nutr* 83(3):674–680
- Lopez-Garcia E, van Dam RM et al (2008) The relationship of coffee consumption with mortality. *Ann Intern Med* 148:904–914
- Lopez-Garcia E, Rodriguez-Artalejo F et al (2009) Coffee consumption and risk of stroke in women. *Circulation* 119:1116–1123
- Lovallo WR, Wilson MF et al (2004) Blood pressure response to caffeine shows incomplete tolerance after short-term regular consumption. *Hypertension* 43:760–765
- Lundsberg LS (1998) Caffeine consumption. In: Spiller JA (ed) *Caffeine*, 1st edn. CRC Press, Boca Raton, FL
- Lynge J, Hellsten Y (2000) Distribution of adenosine A1, A2a and A2b receptors in human skeletal muscle. *Acta Physiol Scand* 169:283–290
- Maia L, de Mendonca A (2002) Does caffeine intake protect from Alzheimer's disease? *Eur J Neurol* 9(4):377–382
- Martin EA, Nicholson WT et al (2006a) Bimodal distribution of vasodilator responsiveness to adenosine due to difference in nitric oxide contribution: implications for exercise hyperemia. *J Appl Physiol* 101(2):492–499
- Martin EA, Nicholson WT et al (2006b) Influences of adenosine receptor antagonism on vasodilator responses to adenosine and exercise in adenosine responders and nonresponders. *J Appl Physiol* 101(6):1678–1684
- Martin EA, Nicholson WT et al (2007) Adenosine transporter antagonism in humans augments vasodilator responsiveness to adenosine, but not exercise, in both adenosine responders and non-responders. *J Physiol* 59(Pt 1):237–245
- Matsuda M, DeFronzo RA (1999) Insulin sensitivity indices obtained from oral glucose tolerance testing: comparison with the euglycemic insulin clamp. *Diabetes Care* 22(9):1462–1470
- McCarthy MF (2005) A chlorogenic acid-induced increase in GLP-1 production may mediate the impacts of heavy coffee consumption on diabetes risk. *Med Hypotheses* 64:848–853
- McLane MP, Black PR et al (1990) Adenosine reversal of in vivo hepatic responsiveness to insulin. *Diabetes* 39(1):62–69
- McLean C, Graham TE (2002) Effects of exercise and thermal stress on caffeine pharmacokinetics in men and eumenorrhic women. *J Appl Physiol* 93:1471–1478
- Meier JJ, Goetze O et al (2004) Gastric inhibitory polypeptide does not inhibit gastric emptying in humans. *Am J Physiol Endocrinol Metab* 286:E621–E625
- Michels KB, Willet WC et al (2005) Coffee, tea, and caffeine consumption and incidence of colon and rectal cancer. *J Natl Cancer Inst* 97(4):282–292
- Mineharu Y, Koizumi A et al (2009) Coffee, green tea, black tea and oolong tea consumption and risk of mortality from cardiovascular disease in Japanese men and women. *J Epidemiol Community Health*, doi:10.1136/jech.2009.097311
- Misra A, Khurana L (2008) Obesity and the metabolic syndrome in developing countries. *J Clin Endocrinol Metab* 93:S9–S30
- Mohr T, Van Soeren M et al (1998) Caffeine ingestion and metabolic responses of tetraplegic humans during electrical cycling. *J Appl Physiol* 85(3):979–985
- Moisey LL, Kacker S et al (2008) Caffeinated coffee consumption impairs blood glucose homeostasis in response to high and low glycemic index meals in healthy men. *Am J Clin Nutr* 87(5):1254–1261
- Moisey LL, Robinson LE et al (2009) Consumption of caffeinated coffee and a high carbohydrate meal affects postprandial metabolism of a subsequent oral glucose tolerance test in young, healthy males. *Br J Nutr* 103(6):833–841



- Montella M, Polesel J et al (2007) Coffee and tea consumption and risk of hepatocellular carcinoma in Italy. *Int J Cancer* 120(7):1555–1559
- Mukamal KJ, Hallqvist J et al (2009) Coffee consumption and mortality after acute myocardial infarction: the Stockholm Heart Epidemiology Program. *Am Heart J* 157:495–501
- Naganuma T, Kuriyama S et al (2007) Coffee consumption and risk of colorectal cancer: a prospective cohort study in Japan. *Int J Cancer* 120:1542–1547
- Nawrot P, Jordan S et al (2003) Effects of caffeine on human health. *Food Addit Contam* 20(1):1–30
- Nkondjock A, Ghadirian P et al (2006) Coffee consumption and breast cancer risk among BRCA1 and BRCA2 mutation carriers. *Int J Cancer* 118(1):103–107
- Norager CB, Jensen MB et al (2006) Metabolic effects of caffeine ingestion and physical work in 75-year old citizens. A randomized, double-blind, placebo-controlled, cross-over study. *Clin Endocrinol* 65:223–228
- Nygaard O, Refsum H et al (1997) Coffee consumption and plasma total homocysteine: the Hordaland Homocysteine Study. *Am J Clin Nutr* 65:136–143
- Olmos V, Bardoni N et al (2009) Caffeine levels in beverages from Argentina's market: application to caffeine dietary intake assessment. *Food Addit Contam A Chem Anal Control Expos Risk Assess* 26(3):275–281
- Palatini P, Ceolotto G et al (2009) CYP1A2 genotype modifies the association between coffee intake and the risk of hypertension. *J Hypertens* 27(8):1594–1601
- Pelluchi C, La Vecchia C (2009) Alcohol, coffee, and bladder cancer risk: a review of epidemiological studies. *Eur J Cancer Prev* 18(1):62–68
- Pencek RR, Battram D et al (2004) Portal vein caffeine infusion enhances net hepatic glucose uptake during a glucose load in conscious dogs. *J Nutr* 134(11):3042–3046
- Pereira MA, Parker ED et al (2006) Coffee consumption and risk of type 2 diabetes mellitus: an 11-year prospective study of 28 812 postmenopausal women. *Arch Intern Med* 166(12):1311–1316
- Petrie HJ, Chown SE et al (2004) Caffeine ingestion increases the insulin response to an oral-glucose-tolerance test in obese men before and after weight loss. *Am J Clin Nutr* 80(1):22–28
- Pizzoli A, Tikhonoff V et al (1998) Effects of caffeine on glucose tolerance: a placebo-controlled study. *Eur J Clin Nutr* 52(11):846–849, Abstract
- Quinlan PT, Lane J et al (2000) The acute physiological and mood effects of tea and coffee: the role of caffeine level. *Pharmacol Biochem Behav* 66(1):19–28
- Ranheim T, Halvorsen B (2005) Coffee consumption and human health – beneficial or detrimental? Mechanisms for effects of coffee consumption on different risk factors for cardiovascular disease and type 2 diabetes mellitus. *Mol Nutr Food Res* 49:274–284
- Rapuri PB, Gallagher C et al (2001) Caffeine intake increases the rate of bone loss in elderly women and interacts with vitamin D receptor genotypes. *Am J Clin Nutr* 74:694–700
- Reissig CJ, Strain ED et al (2009) Caffeinated energy drinks – a growing problem. *Drug Alcohol Depend* 99(1–3):1–10
- Reppert SM, Weaver DR (1991) Molecular cloning and characterization of a rat A1-adenosine receptor that is widely expressed in brain and spinal cord. *Mol Endocrinol* 5(8):1037–1048
- Ricksen NP, Rongen GA et al (2009) Acute and long-term cardiovascular effects of coffee: implications for coronary heart disease. *Pharmacol Ther* 121(2):185–191
- Ricksen NP, Smits P et al (2010) The cardiovascular effects of methylxanthines. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Robinson LE, Spafford et al (2009) Acute caffeine-induced impairment in insulin sensitivity in women with gestational diabetes mellitus. *J Obstet Gynecol Can* 31:304–312
- Robertson D, Wade D et al (1981) Tolerance to the humoral and hemodynamic effects of caffeine in man. *J Clin Invest* 67(4):1111–1117
- Robinson LE, Savani S et al (2004) Caffeine ingestion before an oral glucose tolerance test impairs blood glucose management in men with type 2 diabetes. *J Nutr* 134(10):2528–2533
- Rosner SA, Akesson A et al (2006) Coffee consumption and risk of myocardial infarction among older Swedish women. *Am J Epidemiol* 165:288–293

- Ruhl CE, Everhart JE (2005a) Coffee and caffeine consumption reduce the risk of elevated serum alanine aminotransferase activity in the United States. *Gastroenterology* 128(1):24–32
- Ruhl CE, Everhart JE (2005b) Coffee and tea consumption are associated with a lower incidence of chronic liver disease in the United States. *Gastroenterology* 129(6):1928–1936
- Rush JW, Spriet LL (2001) Skeletal muscle glycogen phosphorylase a kinetics: effects of adenine nucleotides and caffeine. *J Appl Physiol* 91(5):2071–2078
- Salazar-Martinez E, Willet WC et al (2004) Coffee consumption and risk for type 2 diabetes mellitus. *Ann Intern Med* 140(1):1–8
- Sandhu H, Wiesenthal SR et al (1999) Glucagon-like peptide 1 increases insulin sensitivity in depancreatized dogs. *Diabetes* 48(5):1045–1053
- Sasaki H, Takaoka I et al (1987) Effects of sucrose or caffeine ingestion on running performance and biochemical responses to endurance running. *Int J Sports Med* 8(3):203–207
- Sawynok J (2010) Methylxanthines and pain. In: Fredholm BB (ed) *Methylxanthines*. Springer, Heidelberg
- Schulte G, Fredholm BB (2003) Signalling from adenosine receptors to mitogen-activated protein kinases. *Cell Signal* 15(9):813–827
- Shearer J, Farah A et al (2003) Quinides of roasted coffee enhance insulin action in conscious rats. *J Nutr* 133(11):3529–3532
- Shearer J, Sellars EA et al (2007) Effects of chronic coffee consumption on glucose kinetics in the conscious rat. *Can J Physiol Pharmacol* 85:823–830
- Shimazu T, Tsuborno Y et al (2005) Coffee consumption and the risk of primary liver cancer: pooled analysis of two prospective studies in Japan. *Int J Cancer* 116(1):150–154
- Silletta MG, Marfisi RM et al (2007) Coffee consumption and risk of cardiovascular events after acute myocardial infarction: results from the GISS (Gruppo italiano per lo studio della sopravvivenza nell'infarto miocardico) – prevenzione trial. *Circulation* 116:2944–2951
- Smith B, Wingard DL et al (2006) Does coffee consumption reduce the risk of type 2 diabetes in individuals with impaired glucose? *Diabetes Care* 29:2385–2390
- Smits P, Hoffmann H et al (1983) Hemodynamic and humoral effects of coffee after beta 1-selective and nonselective beta-blockade. *Clin Pharmacol Ther* 34(2):153–158
- Smits P, Lenders JW et al (1990) Caffeine and theophylline attenuate adenosine-induced vasodilation in humans. *Clin Pharmacol Ther* 48(4):410–418
- Smits P, Stratman C et al (1991) Dose-dependent inhibition of the hemodynamic response to dipyridamole by caffeine. *Clin Pharmacol Ther* 50(5 Pt 1):529–537
- Stamler J, Vaccaro O et al (1993) Diabetes, other risk factors, and 12-yr cardiovascular mortality for men screened in the Multiple Risk Factor Intervention Trial. *Diabetes Care* 16(2):434–444
- Tan EK, Tan C et al (2003) Dose-dependent protective effects of coffee, tea, and smoking in Parkinson's disease: a study in ethnic Chinese. *J Neurol Sci* 216(1):163–167
- Tanaka K, Hara M et al (2007) Inverse association between coffee drinking and the risk of hepatocellular carcinoma: a case-control study in Japan. *Cancer Sci* 98(2):214–218
- Thom E (2007) The effect of chlorogenic acid enriched coffee on glucose absorption in healthy volunteers and its effect on body mass when used long-term in overweight and obese people. *J Int Med Res* 35:900–908
- Thong FSL, Graham TE (2002) Caffeine-induced impairment of glucose tolerance is abolished by beta-adrenergic receptor blockade in humans. *J Appl Physiol* 92(6):2347–2352
- Thong FSL, Derave W et al (2002) Caffeine-induced impairment of insulin action but not insulin signaling in human skeletal muscle is reduced by exercise. *Diabetes* 51(3):583–590
- Thong FS, Lally JS et al (2007) Activation of the A1 adenosine receptor increases insulin-stimulated glucose transport in isolated rat soleus muscle. *Appl Physiol Nutr Metab* 32(4):701–710
- Töpfer M, Burbeil CE et al (2008) Modulation of insulin release by adenosine A<sub>1</sub> receptor agonists and antagonists in INS-1 cells: the possible contribution of <sup>86</sup>Rb<sup>+</sup> efflux and <sup>45</sup>Ca<sup>+</sup> uptake. *Cell Biochem Funct* 26:833–843

- Tse SYH (1991) Coffee contains cholinomimetic compound distinct from caffeine. I: purification and chromatographic analysis. *J Pharm Sci* 80:665–669
- Tse SYH (1992) Cholinomimetic compound distinct from caffeine contained in coffee. II: Muscarinic actions. *J Pharm Sci* 81:449–452
- Tunicliffe JM, Shearer J (2008) Coffee, glucose homeostasis, and insulin resistance: physiological mechanisms and mediators. *Appl Physiol Nutr Metab* 33:1290–1300
- Tunicliffe JM, Erdman KA et al (2008) Consumption of dietary caffeine and coffee in physically active populations: physiological interactions. *Appl Physiol Nutr Metab* 33:1301–1310
- Tuomilehto J, Hu G et al (2004) Coffee consumption and risk of type 2 diabetes mellitus among middle-aged Finnish men and women. *JAMA* 291(10):1213–1219
- van Dam RM (2008) Coffee consumption and risk of type 2 diabetes, cardiovascular disease, and cancer. *Appl Physiol Nutr Metab* 33:1269–1283
- van Dam RM, Hu FB (2005) Coffee consumption and risk of type 2 diabetes: a systematic review. *JAMA* 294(1):97–104
- van Dam RM, Willett WC et al (2006) Coffee, caffeine, and risk of type 2 diabetes. *Diabetes Care* 29(2):398–403
- van Dijk AE, Olthof MR et al (2009) Acute effects of decaffeinated coffee and the major coffee components chlorogenic acid and trigonelline on glucose tolerance. *Diabetes Care* 32:1023–1025
- Van Dusseldorp M, Katan MB et al (1991) Cholesterol-raising factor from boiled coffee does not pass a paper filter. *Arterioscler Thromb Vasc Biol* 11:586–593
- Van Nieuwenhoven MA, Brummer RM et al (2000) Gastrointestinal function during exercise: comparison of water, sports drink, and sports drink with caffeine. *J Appl Physiol* 89(3):1079–1085
- Van Soeren MH, Nohr T et al (1996) Acute effects of caffeine ingestion at rest in humans with impaired epinephrine responses. *J Appl Physiol* 80:999–1005
- Van Soeren MH, Sathasivam P et al (1993) Caffeine metabolism and epinephrine responses during exercise in users and nonusers. *J Appl Physiol* 75(2):805–812
- Vergauwen L, Hespel P et al (1994) Adenosine receptors mediate synergistic stimulation of glucose uptake and transport by insulin and by contractions in rat skeletal muscle. *J Clin Invest* 93:974–981
- Vergauwen L, Richter EA et al (1997) Adenosine exerts a glycogen-sparing action in contracting rat skeletal muscle. *Am J Physiol* 272(5Pt1):E762–E768
- Villanueva CM, Silverman DT et al (2009) Coffee consumption, genetic susceptibility and bladder cancer risk. *Cancer Causes Control* 20(1):121–127
- Wei Y, Mojsov S (1996) Distribution of GLP-1 and PACAP receptors in human tissues. *Acta Physiol Scand* 157(3):355–357
- World Health Organization (2010) Diabetes. <http://www.who.int/mediacentre/factsheets/fs312/en/print.html>. Accessed 10 Feb 2010
- Wu T, Willett WC et al (2005) Caffeinated coffee, decaffeinated coffee, and caffeine in relation to plasma C-peptide levels, a marker of insulin secretion, in U.S. women. *Diabetes Care* 28:1390–1396
- Yamada M, Sasaki S et al (2009) Estimation of caffeine intake in Japanese adults using 16 d weighed diet records based on a food composition database newly developed for Japanese populations. *Public Health Nutr* 16:1–10
- Yeo SE, Jentjens RL et al (2005) Caffeine increases exogenous carbohydrate oxidation during exercise. *J Appl Physiol* 99(3):844–850
- Zegers MP, Tan FE et al (2001) Are coffee and tea consumption associated with urinary tract cancer risk? *Int J Epidemiol* 30(2):353–362
- Zhang WL, Lopez-Garcia E et al (2009) Coffee consumption and risk of cardiovascular events and all-cause mortality among women with type 2 diabetes. *Diabetologia* 52:810–817

# Index

## A

### Absorption

- caffeine, 35–36, 218
- paraxanthine, 73
- theobromine, 66–67, 205, 211
- theophylline, 55, 58

Abuse potential, 488–490, 492

Accessory proteins, 272, 293

ACE inhibitor, 400

Acetaminophen, 312, 316–318,  
323–325, 477

Acrosome reaction, 355

Additives, 96, 114, 491, 493, 497, 499

### Adenosine

and caffeine, sleep, 40, 213, 338–344

deficiency, 255–257

receptor, 470, 473, 474

#### sleep

basal forebrain adenosine  
concentration, 337

circadian variation, 336–338

genetic variation, ADA, 338

vigilance-state-related variation, 336

Adenosine deaminase (ADA), 107, 338, 461

Adenosine receptors, 102, 415–423, 444–449.

*See also* Neurodegeneration

accessory proteins, 272, 293

antagonism, 102, 163, 206, 255, 258, 261,

269, 270, 311, 315, 323, 375, 376,

382, 413, 415–417, 419, 439,

444–449, 458, 470, 473, 475, 492,

495, 496, 510, 516, 523, 524, 526,

527, 540

A<sub>2A</sub>R, 73, 135, 143, 172–177, 206, 207, 257,

270, 311, 315, 316, 318, 320, 325,

336, 338–341, 343, 375, 376, 382,

384, 394, 397, 415–418, 420, 459,

462, 463, 469, 470, 473, 474, 476,

477, 487, 491, 492, 495, 496, 499

G-protein-coupled receptors, 271–272, 293,  
382, 447, 529

isoforms, 102, 523

A<sub>1</sub>R, 135, 136, 142, 158–172, 206, 215,  
217, 236, 255, 270, 311, 314, 315,  
318, 320–322, 325, 339–340, 356,  
363, 364, 375, 376, 380, 382–384,  
392, 394–398, 401–404, 415–418,  
420, 421, 424, 473, 487

A<sub>2</sub>R, 73, 135, 143, 172–177, 206, 207,  
257, 270, 311, 315, 316, 318,  
320, 325, 336, 338–341, 343, 375,  
376, 382, 384, 394, 397, 399,  
15–418, 420, 459, 462, 463, 469,  
470, 473, 474, 476, 477, 487,  
491, 492, 495, 496, 499, 523

A<sub>2B</sub>R, 315, 316, 320, 323, 415–417, 446,  
447, 458–460, 462

A<sub>3</sub>R, 216, 375–377, 415–418, 445, 447,  
458–460, 473

ADHD, 383

### Adipose tissue

FFA mobilization, 525, 527, 537

lipolysis, 527

Adjuvant, 235, 312, 313, 316, 318, 323–325,  
464, 477

Adolescence, 376, 384

Adrenal medulla, 525–526

### Age

caffeine, 9, 36, 41–42, 46, 48, 51, 253,  
286, 363, 365–366, 383, 384, 393,  
533–534

theophylline, 59, 60, 62–63, 65, 253, 379

Airway smooth muscle (ASM), 105, 439, 442,  
444–447

- Alcohol, caffeine, 46–47, 103, 362, 363, 471, 472, 532
- Alzheimer's disease (AD)  
 A<sub>2A</sub> AR antagonists, 174  
 caffeine-induced beneficial effects, 539–540  
 experimental studies, 282–284  
 human studies, 282–283
- Aminophylline, 60, 145, 252–254, 260, 277, 314, 320, 321, 344, 373, 375, 395, 397, 401, 402, 420, 421, 442, 446, 447, 449, 532
- Amphetamine, 220, 221, 287, 288, 421, 486, 488, 489, 491–494, 501
- Analgesic, 187, 312–314, 316, 318, 321–325, 342, 384
- Anandamide, 202, 222–223
- Angiotensin II, 423, 424
- Antagonists, 143, 144, 151–187, 206, 207, 217, 219, 236, 237, 240, 251, 254, 257, 258, 261, 271, 272, 274, 277, 278, 281–288, 290, 292, 311–313, 315, 316, 318, 320, 322, 340, 356, 380, 392, 395–398, 401–403, 415, 416, 420, 445, 446, 458–460, 464, 474–477, 492, 498–499, 513, 522, 532
- Antiasthmatic, 156, 177, 187, 484
- Antibiotics, fluoroquinolone, 254
- Anticonvulsant, 251, 253–257, 259–261, 313, 322
- Antidepressants, 45, 222, 313, 321–322, 500
- Antioxidants, 202, 203, 260, 427, 511, 521, 530, 535, 537, 538, 540
- Antipsychotics, 45, 271, 500
- Apaxifylline, 163, 164
- Apnea/apnoea, 187, 374–377, 379–385  
 of prematurity, 374, 375, 377, 380–381, 383–385, 450
- Arrhythmias, 167, 253, 417, 421, 449
- Arterial stiffness, 420, 421, 429
- Aspirin, 312, 316–318, 323, 324
- Astemizole, 254
- Asthma, 8, 9, 41, 59, 104, 105, 162, 167, 177, 178, 215, 216, 252, 254, 375, 392, 439–451, 461, 462, 484, 487
- Astrocytes, 237, 240–242, 244–247, 256, 273, 376
- Atherosclerosis, 429
- ATL–802, 158, 178
- ATL–852, 158, 178
- Atropine, 253
- B**
- Barbiturates, 500
- Benzodiazepines, 253, 259, 269, 500
- BG–9719, 163, 164
- BG–9928, 163, 164
- Bioavailability  
 caffeine, 35, 36, 42, 381  
 8-cycloalkylxanthines, 163  
 1,8-disubstituted xanthenes, 179  
 theophylline, 58, 381
- Biotin conjugates, 183, 186
- Birth, 42, 46, 63, 67, 362–366, 374, 376, 378–380, 382, 384, 404
- Blood–brain barrier (BBB), 37, 67, 73, 224, 275–276, 284, 288, 291
- Blood pressure, 219, 220, 268, 332, 376, 396, 399, 414, 417–419, 423–426, 429, 486, 520, 521, 531, 532, 535
- Breast feeding, 379–380, 382, 383
- Bronchitis, 215, 252
- C**
- Cacao (*Theobroma cacao*), 7, 12–15, 18, 25, 204–206, 211
- Café Procope, 4
- Caffeinated energy drinks, 143, 252, 253, 280, 286, 378, 393, 422, 424, 427, 472, 487, 495, 496, 511, 520–532, 535, 538–540
- Caffeine (1,3,7-trimethylxanthine), 152–158, 169, 174, 180, 187, 252–254, 258–261, 470–477. *See also* Neurodegeneration  
 absorption, 35–36, 218  
 animals, 35, 36, 38, 39, 45, 51–53, 56–57, 352, 361, 362, 366  
 biosynthesis, 15, 17–21, 23, 25, 27  
 catabolism, 12, 23–25  
 decaffeinated coffee, 12, 26–27, 40, 342, 378, 421, 422, 424, 427, 472, 518, 520–522, 525, 530, 535, 538, 539  
 degradation, 19, 25  
 discovery, 96  
 distribution, 36–38, 42–45  
 ecological role, 26  
 European multicentre study, 362  
 excretion, 38–39  
 habituation, 520, 535  
 human intake, 268–269  
 humans, 361–363, 366  
 LD50 caffeine, 353, 362  
 long-lasting behavioural effects, 363, 364

- maternal/fetal genotype, 37, 42, 52, 361, 378, 380, 382–384
- metabolism, 41–51
- metabolites and metabolic pathway, 51–57
- molecular targets
  - adenosine receptors, 270–272
  - non-adenosine receptors, 269–270
  - psychostimulant effect, 219, 268, 484, 489,
- neurodegenerative disorder
  - Alzheimer's disease (AD), 281–284, 536, 540
  - Huntington's disease, 289–291
  - multiple sclerosis (MS), 291–292
  - Parkinson's disease (PD), 143, 144, 284–288, 539, 540
  - stroke, 275–279, 281, 293, 532
  - traumatic brain injury, 38, 279–281
- Parkinson's disease, 143, 144, 284–288, 539, 540
- pharmacokinetics, 3–41
- psychopharmacological activity in
  - chocolate, 202–215, 217–219, 225, 226
- risk of developing, 143
- sleep
  - adenosine A<sub>2A</sub> receptors, 340–341
  - adenosine A<sub>1</sub> receptors, 339–340
  - animals, 341
  - CNS and sleep regulation, 338
  - humans, 337, 341–343
  - human sensitivity, 343–344
  - mechanisms, 338–341
  - methylxantines effects, 344–345
  - performance, 343
  - withdrawal syndromes, 344
- sources of variation, pharmacokinetics and metabolism
  - age, 41–42
  - diet and alcohol, 46–47
  - disease, 45
  - drugs, 44–45
  - gender and hormones, 42–43
  - obesity, 43–44
  - physical exercise, 43
  - smoking, 46
- treatment, 365
- Calcineurin inhibitors, 392, 402
- Calcium, 99, 102, 107, 137, 139, 141–144, 206, 217, 237, 238, 260, 268–270, 272–274, 355, 356, 362, 382, 393, 394, 399, 401, 415, 417, 421, 422, 444–447, 460, 523, 539
- Ca<sup>2+</sup> mobilization, 139, 260, 355, 415, 460, 527
- cAMP. *See* Cyclic AMP
- Cancer, 469–477
  - bladder, 538–540
  - breast cancer, 536, 538, 540
  - colorectal cancer, 538, 540
  - liver cancer, 538–540
  - lung, 538–540
- Cannabis, 222, 223, 495, 501
- Capacitation, 355–357, 365
- Carbamazepine, 253, 322–324
- Carbohydrate homeostasis, 512–522, 527, 529–531
- Cardiac function
  - bradycardia, 253, 380
  - cardiac, 257
  - cardiac arrest, 209, 253
  - diabetic cardiac autonomic neuropathy, 253
- Cardiovascular, 378, 383–384
- Cardiovascular disease (CVD)
  - cardiovascular mortality, 421, 428, 532
  - coronary heart disease, 414, 421, 426–429, 532, 535
  - CVD risk, 414, 425–427, 510, 511, 532, 535, 536
  - stroke, 276, 532
- Cardiovascular effects, 215, 350, 362, 384, 413–429, 511
- CD39, 458, 473
- CD73, 291, 458, 473
- CEBPβ, 459
- Cerebral blood flow, 237, 277, 281, 487
- Chemoreceptors, 382, 417, 418
- Child, 59, 60, 63, 64, 76, 174, 258, 324, 362, 375, 381, 384, 402, 446
- China, 1, 2, 5, 6, 50, 96, 511
- Chocolate, 7, 15, 36, 67–69, 71, 201–203, 205, 207–217, 219–226, 345, 365, 366, 380, 424, 484, 488, 489, 511
- Cholesterol, 276, 284, 354, 356, 414, 426, 427, 429, 521, 536
- Cholinergic mechanisms, 316, 334
- Chronic obstructive pulmonary disease (COPD), 104, 105, 215, 458, 461, 462
- Cialis™, 94
- Cirrhosis, 45, 65, 470–472, 475, 477
- Cisplatin, 392, 402–403
- Cizolirtine, 322–324
- Clinical studies, 44, 104, 163, 235, 246, 247, 252, 253, 257, 280, 285–287,

- 312–325, 362, 374, 375, 404, 443, 446, 449, 450, 461, 470, 473–475
- CNS stimulant, 187, 202, 203, 219, 253, 254, 512, 525, 526
- Cocaine wanting, 494
- Cocoa (*Theobroma cocoa*), 201–226
- Coffee, 96, 102, 103, 105, 393, 398, 414, 415, 418, 421, 422, 424–429, 471–473, 475
- acute consumption, 518, 532
- bioactive compounds
- chlorogenic acid (CGA), 427, 475, 521, 522, 528, 530, 535, 537, 538
  - quinides, 521, 522, 529, 535, 537
  - trigonelline, 475, 521
- boiled/unfiltered, 426, 511, 522, 536, 537
- caffeinated, 143, 252, 253, 280, 286, 378, 393, 422, 424, 427, 472, 487, 495, 496, 511, 520–532, 535, 538–540
- Coffea arabica*, 13, 17, 20, 21, 25, 27, 28, 72
- Coffea canephora*, 13, 22, 27, 28
- Coffea eugenioides*, 13, 25
- decaffeinated, 12, 26–27, 40, 342, 378, 421, 422, 424, 427, 472, 518, 520–522, 525, 530, 535, 538, 539
- filtered, 426, 427, 511, 522, 536
- low caffeine coffee, 13, 25, 27, 28, 39
- method of brewing, 426, 521, 536, 537
- regular consumption, 42, 47, 269, 282, 288, 293, 342, 357, 414, 415, 418, 421, 424, 425, 518, 520, 521, 530, 532, 535
- Cognitive dysfunction, 174, 285
- Conditioned place preference (CPP), 488, 493, 499
- Constantinople, 3
- Coronary heart disease, 414, 421, 426–429, 532, 535
- Cortical excitability, 334
- C-peptide, 529, 531
- CPFPX, 185
- CPX, 163, 164
- CREB, 274, 290, 459
- CSC, 167, 175, 186, 274, 278, 287
- Cumulus cell
- DNA methylation pattern, 365
  - nuclear remodelling, 365
  - somatic cell nuclear transfer embryos, 365
- CVD. *See* Cardiovascular disease
- CVT124, 163
- CVT–6883, 157, 178
- Cyclic AMP (cAMP), 393, 472, 473, 475, 523, 525, 527
- Cyclic AMP dependent protein kinase, 96, 109
- Cyclic AMP syn and anti conformations, 109–111
- Cyclic GMP dependent protein kinase, 108, 109, 112
- Cyclic GMP syn and anti conformations, 109–111
- Cyclic nucleotide gated channels, 107–109, 111
- Cycloaddition, 176
- Cyclosporine, 402
- Cytokines, 242–244, 247, 275, 280, 281, 284, 335, 443, 448, 459, 463, 472–476
- D**
- Dantrolene, 141–144
- D2 dopamine receptors, 186, 271, 272, 285, 288
- 9-Deazaxanthines, 169, 170, 179
- Diabetes, 45, 65, 178, 414, 427
- Diabetes, Type II (T2D)
- gestational diabetes, 521
  - global health problem, 511
  - lifestyle intervention, 512
  - relative risks, 427, 520, 531, 534, 535
- Diazepam, 253, 254, 259
- Diet
- caffeine, 46–47, 214, 276, 312, 414, 424–429
  - theobromine, 71, 206, 207, 354
- Dihydroimidazo[2,1-*i*]purinones, 170
- Dimethylxanthines
- paraxanthine, 13, 17, 18, 21, 34, 35, 37, 38, 40–42, 44–53, 55–57, 66, 68, 72–76, 96, 102, 136, 140, 143–145, 152, 206, 207, 280, 393, 416, 484, 522
  - theophylline, 9, 13, 35, 96, 136, 152, 206, 237, 252, 277, 314, 340, 353, 375, 392, 415, 440, 458, 484, 516 (see also Theophylline)
- 1,3-Dipropyl–8-cyclopentylxanthine (DPCPX), 162, 163, 169–171, 184, 185, 258, 278, 287, 290, 322, 395, 396, 398, 401, 403
- Disease
- caffeine, 45, 399, 400, 402, 427–429, 471–477, 531–540
  - theophylline, 65, 401
- Distribution
- caffeine, 36–38
  - paraxanthine, 73
  - theobromine, 67–68

- theophylline, 58–59  
 Diuretics, 9, 102, 167, 203, 217, 218, 391, 393, 395, 402, 420, 423, 441  
 DMPX, 153, 155, 157, 174–177, 186  
 Dopamine receptors, 271, 288, 384, 487–489, 491, 492  
 Doxofylline, 155, 395  
 DPCPX. *See* 1, 3-Dipropyl-8-cyclopentylxanthine; 1,3-Dipropyl-8-cyclopentylxanthine  
 DPSPX, 158  
 Drug-discrimination/discriminative, 212, 491, 493, 497, 498,  
 Drugs  
   caffeine, 44–45  
   theobromine, 71  
   theophylline, 64–65  
 Dyskinesia, 175, 284, 286, 288
- E**
- Ecstasy, 493  
 Electroencephalogram (EEG)  
   homeostatic regulation, 333  
   human vigilance states, 332  
   low EEG frequencies, 333  
 Embryonic development  
   blastocyst rate, 364  
   dibutyl AMP (dbcAMP), 364  
   follicle-stimulating hormone (FSH), 354, 359, 362–364  
   3-isobutyl-1-methylxanthine (IBMX), 360, 361, 364  
   meiotic potential, 364  
 Encapsulated fibroblasts, 257  
 Endothelial dysfunction, 535  
 Energy drinks, 102, 103, 253, 424, 490, 495, 496  
 Enprofylline, 100, 155, 177, 178, 395, 402, 416, 446, 463  
 EPAC, 108, 109, 111, 459  
 Epilepsy, 252–257, 259, 260  
 Epinephrine, 95, 96, 398, 422, 423, 425, 429, 446, 525–528, 531, 532, 537  
 Equilibrative nucleoside transporters, 256, 404, 420  
 Erectile dysfunction, 106  
 Ethanol intoxication, 495, 496, 501  
 Ethiopia (ethiopian), 2–4, 27, 96  
 Euglycemic-hyperinsulinemic clamp, 513, 516–518, 522, 524, 526, 527, 529  
 Excretion  
   caffeine, 38–39  
   paraxanthine, 74  
   theobromine, 68–69  
   theophylline, 59  
 Experimental autoimmune encephalomyelitis (EAE), 291–292
- F**
- Felbamate, 254  
 Fertility, 352, 353, 355–357, 361–364, 378  
 Fetus, 37, 42, 59, 62, 65, 67, 68, 73, 354, 374, 375, 377–378, 384  
 Flow-mediated dilation, 421  
 Fluorescent conjugate, 184  
 Follicle, 359, 360, 362–364  
 Free radical, 260, 521  
 Furosemide, 398
- G**
- Gabapentin, 254  
 GAF, 121  
 Gastric emptying, 40, 58, 205, 530  
 Gastrointestinal tract, glucose absorption, 530, 531  
 Gender  
   caffeine, 42–43  
   theobromine, 70–71  
   theophylline, 63–64  
 Gestational age, 63, 366, 379, 380  
 GIFT  
   IVF/GIFT study, 366  
 Glia, 240, 243, 289, 322  
 Glomerular filtration rate (GFR), 392, 394, 396, 397, 400–404  
 Glomerulosclerosis, 400  
 Glucose tolerance, 513, 518, 520, 526  
 Glutamate release, presynaptic sites, 272–273  
 G-protein-coupled receptors, 271–272, 447, 521  
 Guinea pig spinal cord homogenates (GPSCH), 292  
 Gurana (*Paulinia cupana*), 14, 15, 203
- H**
- Headache, 187, 214, 219–221, 268, 324, 325, 342, 344, 487, 540  
 Heart  
   conduction, 421  
   failure, 163, 392, 399, 423, 450, 532  
   inotropic action, 421, 422, 451  
   ischemia-reperfusion injury, 418, 422  
   rate, 223, 332, 342, 376, 414, 416–418, 424, 486  
   rhythm, 421  
 Hemodynamic response, 417, 422, 532



- Hepatitis, 471–477
- Histone deacetylase, 102, 442, 448, 460–461
- Histone deacetylase activity (HDAC), 102, 442, 444, 448–449
- Homocysteine, 16, 20, 282, 414, 426–427, 429, 536, 537
- Hormones
- caffeine, 42–43
  - theobromine, 70–71
  - theophylline, 63–64
- Huntington's disease
- experimental studies, 289–291
  - human studies, 289
- 5-Hydroxytryptamine, serotonergic, 221, 320
- Hyperalgesia, 240, 242–244, 313, 314, 317, 318, 321, 322
- Hypertension, 106, 167, 218, 221, 284, 399, 425, 426, 532, 535
- Hypoxia, 65, 209, 210, 278, 279, 374, 380, 381, 384, 473, 474, 476
- I**
- IL–10, 459, 461, 475, 476
- Incretin
- glucagon-like peptide–1 (GLP–1), 529
  - glucose-dependent insulinotropic peptide (GIP), 529, 530
- Infant, 36, 38, 40–42, 46, 52, 58, 59, 65, 66, 68, 253, 366, 374–376, 379–382, 384, 394, 404, 450
- Inflammation, 105, 215, 274, 275, 289, 292, 400, 403, 417, 442, 443, 447, 451, 462, 464, 469–477, 535
- Insomnia, 219, 253, 342, 540
- Insulin resistance, 513, 516, 520–527, 530, 531, 540
- Insulin secretion, 529, 531
- Intrinsic, 48, 62, 312, 313, 316, 321, 323–325, 418
- Intrinsic antinociception, 313–316, 320, 325
- Intrinsic antinociceptive, 313, 314, 318, 323
- In vitro* fertilization (IVF), 364–366
- In vitro* maturation, 364
- Ischemia, 236–239, 246, 258, 275, 277–279, 417, 418, 422
- Ischemia-reperfusion, 403–404, 418, 422, 474
- Ischemic stroke, 268, 277
- 3-Isobutyl-1-methylxanthine (IBMX), 98–100, 111, 114–122, 141, 156, 341, 344, 360, 361, 395
- Isomerization, 176
- Isoproterenol, 398
- IVF. *See* In vitro fertilization
- K**
- Ketamine, 498, 499
- (E)-KF17837, 177, 186
- Kinase, 96, 107–109, 111–112, 242, 255, 256, 261, 272–276, 278, 313, 356, 359–361, 364, 365, 396, 459, 460, 473, 523
- KW6002, 175, 185, 186, 285, 286, 288
- L**
- L–97–1, 162
- Lactation, 374, 380, 383, 385
- L-dopa-induced motor complications, 288
- Leucoptera coffeella
- coffee leaf miner, 364
  - egg laying, 350, 364
- Levetiracetam, 254, 260
- Levitra<sup>TM</sup>, 106
- Liver, 470–475, 477
- hepatic glucose output, 510, 516, 528, 529, 531, 540
- Locomotor
- depression, 315
  - stimulation, 315, 325
- M**
- Magnesium, 98, 118, 224, 394, 521
- Magnesium, psychopharmacological activity
- in chocolate, 224
- Male gamete
- sperm capacitation, 356, 357, 365
  - spontaneous acrosome reaction, 365
- Mammalian oocytes maturation, 357–361, 364
- Manganese, 98
- MAO-B, 168, 175
- Mast cells, 442, 443, 445–448, 458, 460, 462, 463
- Maté (*Ilex paraguariensis*), 7–8, 14, 15, 25, 203, 484, 487, 511
- Maturation, 63, 351, 352, 355, 364, 375–377
- Mecca, 3
- Medullary blood flow, 397, 401
- Meiosis
- oocytes, 359, 360
  - spermatocytes, 351, 352, 357
- Meiotic competence
- adenylyl cyclase (AC), 359, 360, 364
  - G-protein linked receptor, 359
  - heat shock transcription factor 1, 359
  - oocyte plasma membrane, 359
- Metabolism
- caffeine, 39–51, 498, 536, 537

- paraxanthine, 75–76
- theobromine, 70, 205–206
- theophylline, 25, 61–65
- Metabolites and metabolic pathway
  - caffeine, 51–55
  - paraxanthine, 76
  - theobromine, 71–72
  - theophylline, 65–66
- Methotrexate, 464
- Methylphenidate, 498, 501
- Methyluric acid, theacrine (1,3,7,9-tetramethyluric acid), 13
- Microglia, 237, 240, 242, 246, 247, 278, 317, 318, 325
- Midaxifylline, 163
- Mirodenafil–SLX2101, 100, 106
- Mocha, 3, 5
- Monocytes, 375, 376, 443, 459, 461, 462, 473, 487
- Morphine, 12, 243–244, 312, 316, 318–321, 323, 325, 499
- MRE–2029-F20, 178
- MRS1754, 178
- MSX–2, 175–177
- MSX–3, 176
- MSX–4, 176
- Multiple sclerosis (MS)
  - animal studies, 291–292
  - human studies, 291
- Muscle cells, excitation coupling, 137
- Myocardium, 417, 422
- N**
- Na/HCO<sub>3</sub> cotransport, 396
- Na/H exchange, 396
- Natriuresis, 394–396, 398
- Naxifylline, 163
- Neonatal, 36, 46, 52, 62, 63, 66, 208, 237, 279, 382, 383, 461
- Neurodegeneration. *See also* Caffeine
  - acute degeneration, 268
  - chronic degeneration, 268
  - neuroprotection, caffeine and adenosine receptors
    - blood–brain barrier (BBB), 275–276
    - cellular survival signals, postsynaptic sites, 273–274
    - glutamate release, presynaptic sites, 272–273
  - neuroinflammation control, CNS, 274–275
  - neurotrophic factors, 275
- Neurons
  - release neurotransmitters, 137, 142, 143, 334
  - survival, 137, 143–145, 270, 278, 290
- Neuropathic pain, 240–243, 245, 321, 322
- Neuroprotective, 143, 144, 174, 236, 246, 257, 261, 273, 275, 276, 278–283, 285–287, 292, 293, 380, 381
- Neutrophils, 274, 376, 443, 444, 458–460, 462, 472
- Newborn, 38, 41, 42, 63, 66, 67, 187, 279, 374, 376, 379, 394, 487
- Nicotine, 12, 36, 142, 216, 489, 497–498, 501
- Nitric oxide (NO) synthase, 260, 275
- N*-methyltransferase
  - caffeine synthase (SAM:theobromine *N*-methyltransferase), 12, 16–18, 20–23, 28
  - evolutionary relationship, 21–23
  - 7-methylxanthosine synthase (SAM:xanthosine *N*-methyltransferase), 16, 17, 21, 22, 28
  - theobromine synthase (SAM:7-methylxanthine *N*-methyltransferase), 16–18, 20, 21, 23
- Non-steroidal anti-inflammatory drugs (NSAIDs), 313, 316–318, 323, 325
- Noradrenaline, noradrenergic, 316, 318, 325, 335
- Norepinephrine, 397, 398, 417, 421–423, 525, 528
- NSAIDs. *See* Non-steroidal anti-inflammatory drugs
- 5′-Nucleotidase, 19, 20, 255, 397, 416
- N<sup>ω</sup>-nitro-L-arginine methyl ester, 401
- O**
- Obesity
  - caffeine, 43–44, 512
  - theophylline, 64
- Opioid receptors, 182, 186, 243, 321
- Opioids, 182, 243, 246, 313, 316, 318, 320, 321
- Oral glucose tolerance test, 513
- Osteoporosis, 537, 539, 540
- Ovarian age
  - antral follicle count, 362, 363
  - follicle-stimulating hormone (FSH), 354, 359, 361–364
  - inhibin B, 362, 363
  - oestradiol, 362, 363
- Oxazolo[3,2-*a*]purinone, 172

- P**
- Pain, 178, 214, 235–247, 311–325
- Pancreatic  $\beta$  cells, 516, 529–531
- Paraxanthine, 152
  - absorption, 73
  - demethylated metabolites, 145
  - distribution, 73
  - excretion, 74
  - metabolism, 75–76
  - metabolites and metabolic pathway, 76
  - pharmacokinetics, 74–75
  - RyR channel, 140, 143–145
  - survival, 143, 145
- Parkinson's disease (PD), 172, 175, 268, 539, 540
  - experimental studies
    - A<sub>2A</sub>R antagonists, 286–287
    - L-dopa-induced motor complications, 288
    - MPTP-induced reduction, 287
    - neurotoxicity, 287–288
  - human studies, 268
    - disease modifying effect, 286
    - motor symptoms, 285–286
- PDE4, 100, 116–118, 121, 360, 400
- PDE5, 100, 101, 106–117, 120–122
- PDE6, 99–101, 106
- PDE11, 122
- Pentoxifylline, 100, 105, 236, 344, 400, 402, 414, 416
- Perinatal
  - intrauterine growth restriction, 378
  - spontaneous abortions, 378
- Perinatal caffeine, adenosine A<sub>1</sub> receptors, 356, 363, 364
- Peripheral resistance, 414, 418, 420
- Peroxisome-proliferator-activated receptor  $\gamma$  (PPAR $\gamma$ ), 444, 448–449
- Pertussis toxin, 396, 460
- PET. *See* Positron emission tomography
- Pharmacokinetics
  - caffeine, 37, 39–47
  - paraxanthine, 74–75
  - theobromine, 69–71, 205–206
  - theophylline, 59–60, 62–65
- Phencyclidine, 498, 499
- Phenobarbital, 253
- Phentolamine, 316
- Phenylethylamine, psychopharmacological
  - activity in chocolate, 202, 220–221
- Phenylethylamines, 220–221, 500
- 8-Phenylethynyl-DMPX, 177
- 8-Phenyltheophylline, 314, 320, 322, 397, 417, 420, 462
- Phenytoin, 253
- Phosphodiesterase (PDE), 93–123, 136, 156, 206, 217, 236, 237, 338, 394, 395, 397–400, 403, 444–445, 447–449, 470, 472, 473, 475
- Phosphodiesterase (PDE) inhibitors, 247, 259, 269, 312, 341, 349, 356, 357, 360, 361, 364, 366, 394, 413, 415, 416, 419, 439, 440, 445, 460, 470, 472, 473, 475
  - cilostamide, 360
  - hypoxanthine, 360
  - 3-isobutyl-1-methylxanthine (IBMX), 98, 114–122, 360, 361, 364
  - milrinone, 360
  - rolipram, 121, 270, 360, 399
- Photoisomerization, 176
- Physical exercise
  - caffeine, 43
  - theophylline, 64
- PKA, cyclin-dependant kinase (Cdk1), 359, 361
- Placenta, 38, 42, 59, 67, 363, 374, 377, 378
- Plasma concentration, 36, 37, 40, 42, 59, 66, 68, 205, 218, 222, 253, 258, 270, 343, 378–381, 461, 511, 526
- Polycystin, 399
- Polymorphism, 47, 49, 52, 61, 76, 281, 286, 289, 338, 340, 343, 344, 532, 535–537, 540
- Porcine oocytes, 360, 361, 364
- Positron emission tomography (PET), 176, 185–186
  - imaging, 185
  - radioligand, 186
- PPAR $\gamma$ . *See* Peroxisome-proliferator-activated receptor  $\gamma$
- Pregnancy, 34, 38, 41, 42, 49, 59, 64, 69–71, 211, 253, 354, 355, 362, 363, 373–385, 521
- Premature, 36, 52, 59, 63, 65, 66, 339, 355, 365, 374, 376, 379–381, 394
- Primidone, 253
- Proconvulsant, 251, 252, 254–259, 261
- Prodrugs, 163, 164, 167, 176, 179
- Propentofylline, 235–247, 283
- Propranolol, 398
- Proteinuria, 400
- PSB–10, 180, 181
- PSB–11, 180, 181
- PSB–50, 179

- PSB-53, 157, 179  
PSB-55, 157, 179  
PSB-63, 171, 172  
PSB-298, 157, 179  
PSB-601, 157, 160  
PSB-603, 157, 179  
PSB-1115, 157, 179, 316, 318  
Psychostimulant effect, 268, 269, 271, 486, 499  
Pulmonary hypertension, 94, 106  
Puromycin, 400  
Pyrimido[1,2,3-*cd*]purinediones, 170  
Pyrimido[2,1-*f*]purinediones, 171, 176
- R**
- Regulation, sleep  
  induction mechanisms, 334–338  
  waking mechanisms, 334  
Renal sympathetic nerve, 397  
Renin, 219, 398–400, 423  
Renin-angiotensin system, 398, 400, 414, 418, 419, 422–424  
Reperfusion, 418, 473  
Respiration, 219, 376, 381, 383–384, 450–451  
Respiratory drive, 380, 382  
Revatio<sup>TM</sup>, 94, 106  
Rheumatoid arthritis, 240, 464  
Rolipram, 121, 270, 360, 399  
Roscovitine, 360  
Ryanodine, 135–146, 260, 394, 399  
Ryanodine receptors (RyR), 446–447  
  endoplasmic reticulum (ER), 136, 138, 141–142  
  sarcoplasmic reticulum (SR), 136, 137, 139–141  
RyR channels  
  activation site, 138, 140  
  desensitization, 140  
  dopamine, 141–145  
  endogenous modulators, 145  
  neurotransmitter release, 137, 142, 143  
  pore region, 138  
  quantal Ca<sup>2+</sup> release, 140, 447  
  receptor subtypes, 137  
  sensitize, 141, 145  
  somatodendritic, 142  
RyR1, congenital myopathy, 138
- S**
- S-adenosyl-L-methionine (SAM)  
  SAM cycle, 19, 20  
  SAM route for caffeine biosynthesis, 19, 20  
Salsolinol, psychopharmacological activity in chocolate, 223–224  
SCH58261, 273–275, 278, 283, 286, 287, 290, 316, 318  
Second meal, 518  
Seizures  
  epilepsy, 252–255  
  febrile seizures, 253  
  sudden unexplained death in epilepsy (SUDEP), 258–259  
  Theophylline-associated seizures (TAS), 253, 258, 259  
Self-administration, 488, 489, 493, 494, 497, 499  
Self-medication, 496, 500  
Serotonin, psychopharmacological activity in chocolate, 219, 221–223  
Sildenafil, 98–100, 105–107, 111, 114, 117, 120–122  
Silk-based polymers, 257  
Skeletal muscle  
  carbohydrate storage, 525  
  glucose uptake, 523–526, 529  
  glycogen synthesis, 524, 525, 528  
Sleep  
  caffeine (*see* Caffeine, sleep)  
  chocolate, 213, 345  
  circadian component, 333  
  homeostatic regulation, 333, 341  
  in humans, 332  
  induction mechanisms, 334–336  
  in mammals, 332  
  non-REM sleep, 332, 337, 339, 342  
  regulation, 333–338  
  REM sleep, 332, 337, 341  
  tea, 345  
  two-process model, 333  
  waking mechanisms, 334  
Sleep apnea  
  central, 450  
  obstructive, 450  
Smoking, 41, 48–50, 143, 286, 362, 363, 378, 497, 498, 532, 539  
  caffeine, 46  
  theobromine, 71  
  theophylline, 65  
Smooth muscle relaxation, 111, 112, 216  
SNS. *See* Sympathetic nervous system  
Species dependence, 169, 174, 319  
Spermatids, 352, 354, 355  
Spermatogenesis  
  spermatidogenesis, 351, 352  
  spermatocytogenesis, 351  
  spermiogenesis, 351, 352

- Spermatozoa, 351, 352, 355–357  
 Sperm capacitation, 356–357  
 Spin labels, 184–185  
 8-SPT, 158  
 Stem cells, 257, 351  
 Stroke  
   experimental studies, 277–279  
   human studies, 276–277  
 Styrylxanthines, 175, 176, 186  
 8-Styrylxanthines, 167, 168, 175–177,  
   185, 186  
 Substance P, 186  
 Sympathetic nerve activity, 418, 422, 423  
 Sympathetic nervous system (SNS), 417, 418,  
   512, 525–527, 531  
 Synthetic methylxanthines, 141, 236
- T**
- Tadalafil, 100, 117, 121, 122  
 T cells, 137, 291, 376, 444, 448, 470–474,  
   476, 487
- T2D  
   gestational diabetes, 521  
   global health problem, 511  
   lifestyle intervention, 512  
   relative risks, 427, 520, 531, 534, 535
- Tea, 2, 3, 5–9, 19–23, 25, 26, 28, 96, 103, 203,  
   205, 212, 219, 226, 283, 345, 365,  
   366, 424, 472, 484, 511, 520, 538  
   *Camellia irrawadiensis*, 14, 18, 22  
   *Camellia sinensis*, 13, 14, 17, 21, 25  
   kekecha (cocoa tea, *Camellia*  
   *ptilophylla*), 14  
   kucha (*Camellia sinensis assamica* var.  
   *kucha*), 14
- Teratogenic/sperm injuring potential  
   caffeine, 352, 378  
   0.5 caffeine, 353  
   male gamete maturation, 355  
   male rats, 354  
   rats, 353  
   roosters, 352  
   theobromine, 353–355  
   theophylline, 353–355  
   ubiquitin-proteasome pathway (UPP), 355
- Testes, 350–352, 354, 355
- Tetrahydro- $\beta$ -carbolines,  
   psychopharmacological activity in  
   chocolate, 223–224
- Tetraplegia, 526, 527
- T helper (Th), 459
- Theobromine (3,7-dimethylxanthine), 12–18,  
   20, 21, 23–25, 27  
   absorption, 66–67  
   angiogenesis inhibition, 209  
   animal poisoning, 209–210, 225  
   cardiovascular effects, 214–215  
   characteristics, 203  
   dental effects, 217–218, 225  
   distribution, 67–68  
   doping in animal racing, 211, 225  
   excretion, 68–69  
   metabolism, 70  
   metabolism in man, 205–206  
   metabolites and metabolic pathway, 71–72  
   natural occurrence, 203  
   pest control, 210  
   pharmacokinetics, 69–70  
   pharmacokinetics in man, 205–206  
   psychopharmacological activity in  
   chocolate, 201–226  
   renal effects, 217  
   respiratory effects, 215–217  
   sources of variation, pharmacokinetics and  
   metabolism, 70–71  
   synthesis and catabolism in *Theobroma*  
   *cacao*, 203–206  
   teratogenesis, 209  
   testicular atrophy, 208  
   thymus gland atrophy, 208  
   toxicity in animals, 208–210  
   uptake in man, 205–207  
   use as medicine, 216, 218
- Theophylline (1,3-dimethylxanthine), 13, 15,  
   23–25, 152–158, 169, 174, 180,  
   187, 252–255, 258, 260, 261, 314,  
   319–321, 440–451  
   absorption, 55, 58  
   distribution, 58–59  
   excretion, 59  
   metabolism, 61–62  
   metabolites and metabolic pathway, 65–66  
   pharmacokinetics, 59–60  
   sources of variation, pharmacokinetics and  
   metabolism, 62–65
- Thermogenesis, 537
- Tolerance, 113, 209, 211, 243–244, 246, 312,  
   342, 343, 400, 418, 422, 424, 429,  
   441, 484, 486–487, 491, 496, 498,  
   513, 518, 520, 526
- Toponafylline, 163
- Traumatic brain injury (TBI), 38, 256, 268, 293  
   experimental studies, 280–281  
   human studies, 280
- Tryptophan, psychopharmacological activity in  
   chocolate, 219, 221–223

Tubuloglomerular feedback (TGF), 397, 398, 420  
Tumor, 237, 244, 274, 291, 418, 459, 470, 476–477  
Tyramine, psychopharmacological activity in chocolate, 219, 221, 223

**U**

Udenafil, 100, 106  
8-Unsubstituted xanthine, 153–156

**V**

Vardenafil, 98–100, 106, 107, 116, 117, 121, 122  
Vascular resistance, 531, 532  
Vascular tone, 396, 397, 414, 417, 419, 420  
Vasoconstriction, 276, 324, 378, 392, 397, 401–403, 420  
Vasodilation, 215, 274, 417–420  
Vasomotor action, 419  
Vasopressin, 393, 394, 399

Venice, 3, 4  
Viagra<sup>TM</sup>, 105  
Vienna, 4  
Volatile solvents, 500

**W**

Water solubility, 156, 158, 163, 170, 174–177  
Water-soluble prodrugs, 176  
Weight-loss, 208, 537, 538, 540  
Withdrawal, 214, 219, 252, 258, 259, 274, 324, 342, 344, 382, 441, 484, 486–488, 498, 500, 520, 540

**X**

XAC, 158, 162, 169, 174, 181, 182, 184–186  
Xanthosine, 15–20, 27, 28, 204

**Z**

Zaprinast, 98–101, 107  
Zinc, 98, 118